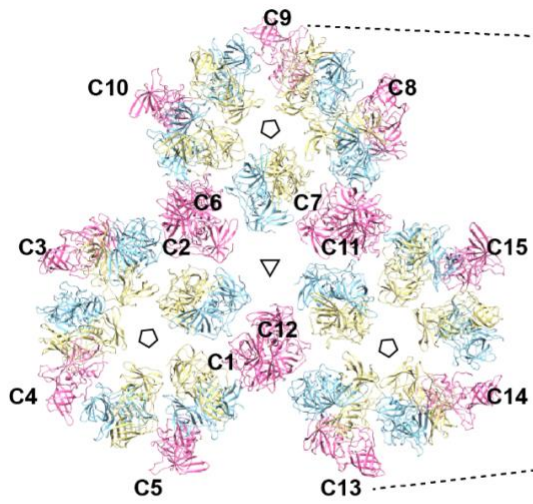


Supplementary Information

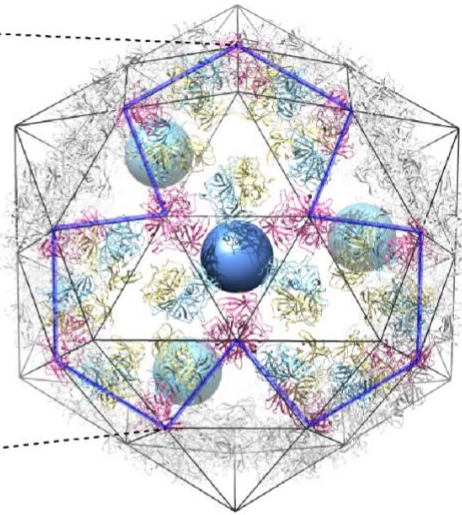
a

Asymmetric unit:
45 VP1 subunits

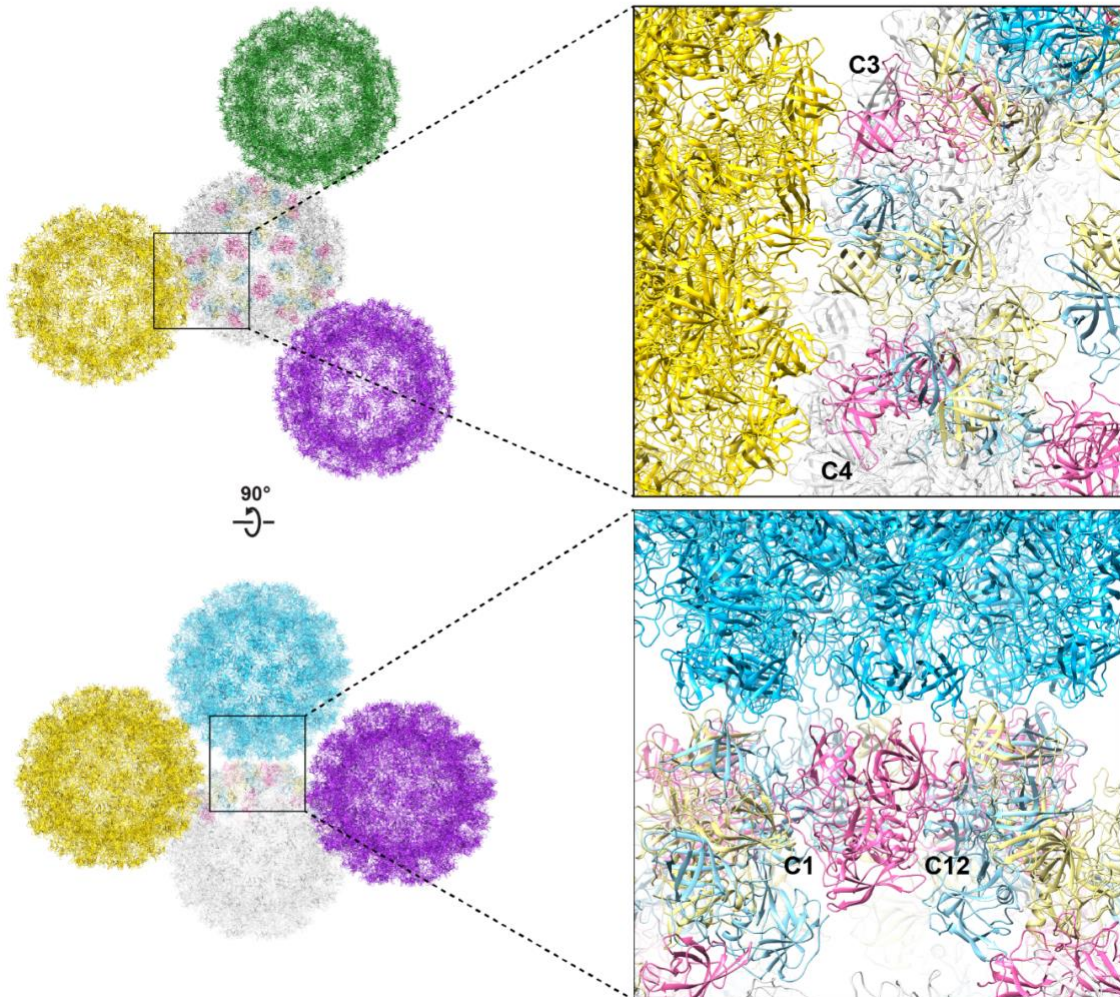


b

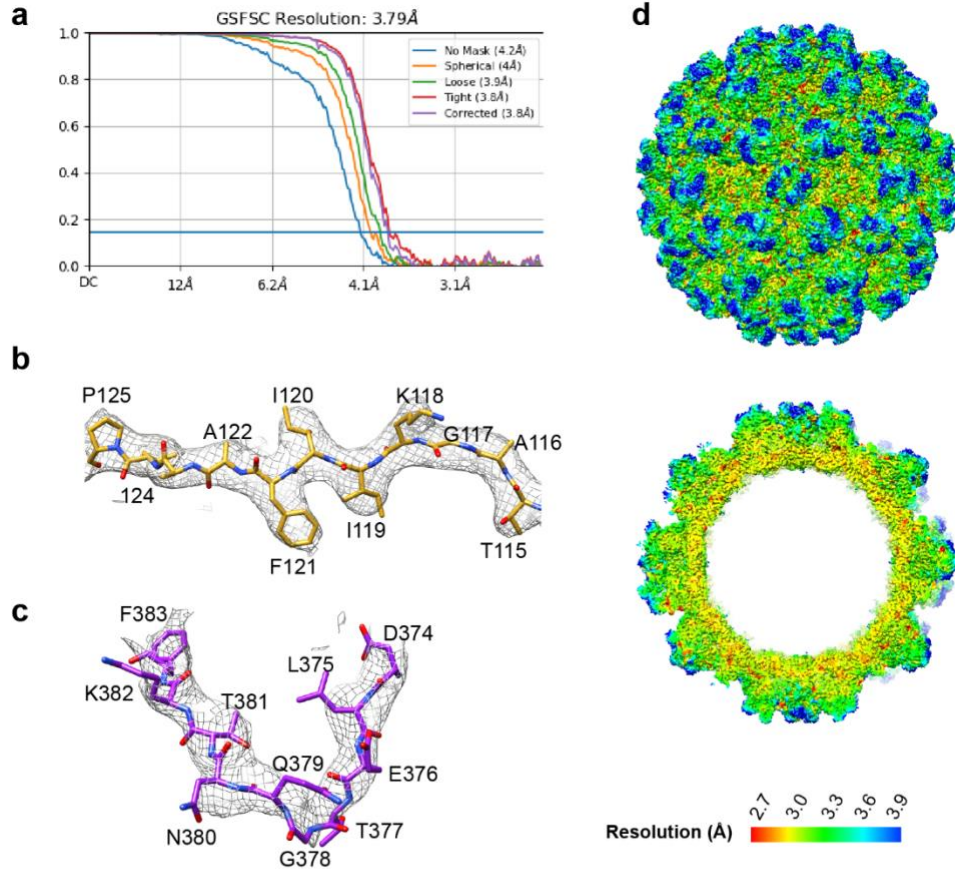
Four asymmetric units:
180 VP1 subunits



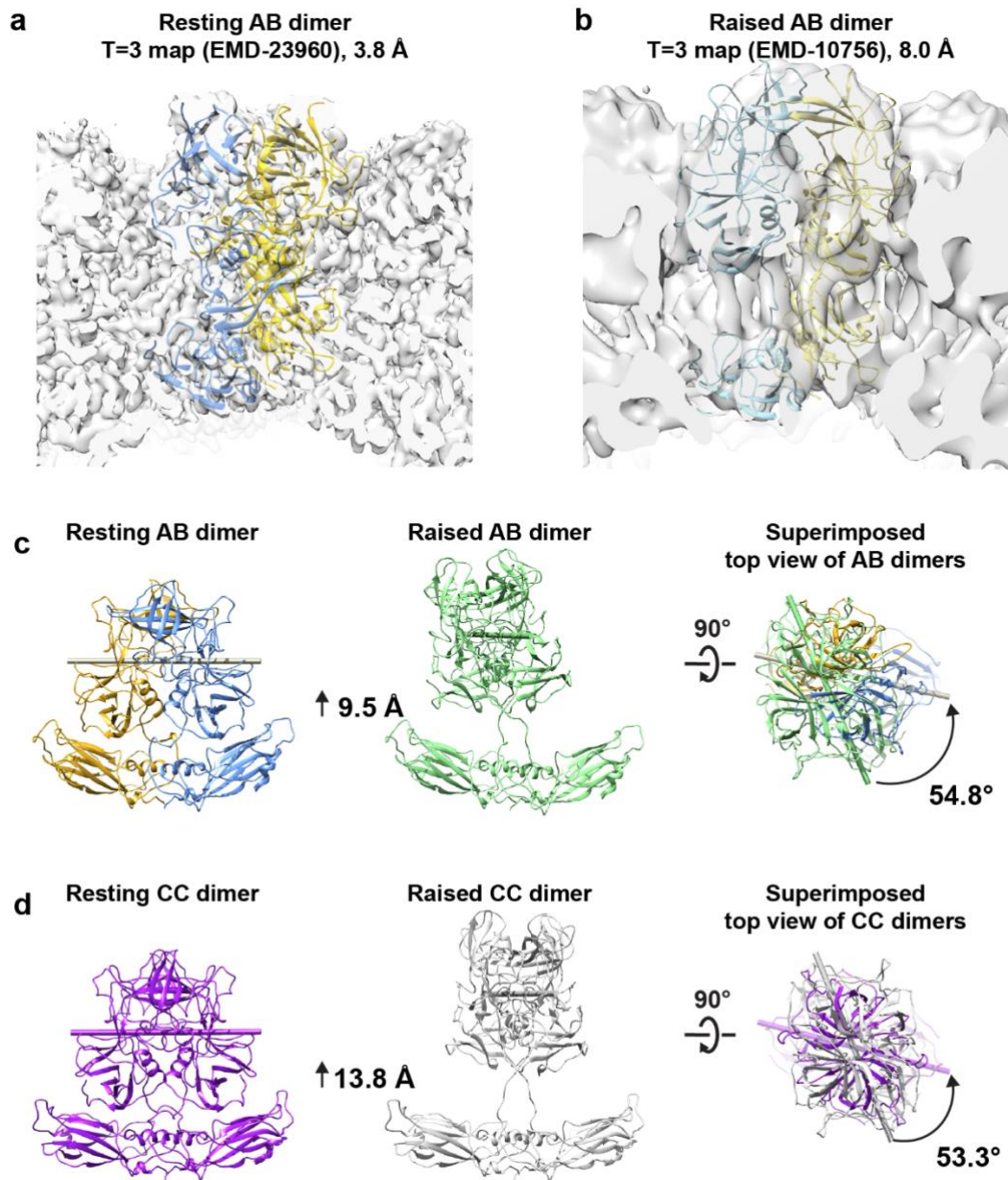
c



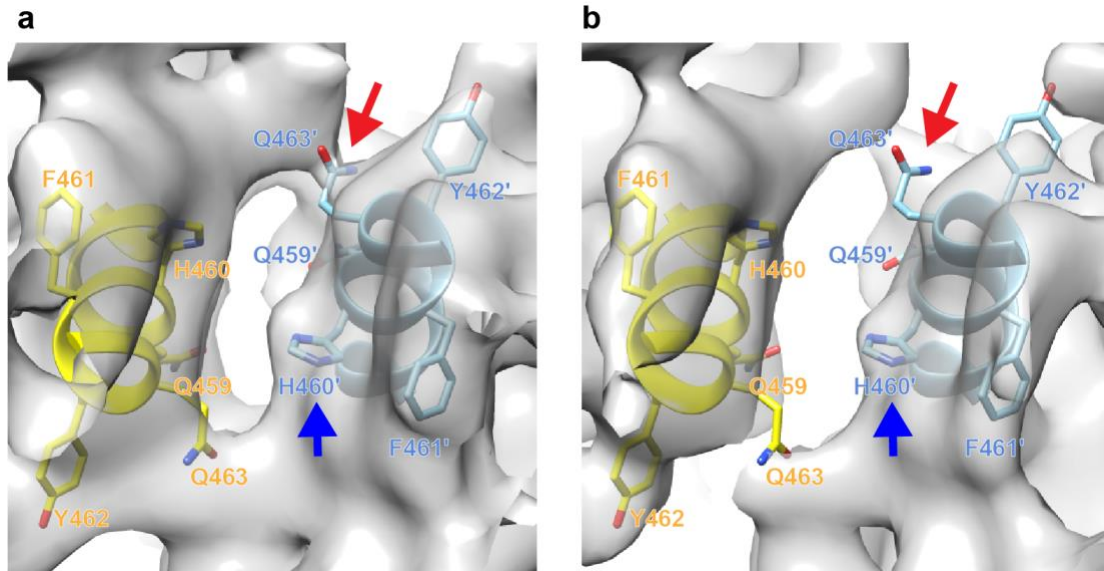
Supplementary Figure 1 | Crystal packing of GII.4 HOV VLPs. **a** The asymmetric unit of GII.4 HOV VLP crystal contains 15 copies of A (yellow), B (blue), and C (pink) chains. For clarity, only C subunits are labeled. The five-fold and three-fold axes are indicated by a pentagon and a triangle, respectively. **b** Four asymmetric units assemble into a VLP particle that consists of 180 copies of VP1. The icosahedron cage is shown with black lines. The symbol of each asymmetric unit is shown as a blue sphere. **c** The crystal packing of VLPs shows the C subunits interacting with the neighboring VLP particles colored in gold, green, purple, and blue.



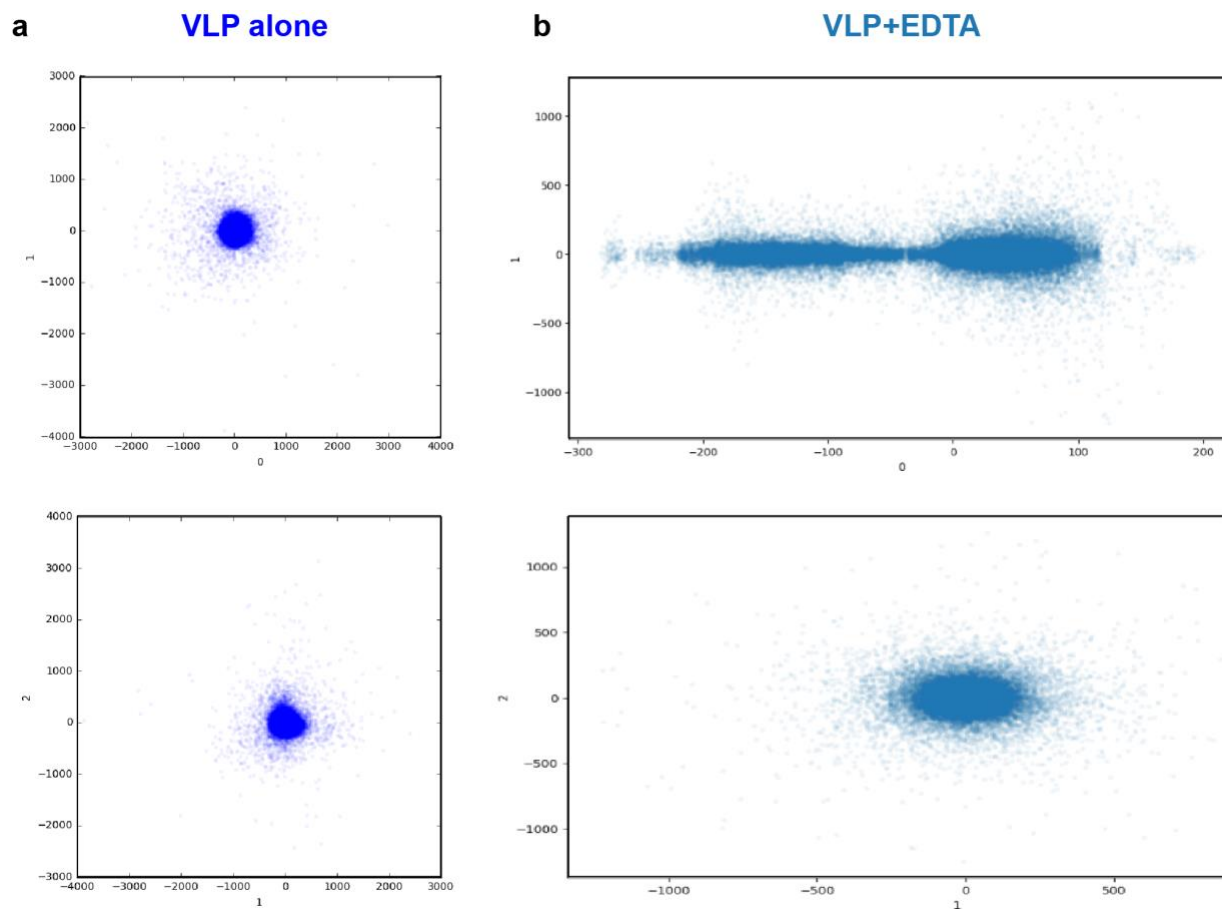
Supplementary Figure 2 | a Fourier shell correlation (FSC) curve indicates the cryo-EM structure's resolution. The resolution estimation of the reconstructions was based on the 0.143 criteria for the comparison between two half-data sets. **b-c Representative section cryo-EM map of VLP.** The density map is shown in gray mesh with the fitted atomic model of VP1. The residues are shown as stick models and labeled. **d Local resolution of the cryo-EM map.**



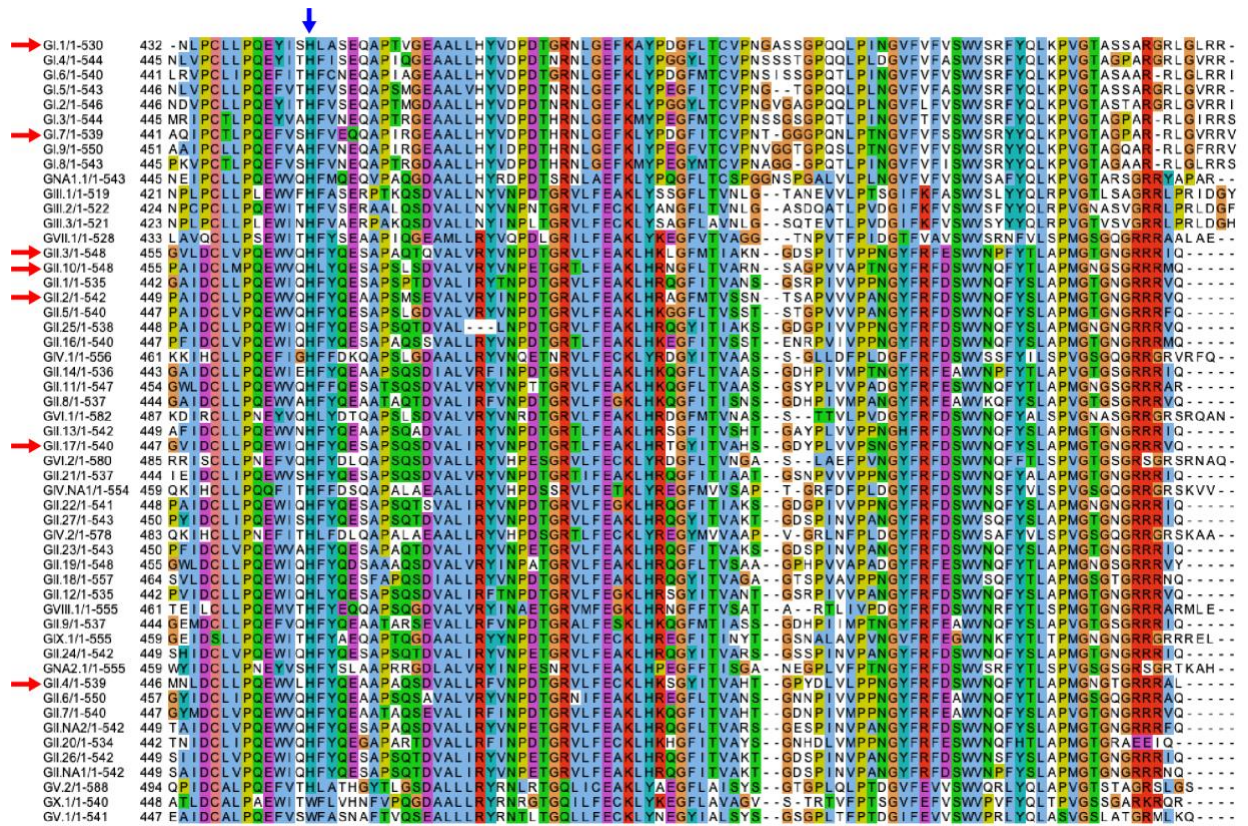
Supplementary Figure 3 | Modeling of GII.4 HOV VLP in the raised conformation. a-b GII.4 HOV VP1 fitted into the 'rising' and 'raised' conformations in the T=3 states. **c-d** Structural comparison of GII.4 HOV AB and CC dimers in both states. The P domain axis was determined by mass weighing using Chimera. The rotation angles and the distance between the 'resting' and 'raised' states are calculated using Chimera.



Supplementary Figure 4 | Density map of GII.4 Minerva (T = 4) VLP. Density at the VP1 dimeric interface around the residue H460 is shown in level 0.1 (a) and 0.14 (b), respectively (PDB ID: 6OUU; EMD-20206). The positions of H460 and Q463 are highlighted by blue and red arrows, respectively. No density between H460 residues was observed.



Supplementary Figure 5 | The 3D variability analysis using CryoSPARC. The reaction coordinate plots for the VLP alone (a) and in the presence of EDTA (b). The axes 0, 1, and 2, correspond to the three different components (eigenvectors) that were solved using 3DVA. The upper panel shows coordinates for 0 vs 1, and the bottom panel for 1 vs 2.



Supplementary Figure 6 | Sequence alignment of HuNoV VP1 from 49 genotypes. Multiple sequence alignment was carried using ClustalW and visualized by Jalview. The conservation of Histidine residue in all the genotypes, except GX.1 and GV.1, shown by the blue arrow at the top. The sequences we selected based on structure availability in Fig. 8 are indicated by red arrows.

Supplementary Table 1 | Human norovirus VLP structures.

| Genotype | Strain | Expression | VP2 | Structure | Resolution | Symmetry | P dimer | PDB or EMDB | Reference |
|-----------------|----------------|---------------------|------------|------------------|-------------------|-----------------|----------------|----------------------------------|-------------------|
| GI.1 | Norwalk | Insect cells | + | Crystal | 3.4 Å | T = 3 | Resting | PDB: 1IHM | Prasad (1999) |
| GI.1 | Norwalk | Tobacco leaves | – | Cryo-EM | 2.9 Å | T = 3 | Resting | PDB: 6OUT EMD-20199/EMD-20205 | Jung (2019) |
| GI.7 | Houston | Insect cells | + | Cryo-EM | 2.9 Å | T = 3 | Resting | PDB: 6OU9 EMD-20197/EMD-20198 | Jung (2019) |
| GII.2 | SMV | Tobacco leaves | – | Cryo-EM | 3.1 Å | T = 3 | Resting | PDB: 6OTF EMD-20195 | Jung (2019) |
| GII.2 | SMV | Tobacco leaves | – | Cryo-EM | 2.7 Å | T = 1 | Resting | PDB: 6OUC EMD-20201/EMD-20202 | Jung (2019) |
| GII.4 | Minerva | Tobacco leaves | – | Cryo-EM | 4.1 Å | T = 4 | Raised | PDB: 6OUU EMD-20206 | Jung (2019) |
| GII.4 | NSW-2012 | Insect cells | – | Cryo-EM | 7.3 Å | T = 4 | Raised | EMD-4550 | Devant (2019) |
| GII.4 | CHDC-1974 | Insect cells | – | Cryo-EM | 6.1 Å | T = 4 | Raised | EMD-4549 | Devant (2019) |
| GII.4 | GII.4c | HEK-293T | – | Cryo-EM | 4.5 Å | T = 4 | Raised | EMD-10758 | Devant (2021) |
| GII.4 | GII.4c | Insect cells | – | Cryo-EM | 4.2 Å | T = 4 | Raised | EMD-10755 | Devant (2021) |
| GII.4 | GII.4c | Insect cells | – | Cryo-EM | 8.0 Å | T = 3 | Raised | EMD-10756 | Devant (2021) |
| GII.4 | GII.4c | Insect cells | – | Cryo-EM | 4.2 Å | T = 1 | Raised | EMD-10757 | Devant (2021) |
| GII.10 | Vietnam 026 | Insect cells | – | Cryo-EM | 10 Å | T = 3 | Raised | EMD-5374 | Hansman (2012) |
| GII.17 | Kawasaki 2014 | Insect cells | – | Cryo-EM | 5.8 Å | T = 3 | Raised | EMD-10759 | Devant (2021) |
| GII.4 | Houston | Insect cells | + | Crystal | 3.0 Å | T = 3 | Resting | PDB: 7K6V | This study |
| GII.4 | Houston | Insect cells | + | Cryo-EM | 3.8 Å | T = 3 | Resting | PDB 7MRY EMD-23960 | This study |

Supplementary Table 2 | X-ray crystallography data collection and refinement statistics.

| | |
|-----------------------------------|-------------------------|
| PDB ID | 7K6V |
| Wavelength (Å) | 0.9795 |
| Resolution range (Å) | 29.99 - 3.0 (3.1 - 3.0) |
| Space group | I 2 2 2 |
| Unit cell | |
| a, b, c (Å) | 420.2, 446.6 464.0 |
| α , β , γ (°) | 90, 90, 90 |
| Unique reflections | 819642 (61796) |
| Multiplicity | |
| Completeness (%) | 95.67 (72.59) |
| Mean I/sigma(I) | 7.0/1.0 |
| Wilson B-factor | 67.97 |
| R-merge (%) | 13.6 (83.5) |
| R-work (%) | 20.71 (31.13) |
| R-free (%) | 24.18 (36.21) |
| Number of non-hydrogen atoms | 176522 |
| macromolecules | 176443 |
| ligands | 76 |
| solvent | 3 |
| Protein residues | 22902 |
| R.M.S. deviations | |
| Bond lengths (Å) | 0.004 |
| Bond angles (°) | 0.77 |
| Ramachandran | |
| Favored (%) | 96.72 |
| Allowed (%) | 3.28 |
| Disallowed (%) | 0 |
| Average B-factor | 67.79 |
| macromolecules | 67.78 |
| ligands | 80.77 |
| solvent | 41.45 |

Statistics for the highest-resolution shell are shown in parentheses.

Supplementary Table 3 | Cryo-EM data collection, reconstruction, and model refinement parameters.

| | HOV VLP (EMDB-23960) (PDB 7MRY) |
|---|---------------------------------------|
| Data collection and processing | |
| Magnification | 30,000 |
| Voltage (kV) | 300 |
| Electron exposure (e ⁻ /Å ²) | 55.96 |
| Defocus range (μm) | 0.6 to 2.6 |
| Pixel size (Å) | 1.24 |
| Symmetry imposed | I2 |
| Map resolution (Å) | |
| FSC threshold | 0.143 |
| Map resolution (Å) | 3.8 |
| Refinement | |
| Initial model used (PDB code) | 7K6V |
| Model resolution (Å) | 3.8 |
| FSC threshold | 0.143 |
| Map sharpening <i>B</i> factor (Å ²) | -210.9 |
| Model composition | |
| Non-hydrogen atoms | 11329 |
| Protein residues | 1460 |
| Ligands | 0 |
| <i>B</i> factors (Å ²) | |
| Protein | 72.34 |
| Ligand | 0 |
| R.m.s. deviations | |
| Bond lengths (Å) | 0.002 |
| Bond angles (°) | 0.529 |
| Validation | |
| MolProbity score | 1.45 |
| Clashscore | 5.47 |
| Poor rotamers (%) | 0 |
| Ramachandran plot | |
| Favored (%) | 97.11 |
| Allowed (%) | 2.89 |
| Disallowed (%) | 0 |