

Supporting Information

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Autonomous Visualization of Damage in Polymers by Metal-Free Polymerizations of Microencapsulated Activated Alkynes

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EXPERIMENTAL SECTION

General Materials and Instruments

1,4-Diazabicyclo[2.2.2]octane (DABCO), propiolic acid, 1,6-hexanediol, 1,3-propanediol, 2,2dibutyl-1,3-propanediol, 4,4'-oxydiphenol, and all other chemicals and reagents were purchased from Meryer, J&K Scientific, or Aldrich and used as commercially received unless stated otherwise. Dichloromethane (DCM), tetrahydrofuran (THF), *N*,*N*-dimethylformamide (DMF), dimethyl sulfoxide (DMSO), and 1,4-dioxane are ultra-dry reagents purchased from J&K Scientific. Poly(vinyl alcohol) 1788 (87.0-89.0% hydrolyzed, mol/mol) was purchased from Aladdin Chemical Ltd. Monomers **1a–1e** were synthesized according to the previous reported procedures.¹⁻³

Weight- (M_w) and number-average molecular weights (M_n) and polydispersity indices (M_w/M_n) of the polymers were estimated on a Waters e2695 gel permeation chromatography (GPC) system equipped with a refractive index detector and calibrated using a set of monodispersed polystyrene standards. THF was used as the eluent. IR spectra were recorded on a Nicolet 6700 FTIR spectrophotometer (KBr disk). ¹H and ¹³C NMR spectra were measured on a Bruker AVANCE III spectrometer in deuterated chloroform (CDCl₃). Chemical shifts were calibrated using CDCl₃ as internal reference at δ 7.26 ppm (¹H NMR) and δ 77.16 ppm (¹³C NMR). High-resolution mass spectra (HRMS) were recorded on a GCT premier CAB048 mass spectrometer. UV spectra were measured on a Shimadzu UV-2450 UV/Vis spectrophotometer. Thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC) measurement was carried on a TA TGA 55 and a TA DSC Q200, respectively, under nitrogen at a heating rate of 10 °C/min. Scanning electron microscopy (JEOL-6390) was applied to observe the morphologies of the microcapsules and scratched areas of polymer films. The SEM samples of microcapsule and coatings were coated with a thin layer of gold before observation. The

photographs of microcapsules and polymer films were acquired using a Canon EOS 70D digital camera under ambient room lighting.

Synthesis and Characterization

Characterization of Polymers. *Characterization Data for PIa*: Brownish red powder; yield: 84% (Table 1, entry 2). $M_n = 21800$; $M_w = 29800$; $M_w/M_n = 1.4$ (GPC for soluble part, polystyrene calibration). FT-IR (KBr disk), v (cm⁻¹): 3005, 2939, 2859, 2220, 1708, 1618, 1463, 1386, 1235, 1169, 958, 744. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.58 (d, J = 12.0 Hz), 6.78 (d, J = 16.0 Hz), 6.47 (d, J = 16.0 Hz), 5.65 (d, J = 12.0 Hz), 4.23–4.13, 1.70–1.63, 1.41. ¹³C NMR (100 MHz, CDCl₃), δ (ppm): 166.29, 164.89, 157.48, 153.35, 135.48, 121.79, 104.35, 87.18, 81.78, 66.42, 65.34, 64.56, 29.84, 28.65, 28.55, 28.52, 28.39, 25.65, 25.62, 25.58, 25.55. *Characterization Data for P1b*: Yellow powder; yield: 84% (Table 1, entry 4). $M_n = 700$; M_w = 1100; $M_w/M_n = 1.6$ (GPC for soluble part, polystyrene calibration). FT-IR (KBr disk), v (cm⁻¹): 3081, 2963, 2904, 2221, 1705, 1619, 1461, 1387, 1231, 1168, 1035, 956, 744. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.58 (d, J = 12.0 Hz), 6.80 (d, J = 16.0 Hz), 6.48 (d, J = 16.0 Hz), 5.67 (d, J = 12.0 Hz), 4.36–4.24, 2.12–2.05. ¹³C NMR (100 MHz, CDCl₃), δ (ppm): 168.09, 164.60, 153.03, 150.76, 135.25, 122.10, 101.65, 85.10, 82.06, 62.77, 61.92, 27.75.

Characterization Data for P1c: Yellow powder; yield: 9% (Table 1, entry 6). $M_n = 4200$; M_w = 6400; $M_w/M_n = 1.5$ (GPC for soluble part, polystyrene calibration). FT-IR (KBr disk), v (cm⁻¹): 3058, 2937, 2866, 2231, 1716, 1621, 1466, 1382, 1235, 1124, 1032, 958, 734. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.56 (d, J = 12.0 Hz), 6.77 (d, J = 16.0 Hz), 6.49 (d, J = 16.0 Hz), 5.66 (d, J = 12.0 Hz), 4.08–3.95, 1.30, 1.21, 0.90. ¹³C NMR data was not obtained due to the low yield and poor solubility of the polymer.

Characterization Data for P1d: Yellow powder; yield: ~100% (Table 1, entry 9). $M_n = 900$; $M_w = 1600$; $M_w/M_n = 1.8$ (GPC for soluble part, polystyrene calibration). FT-IR (KBr disk), v (cm⁻¹): 3069, 2216, 1723, 1619, 1489, 1223, 1172, 1137, 1093, 1008, 952, 838, 508. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.81 (d, J = 12.0 Hz), 7.17–7.08, 7.06, 7.04, 7.01, 6.44 (d, J =

16.0 Hz), 5.92 (d, J = 12.0 Hz), 4.08–3.95, 1.30, 1.21, 0.90. ¹³C NMR data was not obtained due to the poor solubility of the polymer.

Characterization Data for P1e: Brownish red powder; yield: 97% (Table 1, entry 11). FT-IR (KBr disk), v (cm⁻¹): 3016, 2968, 2880, 2231, 1717, 1622, 1465, 1389, 1233, 1163, 1054, 957, 744. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 6.79 (d, J = 16.0 Hz), 6.81 (d, J = 16.0 Hz), 4.26–4.10, 1.31–1.20, 0.93–0.83. ¹³C NMR data was not obtained due to the poor solubility of the polymer.

Model Reaction. To a stirred solution of ethyl propiolate (0.51 mL, 5 mmol) in DCM (4 mL) at 0 °C was added the solution of DABCO (5.6 mg, 0.05 mmol) in DCM (1 mL) under air. The reaction immediately turned into brown and was stirred for 1 h. After reaction, the DABCO catalyst was removed by filtration through a pad of silica gel. Then the solvent was evaporated and the crude reaction mixture was purified by silica gel chromatography using hexane/ethyl acetate mixture (95:5, v/v) as eluent. The diethyl (*E*)-hex-2-en-4-ynedioate product (**M1**) was collected as yellow oil in a NMR yield: 93.8%. The byproduct diethyl 3,3'-oxy(2*E*,2'*E*)-diacrylate (**M2**) was obtained as white solid in an NMR yield of 6.2%. The structural characterization results are summarized as follows.

Characterization Data for M1: ¹H NMR (400 MHz, CDCl₃), δ (ppm): 6.70 (d, J = 16.0 Hz, 1H), 6.39 (d, J = 16.0 Hz, 1H), 4.22–4.15 (m, 4H), 1.26–1.21 (m, 6H). ¹³C NMR (100 MHz, CDCl₃), δ (ppm): 164.12, 152.51, 135.05, 121.10, 86.70, 80.93, 61.96, 60.87, 13.72, 13.56. FT-IR (KBr disk), v (cm⁻¹): 3038, 2981, 2238, 1710, 1618, 1459, 1368, 1243, 1175, 1020, 859. HRMS (CI, CH₄): m/z [M + H]⁺ calcd for C₁₀H₁₂O₄, 196.0736; found, 197.0810.

Characterization Data for M2: ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.57 (d, J = 12.0 Hz, 2H), 5.64 (d, J = 12.0 Hz, 2H), 4.19 (q, J = 7.0 Hz, 4H), 1.28 (t, J = 7.0 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃), δ (ppm): 166.19, 157.37, 104.43, 60.64, 14.37. FT-IR (KBr disk), v (cm⁻¹): 3087,

2989, 2925, 1710, 1608, 1474, 1368, 1330, 1195, 1120, 967, 864, 846, 735, 585. HRMS (CI, CH₄): *m*/*z* [M + H]⁺ calcd for C₁₀H₁₄O₅, 214.0841; found, 215.0912.

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Figure S1. Photographs showing the chromogenic phenomenon upon adding DABCO crystals into the dichloromethane (DCM) solution of activated triyne.

entry	condition	yield (%)	$M_{ m w}{}^b$	$M_{ m w}/M_{ m n}{}^b$	S^{c}
1	adding DABCO solution to 1a solution	81	34 100	1.9	Δ
2	adding 1a solution to DABCO solution	41	18 500	1.3	Δ
3	adding 1a solution to DABCO powder	61	18 200	1.4	Δ
4	adding DABCO solution to 1a powder	70	17 100	1.4	Δ
5	mixing the powder of DABCO and 1a	73	18 000	1.4	Δ
6^d	mixing the powder of DABCO and 1a	96		/ <i>f</i>	×
7^e	mixing the powder of DABCO and 1a	96	/f		×

Table S1. Preliminary Trials on the Polymerization of $1a^a$

^{*a*} Unless otherwise noted, the polymerizations were carried out at 0 °C in anhydrous DCM in air for 1 h. [**1a**] = 0.2 M, [DABCO] = 10 mol% [**1a**]. ^{*b*} Determined by GPC in THF on the basis of a linear polystyrene calibration. ^{*c*} Solubility (S) tested in common organic solvents, such as DCM, chloroform, and THF: Δ = partially soluble, × = insoluble. The GPC results are for the soluble part. ^{*d*} [DABCO] = 50 mol%. ^{*e*} Carried out under room temperature. ^{*f*} Not detected.

entry	solvent	yield (%)	$M_{ m w}{}^b$	$M_{ m w}/M_{ m n}{}^b$	S^c
1^d	DCM	81	34 100 1.9		Δ
2	DCM/water = 1:1 (v/v)	trace			Δ
3	DCM/water = 3:2 (v/v)	trace			Δ
4	THF	32	6 800	1.3	Δ
5	THF/water = $1:1 (v/v)$	3	2 800	1.1	Δ
6	DMF	59	2 900 1.1		Δ
7	DMSO	65	<i> e</i>	2	×
8	1,4-dioxane	gel, ~100	14 600	1.5	Δ
9	hexyl acetate	60	9 000	1.3	Δ
10	ethyl phenylacetate	5	2 800	1.1	Δ
11	MeCN	~100	/ e	2	×
12	toluene	27	2 800	1.1	Δ

Table S2. Effect of Solvent on the Polymerization of $1a^{a}$

^{*a*} Unless otherwise noted, the polymerizations were carried out at 0 °C under air for 1 h by adding DABCO solution to **1a** solution. [**1a**] = 0.2 M, [DABCO] = 10 mol% [**1a**]. ^{*b*} Determined by GPC in THF on the basis of a linear polystyrene calibration. ^{*c*} Solubility (S) tested in common organic solvents, such as DCM, chloroform, and THF: Δ = partially soluble, × = insoluble. The GPC results are for the soluble part. ^{*d*} Data taken from Table S1, entry 2. ^{*e*} Not detected.

entry	time	yield (%)	$M_{ m w}{}^b$	$M_{ m w}/M_{ m n}{}^b$	Sc
1	5 min	23	/	е	Δ
2	15 min	63	/	е	Δ
3	30 min	74	1 000	2.5	Δ
4^d	1 h	84	29 800	1.4	Δ

Table S3. Time Course of the Polymerization of $1a^a$

^{*a*} Unless otherwise noted, the polymerizations were carried out at 0 °C in anhydrous DCM under air by adding DABCO solution to **1a** solution. [**1a**] = 0.2 M, [DABCO] = 15 mol% [**1a**]. ^{*b*} Determined by GPC in THF on the basis of a linear polystyrene calibration. ^{*c*} Solubility (S) tested in common organic solvents, such as DCM, chloroform, and THF: Δ = partially soluble. The GPC results are for the soluble part. ^{*d*} Data taken from Table S3, entry 1. ^{*e*} Not detected.

entry	catalyst	yield (%)	$M_{ m w}{}^b$	$M_{ m w}/{M_{ m n}}^b$	Color ^c
1	DABCO	84	29 800	1.4	
2	DMAP	brownish red solution; u	indesired and uni	dentified product	
3	NEt ₃	deep brownish red s (@ $r.t. + 1$ equiv.	-		
4	pyridine	colorless solution; no polymeric product (@ r.t. + 7 equiv. pyridine: brownish red gel)			-
5	4-picoline	light yellow solu $(@ r.t. + 2 equiv$	tion; no polymer 4 <i>-picoline</i> : browr	ic product nish red gel)	1
6	DBU	yellow solution (@ r.t. + 10 equ no pol	n; no polymeric <i>uv. DBU</i> : yellow lymeric product)	product solution;	2
7	NaOH, Na2CO3, CH3COONa	colorless solutio no po	on with insoluble lymeric product	catalysts;	1

Table S4. Effect of Catalyst Type on the Polymerization of $1a^a$

^{*a*} Unless otherwise noted, the polymerizations were carried out at 0 °C in anhydrous DCM under air for 1 h by adding catalyst to **1a** solution. [**1a**] = 0.2 M, [catalyst] = 15 mol% [**1a**]. The contents in parentheses are the polymerization results obtained under the noted conditions. ^{*b*} Determined by GPC in THF on the basis of a linear polystyrene calibration. The GPC results are for the soluble part. ^{*c*} Photographs showing the appearance color of the reaction mixture after reacting for 1 h. Abbreviation: DMAP = 4-dimethylaminopyridine; DBU = 1,8-diazabicyclo[5.4.0]undec-7-ene; r.t. = room temperature.

entry	monomer	[monomer] (M)	yield (%)	$M_{ m w}{}^b$	$M_{ m w}/{M_{ m n}}^b$	S^c
1	1 a	0.2	84	29 800	1.4	Δ
2	1b	0.2	49	1 900	1.1	Δ
3 ^{<i>d</i>}	1b	0.2	84	1 100	1.6	Δ
4	1c	0.2	trace			
5	1c	0.4	9	6 400	1.5	Δ
6	1d	0.2 or 0.1	gelled within seconds			
7	1d	0.05	gelled within 2 min			
8 ^e	1d	0.05	~100	1 600	1.8	Δ
9	1e	0.2 or 0.1	gelled within seconds			
10	1e	0.05	97	/f		×

Table S5. Polymerization results of monomer $1a-1e^{a}$

^{*a*} Unless otherwise noted, the polymerizations were carried out at 0 °C in anhydrous DCM in the presence of 15 mol% DABCO in air for 1 h. ^{*b*} Determined by GPC in THF on the basis of a linear polystyrene calibration. ^{*c*} Solubility (S) tested in common organic solvents, such as DCM, chloroform, and THF: Δ = partially soluble, × = insoluble. The GPC results are for the soluble part. ^{*d*} Polymerization results obtained at 2 h. ^{*e*} [DABCO] = 5 mol% [1d]. ^{*f*}Not detected.



Figure S2. (A) ¹H NMR spectrum and (B) ¹³C NMR spectrum of model compound M2 in chloroform-*d*.





Figure S4. FT-IR spectra of (A) monomer 1a, (B) model compound M1, (C) model compound M2, and (D) polymer P1a.



Figure S5. FT-IR spectra of (A) monomer 1b, (B) model compound M1, (C) model compound M2, and (D) polymer P1b.



Figure S6. FT-IR spectra of (A) monomer 1c, (B) model compound M1, (C) model compound M2, and (D) polymer P1c.



Figure S7. FT-IR spectra of (A) monomer 1d, (B) model compound M1, (C) model compound M2, and (D) polymer P1d.



Figure S8. FT-IR spectra of (A) monomer 1e, (B) model compound M1, (C) model compound M2, and (D) polymer P1e.



Figure S9. ¹H NMR spectra of (A) polymer P**1a** obtained in air and (B) polymer P**1a** obtained under nitrogen. The ¹H NMR spectra were measured in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S10. ¹H NMR spectra of (A) monomer 1b, (B) model compound M1, (C) model compound M2, and (D) polymer P1b in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S11. ¹H NMR spectra of (A) monomer 1c, (B) model compound M1, (C) model compound M2, and (D) polymer P1c in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S12. ¹H NMR spectra of (A) monomer 1d, (B) model compound M1, (C) model compound M2, and (D) polymer P1d in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S13. ¹H NMR spectra of (A) monomer 1e, (B) model compound M1, (C) model compound M2, and (D) polymer P1e in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S14. ¹H NMR spectrum of polymer P1a in chloroform-*d*. The sample was obtained from the polymerization in THF/water mixture (v/v = 1:1). The solvent peaks are marked with asterisks.



Figure S15. ¹³C NMR spectra of (A) monomer 1a, (B) model compound M1, (C) model compound M2, and (D) polymer P1a in chloroform-*d*. The solvent peaks are marked with asterisks.



Figure S16. ¹³C NMR spectra of (A) monomer 1b, (B) model compound M1, (C) model compound M2, and (D) polymer P1b in chloroform-*d*. The solvent peaks are marked with asterisks.

Scheme S1. Proposed mechanism for the formation of poly(1,3-enyne).





Scheme S2. Proposed mechanism for the formation of copolymers.



Figure S17. TGA thermograms of polymers recorded under nitrogen at a heating rate of 10 °C/min.



Figure S18. (A) DSC thermograms of polymers and the amplified DSC curve of (B) P1b and (C) P1e recorded under nitrogen during the second heating cycle at a heating rate of 10 °C/min.



Table S6. Appearance of Polymers $P1a-P1e^{a}$

^{*a*} The polymeric products were carried out at 0 °C in anhydrous DCM under air for 1 h by adding DABCO catalyst to the solution of monomer. [monomer] = 0.2 M, [DABCO] = 15 mol% [monomer]. ^{*b*} The polymeric product was obtained at a monomer concentration of 0.4 M.



Figure S19. SEM images of intact and ruptured microcapsules containing (A-C) pure EPA solvent, (D-F) alkyne/EPA solution (0.5 wt%), and (H-J) alkyne/EPA solution (5.0 wt%).



Figure S20. SEM images with a wide field of view to show the uniform morphology of the microcapsules.



Figure S21. Size distribution of the microcapsules (sample size n = 200). Inset: quantitative data for the size distribution of the microcapsules shown as mean \pm standard deviation (S.D.).



Figure S22. TGA curves of alkyne/HDI microcapsules (MCs), EPA MCs, shell, pure EPA, and pure alkyne monomer (**1e**) recorded under nitrogen at a heating rate of 10 °C/min.

Effect of alkyne content of microcapsules on damage indication



Figure S23. Photographs of alkyne/EPA microcapsules (MCs) with the alkyne content of (A) 0.5 wt%, (B) 1.0 wt%, and (C) 5.0 wt% without catalyst before crushing, mixing with 50 wt% DABCO catalyst before crushing, and mixing with 50 wt% DABCO catalyst after crushing between glass slides for 1 min. The amount of catalyst was relative to the dosage of the microcapsules.



Effect of catalyst dosage on damage indication

Figure S24. Photographs of alkyne/EPA microcapsules (MCs, alkyne content = 5.0 wt%) without catalyst before crushing, mixing with DABCO catalyst before crushing, and mixing with DABCO catalyst after crushing between glass slides for 1 min. The amount of catalyst relative to the dosage of the microcapsules was 50 wt%, 20 wt% and 10 wt% for panel A, B and C, respectively.



Quantative crushing experiments under different pressure

Figure S25. Photographs of the mixture of alkyne/EPA microcapsules (alkyne content = 5.0 wt%) and DABCO catalyst before and after crushing between two glass slides under different intensity of pressure. The mixture was crushed under the compression of a weight of (A) 200 g, (B) 500 g, and (C) 1 kg. The amount of catalyst relative to the dosage of the microcapsules was 10 wt\%. All these samples share a similar stress area of about 1.5 cm².



Figure S26. UV-vis spectra of alkyne/EPA microcapsules (MCs, alkyne content = 5.0 wt%) without catalyst and the mixture of MCs with DABCO catalyst measured at different scan time. The spectra were measure by fixing and crushing the MCs or the mixture via the compression between two quartz plates. Each scan takes about 1 min.



Figure S27. Photographs of PVA films containing (A) neither microcapsules nor catalyst, and (B) 5 wt% DABCO catalyst before and after being scratched with a utility knife for 10 min.



Figure S28. Photographs of PVA films containing different amount of alkyne/EPA microcapsules (alkyne content = 5.0 wt%) and 10 wt% DABCO catalyst: (A) 15 wt% alkyne/EPA microcapsules and 10 wt% DABCO catalyst, (B) 15 wt% alkyne/EPA microcapsules and 5 wt% DABCO catalyst. (C) 10 wt% alkyne/EPA and 10 wt% DABCO catalyst, and (D) 10 wt% alkyne/EPA microcapsules and 5 wt% DABCO catalyst. The photographs were taken before and after the films being scratched with a utility knife for different time. All the photographs share the same scale bar of 2 cm.



Figure S29. Photographs of PVA films containing 15 wt% alkyne/EPA microcapsules and 5 wt% DABCO catalyst before and after being impacted and compressed by an iron bar for different time.