

Supplementary Information (SI) Guide for

A microphysiological model of human trophoblast invasion during implantation

¹Ju Young Park, ²Sneha Mani, ³Jeremy Clair, ³Heather M. Olson, ³Vanessa L. Paurus, ³Charles K. Ansong, ¹Cassidy Blundell, ¹Rachel Young, ²Jessica Kanter, ⁴Scott Gordon, ¹Alex Y. Yi, ²Monica Mainigi*, ^{1,5,6}Dan Dongeun Huh*

Supplementary Information includes:

Supplementary Figures 1-11

Supplementary Figure 1. Measurement of proliferative EVT population in the implantation-on-a-chip.

Supplementary Figure 2. Analysis of ECM stiffness in the implantation-on-a-chip.

Supplementary Figure 3. Comparison of invasive activity of primary EVT and HTR8/SVneo cells on the 'implantation-on-a-chip' device.

Supplementary Figure 4. Maintenance of the decidualized phenotype of stromal cells in the implantation-on-a-chip.

Supplementary Figure 5. Analysis of ECM composition and stiffness in the presence of different cell types in the hydrogel compartment.

Supplementary Figure 6. Secretomics analysis of effluent samples collected from implantation-on-a-chip in four different culture conditions: i) EVT-mono, ii) EC-mono, iii) CO, and iv) TRI (one-way ANOVA $p < 0.05$).

Supplementary Figure 7. Secretomics analysis of human proteins only. Z scores and the results of t tests of the different proteins identified in the different culture conditions: i) EVT-mono, ii) EC-mono, iii) CO, and iv) TRI (two-sided t-test).

Supplementary Figure 8. Heatmap depicting the human proteins significantly regulated in abundance in the different culture conditions: i) EVT-mono, ii) EC-mono, iii) CO, and iv) TRI

Supplementary Figure 9. List of proteins showing significant increase or decrease between EVT monoculture and coculture of EVT and EC (two-sided t-test).

Supplementary Figure 10. List of proteins showing significant increase or decrease between EC monoculture and coculture of EVT and EC (two-sided t-test).

Supplementary Figure 11. Fourteen significantly upregulated endothelial proteins in the coculture configuration of the implantation-on-a-chip.

Supplementary Figure 12. Twelve significantly upregulated EVT proteins in coculture configuration of the implantation-on-a-chip.

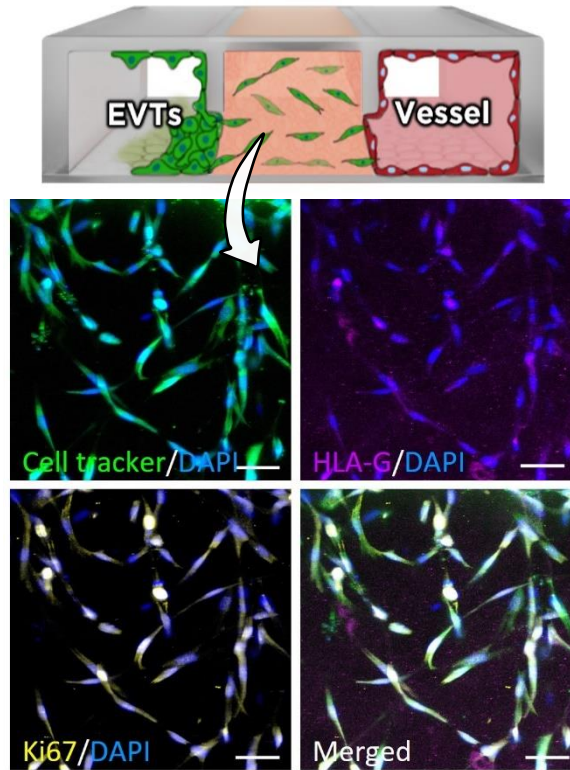
Supplementary Figure 13. Viability and marker expression of DSCs and additional cell types in the implantation-on-a-chip.

Supplementary Table 1. Cells and authentication methods.

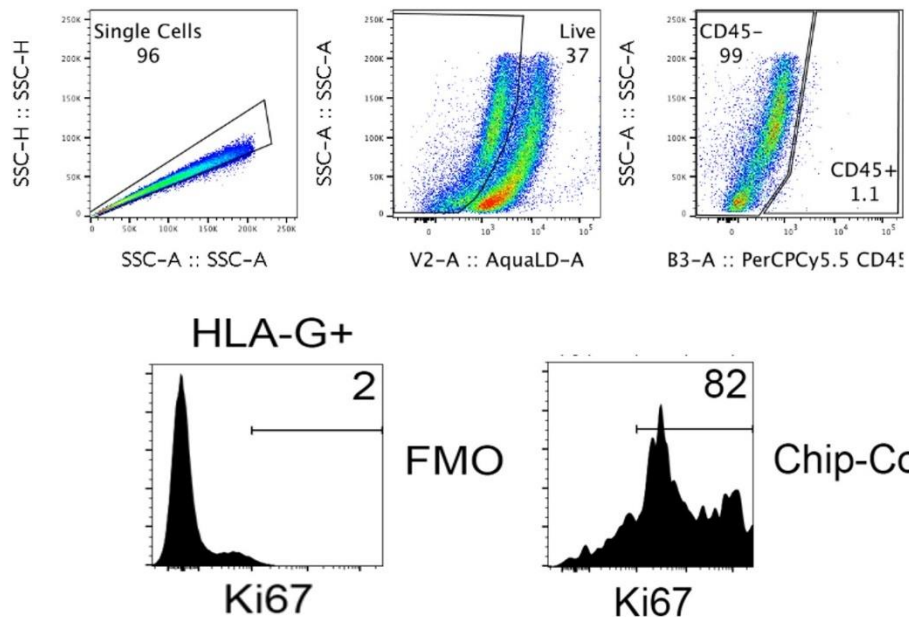
Supplementary Table 2. Primary and secondary antibodies for immunofluorescence staining.

Supplementary Figure 1. Measurement of proliferative EVT population in the implantation-on-a-chip. (a) Immunofluorescence staining of HLA-G (purple) and Ki67 (yellow) in EVT cells migrating in the ECM compartment at Day 6. The representative images are from three independent experiments. Scale bars, 50 μ m. (b) Cells isolated from the device were flow sorted. Analysis of single, live, and CD45-negative cells stained with HLA-G and Ki67 showed that 82% of HLA-G⁺ cells were also Ki67 positive. FMO = fluorescence minus one control.

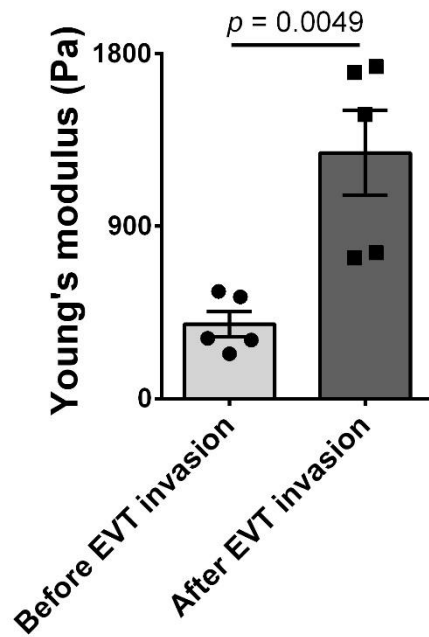
a



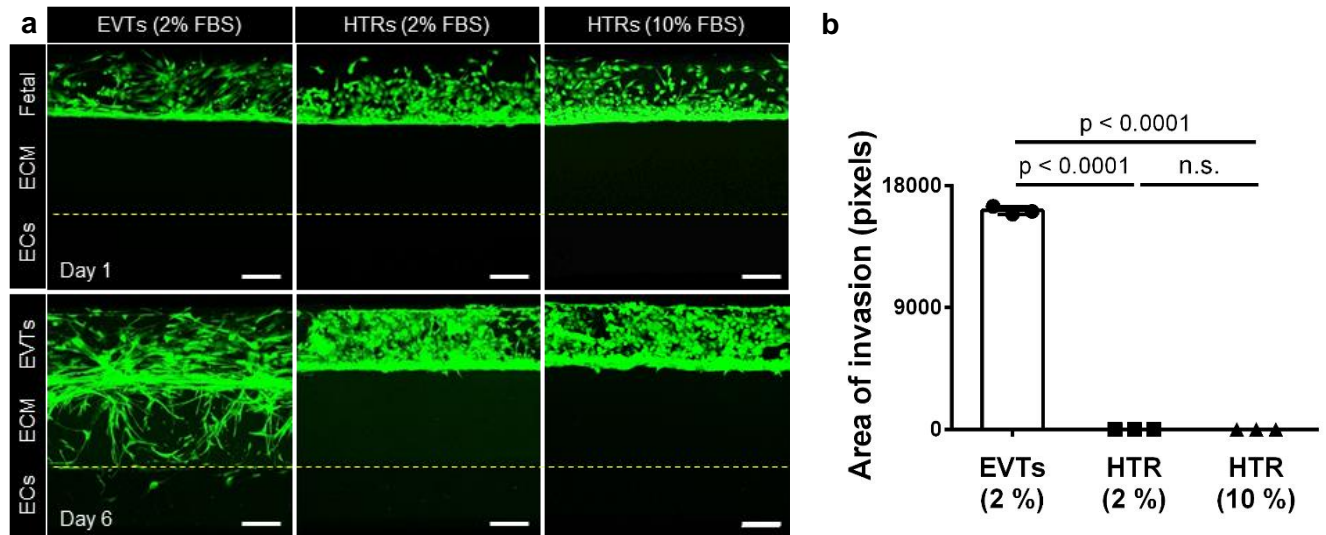
b



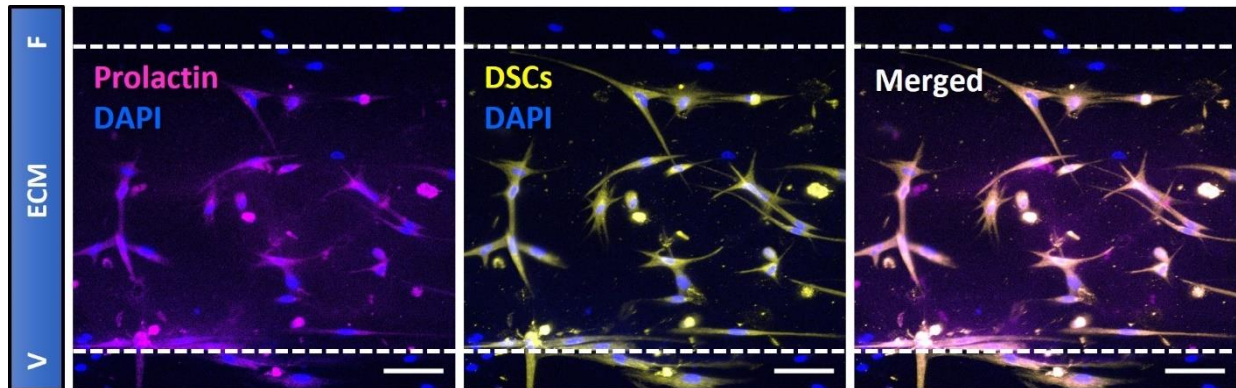
Supplementary Figure 2. Analysis of ECM stiffness in the implantation-on-a-chip. The plot shows the Young's modulus of ECM hydrogel created in the implantation-on-a-chip device prior to EVT invasion (left bar) and after 6 days of device culture (right bar). Data represent mean \pm SD from five independent devices (n=5). P values by unpaired, two-sided t-test.



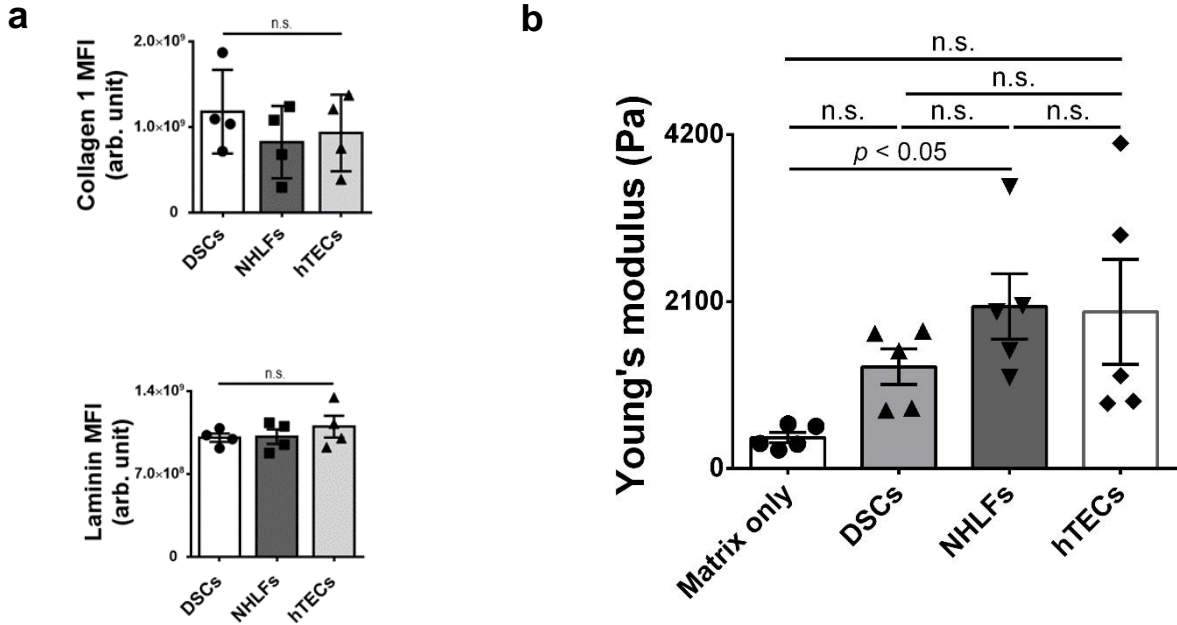
Supplementary Figure 3. Comparison of invasive activity of primary EVT_s and HTR8/SVneo cells on the ‘implantation-on-a-chip’ device. (a) Invasion of primary EVT_s in 2%FBS (first column) was compared to invasion of the HTR8-SVneo cell line in 2% FBS (second column) and 10% FBS (third column) at Day 1 (top panel) and Day 6 (bottom panel). The representative images are from three independent experiments (Scale bars, 200 μm). (b) Area invaded by the cells was quantified at day 6. Data represent mean ± SD from three independent devices (n=3). One-way ANOVA with Tukey’s multiple comparison test. Dashed lines indicate the boundary between the ECM hydrogel and the vascular compartment.



Supplementary Figure 4. Maintenance of the decidualized phenotype of stromal cells in the implantation-on-a-chip. Immunofluorescence staining of prolactin expression in DSCs cultured in the ECM compartment of the implantation-on-a-chip for 6 days without MPA, E_2 , and cAMP. The representative images are from three independent experiments. The dashed lines show the boundary between the ECM compartment and side channels. F and V represent the fetal and vascular chambers, respectively. Scale bars, 100 μ m.



Supplementary Figure 5. Analysis of ECM composition and stiffness in the presence of different cell types in the hydrogel compartment. (a) Quantification of the immunofluorescence of collagen type I and laminin in DSC-, NHLF-, and hTEC-containing devices after 6 days of culture. Data represent mean \pm SD from four independent devices (n=4). One-way ANOVA with Tukey's multiple comparison test. MFI stands for mean fluorescence intensity. (b) Young's modulus of the Matrigel-collagen I mixture hydrogel scaffold containing different cell types after 6 days of device culture. Data represent mean \pm SD from five independent devices (n=5). One-way ANOVA with Tukey's multiple comparison test. n.s.; not significant


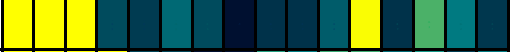

































67	Collagen type XV alpha 1 chain	-	+	-		0.006
68	Ectonucleotide pyrophosphatase/phosphodiesterase family member 2	-	+	-		0.004
69	Proteoglycan 4	+	+	-		0.019
70	Ig-like domain-containing protein	-	+	-		0.007
71	Collagen alpha-2(I) chain	-	+	-		0.049
72	Four and a half LIM domains 1	-	+	-		0.029
73	KIAA1671	-	+	-		0.001
74	Spleen trypsin inhibitor I	-	+	-		0.001
75	Protein tyrosine phosphatase receptor type G	-	+	-		0.000
76	Ig-like domain-containing protein	-	+	-		0
77	Uncharacterized protein	-	+	-		0.037
78	Tyrosine-protein kinase receptor Tie-1	-	+	-		0.000
79	Transforming growth factor beta receptor 3	-	+	-		0.002
80	14-3-3 protein epsilon	+	+	-		0
81	Alpha-1,4 glucan phosphorylase	+	+	-		0.024
82	Glycoprotein 2	-	+	-		0.004
83	Tubulin alpha-4A chain	+	+	-		0
84	Cell adhesion molecule L1 like	-	+	-		0.008
85	Complement factor properdin	-	+	-		0.006
86	Uncharacterized protein	-	+	-		0.043
87	Ig-like domain-containing protein	-	+	-		0.015
88	Uncharacterized protein	-	+	-		0
89	Cathepsin D	-	+	-		0.012
90	Polyubiquitin-B (Fragment)	+	+	-		0.002
91	Histone H3	+	+	-		0.014
92	C-type mannose receptor 2	+	+	-		0.000
93	Dickkopf-related protein 3	+	+	-		0.000
94	Extracellular matrix protein 1	-	+	-		0.038
95	L-lactate dehydrogenase	+	+	-		0.003
96	Ig-like domain-containing protein	-	+	-		0.005
97	Thyroglobulin	-	+	-		0.013
98	Epidermal growth factor receptor	+	+	-		0.031
99	Contactin-1	+	+	-		0.047
100	Complement C1s subcomponent	-	+	-		0.001
101	Vitamin K-dependent protein Z	-	+	-		0.001
102	Uncharacterized protein	-	+	-		0.002

139	Ig-like domain-containing protein	-	+	-		0.008
140	Cadherin-1	-	+	-		0.019
141	Retinol-binding protein 4	-	+	-		0.033
142	Lysosomal-associated membrane protein 2	-	+	-		0.022
143	Follistatin-related protein 1	+	+	-		0.003
144	Annexin A5	+	+	-		0.001
145	Beta-2-microglobulin	-	+	-		0.001
146	Acetyl-CoA acetyltransferase 2	-	+	-		0
147	Biotinidase	-	+	-		0.015
148	Malic enzyme	-	+	-		0.000
149	Ig-like domain-containing protein	-	+	-		0
150	Laminin subunit beta 1	-	+	-		0
151	Calsyntenin-1 (Fragment)	+	+	-		0.008
152	Ig-like domain-containing protein	-	+	-		0.001
153	Platelet-derived growth factor receptor beta	-	+	-		0.034
154	SPARC	+	+	-		0.001
155	Homogentisate 1,2-dioxygenase	+	+	-		0.004
156	Sulfhydryl oxidase	-	+	-		0
157	Uncharacterized protein	-	+	-		0.011
158	Voltage-dependent calcium channel subunit alpha-2/delta-1	+	+	-		0.003
159	Xaa-Pro dipeptidase (Fragment)	+	+	-		0.025
160	Ceruloplasmin	-	+	-		0.022
161	Integrin alpha-2	+	+	-		0.036
162	Ig-like domain-containing protein	-	+	-		0.002
163	Complement C2	-	+	-		0
164	6-phosphogluconate dehydrogenase, decarboxylating (Fragment)	+	+	-		0.037
165	Betaine-homocysteine S-methyltransferase 1	+	+	-		0.004
166	Amine oxidase	-	+	-		0.001
167	Nucleobindin-1 (Fragment)	+	+	-		0.017
168	Metalloproteinase inhibitor 1	-	+	-		0.031
169	Phospholipase A1 member A	-	+	-		0.031
170	Myosin light polypeptide 6	+	+	-		0.042
171	Olfactomedin-like protein 3	+	+	-		0.007
172	3-hydroxyanthranilate 3,4-dioxygenase	-	+	-		0.003
173	Coagulation factor X	-	+	-		0.010
174	Malate dehydrogenase, cytoplasmic	+	+	-		0.006

175	Triosephosphate isomerase	-	+	-																	0.001
176	Thrombospondin-4	-	+	-																	0.006
177	Tubulin alpha-1A chain	+	+	-																	0.008
178	Rho GDP-dissociation inhibitor 1	-	+	-																	0.001
179	Angiotensin-converting enzyme	-	+	-																	0
180	Zinc-alpha-2-glycoprotein	-	+	-																	0.041
181	Insulin-like growth factor II	+	+	-																	0.024
182	IL6ST nirs variant 3	+	+	-																	0.019
183	Serpin A3-2	-	+	-																	0.000
184	Cadherin-11	+	+	-																	0.005
185	VASN protein	-	+	-																	0.027
186	Immunoglobulin superfamily containing leucine-rich repeat protein	-	+	-																	0.015
187	Prolyl endopeptidase FAP	-	+	-																	0.000
188	Alpha-actinin-4	+	+	-																	0.047
189	PCDHGC3 protein	-	+	-																	0.003



27	Cadherin-6	3		0.046	0.597	0.694
28	Adenosine kinase (Fragment)	2		0.094	0.376	0.832
29	ADP/ATP translocase 1	1		0	0.022	0.077
30	Plexin domain-containing protein 2	3		0.008	0.172	0.124
31	Ferritin (Fragment)	1		0.008	0.813	0.138
32	Proteoglycan 4	3		0.064	0.091	0.047
33	14-3-3 protein epsilon	4		0.001	0.223	0.05
34	Alpha-1,4 glucan phosphorylase	3		0.163	0.847	0.262
35	Tubulin alpha-4A chain	12		0	0.129	0.651
36	Polyubiquitin-B (Fragment)	2		0.015	0.275	0.484
37	Histone H3	1		0.019	0.271	0.191
38	C-type mannose receptor 2	3		0.002	0.882	0.269
39	Dickkopf-related protein 3	1		0.005	0.720	0.36
40	L-lactate dehydrogenase	11		0.003	0.208	0.286
41	Epidermal growth factor receptor	1		0.055	0.504	0.189
42	Contactin-1	8		0.019	0.713	0.701
43	Protein kinase C-binding protein NELL2 (Fragment)	2		0.010	0.847	0.418
44	Histone H2B	4		0.023	0.119	0.953
45	Alpha-actinin-1	8		0.285	0.007	0.939
46	Neural cell adhesion molecule 1	4		0.006	0.282	0.292
47	Dextrin	2		0.001	0.155	0.546
48	Cadherin-13	3		0.057	0.478	0.029
49	EGF-containing fibulin-like extracellular matrix protein 2	2		0.026	0.703	0.860
50	Rab GDP dissociation inhibitor beta	6		0.004	0.749	0.234
51	Follistatin-related protein 1	4		0.031	0.324	0.836
52	Annexin A5	2		0	0.617	0.878
53	Calsyntenin-1 (Fragment)	1		0.01	0.117	0.962
54	SPARC	13		0.001	0.012	0.019
55	Homogentisate 1,2-dioxygenase	5		0.024	0.433	0.384
56	Voltage-dependent calcium channel subunit alpha-2/delta-1	10		0.008	0.335	0.828
57	Xaa-Pro dipeptidase (Fragment)	1		0.010	0.439	0.180
58	Integrin alpha-2	1		0.023	0.258	0.852
59	6-phosphogluconate dehydrogenase, decarboxylating (Fragment)	1		0.124	0.121	0.242

60	Betaine--homocysteine S-methyltransferase 1	3		0.089	0.095	0.046
61	Nucleobindin-1 (Fragment)	1		0.003	0.010	0.247
62	Myosin light polypeptide 6	3		0.293	0.004	0.207
63	Olfactomedin-like protein 3	1		0.003	0.668	0.907
64	Malate dehydrogenase, cytoplasmic	2		0.088	0.068	0.066
65	Tubulin alpha-1A chain	12		0.024	0.575	0.899
66	Insulin-like growth factor II	2		0.958	0.070	0.158
67	IL6ST nirs variant 3	1		0.294	0.098	0.401
68	Cadherin-11	2		0.456	0.848	0.016
69	Alpha-actinin-4	8		0.143	0.181	0.154



Supplementary Figure 8. Heatmap depicting the human proteins significantly regulated in abundance in the different culture conditions: i) EVT-mono, ii) EC-mono, iii) CO, and iv) TRI

No.	Description from fasta	EVT-mono	EC-mono	CO	TRI
1	Alpha-1,4 glucan phosphorylase	Yellow	Dark Blue	Yellow	Dark Blue
2	Phosphoglucomutase-1	Yellow	Dark Blue	Dark Blue	Dark Blue
3	Proteoglycan 4	Green	Green	Dark Blue	Green
4	Histone H4	Yellow	Dark Blue	Dark Blue	Dark Blue
5	Neogenin	Yellow	Dark Blue	Dark Blue	Dark Blue
6	Protein disulfide-isomerase	Yellow	Dark Blue	Dark Blue	Dark Blue
7	Interleukin-1 receptor accessory protein (Fragment)	Green	Dark Blue	Dark Blue	Dark Blue
8	Lactoylglutathione lyase	Yellow	Dark Blue	Dark Blue	Yellow
9	Plastin-2	Yellow	Dark Blue	Dark Blue	Dark Blue
10	Vasodilator-stimulated phosphoprotein	Yellow	Dark Blue	Dark Blue	Dark Blue
11	Cadherin-6	Yellow	Dark Blue	Dark Blue	Dark Blue
12	Adenosine kinase (Fragment)	Yellow	Dark Blue	Dark Blue	Dark Blue
13	ADP/ATP translocase 1	Yellow	Dark Blue	Dark Blue	Dark Blue
14	Plexin domain-containing protein 2	Yellow	Dark Blue	Dark Blue	Dark Blue
15	Ferritin (Fragment)	Yellow	Dark Blue	Dark Blue	Dark Blue
16	14-3-3 protein epsilon	Yellow	Dark Blue	Dark Blue	Dark Blue
17	Tubulin alpha-4A chain	Yellow	Dark Blue	Dark Blue	Dark Blue
18	Fructose-bisphosphate aldolase	Dark Blue	Dark Blue	Dark Blue	Dark Blue
19	Translationally-controlled tumor protein	Dark Blue	Dark Blue	Dark Blue	Dark Blue
20	Tubulin beta chain	Dark Blue	Dark Blue	Dark Blue	Dark Blue
21	Collagen alpha-2(I) chain	Dark Blue	Dark Blue	Dark Blue	Dark Blue
22	Receptor-type tyrosine-protein phosphatase S	Dark Blue	Dark Blue	Dark Blue	Dark Blue
23	Collagen alpha-1(IV) chain (Fragment)	Dark Blue	Dark Blue	Dark Blue	Dark Blue
24	Serine protease HTRA1	Dark Blue	Dark Blue	Dark Blue	Dark Blue
25	Proteasome subunit alpha type-2 (Fragment)	Dark Blue	Dark Blue	Dark Blue	Dark Blue
26	Cell adhesion molecule 1	Dark Blue	Dark Blue	Dark Blue	Dark Blue
27	Peroxiredoxin-1 (Fragment)	Dark Blue	Dark Blue	Dark Blue	Dark Blue
28	Proteasome subunit beta type-2	Dark Blue	Dark Blue	Dark Blue	Dark Blue
29	Histone H2A	Dark Blue	Dark Blue	Dark Blue	Dark Blue
30	Thioredoxin reductase 1, cytoplasmic	Dark Blue	Dark Blue	Dark Blue	Dark Blue
31	Collagen alpha-1(VI) chain	Dark Blue	Dark Blue	Dark Blue	Dark Blue
32	Neurobeachin	Yellow	Dark Blue	Dark Blue	Dark Blue
33	Carboxylic ester hydrolase	Dark Blue	Dark Blue	Dark Blue	Dark Blue
34	Isocitrate dehydrogenase [NADP] cytoplasmic	Dark Blue	Dark Blue	Dark Blue	Dark Blue
35	Tropomyosin alpha-4 chain	Dark Blue	Dark Blue	Dark Blue	Dark Blue
36	Alpha-actinin-1	Dark Blue	Dark Blue	Dark Blue	Dark Blue
37	Neural cell adhesion molecule 1	Dark Blue	Dark Blue	Dark Blue	Dark Blue
38	Annexin A5	Dark Blue	Dark Blue	Dark Blue	Dark Blue

Supplementary Figure 9. List of proteins showing significant increase or decrease between EVT monoculture (EVT-mono) and coculture of EVTs and ECs (CO) (two-sided t-test).

Uniquely expressed proteins in CO against EVT-mono													
No.	Protein name	Peptide Hits	EVT-mono				CO				P value	Known role in implantation/placental development?	Ref.
			1	2	3	4	1	2	3	4			
1	Neural cell adhesion molecule 1	4									0.000	Yes	S14
2	Annexin A5	2									0.021	Yes	S15
3	Protein S100-A9	3									0.100	Yes	S16
4	Actin, cytoplasmic 1	27									0.286	Yes	S17
5	Alpha-2-macroglobulin	11									0.312	Yes	S18
6	Golgi-associated plant pathogenesis-related protein 1	1									0.352	No	
7	Thioredoxin	3									0.785	Yes	S19
Up-regulated proteins in CO against EVT-mono													
No.	Protein name	Peptide Hits	EVT-mono				CO				P value	Known role in implantation/placental development?	Ref.
			1	2	3	4	1	2	3	4			
1	Annexin A5	2									0	Yes	S15
2	Calsyntenin-1 (Fragment)	1									0.01	No	
3	Follistatin-related protein 1	4									0.031	Yes	S20
4	Integrin alpha-2	1									0.023	Yes	S21
5	Neural cell adhesion molecule 1	4									0.006	Yes	S14
6	Triosephosphate isomerase	7									0.002	No	
7	SPARC	13									0.001	Yes	S22
8	Protein kinase C-binding protein NELL2 (Fragment)	2									0.010	No	
9	Histone H2B	4									0.023	Yes	S23
10	Moesin	4									0.003	Yes	S24
11	C-type mannose receptor 2	3									0.002	Yes	S25
12	Dextrin	2									0.001	No	
13	Histone H3	1									0.019	Yes	S26
14	Dickkopf-related protein 3	1									0.005	Yes	S27
15	Thioredoxin reductase 1, cytoplasmic	3									0.027	Yes	S28
17	Rab GDP dissociation inhibitor beta	6									0.004	No	

22	Voltage-dependent calcium channel subunit alpha-2/delta-1	10									0.008	No	
24	Polyubiquitin-B (Fragment)	2									0.015	Yes	S29
25	Isocitrate dehydrogenase [NADP] cytoplasmic	4									0.015	Yes	S30
26	Proteasome subunit alpha type	2									0.005	No	
27	L-lactate dehydrogenase	11									0.003	Yes	S31
28	Collagen alpha-1(IX) chain	2									0.004	Yes	S32
29	Tubulin alpha-1A chain	12									0.024	Yes	S33
30	Contactin-1	8									0.019	Yes	S34
31	DnaJ homolog subfamily C member 13	1									0.046	No	
15	Olfactomedin-like protein 3	1									0.003	No	
16	EGF-containing fibulin-like extracellular matrix protein 2	11									0.026	No	
17	Proteasome subunit alpha type-2 (Fragment)	1									0.043	No	
18	Homogentisate 1,2-dioxygenase	5									0.024	No	
19	Nucleobindin-1 (Fragment)	1									0.003	No	
20	Carboxylic ester hydrolase	1									0.017	No	
21	Xaa-Pro dipeptidase (Fragment)	1									0.010	No	

Down-regulated proteins in CO against EVT-mono													
No.	Protein name	Peptide Hits	EVT-mono				CO				P value	Known role in implantation /placental development?	Ref.
			1	2	3	4	1	2	3	4			
1	Cadherin-6	3									0.046	Yes	S35
2	Metalloproteinase inhibitor 1	5									0.019	Yes	S36
3	Plexin domain-containing protein 2	3									0.008	No	
4	Ferritin (Fragment)	1									0.008	Yes	S37
5	Basement membrane-specific heparan sulfate proteoglycan core protein	16									0.047	Yes	S38
6	14-3-3 protein epsilon	4									0.001	Yes	S39
7	Histone H4	3									0.009	Yes	S40
8	Vasodilator-stimulated phosphoprotein	2									0.041	Yes	S41
9	Protein disulfide-isomerase	5									0.001	Yes	S42
10	Neogenin	4									0	Yes	S43
11	Lactoylglutathione lyase	1									0.001	Yes	S44

12	ADP/ATP translocase 1	1							0	No	
13	Interleukin-1 receptor accessory protein (Fragment)	3							0.011	Yes	S45
14	Plastin-2	4							0.001	Yes	S46
15	Tubulin alpha-4A chain	12							0	No	











Supplementary Figure 10. List of proteins showing significant increase or decrease between EC monoculture (EC-mono) and coculture of EVT and EC (CO) (two-sided t-test).

Up-regulated proteins in CO against EC-mono													
No.	Protein name	Peptide Hits	EC-mono				CO				P value	Known role in implantation/placental development?	Ref.
			1	2	3	4	1	2	3	4			
1	Serine protease HTRA1	2	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Yellow	Yellow	Yellow	0.000	Yes	S47
2	Peroxiredoxin-1 (Fragment)	3	Dark Blue	Dark Blue	Dark Blue	Light Green	Yellow	Light Green	Light Green	Light Green	0.012	Yes	S48
3	Receptor-type tyrosine-protein phosphatase S	1	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0	Yes	S49
4	Translationally-controlled tumor protein	2	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.001	Yes	S50
5	Collagen alpha-1(IV) chain (Fragment)	3	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.001	Yes	S51
6	Tubulin beta chain	13	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.000	Yes	S52
7	Collagen alpha-2(I) chain	10	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.002	Yes	S53
8	Golgi-associated plant pathogenesis-related protein 1	1	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.025	No	
9	Tropomyosin alpha-4 chain	3	Light Green	Light Green	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.015	Yes	S54
10	Cell adhesion molecule 1	1	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.009	Yes	S55
11	Fructose-bisphosphate aldolase	5	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.000	Yes	S56
12	Triosephosphate isomerase	7	Light Green	Light Green	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.048	No	
13	Antithrombin-III	11	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.015	Yes	S57
14	SPARC	13	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.012	Yes	S22
15	ADP/ATP translocase 1	1	Light Green	Light Green	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.022	No	
16	Thioredoxin reductase 1, cytoplasmic	1	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.002	Yes	S28
17	Reticulocalbin-3	5	Light Green	Light Green	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.028		S58
18	Phosphoglucomutase-1	2	Dark Blue	Dark Blue	Light Green	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.030	Yes	S59
19	Isocitrate dehydrogenase [NADP] cytoplasmic	4	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.004	Yes	S30
20	Nidogen-1	7	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.003	Yes	S60
21	Biglycan	3	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.013	Yes	S61
22	Proteasome subunit alpha type	2	Dark Blue	Light Green	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.015	No	
23	Collagen alpha-1(VI) chain	20	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.027	Yes	S51
24	Proteasome subunit alpha type-2 (Fragment)	1	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.019	No	
25	Alpha-actinin-1	8	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.007	No	
26	Carboxylic ester hydrolase	1	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.006	No	
27	GTP-binding nuclear protein Ran	2	Light Green	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green	0.025	No	

28	Nucleobindin-1 (Fragment)	1		0.010	No
----	---------------------------	---	--	-------	----

Down-regulated proteins in CO against EC-mono

No.	Protein name	Peptide Hits	EC-mono				CO				P value	Known role in implantation/placental development?	Ref.
			1	2	3	4	1	2	3	4			
1	Myosin light polypeptide 6	3									0.004	Yes	S39



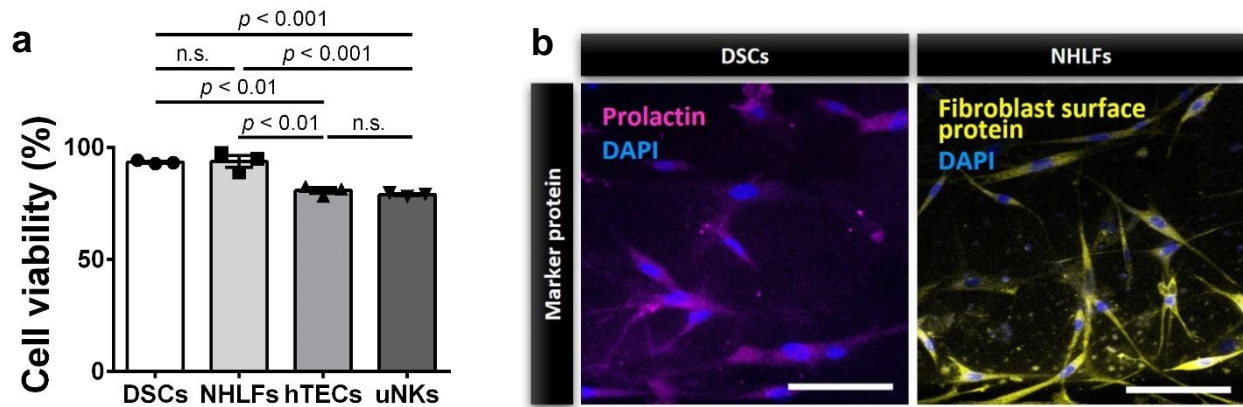
Supplementary Figure 11. Fourteen significantly upregulated endothelial proteins in the coculture configuration of the implantation-on-a-chip.

Endothelial Proteins										
No.	Protein Name	Peptide Hits	EC-Mono				Co			
			1	2	3	4	1	2	3	4
1	Thrombospondin-4	5	Grey	Dark Blue	Grey	Dark Blue	Light Green	Light Green	Light Green	Light Green
2	Pantetheinase	3	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Yellow	Light Green	Light Green	Light Green
3	Coagulation factor X	4	Dark Blue	Grey	Grey	Dark Blue	Light Green	Light Green	Light Green	Light Green
4	Apolipoprotein B-100	6	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
5	Insulin-like growth factor-binding protein 2	10	Dark Blue	Light Green	Grey	Grey	Light Green	Light Green	Light Green	Light Green
6	Rab GDP dissociation inhibitor alpha	12	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
7	Parkinson disease protein 7	4	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Yellow	Light Green	Light Green
8	Collagen alpha-1(VI) chain	37	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
9	Apolipoprotein C-III	1	Grey	Light Green	Grey	Grey	Light Green	Light Green	Light Green	Light Green
10	Collagen alpha-1(III) chain	19	Grey	Light Green	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
11	Collagen alpha-3(VI) chain	4	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Dark Blue	Light Green	Light Green
12	Prolow-density lipoprotein receptor-related protein 1	52	Grey	Grey	Grey	Dark Blue	Light Green	Light Green	Light Green	Light Green
13	Retinol-binding protein	5	Grey	Dark Blue	Grey	Grey	Light Green	Light Green	Light Green	Light Green
14	Reticulocalbin-3	8	Grey	Grey	Grey	Dark Blue	Light Green	Light Green	Light Green	Light Green

Supplementary Figure 12. Twelve significantly upregulated EVT proteins in coculture configuration of the implantation-on-a-chip.

EVT Proteins										
No.	Protein Name	Peptide Hits	EVT-Mono				Co			
			1	2	3	4	1	2	3	4
1	Thrombospondin-4	5	Light Blue	Dark Blue	Light Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
2	Pantetheinase	3	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
3	Coagulation factor X	4	Dark Blue	Light Blue	Light Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
4	Collagen alpha-1(II) chain	4	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
5	Apolipoprotein B-100	6	Dark Blue	Dark Blue	Light Blue	Light Blue	Light Green	Light Green	Light Green	Light Green
6	von Willebrand factor	122	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
7	Thrombospondin-1	13	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
8	Insulin-like growth factor-binding protein 2	10	Dark Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
9	Contactin-1	7	Light Blue	Dark Blue	Light Blue	Light Blue	Light Green	Light Green	Light Green	Light Green
10	Coactosin-like protein	5	Light Blue	Dark Blue	Dark Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green
11	Parkinson disease protein 7	4	Dark Blue	Dark Blue	Dark Blue	Light Blue	Light Green	Light Green	Light Green	Light Green
12	Dipeptidyl peptidase 2	9	Light Blue	Light Blue	Light Blue	Dark Blue	Light Green	Light Green	Light Green	Light Green

Supplementary Figure 13. Viability and marker expression of DSCs and additional cell types in the implantation-on-a-chip. (a) Quantification of the viability of DSCs and additional cell types used in the implantation-on-a-chip after 6 days of device culture. Data were obtained from LIVE/DEAD viability assay using Calcein-AM and ethidium homodimer-1 (EthD-1). Data represent mean \pm SD from three independent devices (n=3). One-way ANOVA with Tukey's multiple comparison test. (b) Immunofluorescence staining of cell-type-specific marker expression by DSCs and NHLFs used in the implantation-on-a-chip after 6 days of device culture. The representative images are from three independent experiments. Scale bars, 100 μ m.



Supplementary Table 1. Cells and authentication methods.

Cells	Vendor	Catalog number	Authentication method
HTR-8/SVneo	ATCC	CRL-3271	STR profiling in July 2016;showed a distinctly human profile (TH01: 6, 9,3;D5S818: 12;D13S317: 9, 12;D7S820: 12;D16S539: 13;CSF1PO: 12;Amelogenin: X;vWA: 13, 18;TPOX:8).
Human endometrial microvascular endothelial cells (HEMEC)	ScienCell	7010	Phenotypic and immunofluorescent characterization by company in March 2015; correct morphology and von Willebrand Factor VIII expression. HEMECs were tested negative for HIV, HBV, HCV, mycoplasma, bacteria, and fungi.
Human lung microvascular endothelial cells (HMVEC-L)	Lonza	CC-2527	Phenotypic and immunofluorescent characterization by company using von Willebrand Factor VIII.
Human brain microvascular endothelial cells (HBMVEC)	AngioPorte-omie	CAP-0002	Immunofluorescent characterization by company with antibodies specific to von Willebrand Factor VIII, Di-I-Acetylated-low density lipoprotein, and PECAM1.
Normal human lung fibroblasts	Lonza	CC-2512	Immunofluorescent characterization by company with negative expression for Factor VIII, Cytokeratin 18, and Cytokeratin 19.
Human tracheal epithelial cells	PromoCell	C-12644	Flow cytometric characterization by company using positive expression of Cytokeratin.

Supplementary Table 2. Primary and secondary antibodies for immunofluorescence staining.

Primary antibody [Clone]	Species	Dilution	Product information	Secondary antibody	Dilution
CD31 [P2B1]	Mouse	1:200	Abcam #ab24590	Goat Anti-Mouse IgG (H+L) (Alexa Fluor® Plus 555)	1:200
VE-Cadherin [D87F2]	Rabbit	1:200	Cell Signaling #2500	Goat Anti-Rabbit IgG H&L (Alexa Fluor® 647)	1:200
Cleaved Caspase-3 [Asp175]	Rabbit	1:200	Cell Signaling #9661	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:200
Ki67 [37C7-12]	Mouse	1:200	Abcam #ab245113	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:200
Cytokeratin [EPR17078]	Rabbit	1:100	Abcam #ab181598	Goat Anti-Rabbit IgG H&L (Alexa Fluor® 647)	1:200
HLA G [MEM-G/9]	Mouse	1:100	BioRad #MCA2044	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:100
VCAM-1/CD106 [6G9]	Mouse	1:200	Novus Biolog- icals #NBP1- 47491	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:200
Complement C4 [JM88-13]	Rabbit	1:200	Thermo Fisher #MA5-32856	Goat Anti-Rabbit IgG H&L (Alexa Fluor® 647)	1:200
Prolactin [PRL02]	Mouse	1:200	Thermo Fisher #MA5-11998	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:200
Fibroblast Surface Protein [1B10]	Mouse	1:200	Abcam #ab11333	Goat Anti-Mouse IgG H&L (Alexa Fluor® 647)	1:200

Supplementary References

1. Blankenship, T. N. & King, B. F. Macaque intra-arterial trophoblast and extravillous trophoblast of the cell columns and cytotrophoblastic shell express neural cell adhesion molecule (NCAM). *Anat Rec* **245**, 525-531, doi:10.1002/(SICI)1097-0185(199607)245:3<525::AID-AR9>3.0.CO;2-Q (1996).
2. Southcombe, J. H., Redman, C. W., Sargent, I. L. & Granne, I. Interleukin-1 family cytokines and their regulatory proteins in normal pregnancy and pre-eclampsia. *Clin Exp Immunol* **181**, 480-490, doi:10.1111/cei.12608 (2015).
3. Qin, F., Tian, J., Zhou, D. & Chen, L. Mst1 and Mst2 kinases: regulations and diseases. *Cell Biosci* **3**, 31, doi:10.1186/2045-3701-3-31 (2013).
4. Menkhorst, E. M. et al. Decidual-secreted factors alter invasive trophoblast membrane and secreted proteins implying a role for decidual cell regulation of placentation. *PLoS One* **7**, e31418, doi:10.1371/journal.pone.0031418 (2012).
5. Sferruzzi-Perri, A. N., Sandovici, I., Constancia, M. & Fowden, A. L. Placental phenotype and the insulin-like growth factors: resource allocation to fetal growth. *J Physiol* **595**, 5057-5093, doi:10.1113/JP273330 (2017).
6. Liu, B., Liu, L., Cui, S., Qi, Y. & Wang, T. Expression and significance of microRNA-126 and VCAM-1 in placental tissues of women with early-onset preeclampsia. *J Obstet Gynaecol Res* **47**, 2042-2050, doi:10.1111/jog.14732 (2021).
7. Giudice, L. C. et al. Identification and regulation of the IGFBP-4 protease and its physiological inhibitor in human trophoblasts and endometrial stroma: evidence for paracrine regulation of IGF-II bioavailability in the placental bed during human implantation. *J Clin Endocrinol Metab* **87**, 2359-2366, doi:10.1210/jcem.87.5.8448 (2002).
8. Taracha, A., Kotarba, G. & Wilanowski, T. Neglected Functions of TFCP2/TFCP2L1/UBP1 Transcription Factors May Offer Valuable Insights into Their Mechanisms of Action. *Int J Mol Sci* **19**, doi:10.3390/ijms19102852 (2018).
9. Altmae, S. et al. Endometrial gene expression analysis at the time of embryo implantation in women with unexplained infertility. *Molecular human reproduction* **16**, 178-187, doi:10.1093/molehr/gap102 (2010).
10. Flood-Nichols, S. K. et al. Elevated ratio of maternal plasma ApoCIII to ApoCII in preeclampsia. *Reprod Sci* **18**, 493-502, doi:10.1177/1933719110390390 (2011).
11. Wen, H., Hu, Y., Chen, L., Zhao, L. & Yang, X. miR-338-5p Targets Epidermal Growth Factor-Containing Fibulin-Like Extracellular Matrix Protein 1 to Inhibit the Growth and Invasion of Trophoblast Cells in Selective Intrauterine Growth Restriction. *Reprod Sci* **27**, 1357-1364, doi:10.1007/s43032-020-00160-3 (2020).
12. Pierzchalski, K. et al. Retinoic acid biosynthesis is impaired in human and murine endometriosis. *Biology of reproduction* **91**, 84, doi:10.1095/biolreprod.114.119677 (2014).
13. Altmae, S., Kallak, T. K., Friden, B. & Stavreus-Evers, A. Variation in hyaluronan-binding protein 2 (HABP2) promoter region is associated with unexplained female infertility. *Reprod Sci* **18**, 485-492, doi:10.1177/1933719110388849 (2011).
14. Blankenship, T. N. & King, B. F. Macaque intra-arterial trophoblast and extravillous trophoblast of the cell columns and cytotrophoblastic shell express neural cell adhesion molecule (NCAM). *Anat Rec* **245**, 525-531, doi:10.1002/(SICI)1097-0185(199607)245:3<525::AID-AR9>3.0.CO;2-Q (1996).
15. Degrelle, S. A., Gerbaud, P., Leconte, L., Ferreira, F. & Pidoux, G. Annexin-A5 organized in 2D-network at the plasmalemma eases human trophoblast fusion. *Sci Rep* **7**, 42173, doi:10.1038/srep42173 (2017).

16. Nair, R. R., Khanna, A. & Singh, K. Role of inflammatory proteins S100A8 and S100A9 in pathophysiology of recurrent early pregnancy loss. *Placenta* **34**, 824-827, doi:10.1016/j.placenta.2013.06.307 (2013).
17. Thie, M. et al. Adhesiveness of the free surface of a human endometrial monolayer for trophoblast as related to actin cytoskeleton. *Mol Hum Reprod* **3**, 275-283, doi:10.1093/molehr/3.4.275 (1997).
18. Esadeg, S., He, H., Pijnenborg, R., Van Leuven, F. & Croy, B. A. Alpha-2 macroglobulin controls trophoblast positioning in mouse implantation sites. *Placenta* **24**, 912-921, doi:10.1016/s0143-4004(03)00148-6 (2003).
19. Tonissen, K. et al. Site-directed mutagenesis of human thioredoxin. Identification of cysteine 74 as critical to its function in the "early pregnancy factor" system. *J Biol Chem* **268**, 22485-22489 (1993).
20. Mouillet, J. F. et al. The expression and post-transcriptional regulation of FSTL1 transcripts in placental trophoblasts. *Placenta* **36**, 1231-1238, doi:10.1016/j.placenta.2015.09.005 (2015).
21. Lee, C. Q. E. et al. Integrin alpha2 marks a niche of trophoblast progenitor cells in first trimester human placenta. *Development* **145**, doi:10.1242/dev.162305 (2018).
22. Jiang, Y. et al. Downregulation of SPARC expression inhibits the invasion of human trophoblast cells in vitro. *PLoS One* **8**, e69079, doi:10.1371/journal.pone.0069079 (2013).
23. Kohan-Ghadr, H. R., Kadam, L., Jain, C., Armant, D. R. & Drewlo, S. Potential role of epigenetic mechanisms in regulation of trophoblast differentiation, migration, and invasion in the human placenta. *Cell adhesion & migration* **10**, 126-135, doi:10.1080/19336918.2015.1098800 (2016).
24. Matsumoto, H., Daikoku, T., Wang, H., Sato, E. & Dey, S. K. Differential expression of ezrin/radixin/moesin (ERM) and ERM-associated adhesion molecules in the blastocyst and uterus suggests their functions during implantation. *Biology of reproduction* **70**, 729-736, doi:10.1095/biolreprod.103.022764 (2004).
25. Ono, Y. et al. CD206+ M2-Like Macrophages Are Essential for Successful Implantation. *Front Immunol* **11**, 557184, doi:10.3389/fimmu.2020.557184 (2020).
26. Jang, C. W., Shibata, Y., Starmer, J., Yee, D. & Magnuson, T. Histone H3.3 maintains genome integrity during mammalian development. *Genes Dev* **29**, 1377-1392, doi:10.1101/gad.264150.115 (2015).
27. Wu, Y. et al. Serum biomarker analysis in patients with recurrent spontaneous abortion. *Mol Med Rep* **16**, 2367-2378, doi:10.3892/mmr.2017.6890 (2017).
28. Jakupoglu, C. et al. Cytoplasmic thioredoxin reductase is essential for embryogenesis but dispensable for cardiac development. *Mol Cell Biol* **25**, 1980-1988, doi:10.1128/MCB.25.5.1980-1988.2005 (2005).
29. Wang, H. M. et al. Effect of ubiquitin-proteasome pathway on mouse blastocyst implantation and expression of matrix metalloproteinases-2 and -9. *Biol Reprod* **70**, 481-487, doi:10.1095/biolreprod.103.021634 (2004).
30. Rosario, F. J. et al. Characterization of the Primary Human Trophoblast Cell Secretome Using Stable Isotope Labeling With Amino Acids in Cell Culture. *Front Cell Dev Biol* **9**, 704781, doi:10.3389/fcell.2021.704781 (2021).
31. Auerbach, S. & Brinster, R. L. Lactate dehydrogenase isozymes in the early mouse embryo. *Exp Cell Res* **46**, 89-92, doi:10.1016/0014-4827(67)90411-9 (1967).
32. Hannan, N. J. & Salamonsen, L. A. CX3CL1 and CCL14 regulate extracellular matrix and adhesion molecules in the trophoblast: potential roles in human embryo implantation. *Biol Reprod* **79**, 58-65, doi:10.1095/biolreprod.107.066480 (2008).

33. Shukla, V. et al. Microtubule depolymerization attenuates WNT4/CaMKIIalpha signaling in mouse uterus and leads to implantation failure. *Reproduction* **158**, 47-59, doi:10.1530/REP-18-0611 (2019).
34. Tapia, A., Salamonsen, L. A., Manuelpillai, U. & Dimitriadis, E. Leukemia inhibitory factor promotes human first trimester extravillous trophoblast adhesion to extracellular matrix and secretion of tissue inhibitor of metalloproteinases-1 and -2. *Hum Reprod* **23**, 1724-1732, doi:10.1093/humrep/den121 (2008).
35. Zhou, W., Santos, L. & Dimitriadis, E. Characterization of the role for cadherin 6 in the regulation of human endometrial receptivity. *Reprod Biol Endocrinol* **18**, 66, doi:10.1186/s12958-020-00624-w (2020).
36. Zhu, J. Y., Pang, Z. J. & Yu, Y. H. Regulation of trophoblast invasion: the role of matrix metalloproteinases. *Rev Obstet Gynecol* **5**, e137-143 (2012).
37. Ng, S. W., Norwitz, S. G. & Norwitz, E. R. The Impact of Iron Overload and Ferroptosis on Reproductive Disorders in Humans: Implications for Preeclampsia. *Int J Mol Sci* **20**, doi:10.3390/ijms20133283 (2019).
38. Chui, A. et al. The expression of placental proteoglycans in pre-eclampsia. *Gynecol Obstet Invest* **73**, 277-284, doi:10.1159/000333262 (2012).
39. Gonzalez, R., Mohan, H. & Unniappan, S. Nucleobindins: bioactive precursor proteins encoding putative endocrine factors? *Gen Comp Endocrinol* **176**, 341-346, doi:10.1016/j.ygcen.2011.11.021 (2012).
40. Ghule, P. N. et al. Maternal expression and early induction of histone gene transcription factor Hinfp sustains development in pre-implantation embryos. *Dev Biol* **419**, 311-320, doi:10.1016/j.ydbio.2016.09.003 (2016).
41. Kayisli, U. A., Selam, B., Demir, R. & Arici, A. Expression of vasodilator-stimulated phosphoprotein in human placenta: possible implications in trophoblast invasion. *Mol Hum Reprod* **8**, 88-94, doi:10.1093/molehr/8.1.88 (2002).
42. Fernando, S. R. et al. Expression of membrane protein disulphide isomerase A1 (PDIA1) disrupt a reducing microenvironment in endometrial epithelium for embryo implantation. *Exp Cell Res* **405**, 112665, doi:10.1016/j.yexcr.2021.112665 (2021).
43. Zhong, S., Zou, L., Zhao, Y., Hu, B. & Xie, H. Effect of different concentrations of neogenin on proliferation, apoptosis and related proliferative factors in human trophoblasts. *J Huazhong Univ Sci Technolog Med Sci* **30**, 500-504, doi:10.1007/s11596-010-0457-x (2010).
44. Aouache, R., Biquard, L., Vaiman, D. & Miralles, F. Oxidative Stress in Preeclampsia and Placental Diseases. *Int J Mol Sci* **19**, doi:10.3390/ijms19051496 (2018).
45. Southcombe, J. H., Redman, C. W., Sargent, I. L. & Granne, I. Interleukin-1 family cytokines and their regulatory proteins in normal pregnancy and pre-eclampsia. *Clin Exp Immunol* **181**, 480-490, doi:10.1111/cei.12608 (2015).
46. Popovici, R. M. et al. Gene expression profiling of human endometrial-trophoblast interaction in a coculture model. *Endocrinology* **147**, 5662-5675, doi:10.1210/en.2006-0916 (2006).
47. Ajayi, F. et al. Elevated expression of serine protease HtrA1 in preeclampsia and its role in trophoblast cell migration and invasion. *Am J Obstet Gynecol* **199**, 557 e551-510, doi:10.1016/j.ajog.2008.04.046 (2008).
48. Morita, K. et al. Peroxiredoxin as a functional endogenous antioxidant enzyme in pronuclei of mouse zygotes. *J Reprod Dev* **64**, 161-171, doi:10.1262/jrd.2018-005 (2018).
49. Biase, F. H. et al. Fine-tuned adaptation of embryo-endometrium pairs at implantation revealed by transcriptome analyses in *Bos taurus*. *PLoS Biol* **17**, e3000046, doi:10.1371/journal.pbio.3000046 (2019).

50. Kalra, S. K., Ratcliffe, S. J., Coutifaris, C., Molinaro, T. & Barnhart, K. T. Ovarian stimulation and low birth weight in newborns conceived through in vitro fertilization. *Obstetrics and gynecology* **118**, 863-871, doi:10.1097/AOG.0b013e31822be65f (2011).
51. Shi, J. W. et al. Collagen at the maternal-fetal interface in human pregnancy. *Int J Biol Sci* **16**, 2220-2234, doi:10.7150/ijbs.45586 (2020).
52. Schatten, G., Simerly, C. & Schatten, H. Microtubule configurations during fertilization, mitosis, and early development in the mouse and the requirement for egg microtubule-mediated motility during mammalian fertilization. *Proc Natl Acad Sci U S A* **82**, 4152-4156, doi:10.1073/pnas.82.12.4152 (1985).
53. Kim, T. H. et al. Loss of HDAC3 results in nonreceptive endometrium and female infertility. *Sci Transl Med* **11**, doi:10.1126/scitranslmed.aaf7533 (2019).
54. Paule, S., Li, Y. & Nie, G. Cytoskeletal remodelling proteins identified in fetal-maternal interface in pregnant women and rhesus monkeys. *J Mol Histol* **42**, 161-166, doi:10.1007/s10735-011-9319-5 (2011).
55. Wang, Y., Ma, C. H. & Qiao, J. [Differential expression of microRNA in eutopic endometrium tissue during implantation window for patients with endometriosis related infertility]. *Zhonghua Fu Chan Ke Za Zhi* **51**, 436-441, doi:10.3760/cma.j.issn.0529-567X.2016.06.007 (2016).
56. Myers, J. E. et al. Integrated proteomics pipeline yields novel biomarkers for predicting preeclampsia. *Hypertension* **61**, 1281-1288, doi:10.1161/HYPERTENSIONAHA.113.01168 (2013).
57. Bashiri, A., Halper, K. I. & Orvieto, R. Recurrent Implantation Failure-update overview on etiology, diagnosis, treatment and future directions. *Reprod Biol Endocrinol* **16**, 121, doi:10.1186/s12958-018-0414-2 (2018).
58. Ma, X. et al. Decidual cell polyploidization necessitates mitochondrial activity. *PLoS One* **6**, e26774, doi:10.1371/journal.pone.0026774 (2011).
59. Castel, G. et al. Induction of Human Trophoblast Stem Cells from Somatic Cells and Pluripotent Stem Cells. *Cell Rep* **33**, 108419, doi:10.1016/j.celrep.2020.108419 (2020).
60. Bilban, M. et al. Identification of novel trophoblast invasion-related genes: heme oxygenase-1 controls motility via peroxisome proliferator-activated receptor gamma. *Endocrinology* **150**, 1000-1013, doi:10.1210/en.2008-0456 (2009).
61. San Martin, S. & Zorn, T. M. The small proteoglycan biglycan is associated with thick collagen fibrils in the mouse decidua. *Cell Mol Biol (Noisy-le-grand)* **49**, 673-678 (2003).