

Supplemental Figure S1. Cell viability images and quantification under Gln deprivation *Related to Figure 1*

(A) Representative brightfield images of cell viability and confluency of D5 mesoderm, D3 endoderm, and D5 ectoderm grown in GIn-supplemented (Ctr), GIn-free (-GIn), or glutaminolysis-inhibited (+GLS1i) conditions. Scale bar indicates 500 µm.

(B) Cell viability of UCLA1-derived D5 mesoderm (n=2) and ectoderm cells relative to Ctr.

(C) Immunoblot and densitometry quantification of phosphorylated pyruvate dehydrogenase (PDH) and total PDH expression after 24h 1 mM DCA treatment (n=2).

(D-F) Volcano plots of differential metabolite abundance in (D) -Gln vs Ctr, (E) GLS1i vs Ctr, and (F) -Gln vs GLS1i conditions, quantified by UHPLC-MS. Differential fold change is plotted against raw p values for each metabolite comparison.

Data represent mean \pm SD of n \geq 3 biological replicates unless indicated otherwise. $*p \leq 0.05$; $**p \leq 0.01$; $***p \leq 0.001$. The p values were determined by (**B**) one-way ANOVA with correction for multiple comparisons, or (**D-F**) using the Student's t test



Supplemental Figure S2. Tri-lineage metabolomics profiles in GIn-free media

Related to Figure 2

(A-B) Immunoblot of GS expression in (A) UCLA1-derived ectoderm and (B) H9-derived mesoderm and endoderm cells differentiated in GIn-free media.

(C) Intracellular amounts of Gln, α -KG, ¹³C₅-Gln, and ¹³C₅- α -KG derived from ¹³C₅-glutamic acid in D3 mesoderm, endoderm, and ectoderm cells grown in Gln-supplemented or Gln-free media.

(D) Percentage of (Left) D5 mesoderm (SNAI2/SLUG⁺) and (Right) D3 endoderm (SOX17⁺) cells grown in GIn-supplemented or GIn-free conditions with 50 μM DON or 1 mM MSO.

(E) Verification of intracellular uptake and conversion to related metabolites upon respective metabolite supplementations (denoted in red), quantified by UHPLC-MS. Intensity values represent area under the curve (arbitrary units).

(F) (Left) Cell viability of UCLA1-derived D5 ectoderm cells differentiated in Gln-starvation and supplemented with GlcN6P, dm- α -KG, nucleosides, or GSH. (Right) Percentage of D5 ectoderm (PAX6⁺MAP2B⁺) cells grown in Gln-starvation with added GlcN6P or both GlcN6P and nucleosides (nuc). Data represent mean ± SD of n = 3 biological replicates. *p ≤ 0.05; **p ≤ 0.01; ***p ≤ 0.001; ****p ≤ 0.0001. The p values were determined using (C, D, F) one-way ANOVA with correction for multiple comparisons or (E) Student's t test.



Supplemental Figure S3. Cell cycle distribution, cell proliferation, and nascent protein synthesis during GIn repletion following 14h GIn-starvation

Related to Figure 3

(A) Flow cytometry cell cycle distribution of ectoderm cells differentiated in continuous Gln-starvation for D1 (24h), D2 (48h), and D3 (72h), colored *p symbols represent corresponding cell cycle stage with significant difference across indicated conditions.

(B) qRT-PCR analysis of *MAP2B*, *PAX6*, and *OTX2* expression in D1 (24h) ectoderm cells grown in GInstarvation.

(C) Flow cytometry cell cycle distribution of D5 ectoderm cells after initial 24h Gln-starvation (-G+M \rightarrow Ctr). (D) (Left) Cell viability and (Right) percentage of UCLA1-derived D5 ectoderm (PAX6⁺MAP2B⁺) cells after initial 24h Gln-starvation (G+M \rightarrow Ctr).

(E) Percentage of pluripotent (OCT4⁺SOX2⁺) and endoderm/mesoderm (SOX17⁺CD34⁺) biomarkers in D5 ectoderm cells after initial 24h Gln-starvation (-G+M \rightarrow Ctr).

(F) qRT-PCR analysis of *GLUL* (encoding GS) in H9 hPSCs expressing shRNA targeting *GLUL* (shRNA *GLUL*1, clone 1; shRNA *GLUL*2, clone 2) or a non-targeting control (NTC).

(G) Representative brightfield images of H9-derived ectoderm cells expressing shRNA targeting *GLUL* (shRNA *GLUL*) or a non-targeting control (NTC) grown continuously in Gln-free media at D1 (24h) and D5 (120h). Scale bar indicates 500 µm.

(H) Cell viability and percentage of ectoderm (PAX6⁺MAP2B⁺, PAX6⁺NESTIN⁺) and pluripotent (OCT4⁺NANOG⁺, OCT4⁺TRA1-81⁺, OCT4⁺SOX2⁺) biomarkers in D5 ectoderm cells after initial 14h GInstarvation (-G+M \rightarrow Ctr) or metabolite-supplemented GIn-starvation (-G+M+GlcN-6P+nuc \rightarrow Ctr).

(I) Cell proliferation (EdU+ staining) in D3 and D5 ectoderm cells after initial 14h Gln-starvation. (J) Nascent protein synthesis (relative AHA staining) in D3 and D5 ectoderm cells after initial 14h Gln-starvation.

Data represent mean \pm SD of n \geq 3 biological replicates. $*p \leq 0.05$; $**p \leq 0.01$; $***p \leq 0.001$; $****p \leq 0.001$; $****p \leq 0.0001$. The p values were determined by (**A**, **C**) two-way ANOVA, (**B**) Student's t test, (**D-F**, **H-J**) or one-way ANOVA with correction for multiple comparisons.



Supplemental Figure S4. mTORC1 inactivation under GIn-starvation

Related to Figure 4

(A) (Left) Flow cytometry quantification and (Right) representative immunoblot of mTORC activation (pS6K1-Thr389) immediately after 24h Gln-starved ectoderm differentiation.

(B) Flow cytometry traces and quantification of %FLAG⁺ cells in H9-derived Empty Vector, WT-Raptor, and Raptor-Rheb15 hPSC lines.

(C) (Left) Flow cytometry quantification and (Right) representative immunoblot of mTORC activation (pS6K1-Thr389) immediately after 14h Gln-starved ectoderm differentiation in H9-derived WT-Raptor or Raptor-Rheb15 hPSC lines.

(D) Flow cytometry cell cycle distribution of H9-derived D5 WT-Raptor or Raptor-Rheb15 ectoderm cells after initial 14h GIn-starvation (-G+M→Ctr).

(E) Cell viability of H9-derived D5 WT-Raptor or Raptor-Rheb15 ectoderm cells after initial 14h GInstarvation (-G+M \rightarrow Ctr).

(F) (Left) Cell viability and (Right) percentage of UCLA1-derived D5 WT-Raptor or Raptor-Rheb15 ectoderm (PAX6⁺MAP2B⁺FLAG⁺) cells after initial 14h GIn-starvation (-G+M→Ctr).

(G) Percentage of UCLA1-derived D5 WT-Raptor or Raptor-Rheb15 ectoderm (PAX6⁺MAP2B⁺FLAG⁺, SOX2⁻OCT4⁺FLAG⁺) cells after initial 14h GIn-starvation with increasing concentrations of exogenous GIn supplementation.

(H) Cell viability of (Left) H9 or (Right) UCLA1-derived D5 ectoderm cells after initial 14h GIn-starvation with increasing concentrations of exogenous GIn supplementation.

(I) (Left) Cell viability and (Right) percentage of pluripotent (OCT4⁺NANOG⁺, OCT4⁺SOX2⁺, OCT4⁺TRA1-81⁺) biomarkers in H9-derived D5 ectoderm cells after initial (-G+M→Ctr), intermediate (Ctr→-G+M→Ctr), or late (Ctr→-G+M) 14h Gln-starvation.

(J) Cell viability in (Left) H9 or (Right) UCLA1-derived D5 ectoderm differentiation after initial 14h GInstarvation, Arg deprivation, or Leu deprivation in WT-Raptor or Raptor-Rheb15 hPSC lines.

(K) Representative immunofluorescence images of mTORC activation in H9-derived ectoderm cells grown for 14h in GIn-supplemented (Ctr), Arginine-free (-Arginine), Leucine-free (-Leucine), or 200 nM rapamycin treatment (Ctr+Rapa). DAPI (blue, nucleus); ACTIN (green, cytoskeleton); pS6K1-Thr389 (yellow, mTORC1 activation). Scale bar indicates 50 µm.

(L) Percentage of pluripotent (OCT4⁺NANOG⁺, OCT4⁺SOX2⁺, OCT4⁺TRA1-81⁺) biomarkers in H9derived WT-Raptor or Raptor-Rheb15 D5 mesoderm and D3 endoderm cells.

(M) Representative immunofluorescence images of mTORC activation in H9-derived (Left) mesoderm and (Right) endoderm cells grown for 24h in Gln-supplemented (Ctr), Gln-free (-Gln), Gln-starvation (-G+M), or 200 nM rapamycin treatment (Ctr+Rapa). DAPI (blue, nucleus); ACTIN (green, cytoskeleton); pS6K1-Thr389 (yellow, mTORC1 activation). Scale bar indicates 50 µm.

Data represent mean \pm SD or (**A**, **C**) min and max of $n \ge 3$ biological replicates. $*p \le 0.05$; $**p \le 0.01$; $***p \le 0.001$; $***p \le 0.0001$. The p values were determined by (**A-C**, **E**, **H-J**) one-way ANOVA, (**D**, **F-G**) two-way ANOVA with correction for multiple comparisons, or (L) Student's t test.



Supplemental Figure S5. Immunofluorescence images for biological and technical controls *Related to Figure 3 and 4*

(A) Representative immunofluorescence images of pluripotent biomarkers in (Top) H9 hPSCs grown in mTESR, and H9-derived (Middle) D3 endoderm and (Bottom) D5 mesoderm cells grown in GIn-supplemented (Ctr), GIn-free (-GIn), or GIn-repletion follow initial 14h GIn-starvation (-G+M→Ctr). OCT4 (magenta, pluripotent); NANOG (cyan, pluripotent); SOX2 (green, pluripotent); DAPI (blue, nucleus). Scale bar indicates 50 μm.

(B) Representative immunofluorescence images of tri-lineage biomarkers in (Top) H9 hPSCs grown in mTESR, and H9-derived (Middle) D3 endoderm and (Bottom) D5 ectoderm cells grown in Gln-supplemented (Ctr), Gln-free (-Gln), or Gln-repletion follow initial 14h Gln-starvation (-G+M→Ctr). TBXT (cyan, mesoderm); SOX17 (green, endoderm); PAX6 (magenta, ectoderm); DAPI (blue, nucleus). Scale bar indicates 50 µm.

(C) Representative immunofluorescence images of pluripotent biomarkers in H9-derived D5 (Left) WT-Raptor or (Right) Raptor-Rheb15 ectoderm cells grown in Gln-supplemented (Ctr) or Gln-repletion follow initial 14h Gln-starvation (-G+M→Ctr). DAPI (blue, nucleus;) NANOG (cyan, pluripotent); OCT4 (magenta, ectoderm); FLAG (grey, transduced cell line reporter). Scale bar indicates 50 µm.



phospho-S6K1 (Thr389)-PE-Texas Red phospho-S6K1 (Thr389)-PE-Texas Red

Supplemental Figure S6. Flow cytometry gating strategy
Related to Figure 3 and 4
(A) Gating strategy for flow cytometry analysis. This figure reflects gating strategy used for flow cytometry quantifications in Figures 1-4, S1-S4.