

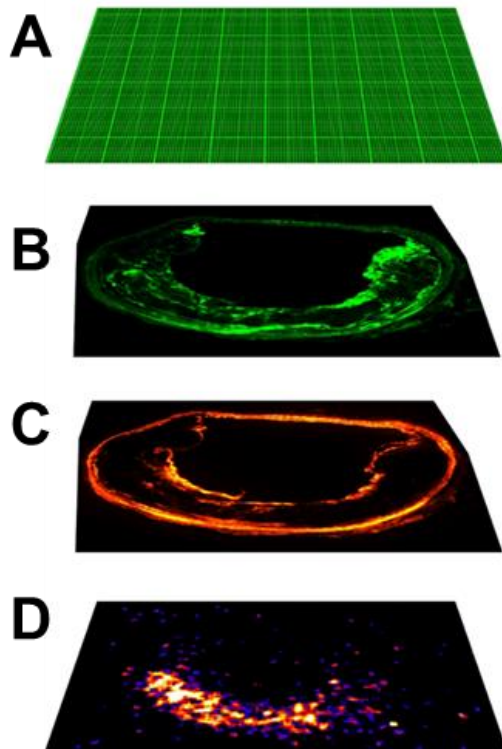
## Supplemental Tables

**Supplemental Table 1.** List of reagents used in the study.

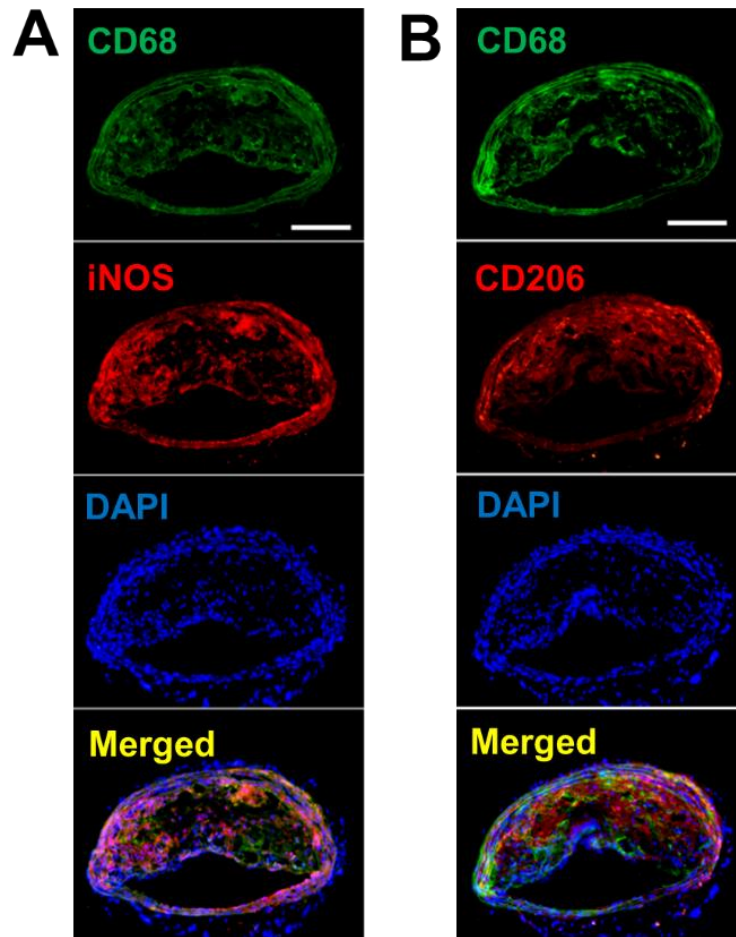
Reagent	Source	Catalog number
<sup>14</sup> C-acetate	Perkin - Elmer	NEC553250UC
Anti-mouse $\alpha$ -smooth muscle actin ( $\alpha$ SMA, clone: 1A4), Cy3-conjugated	Sigma	C6198
Anti-mouse CD68 (clone: FA11)	BioRad	MCA1957
Anti-mouse CD206	BioRad	MCA2235T
Anti-mouse iNOS	Abcam	ab3523
Anti-rat IgG, Cy2-labeled, Host: donkey	Jackson ImmunoResearch	712-225-150
DMEM/F12	Gibco	11320-033
Fetal bovine serum (FBS)	Gibco	10082147
Glucose	Gibco	A24940-01
Glutamine	ThermoFisher	25030081
HEPES buffer	Gibco	15630-080
LPS from <i>E. coli</i> O111:B4	Millipore	437627
Non-essential amino acids	Gibco	11140-050
Movat's pentachrome	ScyTek Laboratories	MPS-2
Optimal cutting temperature (OCT)	Fisher HealthCare	4585
RPMI-1640 (+L-glutamine, -D-glucose)	Gibco	11879-020
Penicillin	Gibco	15070-063
PicoGreen reagent	Invitrogen	P11496
ProLong™ Gold Antifade with (DAPI)	Invitrogen	P36931
QuantiTect® Reverse Transcription Kit	QIAGEN	205313
Recombinant human interferon- $\gamma$	PeproTech	300-02
Recombinant human IL-4	PeproTech	200-04
Recombinant murine interferon- $\gamma$	PeproTech	315-05
Recombinant murine IL-4	PeproTech	214-14
Sodium pyruvate	Gibco	11360-070
Streptomycin	Gibco	15070-063
Thioglycolate	Sigma	T9032-500G
TRIzol® reagent	Invitrogen	15596026

**Supplemental Table 2.** List of TaqMan primers used for quantitative RT-PCR.

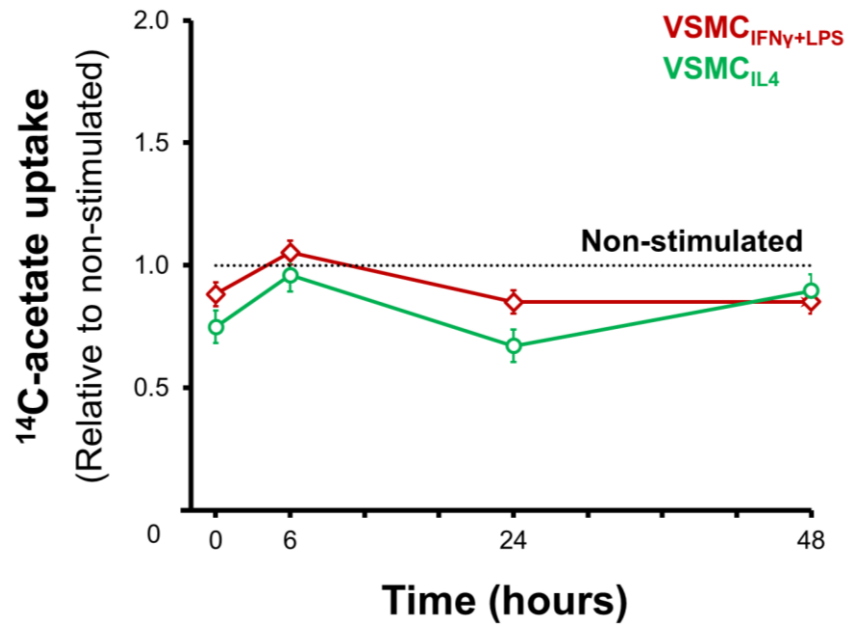
Species	Assay name	Gene full name	Assay ID
Mouse	<i>Arg1</i>	Arginase, liver	Mm00475988_m1
	<i>Arg2</i>	Arginase type II	Mm00477592_m1
	<i>Cd36</i>	CD36 molecule	Mm01135198_m1
	<i>Chi3l3</i>	Chitinase-like 3	Mm00657889_mH
	<i>Cxcl9</i>	Chemokine (C-X-C motif) ligand 9	Mm00434946_m1
	<i>Cxcl10</i>	Chemokine (C-X-C motif) ligand 10	Mm00445235_m1
	<i>Il1b</i>	Interleukin 1	Mm00434228_m1
	<i>Nos2</i>	Nitric oxide synthase 2, inducible	Mm00440502_m1
	<i>Rn18s</i>	18S ribosomal RNA	Mm03928990_g1
	<i>Tfrc</i>	Transferrin receptor	Mm00441941_m1
	<i>Tgfb1</i>	Transforming growth factor, $\beta$ 1	Mm01178820_m1
	<i>Tnf</i>	Tumor necrosis factor	Mm00443258_m1
	<i>Slc16a1</i>	Solute carrier family 16 (monocarboxylic acid transporters), member 1	Mm01306379_m1
	<i>Slc16a3</i>	Solute carrier family 16 (monocarboxylic acid transporters), member 3	Mm00446102_m1
<i>Slc16a7</i>	Solute carrier family 16 (monocarboxylic acid transporters), member 7	Mm00441442_m1	
Human	IL1B	Interleukin 1 $\beta$	Hs01555410_m1
	NOS2	Nitric oxide synthase 2	Hs01075529_m1
	MRC1	Mannose receptor C-type 1	Hs00267207_m1
	CD163	CD163 molecule	Hs00174705_m1
	CD68	CD68 molecule	Hs_00154355_m1
	18S	Eukaryotic 18S rRNA	Hs99999901_s1



**Supplemental Figure 1: Quantification of CD68<sup>+</sup> and αSMA<sup>+</sup> areas in plaque regions with different levels of <sup>14</sup>C-acetate uptake.** A 5x5 μm grid (A) was applied onto overlaid CD68 and αSMA (B and C) immunostainings and high-resolution autoradiography (D) images. <sup>14</sup>C-acetate intensity data were in each 5x5 μm box were ranked from the lowest to the highest and assigned a quartile (Q1 = lowest, Q4 = highest). CD68<sup>+</sup> and αSMA<sup>+</sup> percent areas were determined in each quartile of <sup>14</sup>C-acetate uptake.



**Supplemental Figure 2: Presence of M1-like and M2-like polarized macrophages in murine brachiocephalic plaques.** Both M1-like macrophages, identified as CD68<sup>+</sup>/iNOS<sup>+</sup> (**A**), and M2-like macrophages, identified as CD68<sup>+</sup>/CD206<sup>+</sup> (**B**), are present in murine brachiocephalic plaques. Scale bars represent 200 μm.



**Supplemental Figure 3: Similar uptake of <sup>14</sup>C-acetate in VSMCs stimulated by IFN- $\gamma$  + LPS or IL-4.** Stimulation of VSMCs with IFN- $\gamma$  + LPS or IL-4 does not significantly alter the uptake of <sup>14</sup>C-acetate over 48 hours.