

Supporting information accompanying research article titled

Glucose-6-phosphatase catalytic subunit 2 negatively regulates glucose oxidation and insulin secretion in pancreatic β -cells

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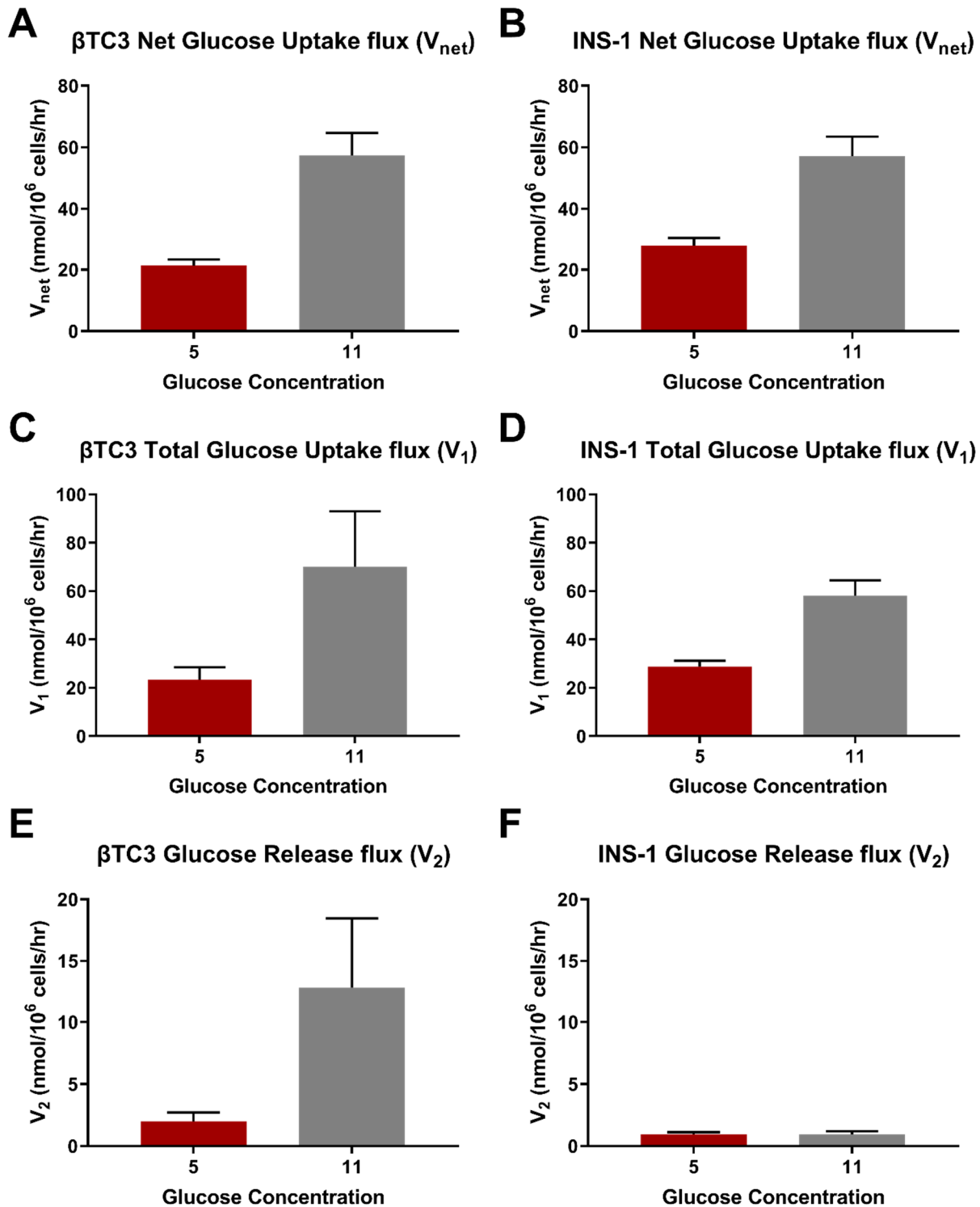


Figure S1: Measurement of glucose uptake and release fluxes in β TC3 and INS-1 832/13 rat insulinoma cells

Measurement of (A-B) net glucose uptake flux (v_{net}), (C-D) total glucose uptake flux (v_1), and (E-F) total glucose release flux (v_2) at 5 and 11 mM glucose concentrations in β TC3 and INS-1 832/13 cells. Data represent means \pm SEM (n=3).

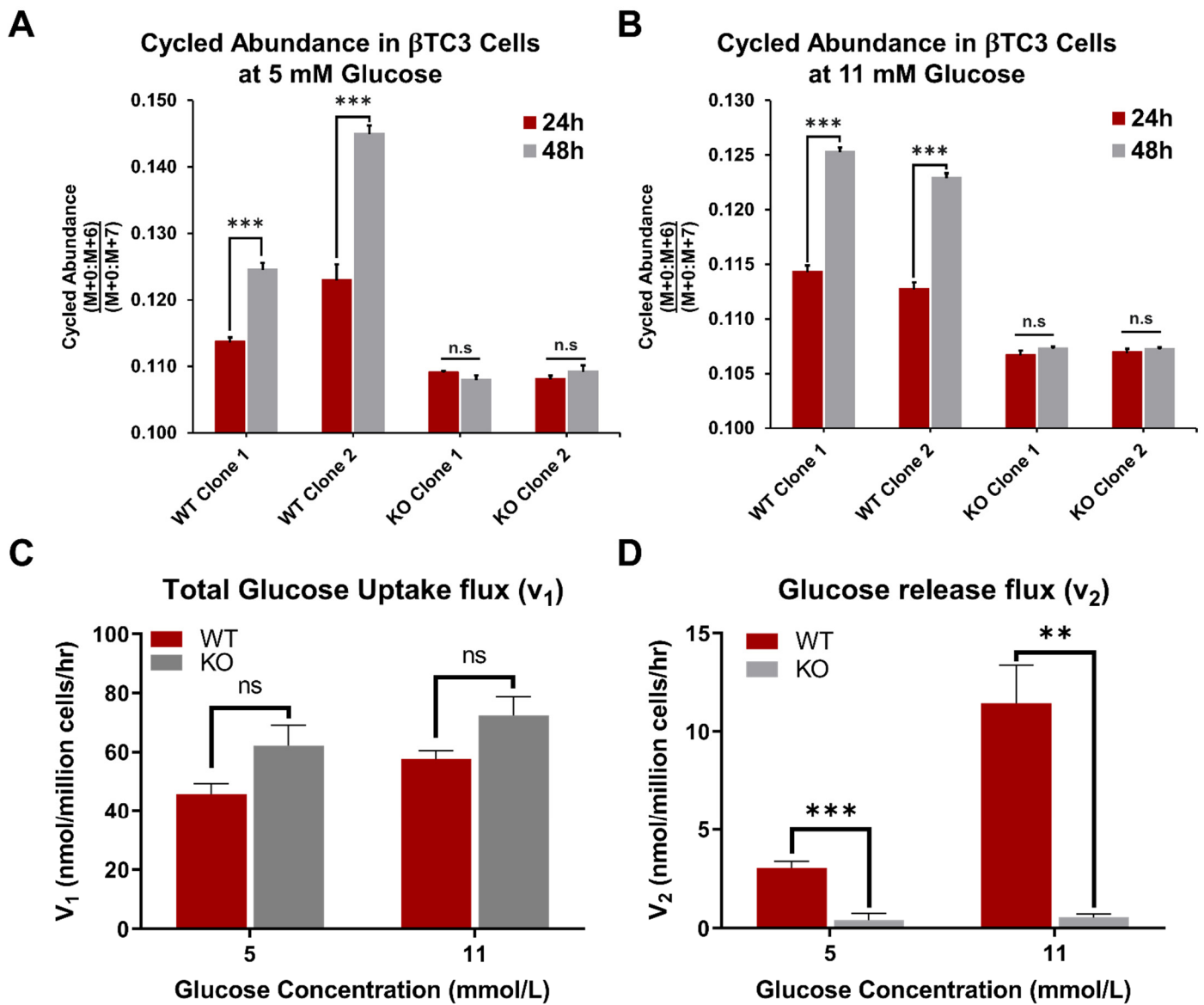


Figure S2: Cycled abundance in two biological isolates of *G6pc2* WT and KO β TC3 single cell clones at 5 and 11 mM glucose concentrations

Cycled abundance, which is the ratio M+0:M+6 isotopomers to M+0:M+7 isotopomers, measured in cell media taken at 24h and 48h after incubation with [1,2,3,4,5,6,6- 2 H $_7$]glucose at (A) 5 mM and (B) 11 mM glucose concentrations. Data represent means \pm SEM, *** p <0.01 (n=3). (C) Total glucose uptake flux (v_1) and (D) Total glucose release flux (v_2) measured in β TC3 *G6pc2* WT and KO cells. Data represent means \pm SEM (n=3).

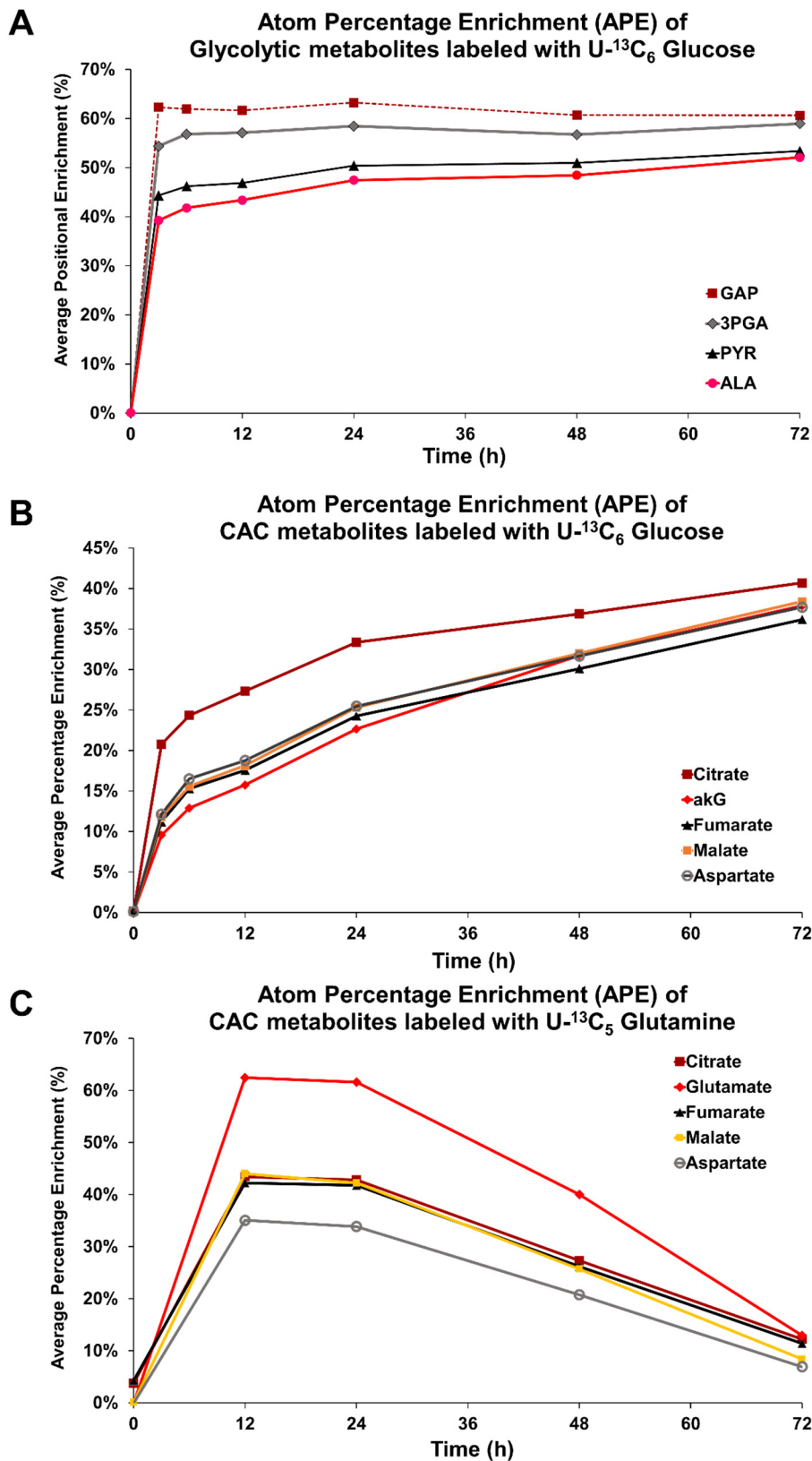
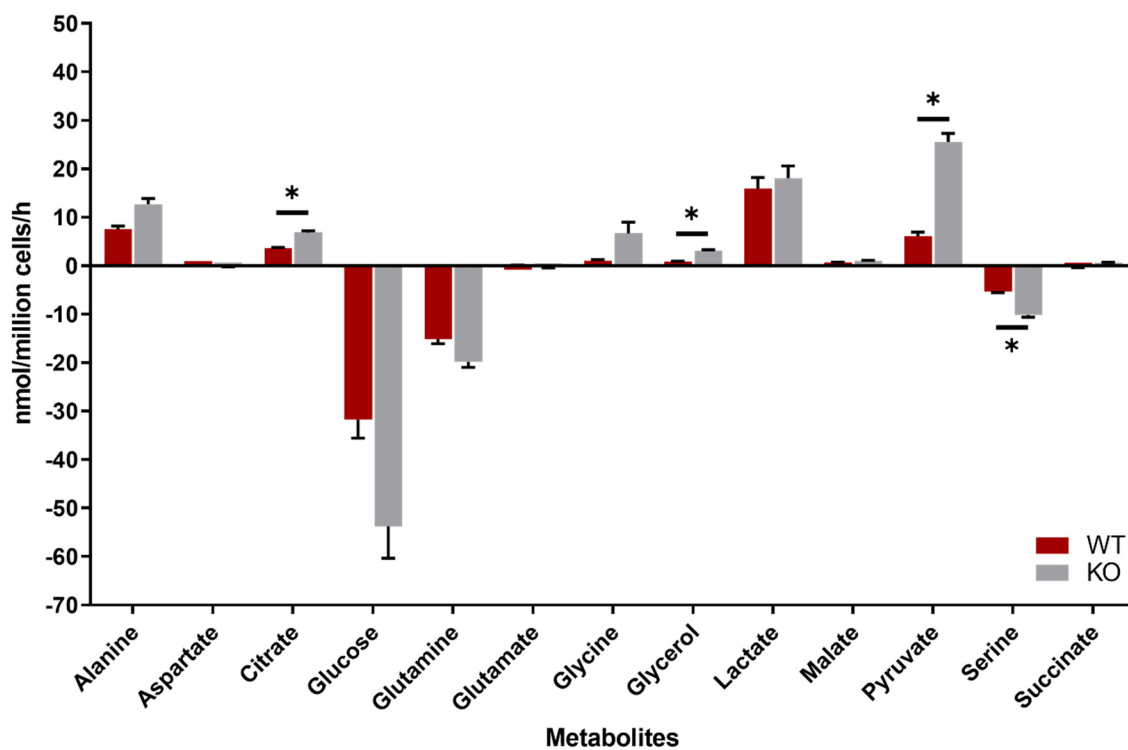


Figure S3: Atom percentage enrichment (APE) in metabolites over time to determine isotopic steady state in isotope labelling experiments

A) APE in glycolytic metabolites reaches steady state within 24h when labelled with 11 mM [U-¹³C₆]glucose.
 B) APE in CAC metabolites continues to increase in enrichment over 72h when labelled with 11 mM [U-¹³C₆]glucose.
 C) APE in CAC metabolites reaches steady state between 12 to 24h when labelled with 2 mM [U-¹³C₅]glutamine.

A Absolute Extracellular Fluxes - 5 mM Glucose



B Absolute Extracellular Fluxes - 11 mM Glucose

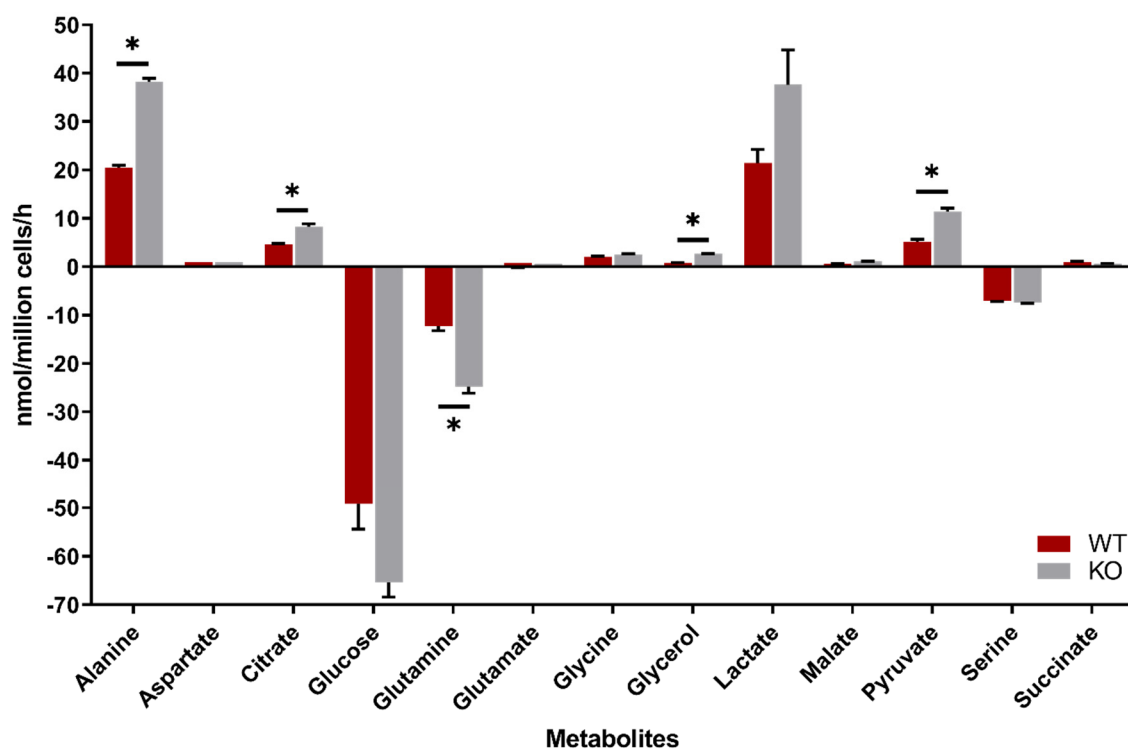


Figure S4: Extracellular uptake and excretion rates in *G6pc2* WT and KO β TC3 cells

Extracellular uptake and excretion measured in *G6pc2* WT and KO β TC3 cells incubated at (A) 5 and (B) 11 mM glucose concentrations. Positive values represent excretion fluxes while negative values indicate net uptake of metabolite. Data represent means \pm SEM, * p < 0.05 (n=3).

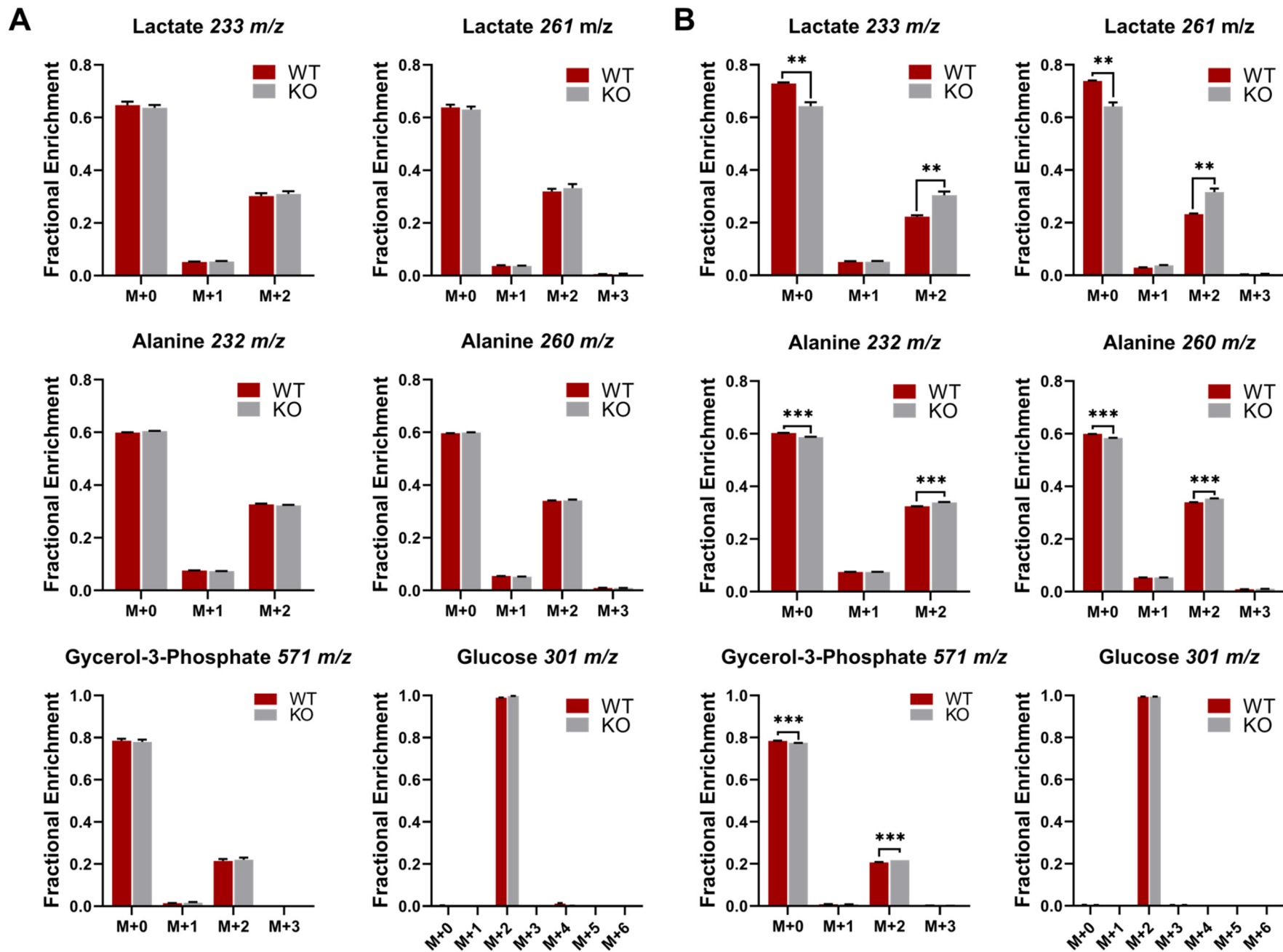


Figure S5: Enrichment of glycolytic metabolites in *G6pc2* WT and KO β TC3 cells labeled with $[1,2-^{13}\text{C}_2]$ glucose

Intracellular enrichment in glycolytic metabolites measured in *G6pc2* WT and KO β TC3 cells incubated with (A) 5 and (B) 11 mM $[1,2-^{13}\text{C}_2]$ glucose. Data represent means \pm SEM, *** p <0.01, ** p <0.05 (n =3).

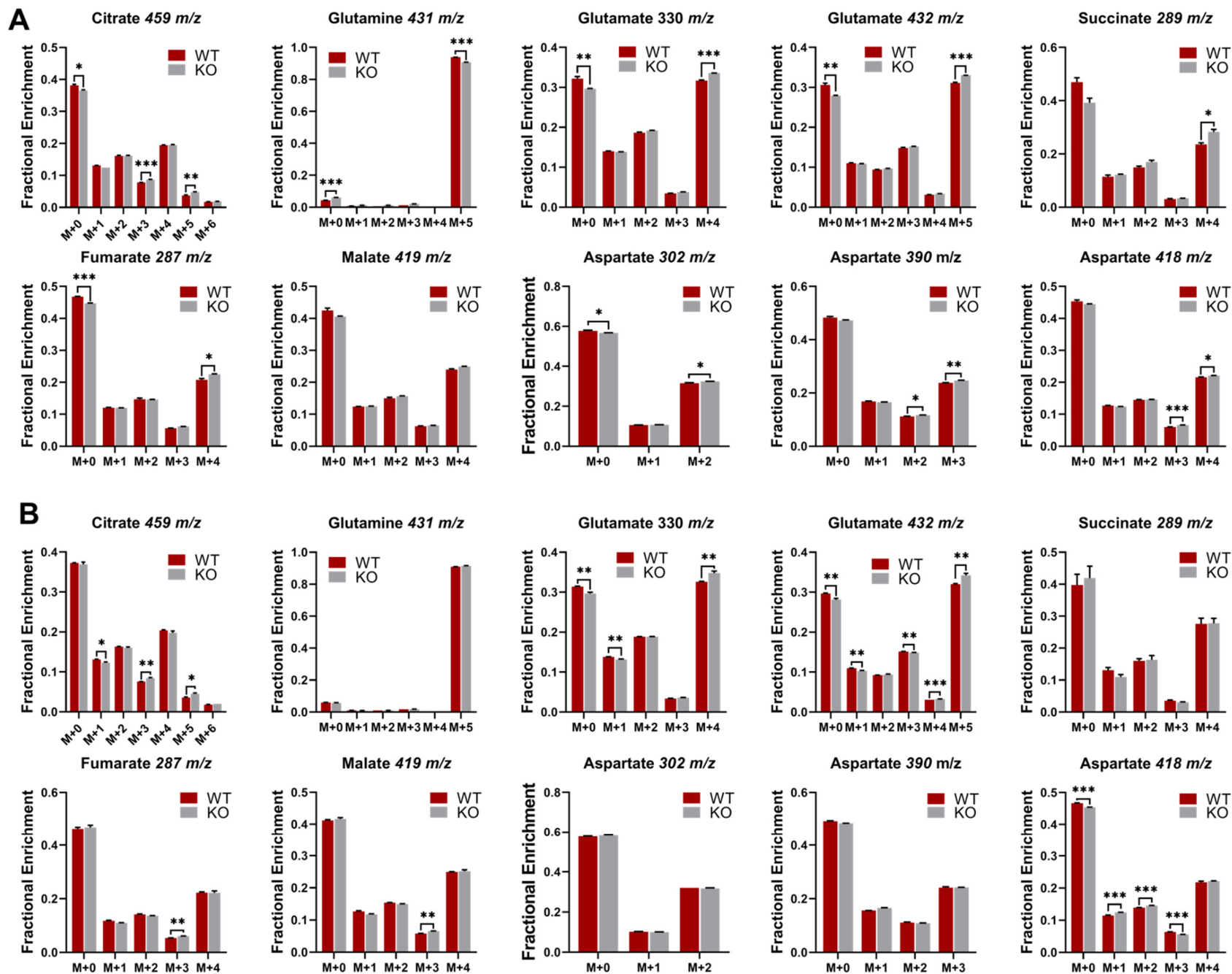


Figure S6: Enrichment of CAC metabolites in *G6pc2* WT and KO β TC3 cells labeled with 2 mM $[U-^{13}C_5]$ glutamine
 Intracellular enrichment in CAC metabolites measured in *G6pc2* WT and KO β TC3 cells incubated with (A) 5 and (B) 11 mM glucose. Data represent means \pm SEM, *** p <0.01, ** p <0.05, * p <0.01 (n =3).

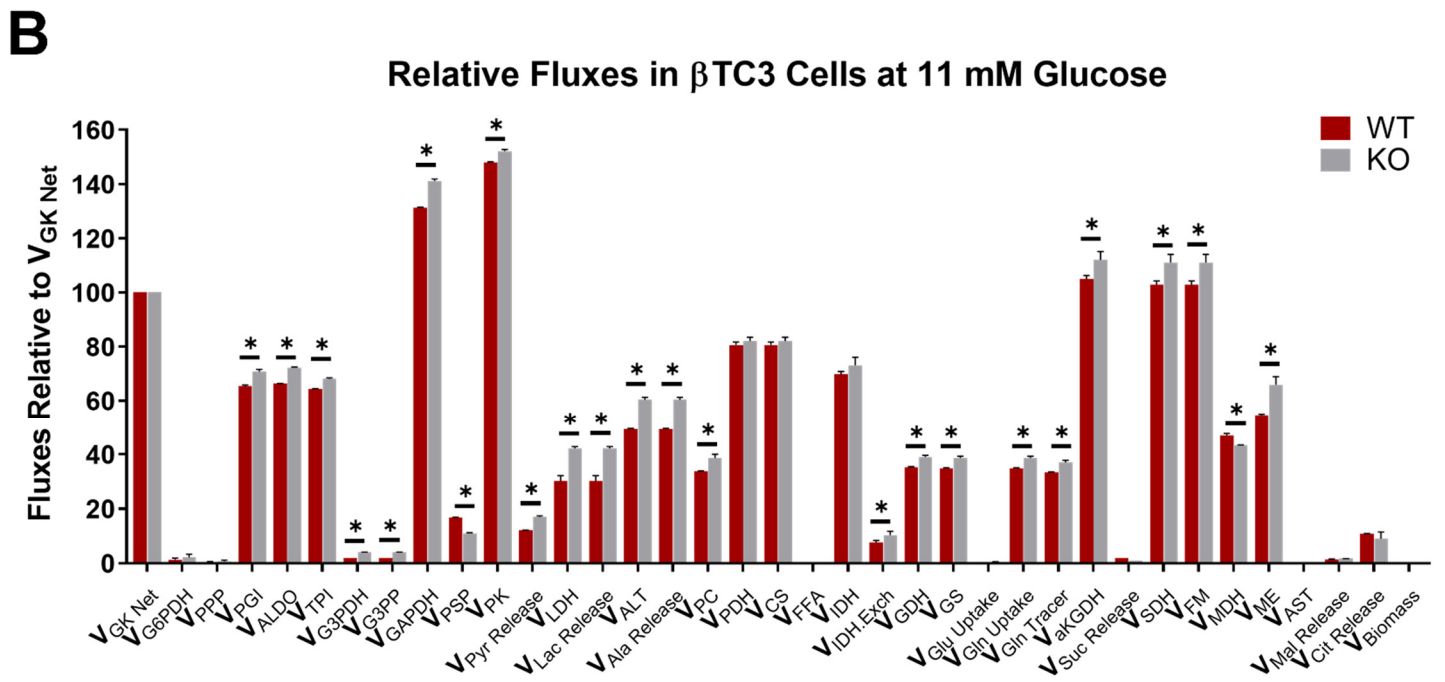
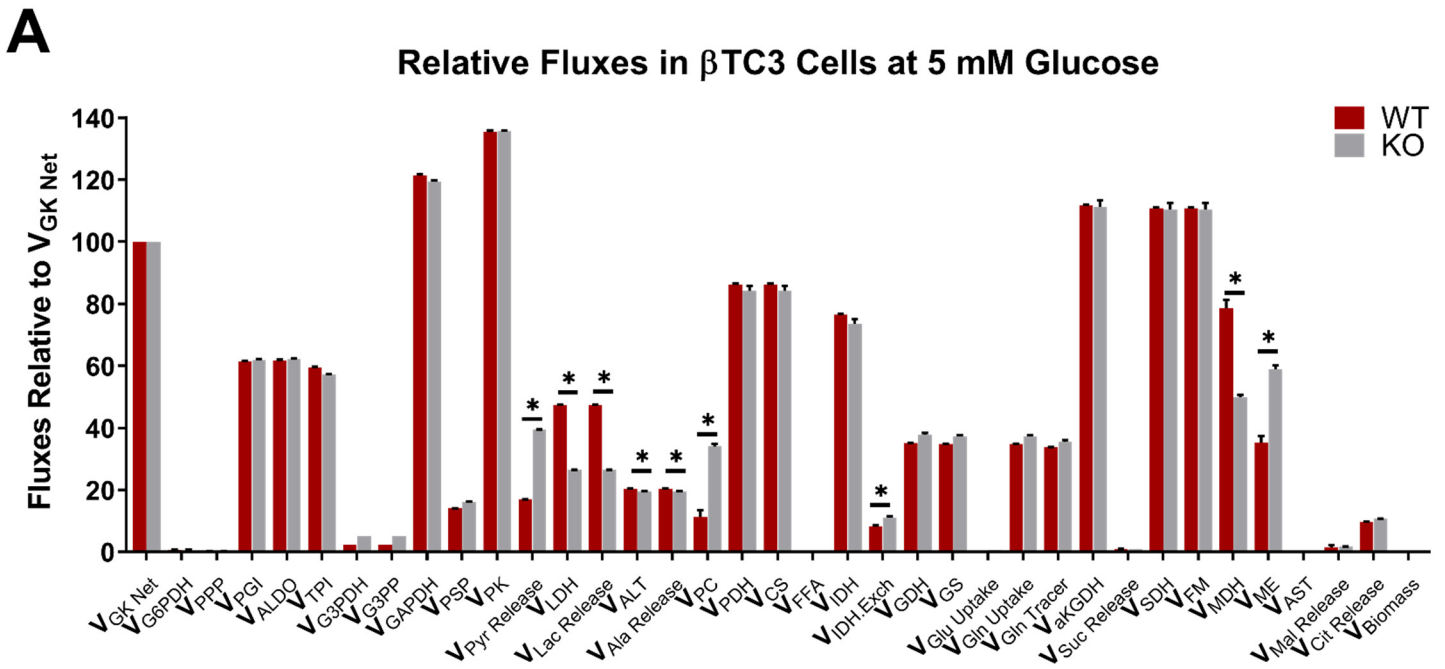


Figure S7: Metabolic fluxes relative to net glucose uptake in *G6pc2* WT and KO β TC3 cells
 Metabolic fluxes relative to net glucose uptake ($V_{GK\ Net}$) in *G6pc2* WT and KO β TC3 cells at (A) 5 mM and (B) 11 mM (right) glucose concentrations estimated using MFA. Data represent means \pm SEM, * $p < 0.05$ (n=3)

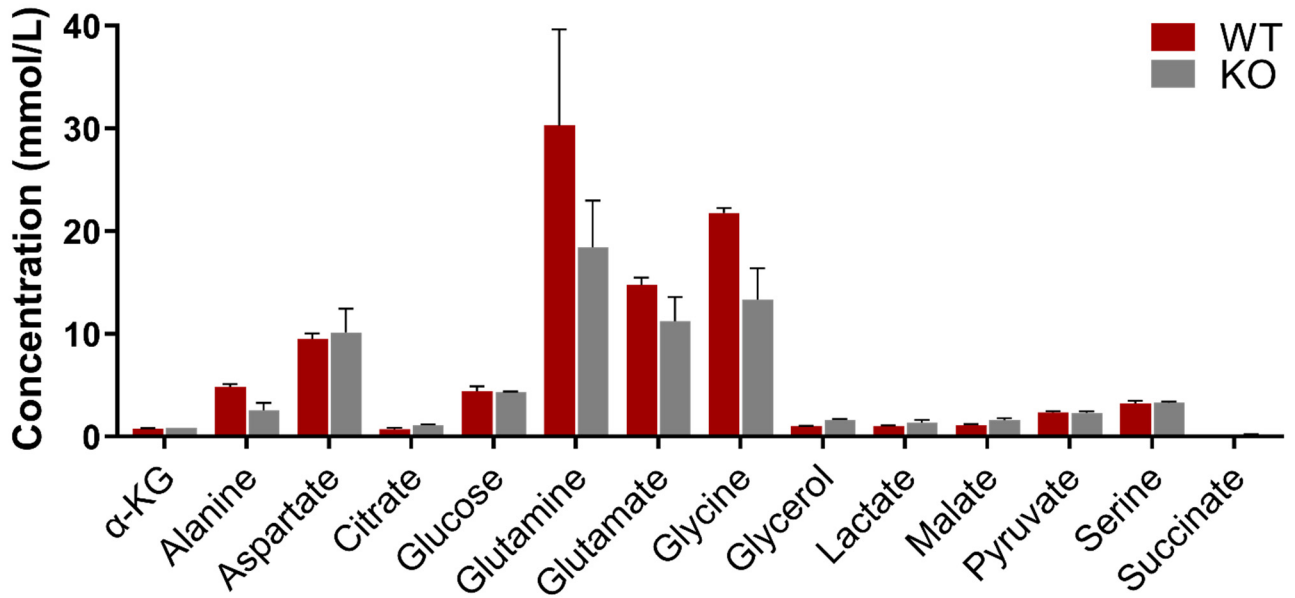
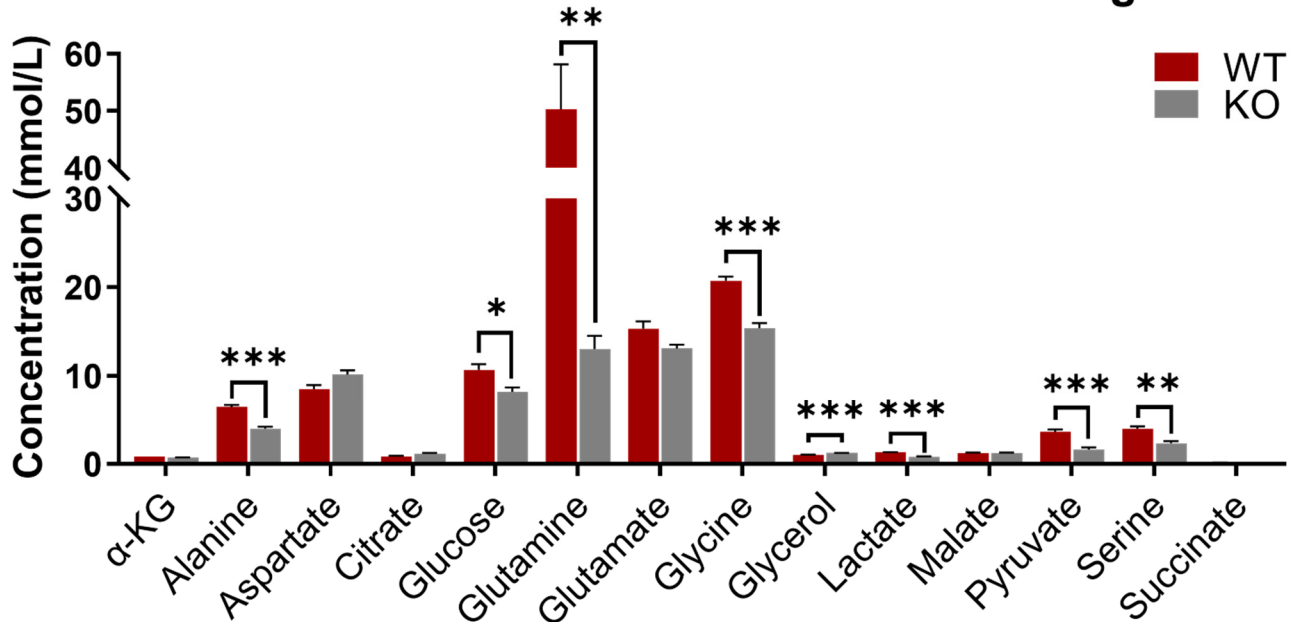
A**Intracellular metabolite concentration at 5 mM glucose****B****Intracellular metabolite concentration at 11 mM glucose**

Figure S8: Intracellular metabolite abundance in *G6pc2* WT and KO β TC3 cells labeled with 2 mM [U- $^{13}\text{C}_5$]glutamine

Intracellular metabolite abundance measured in *G6pc2* WT and KO β TC3 cells incubated with (A) 5 and (B) 11 mM glucose. Data represent means \pm SEM, *** p <0.01, ** p <0.05, * p <0.10 (n =3).

mRNA expression of genes associated with redox balance

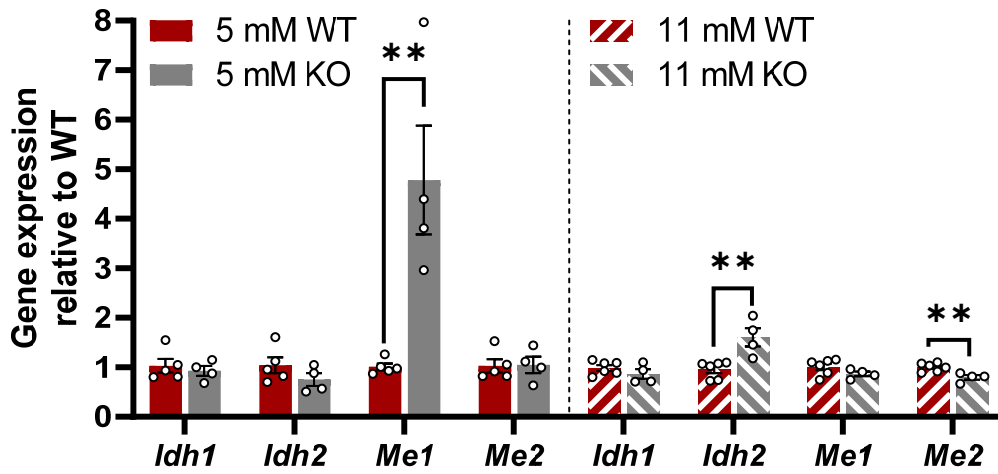


Figure S9: Effects of the loss of *G6pc2* on expression of genes regulating redox control. mRNA expression of genes regulating redox metabolism. Data represent means \pm SEM (n=3) relative to expression of the WT, **p<0.05 (n=3).

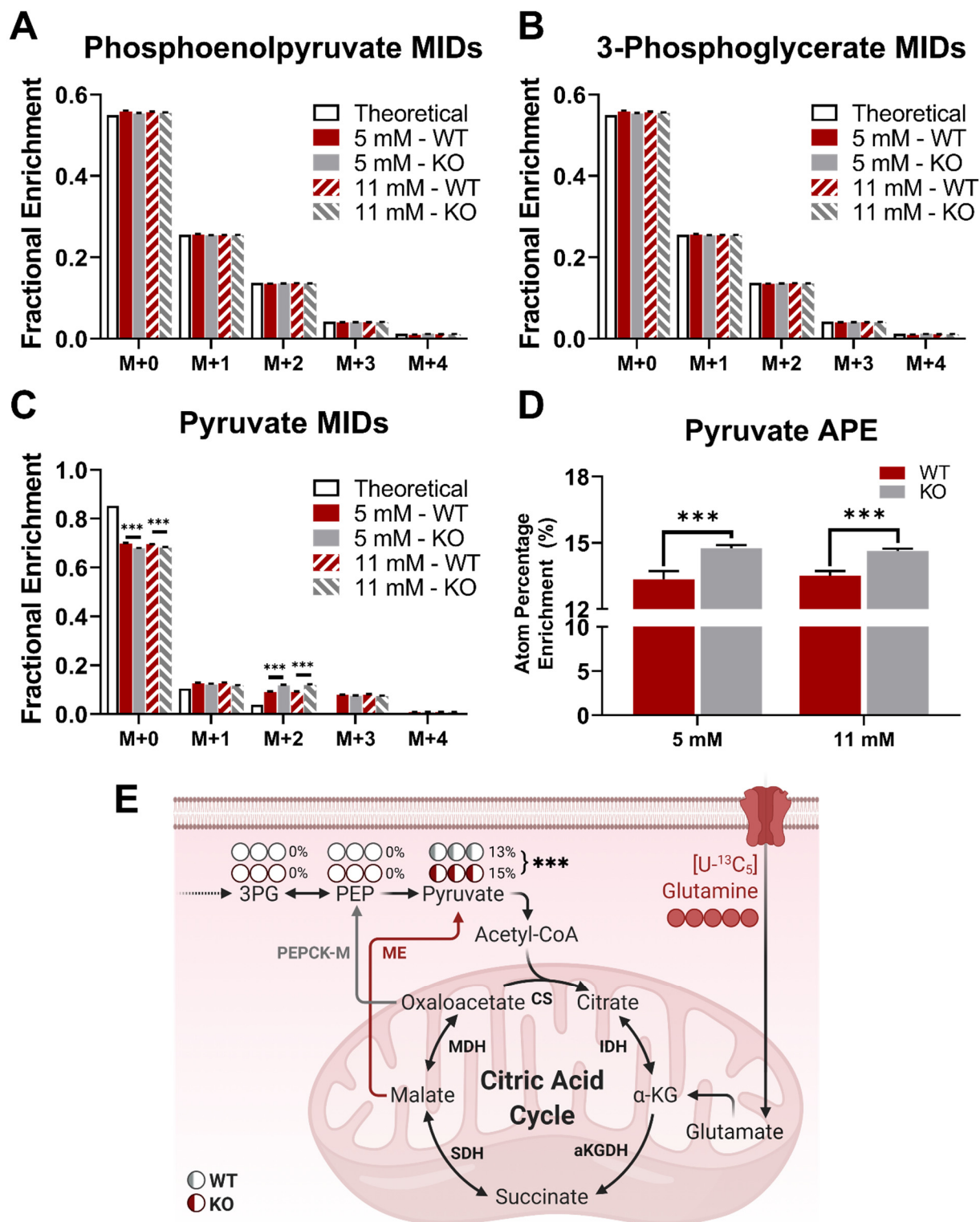


Figure S10: Enrichment patterns in glycolytic metabolites after incubation with 2 mM [U-¹³C₅]glutamine Intracellular enrichment in (A) phosphoenolpyruvate (PEP) (B) 3-phosphoglycerate (3PG), and (C) pyruvate, uncorrected for background abundance in *G6pc2* WT and KO βTC3 cells at 5 and 11 mM glucose concentrations incubated with 2 mM [U-¹³C₅]glutamine for 24h. The theoretical values represent the unlabeled MIDs for each metabolite. (D) APE of pyruvate at 5 and 11 mM glucose concentrations (E) Schematic showing pathways that can enrich 3PG, PEP and pyruvate when [U-¹³C₅]glutamine is used as tracer. Only pyruvate showed isotopic enrichment while PEP and 3PG had negligible ¹³C incorporation, suggesting that PEPCK-M is inactive at steady state in βTC3 cells. Data represent means±SEM, ****p*<0.01 (n=3).

Table S1. Pancreatic β -cell metabolic reaction network for ^{13}C MFA. (Related to Fig. 4-6 and Table S2)
 Network maps of β -cell metabolism track carbon atoms through model reactions. Metabolites used to regress fluxes in both compartments are shown in Table S2. Unenriched sources and sinks and “ CO_2 ” are annotated as “.source” and “.sink”, respectively. ^{13}C isotopes are introduced into model reactions as “.tracer”. Extracellular metabolites are designated as “.ext”.

Glycolysis	
$V_{\text{Gluc.source}}$	Gluc.source (ABCDEF) \rightarrow Gluc (ABCDEF)
V_{GK}	Gluc (ABCDEF) \rightarrow G6P (ABCDEF)
V_{PGI}	G6P (ABCDEF) \rightarrow F6P (ABCDEF)
V_{ALDO}	F6P (ABCDEF) \rightarrow DHAP (CBA) + GAP (DEF)
V_{TPI}	DHAP (ABC) \leftrightarrow GAP (ABC)
V_{G3PDH}	DHAP (ABC) \rightarrow G3P (ABC)
V_{G3PP}	G3P (ABC) \rightarrow Glycerol.ext (ABC)
V_{GAPDH}	GAP (ABC) \rightarrow 3PG (ABC)
V_{PSP}	Ser (ABC) \rightarrow 3PG (ABC)
V_{PK}	3PG (ABC) \rightarrow Pyr (ABC)
$V_{\text{Pyr.exch}}$	Pyr (ABC) \leftrightarrow Pyr.ext (ABC)
$V_{\text{Pyr.release}}$	Pyr.ext (ABC) \rightarrow Pyr.sink (ABC)
V_{LDH}	Pyr (ABC) \leftrightarrow Lac (ABC)
$V_{\text{Lac.exch}}$	Lac (ABC) \leftrightarrow Lac.ext (ABC)
$V_{\text{Lac.release}}$	Lac.ext (ABC) \rightarrow Lac.sink (ABC)
V_{ALT}	Pyr (ABC) \leftrightarrow Ala (ABC)
$V_{\text{Ala.exch}}$	Ala (ABC) \leftrightarrow Ala.ext (ABC)
$V_{\text{Ala.release}}$	Ala.ext (ABC) \rightarrow Ala.sink (ABC)
V_{PC}	Pyr (ABC) + CO_2 (D) \rightarrow Oac (ABCD)
V_{PDH}	Pyr (ABC) \rightarrow AcCoA (BC) + CO_2 (A)
Pentose Phosphate Pathway	
V_{G6PDH}	G6P (ABCDEF) \rightarrow P5P (BCDEF) + CO_2 (A)
V_{TK1}	S7P (ABCDEFGF) + GAP (HIJ) \leftrightarrow F6P (ABCHIJ) + E4P (DEFG)
V_{PPP}	P5P (ABCDE) + P5P (FGHIJ) \leftrightarrow S7P (ABFGHIJ) + GAP (CDE)
V_{TK2}	P5P (ABCDE) + E4P (FGHI) \leftrightarrow F6P (ABFGHI) + GAP (CDE)
Citric Acid Cycle	
V_{CS}	Oac (ABCD) + AcCoA (EF) \rightarrow Cit (DCBFEA)
$V_{\text{Fat.entry}}$	FA (AB) \rightarrow AcCoA (AB)
V_{IDH}	Cit (ABCDEF) \leftrightarrow α -kg (ABCDE) + CO_2 (F)
V_{GDH}	Glu (ABCDE) \leftrightarrow α -kg (ABCDE)

V_{GS}	Gln (ABCDE) \leftrightarrow Glu (ABCDE)
$V_{Glu.entry}$	Glu.ext (ABCDE) \leftrightarrow Glu (ABCDE)
$V_{Glu.source}$	Glu.source (ABCDE) \rightarrow Glu.ext (ABCDE)
$V_{Gln.source}$	Gln.source (ABCDE) \rightarrow Gln.ext (ABCDE)
$V_{Gln.entry}$	Gln.ext (ABCDE) \rightarrow Gln (ABCDE)
$V_{\alpha KGDH}$	α -kg (ABCDE) \rightarrow Suc (BCDE) + CO ₂ (A)
$V_{Suc.release}$	Suc (ABCD) \rightarrow Suc.ext (ABCD)
V_{SDH}	Suc (ABCD) \leftrightarrow Fum (ABCD)
V_{FM}	Fum (ABCD) \leftrightarrow Mal (ABCD)
V_{MDH}	Mal (ABCD) \leftrightarrow Oac (ABCD)
V_{ME}	Mal (ABCD) \leftrightarrow Pyr (ABC) + CO ₂ (D)
V_{AST}	Oac (ABCD) \leftrightarrow Asp (ABCD)
$V_{Asp.exch}$	Asp (ABCD) \leftrightarrow Asp.ext (ABCD)
$V_{Asp.source}$	Asp.source (ABCD) \rightarrow Asp.ext (ABCD)
$V_{Mal.release}$	Mal \rightarrow Mal.sink
$V_{Cit.release}$	Cit \rightarrow Cit.sink
Isotope uptake, CO₂ recycling and biomass equation	
$V_{Gluc.tracer}$	Gluc.tracer (ABCDEF) \rightarrow Gluc (ABCDEF)
$V_{Gln.tracer}$	Gln.tracer (ABCDE) \rightarrow Gln.ext (ABCDE)
$V_{CO2.source}$	CO ₂ .source (A) \rightarrow CO ₂ (A)
$V_{CO2.sink}$	CO ₂ (A) \rightarrow CO ₂ .sink (A)
$V_{Biomass}$	1389*G6P \rightarrow Biomass

Table S2. GC-MS fragment ions of measured metabolites regressed using the metabolic model for MFA. (Related to Fig. 4-6, S3, S5-7 and Table S1)

Metabolite	<i>m/z</i>	Derivative Formula	Carbons						
3-Phosphoglycerate	585	C ₂₃ H ₅₄ O ₇ Si ₄ P	C1	C2	C3				
Alanine	260	C ₁₁ H ₂₆ O ₂ NSi ₂	C1	C2	C3				
Alanine	232	C ₁₀ H ₂₆ ONSi ₂		C2	C3				
Aspartate	302	C ₁₄ H ₃₂ O ₂ NSi ₂	C1	C2					
Aspartate	390	C ₁₇ H ₄₀ O ₃ NSi ₃		C2	C3	C4			
Aspartate	418	C ₁₈ H ₄₀ O ₄ NSi ₃	C1	C2	C3	C4			
Citrate	459	C ₂₀ H ₃₉ O ₆ Si ₃	C1	C2	C3	C4	C5	C6	
Fumarate	287	C ₁₂ H ₂₃ O ₄ Si ₂	C1	C2	C3	C4			
Glucose	301	C ₁₄ H ₂₁ O ₇	C1	C2	C3	C4	C5	C6	
Glutamate	432	C ₁₉ H ₄₂ O ₄ NSi ₃		C2	C3	C4	C5		
Glutamate	330	C ₁₆ H ₃₆ O ₂ NSi ₂	C1	C2	C3	C4	C5		
Glutamine	431	C ₁₉ H ₄₃ O ₃ N ₂ Si ₃	C1	C2	C3	C4	C5		
Glycerol	377	C ₁₇ H ₄₁ O ₃ Si ₃	C1	C2	C3				
Glycerol-3-Phosphate	571	C ₂₀ H ₅₁ O ₆ Si ₄ P	C1	C2	C3				
Lactate	261	C ₁₁ H ₂₅ O ₃ Si ₂	C1	C2	C3				
Lactate	233	C ₁₀ H ₂₅ O ₂ Si ₂		C2	C3				
Malate	419	C ₁₈ H ₃₉ O ₅ Si ₃	C1	C2	C3	C4			
Phosphoenolpyruvate	453	C ₁₇ H ₃₈ O ₆ Si ₃ P	C1	C2	C3				
Pyruvate	174	C ₆ H ₁₂ O ₃ NSi	C1	C2	C3				
Serine	390	C ₁₇ H ₄₀ O ₃ NSi ₃	C1	C2	C3				
Succinate	289	C ₁₂ H ₂₅ O ₄ Si ₂	C1	C2	C3	C4			