Supplementary Material

Supplementary Table 1. A detailed summary of study characteristics included in the systematic review

Study	Study Design	Cohort(s)	Sample size	Tissue	Proposed blood biomarker(s)	Analytical technique	Statistics	Type of biomarker(s)
McGuinness et al., 2015 (1)	Longitudi nal study (2-year follow-up)		N = 182 MCI (97), CN (85)	Platelet	BACE1	BMG Labtech FLUOstar OPTIMA	AUC=0.64 (p=0.04)	Prognostic
			N = 817					
Burnham et r al., 2016 (2) r f	Longitudi nal study		CN (585), MCI (74), AD (158)	Plasma	Aβ42 + CXCL13 +	Luminex human	Sn=80%; Sp=82%	Diagnostic
	(34- month follow-up)	AIDE	Validation group: CN (227), MCI (11), AD (5)		+ VCAM-1	pane	Validation cohort: Sn=79%; Sp=76%	Diagnostic
lanalidza at		BioFIND	N = 719				Aβ42: AUC (CSF) = 0.655, AUC (PET) = 0.604	
Janelidze et al., 2016 (3)	CC	ER	CN (274), SCD (174), MCI (214), AD (57)	Plasma	Αβ40, Αβ42	Simoa platform	Aβ42/Aβ40 ratio: AUC = 0.683, AUC (PET) = 0.621	Diagnostic
Jammeh et al., 2016 (4)	СС	ADNI	N = 157 AD (106), CN (51)	Plasma	A1M + A2M + C3 + AAT + APOE + PPP	-	AUC=0.85, Sn=85.4%, Sp=78.6%, accuracy = 83.6%	Diagnostic
Yu et al., 2016 (5)	СС		N = 90 AD (50), CN (40)	Serum	VEGF + sCD40L	Immunomagnetic beads assay	AUC=0.58 (95% CI: 0.775-0.941)	Diagnostic
Zheng et al., 2016 (6)	Longitudi nal study (2-years follow-up)		N = 254 CN (90), MCI-AD (76), MCI-MCI (88)	Serum	HCY + BDNF + APOEε4	ELISA and microparticle- enzyme- immunoassay	Sn=85.0%, Sp=86.0%, PPV=0.93, NPV=0.73%	Predictive for early diagnosis
An et al., 2017 (7)	CC		N = 171 AD (24), CN (29)	Plasma	Aβ oligomers	Multimer detection system	Sn=83.3%, Sp=90.0%	Diagnostic

Goudey et al., 2017 (8)		ADNI	N = 566 Training: MCI (198), AD (102), CN (58) Validation: MCI (198), AD (10)	Plasma	Aβ42 + APOE + CgA + EOT3 + APOEε4 status	-	AUC=0.84, Sn=78%, Sp=0.73%	Predictive of CSF Aβ42 status
Janel et al., 2017 (9)	сс		N = 140 Cohort (M): CN (20), AD (69) Cohort (P): CN (25), AD (26)	Plasma	DYRK1A + BDNF + HCY	Solid-phase immobilized epitope- immunoassay, ELISA, and fluorimetric high- performance liquid chromatography	AUC=0.933; Sn=95.2%; Sp=88.9%	Diagnostic
Mattsson et al., 2017 (10)	Longitudi nal study (6.5- years follow up)	ADNI	N = 570 CN (193), MCI (197), AD (180)	Plasma	NFL	Ultrasensitive single- molecule array (Simoa) platform	AUC=0.87	Diagnostic and prognostic
Mohd Hasni et al., 2017 (11)	СС		N = 78 AD (39), CN (39)	Serum	IL-13 + CXCL10	Procarta Multiplex Cytokine and ELISA assay	AUC=1 (95% CI); Sp=100%; Sn=100%	Diagnostic
Pedrini et al., 2017 (12)	Longitudi nal study (18- and 54- months follow-up)	AIBL	N = 665 18 months: CN (559), MCI (39), AD (67) 54 months: CN (528), MCI (51), AD (86)	Plasma	IL-10 + IL-12/23p40	Multiplex assay panel	18 months: AUC=0.802 54 months: AUC=0.805	Early diagnosis
Popp et al., 2017 (13)	СС		N = 120 CN (48), MCI (72)	Serum	bFGF + CRP + IL-16 + sFLT-1 + sICAM-1 + Tie-2 + VEGF-C + VEGF-D	V-Plex Neuroinflammation Panel 1	AUC=0.89 (0.81 - 0.95)	Classification for AD pathology
Tateno et al., 2017 (14)	СС		N = 117 ApoE4 (28), non- ApoE4 (89)	Plasma	Αβ42/Αβ40 + ΑΡΟΕε4	Sandwich ELISA assay	APOEε4 (+): AUC=0.519, Sn=63.6%, Sp=52.9% APOEε4 (-): AUC=0.648,	Diagnostic

							Sn=92.9%, Sp=45.9%, PPV=44.1%, NPV=93.3%	
Wang et al., 2017 (15)	СС		N = 61 AD (24), CN (37)	Plasma	Aβ oligomers	Multimer detection system	AUC=0.844, Sn=78.3%, Sp=86.5%	Diagnostic
Yu et al., 2017 (16)	СС		N = 312 AD (156), CN (156)	Serum	IL-13 + IL-1α + CXCL10 + IL-3 + TNFα	Immunomagnetic beads assay	AUC=0.58 (95% CI: 0.775-0.941)	Diagnostic
Chen et al., 2018 (17)	CC		N = 126 AD (96), CN (30)	Serum	ΑΡΡ + NCAM + Αβ40 + Αβ42	ELISA assay	AUC=0.997, Sn=98.5	Diagnostic and prognostic
Cheng et al., 2018 (18)	СС		N = 199 AD (98), CN (101)	Plasma	BDNF + AGT + IGFBP-2 + OPN + cathepsin D + SAP + C4 + TTR	Luminex xMAP	AUC= 0.958, Sn=86.7%, Sp=88.1%, Accuracy=87.4%	Diagnostic
de Rojas et al., 2018 (19)	СС	FACEHB I	N = 200	Plasma	Αβ42/Αβ40	Sandwich ELISA assay	AUC=0.681, Sn=83%, Sp=59.1%	Predictive of amyloid PET
Eke et al., 2018 (20)	CC	ADNI	N = 166 AD (108), CN (58)	Plasma	A1M + A2M + C3 + IgM + TNC	-	AUC=0.89, Sn=86.5%, Sp=82.1%, Accuracy=85%	Diagnostic
Lewczuk et			N = 140				Age (-): AUC=0.853, Sn=84%, Sp=0.78%, accuracy=82%	Diagnostic
al., 2018 (21)	CC		MCI (25), AD (33), CN (41)	Plasma	NFL	Simoa assay	Age (50-80): AUC=0.920, Sn=61%- 91%, Sp=89%- 20%, accuracy=84%-80%	(nonspecific)
Mielke et al.,	<u> </u>	MCSA,	N = 269	Plaama	p-tau181, p-tau181 +	MSD	AUC (p-tau181) = 0.803	Predictor of
2018 (22)		ADRC	CN (172), MCI (57), AD (40)	riasilia	ΑΡΟΕε4+	IVIOU	AUC (p-tau181 + ΑΡΟΕε4+) = 0.860	brain Aβ
Nabers et al., 2018 (23)			N = 385					
	Longitudi	BioFIND FR	BioFINDER:	Plasma	Aβ secondary	Vertex 70V FTIR- spectrometer	AUC (BioFINDER) = 0.78 , Sn = 69% , Sn =	Diagnostic
	nal study	ESTHER	PET+ (36), PET- (37)		structure		86%	Diagnostic

			ESTHER: AD (65), CN (247)				AUC (ESTHER) = 0.80, Sn = 71%, Sp =			
			,				JI/0			
Nakamura		NCCC				Immunoprecipitation	Discovery: AUC=0.967	Dradiative of		
et al., 2018	CC	AIBL	RCGG = 121, ABL = 252	Plasma	AB42 + APP/AB42 + AB42/AB40	coupled with mass	Validation: AUC=0.941	amvloid PET		
(24)					· + ·· + · •	spectrometry	Accuracy=90%			
Chan at al	Longitudi		N = 224			Synthetic				
Sheh et al., 2018 (25)	(3-years		Probable AD (75),	Plasma	BACE1	substrate ELISA and	Sn = 84%, Sp = 88%	Prognostic		
	follow-up)		MCI (96), ND (53)			Western blot				
	المحمنانيطن	SCIENC								
Verberk et	nal study	and		D.	A O A O / A O A O	Simoa Human	AUC=0.83 (95% CI	5		
al., 2018	(3 ± 2	Amsterda	N = 248	Plasma	APOFε4 status	Neurology 3-Plex A	0.77-0.89), Sn=76%,	Prescreener, diagnostic		
(26)	years follow-up)	M Dementia				assay kit	Sp=75%	alagileene		
	ronow up)	Cohort								
Winston et			N = 137				Aβ42: Sn=80.6%			
Winston et al., 2018 ((27)	СС		CN (76), MCI (61)	Exosomes	AB42, NRGN, SYP, SYT, and SYNPO	ELISA assay	NRGN, SYP, SYT, and SYNPO: Sn=100%	Diagnostic		
Adliardi et			N - 42							
al., 2019	CC		N = 42	Exosomes	SNAP-25	Western Blot and	AUC=0.82, Sn=87.5%, Sn=70.6%	Diagnostic		
(28)			AD (24), CN (17)				0p=10.070			
Baldacci et	00		N = 78	550			AUC (α-syn/Aβ) = 0.76	Diamantia		
(29)	CC		Early AD (39), CN (39)	RBC	α-syn/Ap, α-syn/tau	Immunoassays	AUC (α -syn/tau) = 0.72	Diagnostic		
Boccardi et			N = 289							
al., 2019 (30)	CC		HC (87), MCI (73), AD (129)	Plasma	IFNa-2, IL-1, TNFa		AUC = 0.6524	Prognosis		
							AUC (ADAM10) = 0.9			
Bram et al., 2019 (31)			N = 40		ADAM10 + BACE1 +		Sn = 88.90%, Sp = 78.90%			
	CC		AD (20), CN (20)	Platelet	ADAM10 + BACE1 + , PSEN1	Western blot	AUC (PSEN1) = 0.80.	Diagnostic		
								FJENT		Sn = 77.80%, Sp = 83.30%

							AUC (ADAM10 + BACE1 + PSEN1) = 0.9	
Chatterjee et al., 2019 (32)	CC	KARVIA H	N = 95 SMC (72), CN (23)	Plasma	Aβ42/Aβ40 + APOEε4 status	Ultrasensitive Simoa assay	AUC=0.78	Predictive of amyloid PET
			N = 151				Discovery cohort:	
			Discovery cohort:				MCI vs. Con: AUC=0.79	
Chen et al		HABS,	CN (19), MCI (21), AD (25)		_	ELISA and Simoa	AD vs. Con: AUC=0.75	
2019 (33)	CC	UCL, UCSD	Validation cohort:	Plasma	Tau	assay	Validation cohort:	Diagnostic
			CN (41), MCI (22), AD (23)				MCI vs. Con: AUC=0.88	
							AD vs. Con: AUC=0.96	
Guzmán- Martínez et	СС	FACEHB	N = 51	Platelet	Alz-tau® (HMW/LMW	SDS-PAGE and	AUC=0.756, Sn=71.43%,	Diagnostic
al., 2019 (34)		-	AD (36), CN (15)		tau ratio)	vvestern Blot	Sp=69.23%	Ũ
Han et al., 2019 (35)	сс	KBASE	N = 407 CN (241), MCI	Plasma	AChE	ELISA and colorimetric assay	AChE + ApoE ε4: AUC=0.728-0.798 AChE activity + ApoE	Predictor of cerebral Aβ deposition in cognitively
			(103), AD (63)				ε4: AUC=0.727-0.768	normal
	L on situdi						pAD: AUC=0.732	
lulita et al.,	nal study	LEAD	N = 107 pAD (28) MCI (30)	Plasma	Αβ40 + Αβ42 + MMP- 1 + MMP-3 + IL-8 +	Multi-Spot V-PLEX	MoCA decline: AUC= 0.751	Diagnosis,
2019 (36)	(3 years follow-up)		SMI (30), CN (19)		IL-10 + TNF- α		CAMCOG decline: AUC= 0.844	prognosis
Jia et al., 2019 (37)			N = 298			Neuronal-derived	Discovery cohort:	Predictive of
	CC		Discovery stage: AD (28), aMCI (25), CN (29)	Exosomes	Aβ42 + T-tau + P- T181-tau	exosomes immunoprecipitation and ELISA	Validation cohort: AUC=0.85-0.98	T-tau, and P- T181-tau status

			Validation stage: AD (73), aMCI (71), CN (72)					
							BLSA cohort:	
							Training set (n=161): AUC=0.896, Sn=81.8%, Sp=85.8%	
Kapogiannis	Longitudi	51.04	N = 414 BLSA cohort: MCI		nEV concentration + nEV mean diameter +	Neuronal-enriched	Validation set (n=80): AUC=0.80, Sn=55.6%, Sp=88.7%	
et al., 2019	nal study (2 years	BLSA, JHADRC	to AD (128), CN (222)	nEV	nEV proteins (Itau, p- tau181, p-tau231, pY-	extracellular vesicles immunoprecipitacion	JHADRC cohort:	Diagnostic
(38)	follow-up)		JHADRC cohort: AD (35), CN (29)		IRS-1, and pSer312- IRS-1)	and ELISA	Training set: AUC=0.989, Sn=100%, Sp=94.7%	
							Validation set: AUC=0.767, Sn=91.7%, Sp=60%	
			N = 84				Αβ42/Αβ40:	
Li et al., 2019 (39)	CC		AD (53), MCI (22), CN (9)	Plasma	Αβ42, Αβ40, T-tau	Simoa assay	AUC=0.77, Sn=82%, Sp=64%	Diagnostic
			N = 63				AUC (apoA-I) = 0.77	
Liu et al.,	CC	l iu et al	CN (10), EMCI	Serum	apoA-I + aC3/nC3 +	Turbidimetric	AUC (aC3/nC3) = 0.79	MCI
2019 (40)			(26), LMCI (23), AD (4)	Corum	TTR	ELISA assay	AUC = 0.89, Sn=83%, Sp= 90%	prognosis
	Longitudi						AUC (6 months) = 0.688	
Liu et al., 2019 (41)	nal study (2-year		N = 92	Serum	ACT	ELISA and Western blot	AUC (12 months) = 0.839	aMCI diagnosis
	follow-up)						AUC (24 months) = 0.887	
Meng et al., 2019 (42)	CC		N = 58 AD (30), CN (28)	Plasma	Aβ oligomers	Multimer detection system	AUC=0.89, Sn=82.1%, Sp=90.0%	Diagnostic

Morgan et al., 2019 (43)	сс	AddNeur oMed, EMIF-AD MBD, DCR	N = 1580 Discovery cohort: CN (259), MCI (199), AD (262) Replication cohorts: AD (193), MCI (494), CN (600)	Plasma	FB + FH + sCR1, MCP1 + EOT1 sCR1 + MCP1 + EOT1 FB + FH + age	Singleplex and multiplex assay, ELISA and V-plex electrochemiluminesc ence immunoassay	AD vs. Con: AUC (discovery/replication cohort) =0.74/0.81 AD vs. MCI: AUC (discovery/replication cohort) =0.74/0.67 MCI to AD: AUC=0.71	Diagnostic and prognostic
Nabers et al., 2019 (44)	СС	BioFIND ER, ESTHER	N = 100 CN (61), AD (39)	Plasma	Aβ secondary structure	Spectrometry	BioFINDER: AUC=0.78, Sn=69%, Sp=86% ESTHER: AUC=0.80, Sn=71%, Sp=91%	Diagnostic
Palmqvist et al., 2019 (45)	сс	BioFIND ER and a German biomarke r study for validation	N = 1079 CU, MCI, AD	Plasma	Ab42/Ab40, APOE+4 status, tau, NF-L	Elecsys immunoassays (Roche Diagnostics)	Cohort 1: 0.80–0.87; Cohort 2: 0.86	Predictive of CSF Ab status
Park et al., 2019 (46)	СС		N = 76 CN (52), MCI (9), AD (15)	Plasma	T-tau/Aβ42	Simoa assay and xMAP technology	AUC=0.89, Sn=80%, Sp=91.43%	Predictive of tau pathology
Park et al., 2019 (47)	сс	KBASE	N = 254 CN (107), MCI (107), AD (40)	Plasma	Set I: LGALS3BP + ACE + gal-3 Set II: POSTN + ACE + VE-cadherin Proposed biomarkers: LGALS3BP, CDH5, ACE, POSTN	Mass spectrometry and ELISA	Set I: AUC (PiB- vs. PiB+) =0.871, Sn=79%, Sp=84% Set II: AUC (for MCI) = 0.836, Sn=68%, Sp=90%	Predictive of Ab deposition
Pérez- Grijalba et al., 2019 (48)	СС	AB255 study	N = 59 CN (39), MCI (20)	Plasma	Αβ42/Αβ40	ABtest kit	AUC=0.881, Sn=77.8%, Sp=87.5%, PPV=0.732, NPV=0.900	Predictive of Ab PET, diagnostic
Perrotte et al., 2019 (49)	CC		N = 95	Plasma	ApoJ, Klotho, protein carbonyls, circulating- proteosome	Immunoblotting, ELISA	<i>MCI vs. Con:</i> AUC (ApoJ) = 0.8804	Diagnostic and prognostic

			CN (24), MCI (24),				AUC (Klotho) = 0.8036	
			AD (47)				AD vs. Con:	
							AUC (ApoJ) = 0.8168	
							AUC (Klotho) = 0.7976	
							AUC (protein carbonyls) = 0.7528	
							AUC (circulating- proteosome activity) = 0.8440	
							MCI vs. AD:	
							AUC (protein carbonyls) = 0.7522	
							AUC (circulating- proteosome activity) = 0.8379	
Schindler et al., 2019 (50)	СС		N = 158	Plasma	Aβ42/Aβ40 + APOEε4 status	Immunoprecipitation, Mass spectrometry	AUC=0.94 (95% CI 0.90–0.97)	Diagnostic
Schipke et al., 2019 (51)	СС		N = 160 CN (79), AD (81)	Serum	BDNF + IGF-1 + VEGF + TGF-β 1 + MCP-1 + IL-18	ELISA assays	AUC=0.94, Sn=76%, Sp=95%, Accuracy=85%	Diagnostic
			N = 843				AD vs. Con:	
Whelan et		BioFIND	Control Aβ- (415),	Plasma	OSM. MMP9. HAGH.	Olink [™] ProSeek	AUC=0.94	D : (1)
al., 2019 (52)	CC	ER	Control A β + (142),		CD200, AXIN1, uPA	immunoassay	Prodromal AD vs. Con:	Diagnostic
、 ,			Aβ+ (75), AD (161)				AUC=0.78	
Yao et al			N = 123		GSN. BDNF. TIMP1.	ELISA and Western	AUC (VLDLR) = 0.932	
2019 (53)	CC		AD (54), CN (69)	Serum	VLDLR, APLP2	blot for validation	AUC (TIMP1) = 0.903	Diagnostic
Abate et al., 2020 (54)	СС	InveCe.A b, PharmaC og/E- ADNI	InveCe.Ab: N = 264 PharmaCog/E- ADNI: N = 111	Plasma	Unfolded p53	-	AUC = 0.92	Predictive

Abe et al., 2020 (55)	Multicent er, CC	Abe et al.	N = 259 CN (100), MCI (60), and AD (99)	Serum	FBC + AHSG + FAC + PPC11	BLOTCHIP [®] - MS analysis	MCI vs. Con: AUC=0.662, Sn=72%, Sp=59% MCI vs. AD: AUC=0.672, Sn=77%, Sp=62% AD vs. Con: AUC=0.804, Sn=87%, Sp=65%	Diagnostic
Barthélemy et al., 2020 (56)	СС	SILK	Discovery cohort = 58 Validation cohort = 92	Plasma	p-tau-217, p-tau-181	Immunopurification (IP)-LC-MS	p-tau-217: AUC (discovery) = 0.99 AUC (validation) = 0.92 p-tau-181: AUC (discovery) = 0.98 AUC (validation) = 0.75	
Doecke et al., 2020 (57)	Longitudi nal study (54 months follow-up)	AIBL	Month 18: n = 176 Month 36: n = 169 Month 54: n = 135	Plasma	Αβ42/Αβ40 + ΑΡΟΕε4	ELISA assays	AUC=0.88-0.913 AUC(HC)=0.808-0.898	Diagnostic
Eke et al., 2020 (58)	СС	ADNI	N = 358 CN (58), MCI (198), AD (102)	Plasma	Aβ42 + CgA + EOT3 + APOEε4	-	AUC=0.84, Sn=0.82, Sp=0.62, PPV=0.81, NPV=0.64	Predictive of CSF Aβ42
Ellegaard et al., 2020 (59)	сс		N = 30 AD (10), MCI (10), CN (10)	Plasma	CLEC1B/CCL11 ratio in EVs TGFa/CCL20 ratio in plasma	Proximity extension assay	AUC (CLEC1B/CCL11) = 0.95, 95% CI = 0.86–1, p = 0.001 AUC (TGFa/CCL20) = 0.96, 95% CI = 0.88–1, p = 0.001	Diagnostic

Feinkohl et al., 2020 (60)	СС		N = 93 AD (44), CN (49)	Plasma	Αβ42/Αβ40	ELISA assays	AUC=0.59, Sn=61.2%, Sp=63.6%	Diagnostic
Gao et al., 2020 (61)	СС		N = 30 CN (10), MCI (10), AD (10)	Plasma, Serum	Αβ42/Αβ40	Nanosheet-based sensor system	aMCI vs. Con: AUC=0.83, Sn=70%, Sp=70% AD vs. Con: AUC=0.91, Sn=90%, 90% aMCI vs. AD:	Diagnostic
							AUC=0.75, Sn=80%, Sp=60%	
Janelidze et al., 2020 (62)	сс	BioFIND ER, Arizona Study of Aging and Neurode generativ e Disorders /Brain and Body Donation Program	Cohort 1: 182 Cohort 2: 344 Cohort 3: 63	Plasma	Αβ42/Αβ40 + pTau181	MSD, Elecsys fully- automated immunoassays, Simoa	AUC=0.84 (95% CI = 0.79-0.89)	Diagnostic
Karikari et al., 2020 (63)	СС	TRIAD, BioFIND ER-2, primary care cohort referred from primary care physician s of the Canadian National Health Service	N=1131 TRIAD cohort: 226 BioFINDER-2 cohort: 763 Primary care cohort: 105	Plasma	p-tau181	Ultrasensitive blood immunoassay	AUC = 85.3%; 95% Cl 81.4–89.2%	Diagnostic, prognosis

		for specialist care.						
Palmqvist et al., 2020 (64)	СС	Arizona- based neuropat hology cohort, Swedish BioFIND ER-2 cohort, Colombia n autosom al- dominant AD kindred	N = 1402	Plasma	p-tau217	Immunoassay	AUC = 0.89 and 0.96	Diagnostic
Perrotte et al., 2020 (65)	сс	Recruited from the Memory Clinic of Sherbroo ke	N = 60	Plasma exosomes	p-tau181/t-tau ratio	Luminex assay	Control from AD [AUC of 0.823 (p < 0.05) MCI from AD: AUC of 0.87 (p < 0.05)	Diagnostic, prognosis
							Serum:	
							AUC (T-tau) = 0.675	
							AUC (p-tau202) = 0.5958	
Nam et al.,	Longitudi nal study		N = 76	Serum,	T-tau, p-Tau202, p-	ELISA assays and	AUC (p-tau202/T-tau) = 0.5253	
2020 (66)	(4-year		CN (26), MCI (30), AD (20)	Exosomes	tau202/T-tau	Western blot	Exosomes:	Prognostic
	ioliow up)						AUC (T-tau) = 0.6434	
							AUC (p-tau202) = 0.7333	Prognostic

Sheu et al., 2020 (67)	СС		N = 86 AD (32), CN (54)	Serum	AHI1	Western blot and ELISA assays	AUC=0.7-0.82	Diagnostic
	Longitudi	SCIENC e project					AUC (Αβ misfolding) =0.94 (95% Cl, 0.86- 1.00; 6-year follow-up)	
Stockmann et al., 2020 (68)	nal study (6 years follow-up)	and Amsterda m Dementia	N = 203	Plasma	Aβ misfolding + Aβ42/Aβ40ª	Immuno-infrared- sensor technology and SIMOA	AUC (Αβ42/Αβ40) = 0.92 (95% CI, 0.82- 1.00)	Prognostic
		Cohort					AUC (panel) = 0.99 (95% Cl, 0.99-1.00)	
Teuber- Hanselmann	er- Multi- Teuber- elmann centre litereret		N = 237	_			AUC(MCI)=0.94, Sn = 52.4%, Sp = 99.2%	
et al., 2020 (69)	trans- sectional	Hanselm ann	MCI (21), mild-AD (98), CN (118)	Serum	KLK8	ELISA assay	AUC(AD)=0.83, Sn = 56.1%, Sp = 69.8%	Diagnostic
Verberk et			N - 252				Total (AUC): 0.88 (Sn = 0.82, Sp = 0.86)	Prediction of amyloid PET
al., 2020 (70)	CC		SCD, MCI, AD	Plasma	Ab42/Ab40, GFAP, NF-L		Nondemented (AUC): 0.84 (Sn = 0.70, Sp = 0.86)	status and disease monitoring
			N 00				AUC = 0.8 (MCI), 0.74 (AD), 0.77 (MCI + AD)	
Wang et al., 2020 (71)	Pilot study, CC	Wang et al.	N = 30 MCI (10), AD (10), CN (10)	Plasma	ΑΜΡΚα1	ELISA assay	Sn = 88.89% (MCI), 77.78% (AD), 83.33% (MCI + AD)	Diagnostic
							Sp = 70% (MCI, AD)	
Westwood et al., 2020 (72)	СС	EMIF-AD catalogu e	N = 1866 HC, MCI, AD	Plasma	FCN2, CFI, C4, B2M, Cathepsin D, APOE+4, A1AT	Luminex xMAP, ELISA, and Meso Scale Discovery assavs	AUC = 0.742, Sn = 0.682, Sp = 0.704	Diagnostic
Youn et al., 2020 (73)	СС		N = 104 AD (52), CN (52)	Plasma	Aβ oligomers	Multimer detection system	Sn=100%, Sp=92.31%	Diagnostic
Jia et al., 2020 (74)	Two- stage- sectional study,		N = 739 Discovery stage: AD (28), aMCI (25),	Exosomes	GAP43 + Ng + SNAP-25	ELISA assays	AUC=0.87-0.89	Predictive

longitudin al study	CN (29); Validation stage: AD (73), aMCI (71), CN (72)
	preclinical AD (160), CN (160)
	mutation carriers (59), non-mutation carriers (62)
^a Age, gender, and the pres	ence of APOEε4 allele were included as covariates

References:

- McGuinness B, Fuchs M, Barrett SL, Passmore AP, Johnston JA. Platelet Membrane β-Secretase Activity in Mild Cognitive Impairment and Conversion to Dementia: a Longitudinal Study. *J Alzheimers Dis* (2015) **49**:1095–1103. doi:10.3233/JAD-150795
- Burnham SC, Rowe CC, Baker D, Bush AI, Doecke JD, Faux NG, Laws SM, Martins RN, Maruff P, Macaulay SL, et al. Predicting Alzheimer disease from a blood-based biomarker profile. *Neurology* (2016) 87:1093–1101. doi:10.1212/WNL.00000000003094
- 3. Janelidze S, Stomrud E, Palmqvist S, Zetterberg H, Van Westen D, Jeromin A, Song L, Hanlon D, Tan Hehir CA, Baker D, et al. Plasma β-amyloid in Alzheimer's disease and vascular disease. *Sci Rep* (2016) **6**: doi:10.1038/srep26801
- 4. Jammeh E, Zhao P, Carroll C, Pearson S, Ifeachor E. Identification of blood biomarkers for use in point of care diagnosis tool for Alzheimer's disease IEEE Conference Publication. *2016 38th Annu Int Conf IEEE Eng Med Biol Soc* (2016) **2016**:2415–2418. doi:10.1109/EMBC.2016.7591217
- 5. Yu S, Liu YPY-HY-PH, Liu YPY-HY-PH, Jiao S-SS, Liu L, Wang Y-JJ, Fu W-LL. Diagnostic utility of VEGF and soluble CD40L levels in serum of Alzheimer's patients. *Clin Chim Acta* (2016) **453**:154–159. doi:10.1016/j.cca.2015.12.018
- 6. Zheng L, Kong X, Cui Y, Wei Y, Zhang J, Wei W. Conversion from MCI to AD in patients with the APOE ε4 genotype: Prediction by plasma HCY and serum BDNF. *Neurosci Lett* (2016) **626**:19–24. doi:10.1016/j.neulet.2016.05.018
- 7. An SSA, Lee BS, Yu JS, Lim K, Kim GJ, Lee R, Kim S, Kang S, Park YH, Wang MJ, et al. Dynamic changes of oligomeric amyloid β levels in plasma induced by spiked synthetic Aβ42. *Alzheimer's Res Ther* (2017) **9**:1–10. doi:10.1186/S13195-017-0310-6/FIGURES/4
- 8. Goudey B, Fung BJ, Schieber C, Faux NG. A blood-based signature of cerebrospinal fluid Aβ1–42 status. *bioRxiv* (2017) doi:10.1101/190207
- 9. Janel N, Alexopoulos P, Badel A, Lamari F, Camproux AC, Lagarde J, Simon S, Feraudet-Tarisse C, Lamourette P, Arbones M, et al. Combined assessment of DYRK1A, BDNF and homocysteine levels as diagnostic marker for Alzheimer's disease. *Transl Psychiatry* (2017) **7**:e1154–e1154. doi:10.1038/tp.2017.123
- Mattsson N, Andreasson U, Zetterberg H, Blennow K, Weiner MW, Aisen P, Toga AW, Petersen R, Jack CR, Jagust W, et al. Association of plasma neurofilament light with neurodegeneration in patients with Alzheimer disease. *JAMA Neurol* (2017) 74:557–566. doi:10.1001/jamaneurol.2016.6117
- 11. Mohd Hasni DS, Lim SM, Chin AV, Tan MP, Poi PJH, Kamaruzzaman SB, Majeed ABA, Ramasamy K. Peripheral cytokines, C-X-C motif ligand10 and interleukin-13, are associated with Malaysian Alzheimer's disease. *Geriatr Gerontol Int* (2017)

17:839–846. doi:10.1111/ggi.12783

- Pedrini S, Gupta VB, Hone E, Doecke J, O'Bryant S, James I, Bush AI, Rowe CC, Villemagne VL, Ames D, et al. A bloodbased biomarker panel indicates IL-10 and IL-12/23p40 are jointly associated as predictors of β-amyloid load in an AD cohort. *Sci Rep* (2017) **7**:14057. doi:10.1038/s41598-017-14020-9
- Popp J, Oikonomidi A, Tautvydaitė D, Dayon L, Bacher M, Migliavacca E, Henry H, Kirkland R, Severin I, Wojcik J, et al. Markers of neuroinflammation associated with Alzheimer's disease pathology in older adults. *Brain Behav Immun* (2017) 62:203–211. doi:10.1016/j.bbi.2017.01.020
- Tateno A, Sakayori T, Kim WC, Koeda M, Kumita S, Suzuki H, Okubo Y. Effect of apolipoprotein E phenotype on the association of plasma amyloid β and amyloid positron emission tomography imaging in Japan. *Alzheimer's Dement Diagnosis, Assess Dis Monit* (2017) 9:51–56. doi:10.1016/j.dadm.2017.08.002
- 15. Wang MJ, Yi S, Han J-Y, Park SY, Jang J-W, Chun IK, Kim SE, Lee BS, Kim GJ, Yu JS, et al. Oligomeric forms of amyloid-β protein in plasma as a potential blood-based biomarker for Alzheimer's disease. *Alzheimers Res Ther* (2017) **9**:98. doi:10.1186/s13195-017-0324-0
- 16. Yu S, Liu Y-P, Liu H-L, Li J, Xiang Y, Liu Y-H, Jiao S-S, Liu L, Wang Y, Fu W. Serum Protein-Based Profiles as Novel Biomarkers for the Diagnosis of Alzheimer's Disease. *Mol Neurobiol* (2017) **55**:3999–4008. doi:10.1007/s12035-017-0609-0
- 17. Chen K, Gao T, Bai Z, Yuan Z. Circulating APP, NCAM and Aβ serve as biomarkers for Alzheimer's disease. *Brain Res* (2018) **1699**:117–120. doi:10.1016/j.brainres.2018.08.015
- 18. Cheng Z, Yin J, Yuan H, Jin C, Zhang F, Wang Z, Liu X, Wu Y, Wang T, Xiao S. Blood-Derived Plasma Protein Biomarkers for Alzheimer's Disease in Han Chinese. *Front Aging Neurosci* (2018) **10**:414. doi:10.3389/fnagi.2018.00414
- 19. de Rojas I, Romero J, Rodríguez-Gomez O, Pesini P, Sanabria A, Pérez-Cordon A, Abdelnour C, Hernández I, Rosende-Roca M, Mauleón A, et al. Correlations between plasma and PET beta-amyloid levels in individuals with subjective cognitive decline: the Fundació ACE Healthy Brain Initiative (FACEHBI). *Alzheimers Res Ther* (2018) **10**:119. doi:10.1186/s13195-018-0444-1
- 20. Eke CS, Jammeh E, Li X, Carroll C, Pearson S, Ifeachor E. Identification of Optimum Panel of Blood-based Biomarkers for Alzheimer's Disease Diagnosis Using Machine Learning. in *2018 40th Annual International Conference of the IEEE Engineering in Medicine and Biology Society (EMBC)* (United States: IEEE), 3991–3994. doi:10.1109/EMBC.2018.8513293
- Lewczuk P, Ermann N, Andreasson U, Schultheis C, Podhorna J, Spitzer P, Maler JM, Kornhuber J, Blennow K, Zetterberg H. Plasma neurofilament light as a potential biomarker of neurodegeneration in Alzheimer's disease. *Alzheimers Res Ther* (2018) 10:71. doi:10.1186/s13195-018-0404-9
- 22. Mielke MM, Hagen CE, Xu J, Chai X, Vemuri P, Lowe VJ, Airey DC, Knopman DS, Roberts RO, Machulda MM, et al. Plasma phospho-tau181 increases with Alzheimer's disease clinical severity and is associated with tau- and amyloid-positron emission

tomography. Alzheimer's Dement (2018) 14:989–997. doi:10.1016/j.jalz.2018.02.013

- 23. Nabers A, Perna L, Lange J, Mons U, Schartner J, Güldenhaupt J, Saum K-U, Janelidze S, Holleczek B, Rujescu D, et al. Amyloid blood biomarker detects Alzheimer's disease. *EMBO Mol Med* (2018) **10**: doi:10.15252/emmm.201708763
- 24. Nakamura A, Kaneko N, Villemagne VL, Kato T, Doecke J, Doré V, Fowler C, Li QX, Martins R, Rowe C, et al. High performance plasma amyloid-β biomarkers for Alzheimer's disease. *Nature* (2018) **554**:249–254. doi:10.1038/nature25456
- 25. Shen Y, Wang H, Sun Q, Yao H, Keegan AP, Mullan M, Wilson J, Lista S, Leyhe T, Laske C, et al. Increased Plasma Beta-Secretase 1 May Predict Conversion to Alzheimer's Disease Dementia in Individuals With Mild Cognitive Impairment. *Biol Psychiatry* (2018) **83**:447–455. doi:10.1016/j.biopsych.2017.02.007
- 26. Verberk IMW, Slot RE, Verfaillie SCJ, Heijst H, Prins ND, van Berckel BNM, Scheltens P, Teunissen CE, van der Flier WM. Plasma Amyloid as Prescreener for the Earliest <scp>A</scp> Izheimer Pathological Changes. *Ann Neurol* (2018) **84**:648–658. doi:10.1002/ana.25334
- 27. Winston CN, Goetzl EJ, Baker LD, Vitiello M V., Rissman RA. Growth Hormone-Releasing Hormone Modulation of Neuronal Exosome Biomarkers in Mild Cognitive Impairment. *J Alzheimer's Dis* (2018) **66**:971–981. doi:10.3233/JAD-180302
- Agliardi C, Guerini FR, Zanzottera M, Bianchi A, Nemni R, Clerici M. SNAP-25 in Serum Is Carried by Exosomes of Neuronal Origin and Is a Potential Biomarker of Alzheimer's Disease. *Mol Neurobiol* (2019) 56:5792–5798. doi:10.1007/s12035-019-1501-x
- Baldacci F, Daniele S, Piccarducci R, Giampietri L, Pietrobono D, Giorgi FS, Nicoletti V, Frosini D, Libertini P, Lo Gerfo A, et al. Potential Diagnostic Value of Red Blood Cells α-Synuclein Heteroaggregates in Alzheimer's Disease. *Mol Neurobiol* (2019) 56:6451–6459. doi:10.1007/s12035-019-1531-4
- Boccardi V, Paolacci L, Remondini D, Giampieri E, Poli G, Curti N, Cecchetti R, Villa A, Ruggiero C, Brancorsini S, et al. Cognitive Decline and Alzheimer's Disease in Old Age: A Sex-Specific Cytokinome Signature. J Alzheimer's Dis (2019) 72:911–918. doi:10.3233/JAD-190480
- 31. Bram JM de F, Talib LL, Joaquim HPG, Sarno TA, Gattaz WF, Forlenza OV. Protein levels of ADAM10, BACE1, and PSEN1 in platelets and leukocytes of Alzheimer's disease patients. *Eur Arch Psychiatry Clin Neurosci* (2019) **269**:963–972. doi:10.1007/s00406-018-0905-3
- 32. Chatterjee P, Elmi M, Goozee K, Shah T, Sohrabi HR, Dias CB, Pedrini S, Shen K, Asih PR, Dave P, et al. Ultrasensitive Detection of Plasma Amyloid-β as a Biomarker for Cognitively Normal Elderly Individuals at Risk of Alzheimer's Disease. J Alzheimer's Dis (2019) 71:775–783. doi:10.3233/JAD-190533
- 33. Chen Z, Mengel D, Keshavan A, Rissman RA, Billinton A, Perkinton M, Percival-Alwyn J, Schultz A, Properzi M, Johnson K, et al. Learnings about the complexity of extracellular tau aid development of a blood-based screen for Alzheimer's disease.

Alzheimer's Dement (2019) 15:487–496. doi:10.1016/j.jalz.2018.09.010

- 34. Guzmán-Martínez L, Tapia JP, Farías GA, González A, Estrella M, MacCioni RB. The Alz-tau Biomarker for Alzheimer's Disease: Study in a Caucasian Population. *J Alzheimer's Dis* (2019) **67**:1181–1186. doi:10.3233/JAD-180637
- 35. Han S-HH, Park J-CC, Byun MS, Yi D, Lee JH, Lee DY, Mook-Jung I. Blood acetylcholinesterase level is a potential biomarker for the early detection of cerebral amyloid deposition in cognitively normal individuals. *Neurobiol Aging* (2019) **73**:21–29. doi:10.1016/j.neurobiolaging.2018.09.001
- 36. Iulita MF, Ganesh A, Pentz R, Flores Aguilar L, Gubert P, Ducatenzeiler A, Christie S, Wilcock GK, Cuello AC. Identification and Preliminary Validation of a Plasma Profile Associated with Cognitive Decline in Dementia and At-Risk Individuals: A Retrospective Cohort Analysis. J Alzheimer's Dis (2019) 67:327–341. doi:10.3233/JAD-180970
- 37. Jia L, Qiu Q, Zhang H, Chu L, Du Y, Zhang J, Zhou C, Liang F, Shi S, Wang S, et al. Concordance between the assessment of Aβ42, T-tau, and P-T181-tau in peripheral blood neuronal-derived exosomes and cerebrospinal fluid. *Alzheimers Dement* (2019) **15**:1071–1080. doi:10.1016/j.jalz.2019.05.002
- 38. Kapogiannis D, Mustapic M, Shardell MD, Berkowitz ST, Diehl TC, Spangler RD, Tran J, Lazaropoulos MP, Chawla S, Gulyani S, et al. Association of Extracellular Vesicle Biomarkers with Alzheimer Disease in the Baltimore Longitudinal Study of Aging. *JAMA Neurol* (2019) **76**:1340–1351. doi:10.1001/jamaneurol.2019.2462
- Li W-WW, Shen Y-YY, Tian D-YY, Bu X-L Le, Zeng F, Liu Y-HH, Chen Y, Yao X-QQ, Li H-YY, Chen D-WW, et al. Brain Amyloid-β deposition and blood biomarkers in patients with clinically diagnosed alzheimer's disease. *J Alzheimer's Dis* (2019) 69:169–178. doi:10.3233/JAD-190056
- 40. Liu S, Suzuki H, Ito H, Korenaga T, Akatsu H, Meno K, Uchida K. Serum levels of proteins involved in amyloid-β clearance are related to cognitive decline and neuroimaging changes in mild cognitive impairment. *Alzheimer's Dement Diagnosis, Assess Dis Monit* (2019) **11**:85–97. doi:10.1016/j.dadm.2018.11.003
- 41. Liu S, Pan J, Tang K, Lei Q, He L, Cai X, Li Z. Alpha 1-antichymotrypsin may be a biomarker for the progression of amnestic mild cognitive impairment. *Acta Neurol Belg* (2019) doi:10.1007/s13760-019-01206-3
- 42. Meng X, Li T, Wang X, Lv X, Sun Z, Zhang J, Su F, Kang S, Kim S, An SSA, et al. Association between increased levels of amyloid-β oligomers in plasma and episodic memory loss in Alzheimer's disease. *Alzheimers Res Ther* (2019) **11**:89. doi:10.1186/s13195-019-0535-7
- 43. Morgan AR, Touchard S, Leckey C, O'Hagan C, Nevado-Holgado AJ, Barkhof F, Bertram L, Blin O, Bos I, Dobricic V, et al. Inflammatory biomarkers in Alzheimer's disease plasma. *Alzheimer's Dement* (2019) **15**:776–787. doi:10.1016/j.jalz.2019.03.007
- 44. Nabers A, Hafermann H, Wiltfang J, Gerwert K. Aβ and tau structure-based biomarkers for a blood- and CSF-based two-step

recruitment strategy to identify patients with dementia due to Alzheimer's disease. *Alzheimer's Dement Diagnosis, Assess Dis Monit* (2019) **11**:257–263. doi:10.1016/j.dadm.2019.01.008

- 45. Palmqvist S, Janelidze S, Stomrud E, Zetterberg H, Karl J, Zink K, Bittner T, Mattsson N, Eichenlaub U, Blennow K, et al. Performance of Fully Automated Plasma Assays as Screening Tests for Alzheimer Disease–Related β-Amyloid Status. JAMA Neurol (2019) 76:1060. doi:10.1001/jamaneurol.2019.1632
- 46. Park J-C, Han S-H, Yi D, Byun MS, Lee JH, Jang S, Ko K, Jeon SY, Lee Y-S, Kim YK, et al. Plasma tau/amyloid-β1–42 ratio predicts brain tau deposition and neurodegeneration in Alzheimer's disease. *Brain* (2019) **142**:771–786. doi:10.1093/brain/awy347
- 47. Park JC, Han SH, Lee H, Jeong H, Byun MS, Bae J, Kim H, Lee DY, Yi D, Shin SA, et al. Prognostic plasma protein panel for Aβ deposition in the brain in Alzheimer's disease. *Prog Neurobiol* (2019) **183**:101690. doi:10.1016/j.pneurobio.2019.101690
- 48. Pérez-Grijalba V, Arbizu J, Romero J, Prieto E, Pesini P, Sarasa L, Guillen F, Monleón I, San-José I, Martínez-Lage P, et al. Plasma Aβ42/40 ratio alone or combined with FDG-PET can accurately predict amyloid-PET positivity: a cross-sectional analysis from the AB255 Study. *Alzheimers Res Ther* (2019) **11**:96. doi:10.1186/s13195-019-0549-1
- 49. Perrotte M, Le Page A, Fournet M, Le Sayec M, Rassart É, Fulop T, Ramassamy C. Blood-based redox-signature and their association to the cognitive scores in MCI and Alzheimer's disease patients. *Free Radic Biol Med* (2019) **130**:499–511. doi:10.1016/j.freeradbiomed.2018.10.452
- 50. Schindler SE, Bollinger JG, Ovod V, Mawuenyega KG, Li Y, Gordon BA, Holtzman DM, Morris JC, Benzinger TLS, Xiong C, et al. High-precision plasma β-amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* (2019) **93**:E1647–E1659. doi:10.1212/WNL.00000000008081
- Schipke CG, Günter O, Weinert C, Scotton P, Sigle J-P, Kallarackal J, Kabelitz D, Finzen A, Feuerhelm-Heidl A. Definition and quantification of six immune- and neuroregulatory serum proteins in healthy and demented elderly. *Neurodegener Dis Manag* (2019) 9:193–203. doi:10.2217/nmt-2019-0003
- 52. Whelan CD, Mattsson N, Nagle MW, Vijayaraghavan S, Hyde C, Janelidze S, Stomrud E, Lee J, Fitz L, Samad TA, et al. Multiplex proteomics identifies novel CSF and plasma biomarkers of early Alzheimer's disease. *Acta Neuropathol Commun* (2019) **7**:169. doi:10.1186/s40478-019-0795-2
- 53. Yao F, Zhang K, Zhang Y, Guo Y, Li A, Xiao S, Liu Q, Shen L, Ni J. Identification of blood biomarkers for Alzheimer's disease through computational prediction and experimental validation. *Front Neurol* (2019) **10**:1158. doi:10.3389/fneur.2018.01158
- 54. Abate G, Vezzoli M, Polito L, Guaita A, Albani D, Marizzoni M, Garrafa E, Marengoni A, Forloni G, Frisoni GB, et al. A conformation variant of p53 combined with machine learning identifies alzheimer disease in preclinical and prodromal stages. *J Pers Med* (2021) **11**:1–16. doi:10.3390/jpm11010014

- 55. Abe K, Shang J, Shi X, Yamashita T, Hishikawa N, Takemoto M, Morihara R, Nakano Y, Ohta Y, Deguchi K, et al. A New Serum Biomarker Set to Detect Mild Cognitive Impairment and Alzheimer's Disease by Peptidome Technology. *J Alzheimer's Dis* (2020) **73**:217–227. doi:10.3233/JAD-191016
- 56. Barthélemy NR, Horie K, Sato C, Bateman RJ. Blood plasma phosphorylated-tau isoforms track CNS change in Alzheimer's disease. *J Exp Med* (2020) **217**: doi:10.1084/jem.20200861
- 57. Doecke JD, Pérez-Grijalba V, Fandos N, Fowler C, Villemagne VL, Masters CL, Pesini P, Sarasa M. Total Aβ42/Aβ40 ratio in plasma predicts amyloid-PET status, independent of clinical AD diagnosis. *Neurology* (2020) **94**:E1580–E1591. doi:10.1212/WNL.00000000009240
- 58. Eke CS, Sakr F, Jammeh E, Zhao P, Ifeachor E. A Robust Blood-based Signature of Cerebrospinal Fluid Aβ(42) Status. Annu Int Conf IEEE Eng Med Biol Soc IEEE Eng Med Biol Soc Annu Int Conf (2020) 2020:5523–5526. doi:10.1109/EMBC44109.2020.9175158
- 59. Ellegaard Nielsen J, Sofie Pedersen K, Vestergård K, Georgiana Maltesen R, Christiansen G, Lundbye-Christensen S, Moos T, Risom Kristensen S, Pedersen S. Novel Blood-Derived Extracellular Vesicle-Based Biomarkers in Alzheimer's Disease Identified by Proximity Extension Assay. *Biomedicines* (2020) **8**:199. doi:10.3390/biomedicines8070199
- 60. Feinkohl I, Schipke CG, Kruppa J, Menne F, Winterer G, Pischon T, Peters O. Plasma Amyloid Concentration in Alzheimer's Disease: Performance of a High-Throughput Amyloid Assay in Distinguishing Alzheimer's Disease Cases from Controls. *J Alzheimer's Dis* (2020) **74**:1285–1294. doi:10.3233/JAD-200046
- 61. Gao H, Liu M, Zhao Z, Yang C, Zhu L, Cai Y, Yang Y, Hu Z. Diagnosis of Mild Cognitive Impairment and Alzheimer's Disease by the Plasma and Serum Amyloid-beta 42 Assay through Highly Sensitive Peptoid Nanosheet Sensor. *ACS Appl Mater Interfaces* (2020) **12**:9693–9700. doi:10.1021/acsami.0c00370
- 62. Janelidze S, Mattsson N, Palmqvist S, Smith R, Beach TG, Serrano GE, Chai X, Proctor NK, Eichenlaub U, Zetterberg H, et al. Plasma P-tau181 in Alzheimer's disease: relationship to other biomarkers, differential diagnosis, neuropathology and longitudinal progression to Alzheimer's dementia. *Nat Med* (2020) **26**:379–386. doi:10.1038/s41591-020-0755-1
- 63. Karikari TK, Pascoal TA, Ashton NJ, Janelidze S, Benedet AL, Rodriguez JL, Chamoun M, Savard M, Kang MS, Therriault J, et al. Blood phosphorylated tau 181 as a biomarker for Alzheimer's disease: a diagnostic performance and prediction modelling study using data from four prospective cohorts. *Lancet Neurol* (2020) **19**:422–433. doi:10.1016/S1474-4422(20)30071-5
- 64. Palmqvist S, Janelidze S, Quiroz YT, Zetterberg H, Lopera F, Stomrud E, Su Y, Chen Y, Serrano GE, Leuzy A, et al. Discriminative Accuracy of Plasma Phospho-tau217 for Alzheimer Disease vs Other Neurodegenerative Disorders. *JAMA* (2020) **324**:772–781. doi:10.1001/JAMA.2020.12134

- 65. Perrotte M, Haddad M, Le Page A, Frost EH, Fulöp T, Ramassamy C. Profile of pathogenic proteins in total circulating extracellular vesicles in mild cognitive impairment and during the progression of Alzheimer's disease. *Neurobiol Aging* (2020) **86**:102–111. doi:10.1016/j.neurobiolaging.2019.10.010
- 66. Nam E, Lee Y-B, Moon C, Chang K-A. Serum Tau Proteins as Potential Biomarkers for the Assessment of Alzheimer's Disease Progression. *Int J Mol Sci* (2020) **21**:5007. doi:10.3390/ijms21145007
- 67. Sheu JJ, Yang LY, Sanotra MR, Wang S Te, Lu HT, Kam RSY, Hsu IU, Kao SH, Lee CK, Shieh JCC, et al. Reduction of AHI1 in the serum of Taiwanese with probable Alzheimer's disease. *Clin Biochem* (2020) **76**:24–30. doi:10.1016/j.clinbiochem.2019.11.011
- 68. Stockmann J, Verberk IMW, Timmesfeld N, Denz R, Budde B, Lange-Leifhelm J, Scheltens P, van der Flier WM, Nabers A, Teunissen CE, et al. Amyloid-β misfolding as a plasma biomarker indicates risk for future clinical Alzheimer's disease in individuals with subjective cognitive decline. *Alzheimers Res Ther* (2020) **12**:169. doi:10.1186/s13195-020-00738-8
- Teuber-Hanselmann S, Rekowski J, Vogelgsang J, Von Arnim C, Reetz K, Stang A, Jöckel KH, Wiltfang J, Esselmann H, Otto M, et al. CSF and blood kallikrein-8: A promising early biomarker for Alzheimer's disease. *J Neurol Neurosurg Psychiatry* (2020) **91**:40–48. doi:10.1136/jnnp-2019-321073
- 70. Verberk IMW, Thijssen E, Koelewijn J, Mauroo K, Vanbrabant J, de Wilde A, Zwan MD, Verfaillie SCJ, Ossenkoppele R, Barkhof F, et al. Combination of plasma amyloid beta(1-42/1-40) and glial fibrillary acidic protein strongly associates with cerebral amyloid pathology. *Alzheimers Res Ther* (2020) **12**:118. doi:10.1186/s13195-020-00682-7
- 71. Wang X, Zimmermann HR, Lockhart SN, Craft S, Ma T. Decreased Levels of Blood AMPKα1 but not AMPKα2 Isoform in Patients with Mild Cognitive Impairment and Alzheimer's Disease: A Pilot Study. J Alzheimer's Dis (2020) 76:217–224. doi:10.3233/JAD-191189
- 72. Westwood S, Baird AL, Anand SN, Nevado-Holgado AJ, Kormilitzin A, Shi L, Hye A, Ashton NJ, Morgan AR, Bos I, et al. Validation of Plasma Proteomic Biomarkers Relating to Brain Amyloid Burden in the EMIF-Alzheimer's Disease Multimodal Biomarker Discovery Cohort. *J Alzheimer's Dis* (2020) **74**:213–225. doi:10.3233/JAD-190434
- 73. Youn YC, Lee BS, Kim GJ, Ryu JS, Lim K, Lee R, Suh J, Park YH, Pyun J-M, Ryu N, et al. Blood Amyloid-β Oligomerization as a Biomarker of Alzheimer's Disease: A Blinded Validation Study. *J Alzheimer's Dis* (2020) **75**:493–499. doi:10.3233/jad-200061
- 74. Jia L, Zhu M, Kong C, Pang Y, Zhang H, Qiu Q, Wei C, Tang Y, Wang Q, Li Y, et al. Blood neuro-exosomal synaptic proteins predict Alzheimer's disease at the asymptomatic stage. *Alzheimer's Dement* (2021) **17**:49–60. doi:10.1002/alz.12166