Supplementary information

Widespread occurrence of covalent lysine– cysteine redox switches in proteins

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Widespread occurrence of covalent lysine–cysteine redox switches in proteins

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Supplementary Data

Supplementary Data 1 (caption)

Excel-based data table listing all pdb entries (resolution \leq 2.0 Å) with close contacts between cysteine and lysine sidechains (N_{Lys -} S_{Cys} interatomic distance < 3 Å). The table contains the corresponding pdb entry, resolution, sequence positions of the lysine and cysteine residues, protein name and source (organism), cellular localisation as well as identification and classification of NOS and/or SONOS crosslinks.

Supplementary Figures

Supplementary Fig. 1. Dependence of the N-S distance on the dielectric constant for the three model situations of Lys-Cys binding. The calculations were carried out for a single sampled conformer at the B3LYP-D3(BJ)/def2-SVP level of theory applying different dielectric constants with the use of the SMD module.

Supplementary Fig. 2. Geometric properties of NOS bridges. Pair plots for four selected structural variables of the NOS bond: N-S distance (dNS), NOS angle (Angle N-O-S), N-O distance (dNO) and O-S distance (dOS). The data used corresponds to the larger set of structures obtained with the def2-SVP basis set (the same as panels a and b). Several variables correlate strongly. The bimodal distribution of the N-S distances is due to steric constraints between the sulfur atom and the lysine chain, with an interplay between the N-O and the O-S bond distances. The N-O-S angle, as it should be expected, strongly correlates with the N-S distance. The results are largely independent of the distance between the α-carbons of the two chains.

Supplementary Fig. 3. Top: histogram of N-S distances (normalised for each set individually), from model calculations on isolated Lysine and Cysteine residues (alpha-carbon distance of 8 Å). The data includes the NOS data (just as listed in Supplementary Fig. 1) adding the two tautomers for Lys(NH₂)/Cys(SH) and Lys(NH₃+)/Cys(S-), the latter data obtained with the use of the SMD continuum model with water as solvent. The geometries were optimised at the B3LYP-D3(BJ)/def2-TZVPP level of theory. Bottom: relative energy differences for the two non-covalent bonded sets, showing that the most stable structures are in the combination Lys(NH₂)/Cys(SH) (about 2 kcal/mol lower), even with such a high dielectric constant. The Lys (NH_3^+) /Cys (S) minima are not obtained for the models in vacuum.

Supplementary Fig. 4. Histogram showing the distribution of detected proteins (non-redundant counts) containing lysine-cysteine crosslinks in dependence from the distance in sequence between the lysine and cysteine residues.

Supplementary Fig. 5. Sequence conservation of NOS bridge residues identified in human proteins or from model organisms.

Supplementary Fig. 6. Sequence conservation of NOS bridge residues identified in proteins from pathogens.

Supplementary Fig. 7. Occurrence and biological functions of proteins containing NOS and SONOS redox switches. Functions of proteins originating from human and plant pathogen are highlighted in blue color, representative examples of relevant species are shown alongside. Cellular functions of human proteins are highlighted in red color, functional subclasses and representative protein families are listed below. Specific information about all proteins regarding origin, biological function, type of NOS/SONOS redox switch, suggested mechanism of the redox switch and potential relevance in disease states is compiled in Supplementary Tables 2 & 3.

Supplementary Fig. 8. Energy penalty for the constrained N-S-distance of 2.7 Å in NHS (H-bond interaction). Top: electronic energy of the lowest conformer of NHS without the constraint. Bottom: lowest energy conformer of NHS with a constrained N-S distance of 2.7 Å. The electronic energy of the nonconstrained conformer is taken as reference.

Supplementary Fig. 9. Summary of the thermochemistry in SONOS bond formation using a cluster model of the COVID-19 main protease SONOS site. The calculations were based on the reduced cluster from COVID-19 main protease (PDB code: 7JR4, rebuilt without covalent NOS or SONOS bridge), a structure including only the Cys22, Cys44 and Lys61 residues and truncating the α -carbons (as terminal methyl groups) was taken for the calculation. The starting material contained additionally two oxygen molecules, which were turned into the two NOS linkages and water molecules via two steps. The structures were optimised using B3LYP-D3(BJ)/def2-SVPD to obtain the free energy corrections under standard state conditions (*T*=298.15 K). The electronic energies were refined using B3LYP-D3(BJ)/def2- TZVPD. Calculations at both levels were carried out with the Gaussian16-A.03 program package under the application of the SMD solvation model (water as solvent). The most thermodynamically stable structure (the SONOS linked cluster) was taken as reference for the energies.

Supplementary Tables

Supplementary Table 1. X-ray crystallographic data collection and refinement statistics.

* Values in parentheses are for the highest-resolution shell.

Supplementary Table 2. NOS/SONOS bridges in proteins from human and plant pathogens. Information is provided for the protein identity, the origin of the protein, the biological function of the protein, the detected switch type (NOS or SONOS) with residues involved and relevant pdb codes, the suggested mechanism of the redox switch and relevant diseases associated with the identified species. Proteins identified based on structural homology are highlighted in gray shading. The complete list of all proteins with detected NOS bridges is provided in Supplementary Data 1.

Supplementary Table 3. Human proteins (or from animal models) with NOS bridges classified according to their cellular function. Information is provided for the protein identity, the origin of the protein, the biological function of the protein, the detected switch type (NOS or SONOS) with residues involved and relevant pdb codes, the suggested mechanism of the redox switch and relevant diseases associated with the identified species. Proteins identified based on sequence homology are highlighted in gray shading. In case the identified protein originates from an animal model system, a putative sequence conservation of the NOS residues in the human orthologue is indicated. The complete list of all proteins with detected NOS bridges is provided in Supplementary Data 1.

Supplementary Table 4. Steady-state kinetic analysis of *Ng*TAL wild-type and variant Glu93Gln under oxidizing (w/o DTT) and reducing (w/ 1 mM DTT) conditions.

Oxidizing conditions, without DTT; reducing conditions, with 1 mM DTT. We estimated k_{cat} , apparent K_M for the substrate D-fructose 6-phosphate (F6P) and catalytic efficiency (k_{cat}/K_M) in a continuous spectrophotometric assay for the conversion of F6P + E4P \rightarrow S7P + G3P as detailed in Methods section. In case of the oxidized, we observed a pronounced lag phase that suggested a catalytic activation under turnover conditions. We thus provide both the steady-state activities after full activation as well as basal activities at $t = 0$ before activation sets in. The x-*fold* change for k_{cat} and K_M for the reduced enzyme relative to the oxidized form at steady state is indicated. All measurements were carried out in triplicate and are shown as mean ± s.d.

^a In case of catalytic activation, progress curves were fitted with eq 1 as detailed in the Methods section.