

SUPPORTING INFORMATION

In silico analysis of SARS-CoV-2 proteins as targets for clinically available drugs

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3. Supporting References

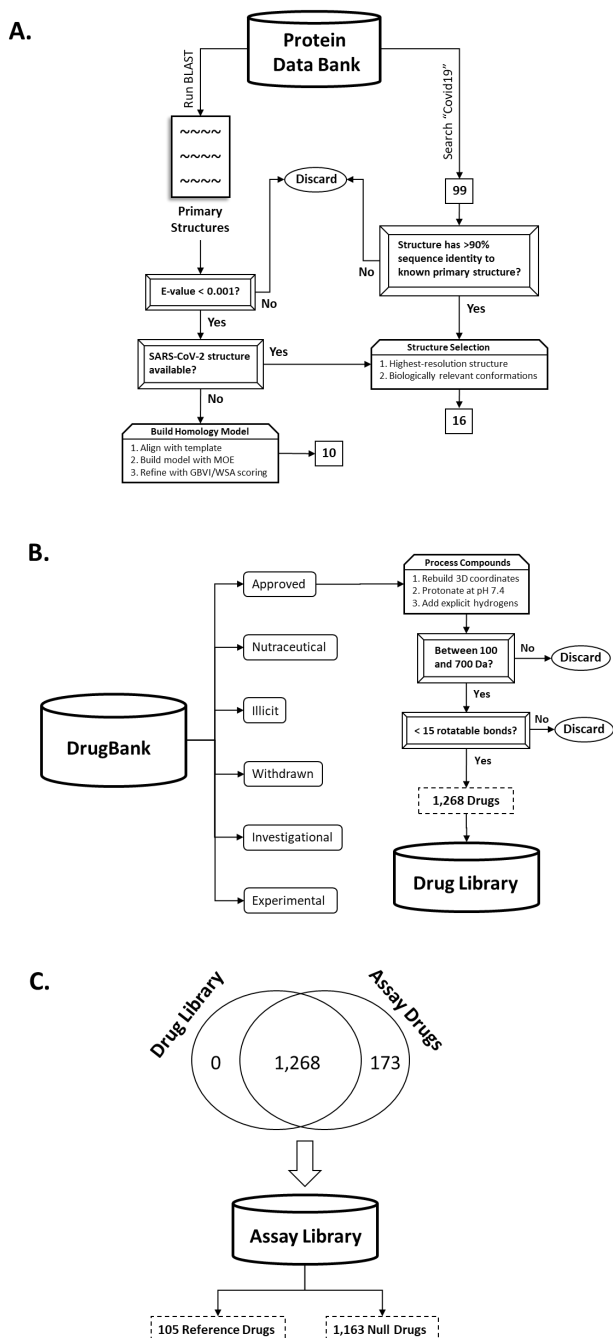


Fig. S1. Flowchart for preparation of SARS-CoV-2 protein structures and drug library for docking. (A) SARS-CoV-2 protein structures were selected from available crystal structures, and where unavailable, homology models were created from related coronaviruses. **(B)** FDA-approved drugs were extracted from DrugBank and prepared for downstream docking experiments. **(C)** From the assay library, 1,268 FDA-approved drugs overlapped with our *in silico* drug library, out of which 105 small molecule drugs had $IC_{50} \leq 2 \mu M$ against Huh7 cells. These were termed 'Reference Drugs'. The flowcharts were prepared using PowerPoint software.

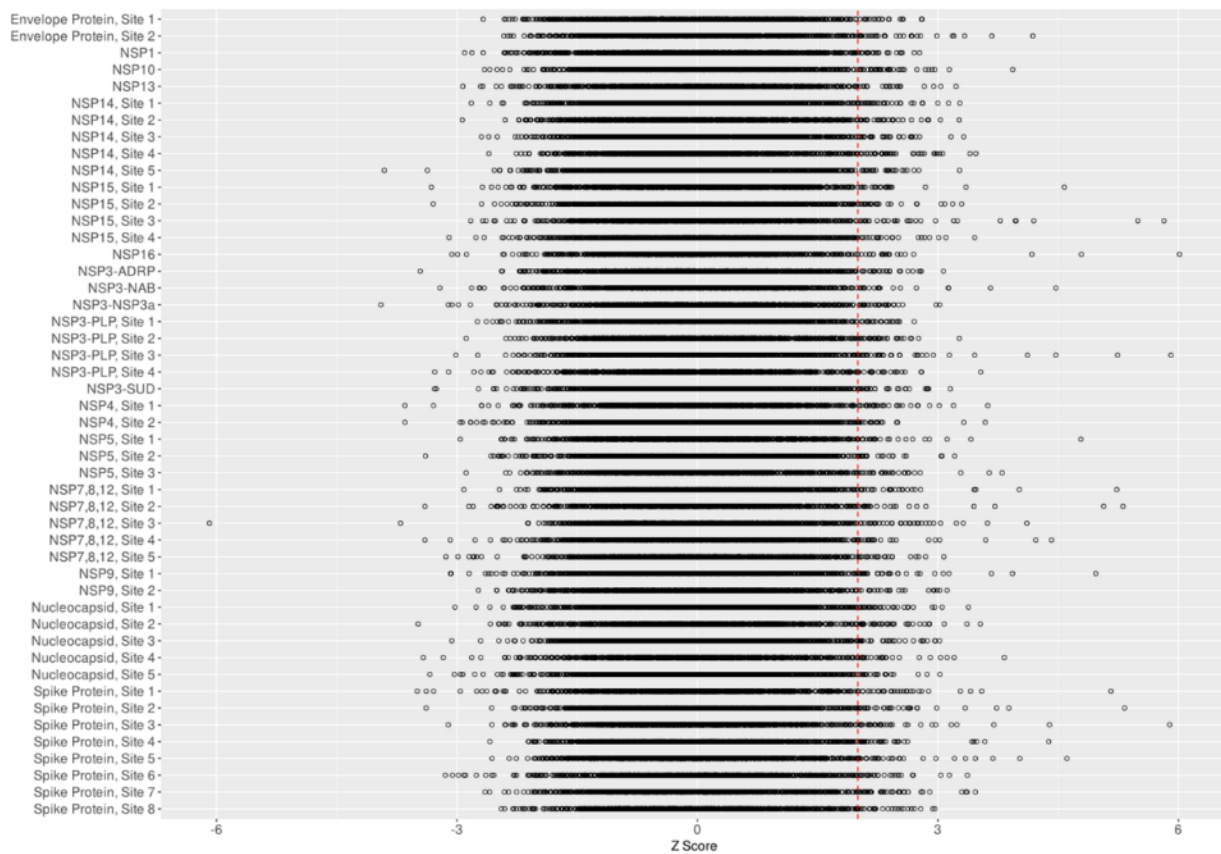


Fig. S2. Z score distributions of drugs for each predicted site. Each black open circle represents a single FDA-approved drug. The dashed red line depicts the Z score cutoff at 2. The plot was generated with R language using the ggplot2 package.

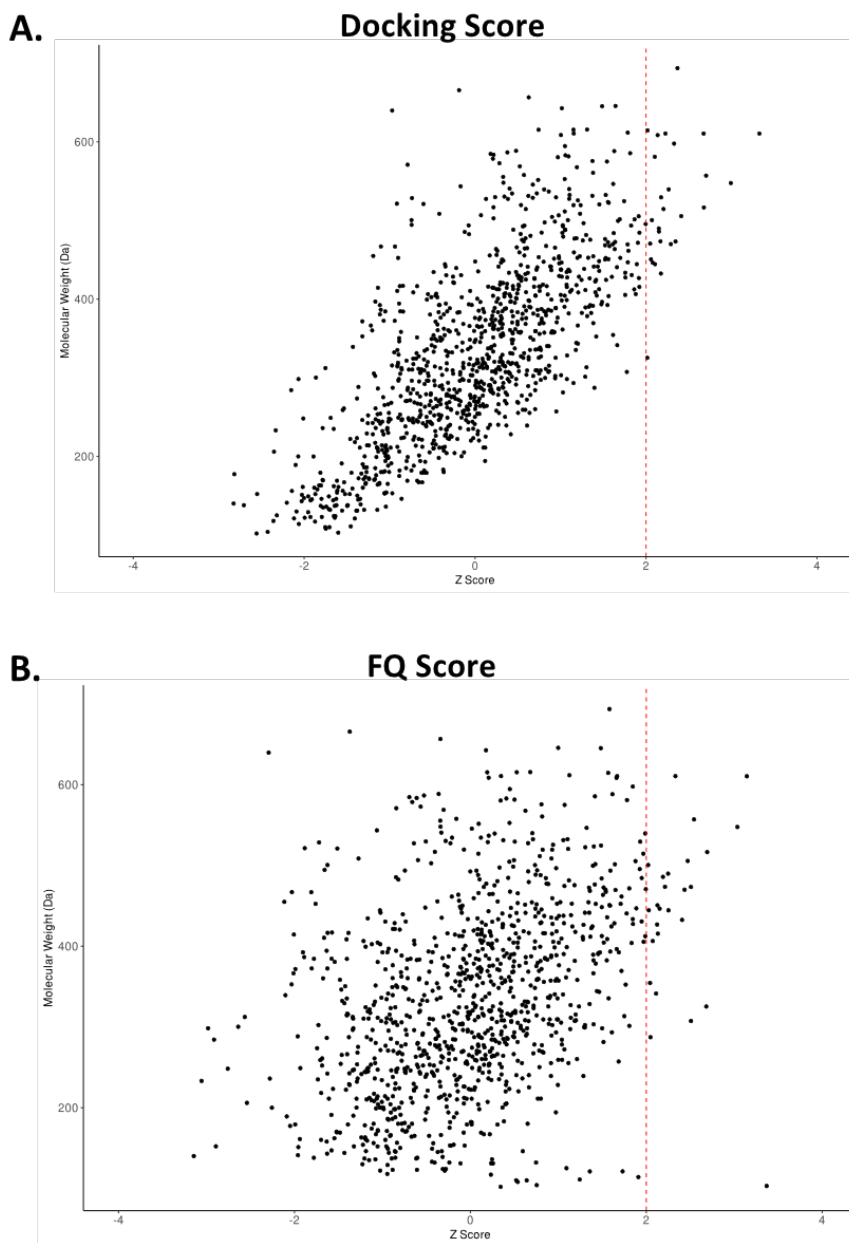


Fig. S3. Molecular weight dependence of scoring methods. The prioritized drugs from site 6 of the spike protein are shown using different scoring methods. (A.) A strong positive correlation was observed between the Z score and molecular weight for the prioritized drugs when using the docking score alone. (B.) However, utilizing the FQ score minimized the positive correlation. The dashed red line represents the Z score cutoff at 2. Plots were generated with R programming language and the ggplot2 package.

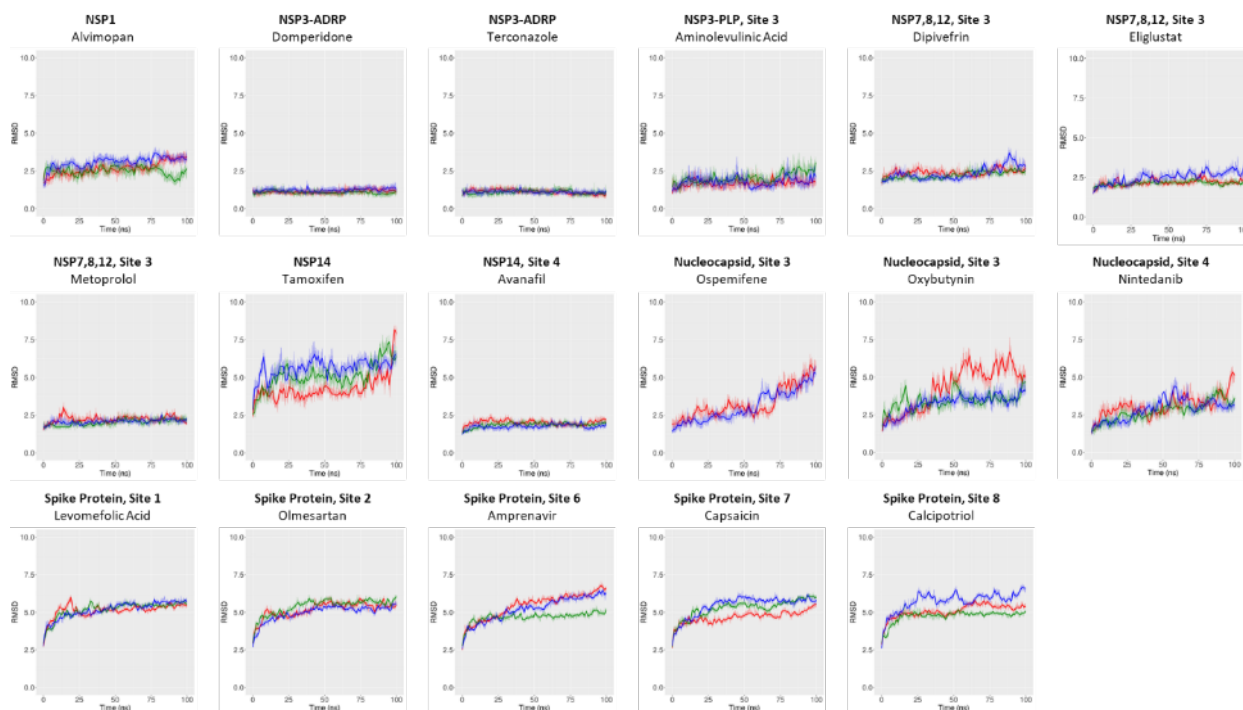


Fig. S4. Protein RMSDs from molecular dynamics simulations trajectories. RMSD calculations were performed using the peptide backbone (C, C α , N). Runs 1, 2, and 3 are colored red, green, and blue, respectively. Systems with larger RMSD display the cases where drugs migrated to symmetry-related sites. Plots were generated with R programming language and the ggplot2 package.

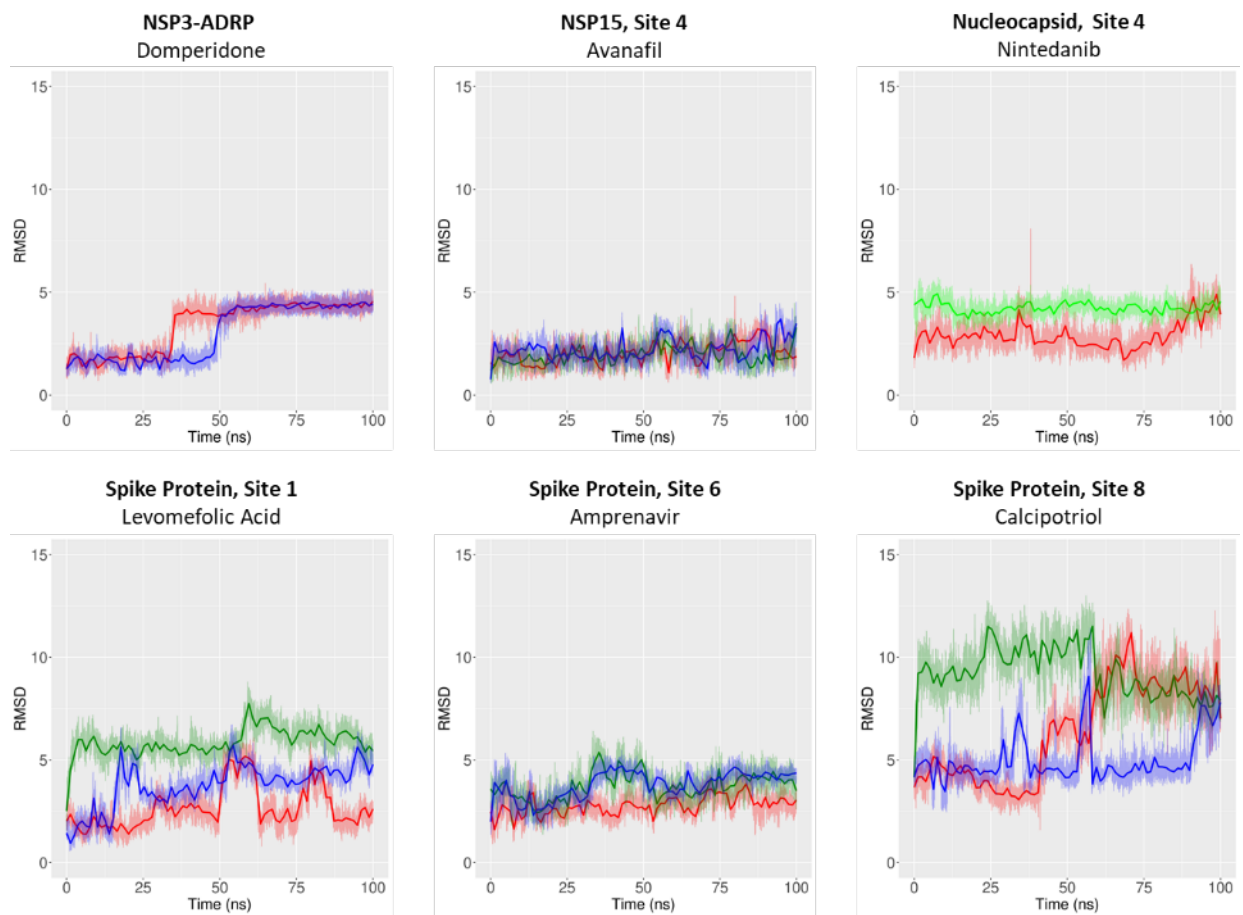


Fig. S5. Drug RMSDs from trajectories of successful systems. RMSD calculations were performed using the heavy atoms (i.e., non-hydrogen atoms) of the drug. Runs 1, 2, and 3 are colored red, green, and blue, respectively. Plots were generated with R programming language and the ggplot2 package.

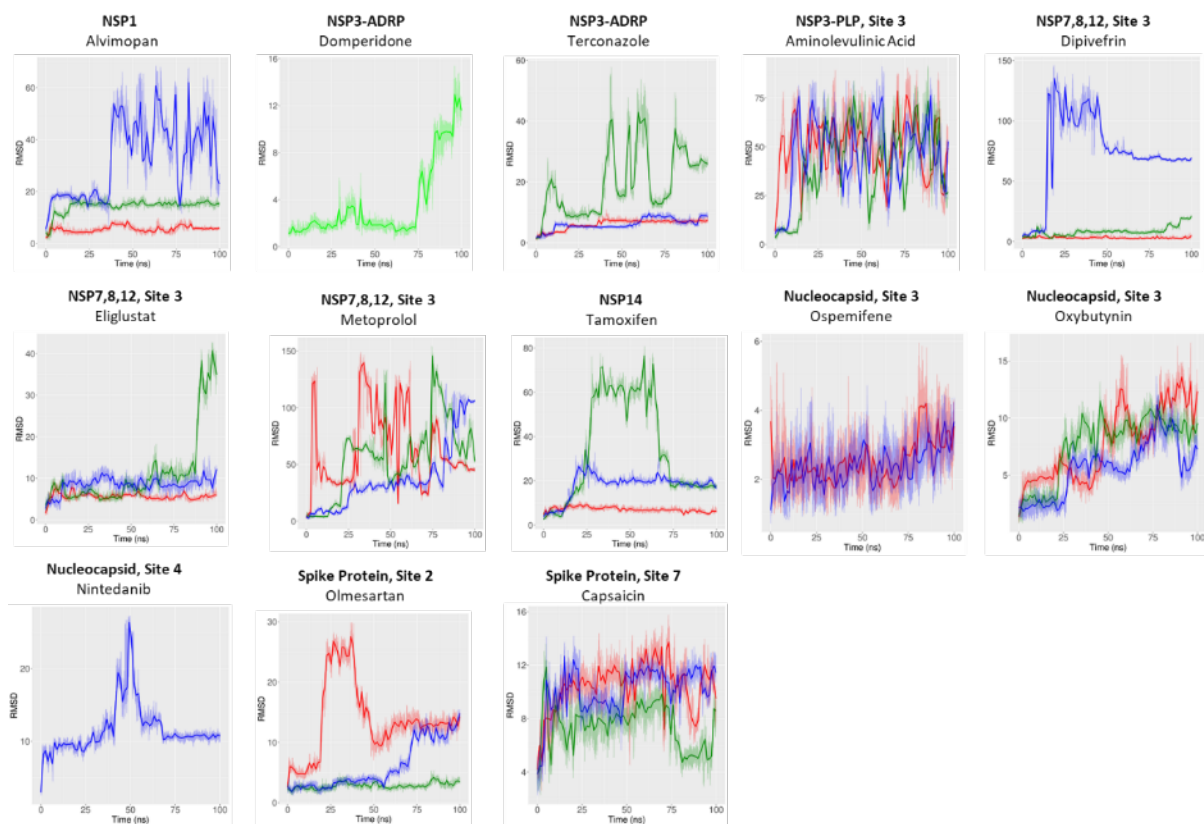


Fig. S6. Drug RMSDs from trajectories of failed systems. RMSD calculations were performed using the heavy atoms (i.e., non-hydrogen atoms) of the drug. Runs 1, 2, and 3 are colored red, green, and blue, respectively. Plots were generated with R programming language and the ggplot2 package.

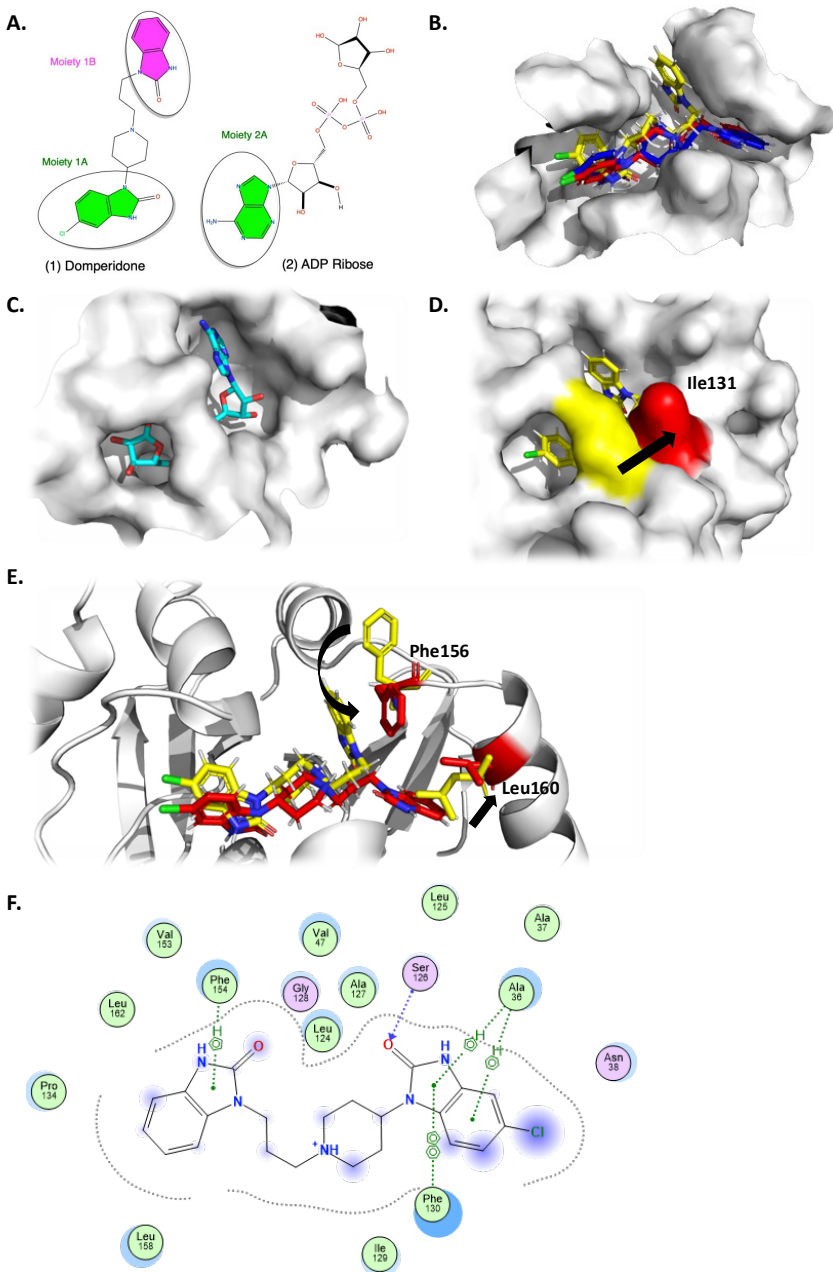


Fig. S7. Domperidone is predicted to displace ADP ribose from the ADRP domain of NSP3. (A) The chemical structures of domperidone (1) and ADP ribose (2), with Moiety 1. (B) The final snapshots from runs 1 and 3 (red and blue, respectively) are displayed in comparison with the original docked pose (yellow). (C) ADP ribose (cyan) binds to the active site of the ADRP domain of NSP3 (PDB: 6WOJ). (D) Upon achieving a stable interaction, domperidone was observed to cause Ile131 to move outwards, opening up the 'clamp' seen over the ribose on ADP ribose prior to hydrolysis. (E) Furthermore, Leu160 moves away to reveal a cryptic site that allows for the

accommodation of the non-chlorinated 1,3-benzodiazol-2-one group of domperidone, which is further stabilized with Phe156. For both (**D**) and (**E**), the original docked system (yellow) was compared with the final frame of run 1 (red). (**F**) A 2D interaction map of domperidone bound in the system with final frame of first MD simulation replicate. Graphics were generated using PyMOL and MOE.

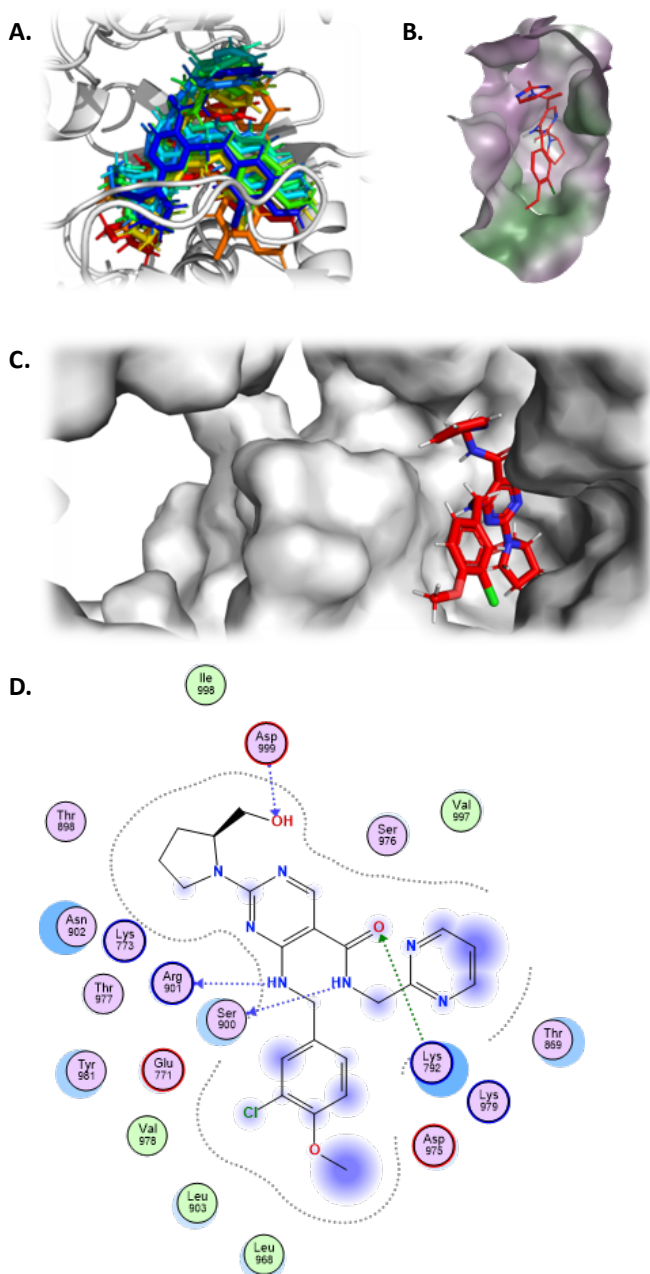


Fig. S8. Avanafile interacts with an allosteric site on NSP15. (A.) Representative frames from run 1 are shown in 10 ns increments (blue → green → red). (B.) Hydrophobic surface representation of the putative allosteric site to which avanafile binds was determined using MOE, where green and purple denote hydrophobicity and hydrophilicity, respectively. (C.) Avanafile is situated in an exposed pocket that is readily accessible to the exterior of the protein via a solvent channel. (D.) A 2D interaction map of avanafile with the system with final frame of first MD simulation replicate. Molecular graphics were generated using PyMOL and MOE.

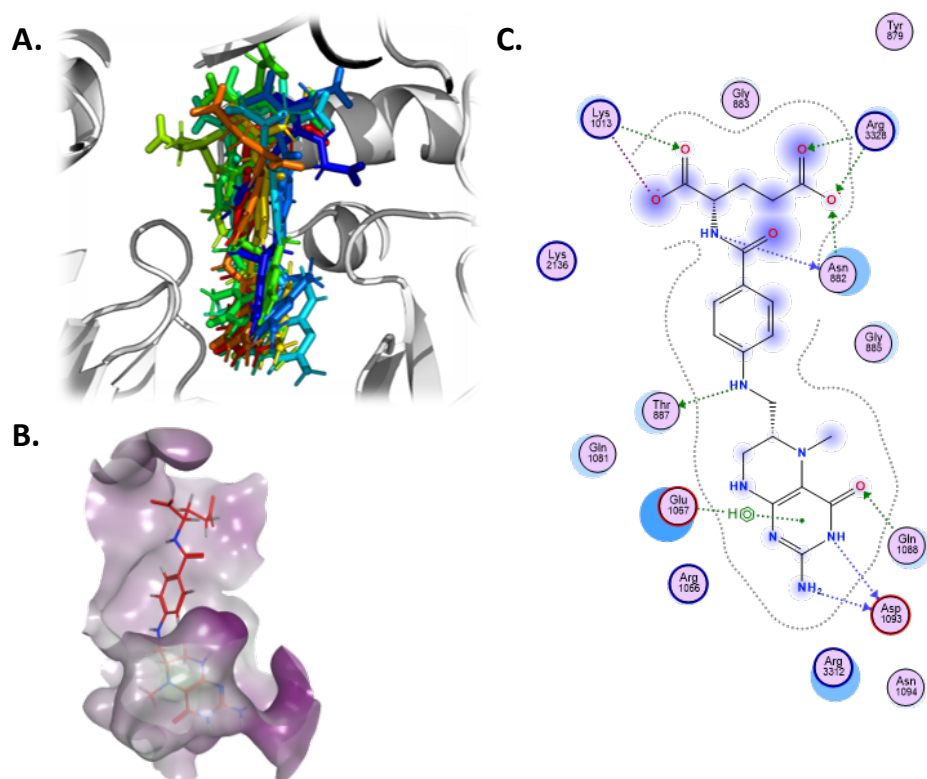


Fig. S9. Levomefolic acid interacts with a putative allosteric site formed between the trimeric spike protein complex. (A.) Representative frames from run 1 are shown in 10 ns increments (blue → green → red). **(B.)** Hydrophobic surface representation of the putative allosteric site to which levomefolic acid binds was determined using MOE, where green and purple denote hydrophobicity and hydrophilicity, respectively. **(C.)** A 2D interaction map of levomefolic acid with the system with final frame of first MD simulation replicate. Molecular graphics were generated using PyMOL and MOE.

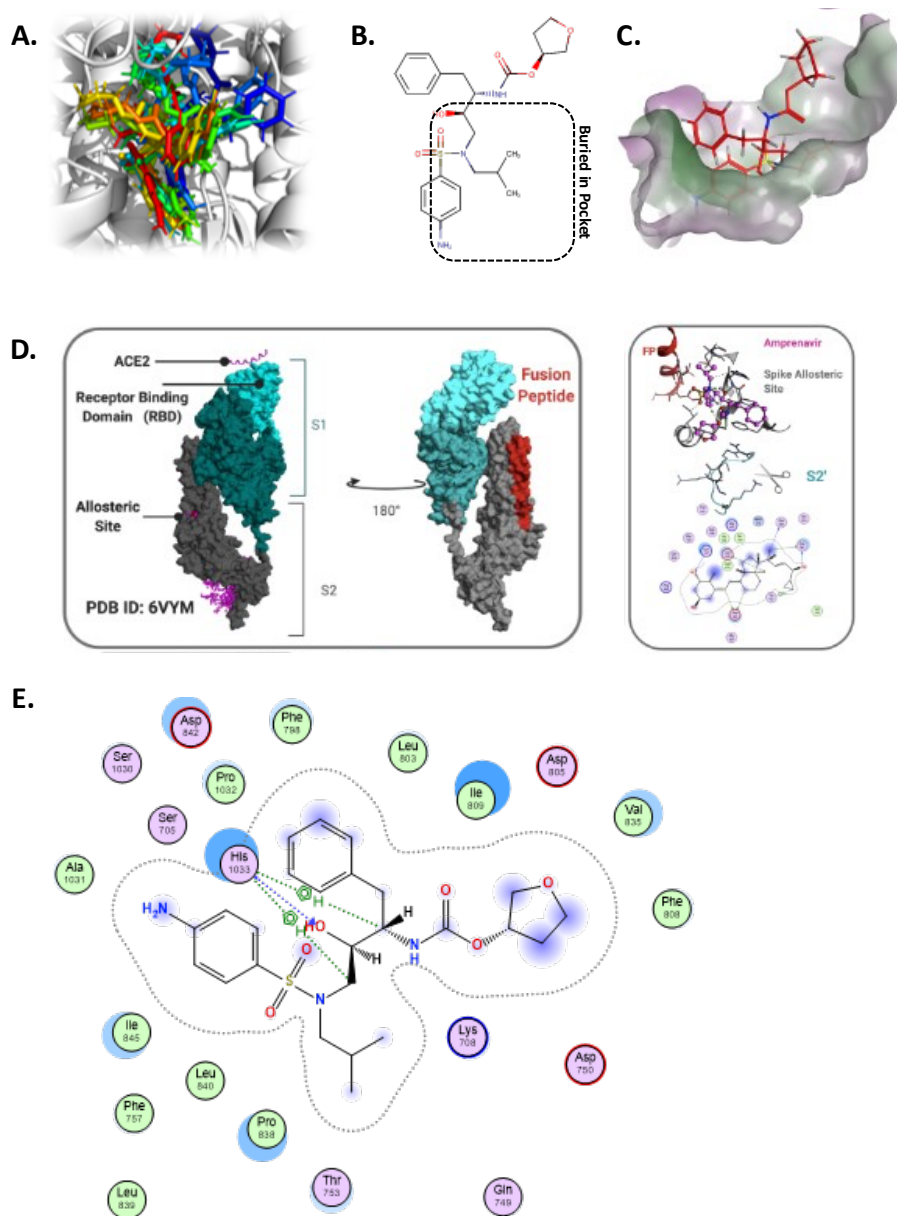


Fig. S10. Amprenavir interacts stably with a putative allosteric site on the spike protein. (A.) Representative frames from run 1 are shown in 10 ns increments (blue → green → red). (B.) Part of amprenavir remains buried in the predicted site throughout the simulations. MarvinSketch was used for drawing and displaying the 2D chemical structure, MarvinSketch 18.18.0, ChemAxon (<https://www.chemaxon.com>). (C.) Hydrophobic surface representation of the putative allosteric site to which amprenavir binds was determined using MOE, where green and purple denote hydrophobicity and hydrophilicity, respectively. (D.) (Left) Single monomer of homotrimer S protein. Domains colored RBD (Cyan), S1 (cyan marine). (Right) Amprenavir to spike allosteric site. 2D generation map of docked pose. Created with biorender.com. (E.) A 2D interaction map of amprenavir with the system with final frame of first MD simulation replicate. Molecular graphics were generated using PyMOL and MOE.

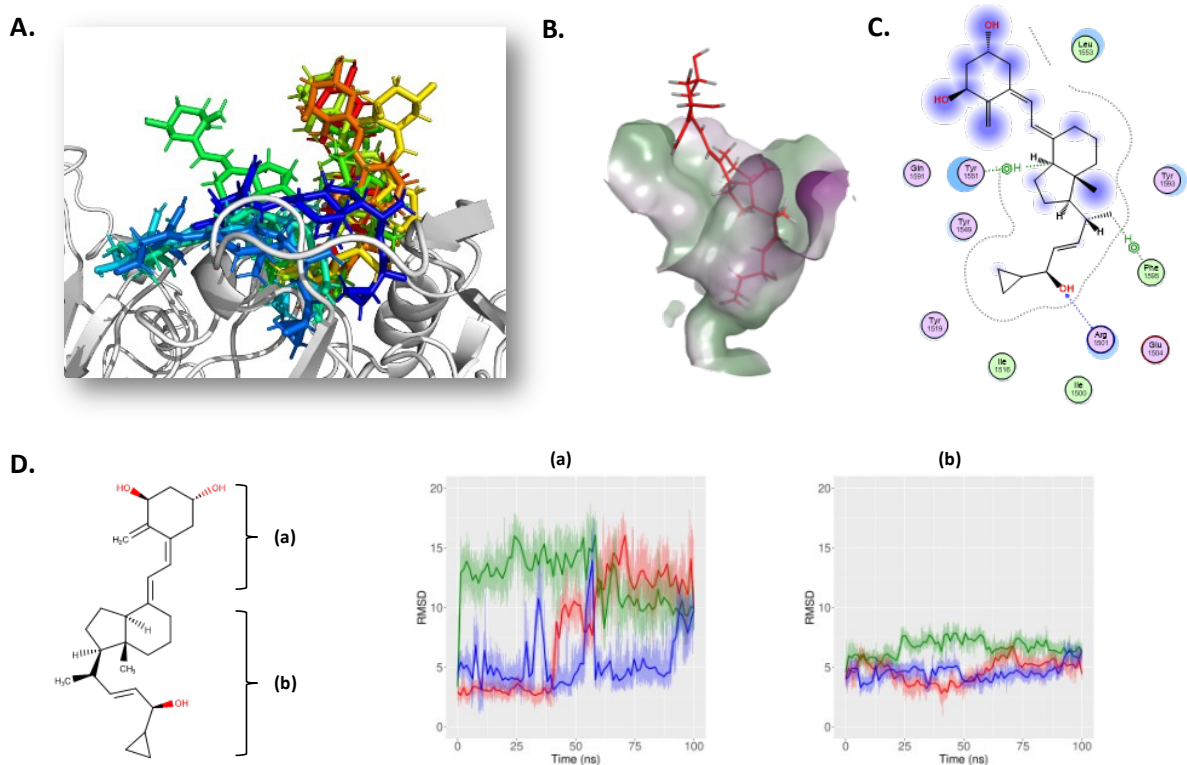


Fig. S11. Calcipotriol maintains a stable interaction with the RBD of the spike protein. (A.) Representative frames from run 1 are shown in 10 ns increments (blue → green → red). (B.) Hydrophobic surface representation of the putative allosteric site to which amprenavir binds was determined using MOE, where green and purple denote hydrophobicity and hydrophilicity, respectively. (C.) A 2D interaction map of calcipotriol with the system. (D.) Calcipotriol contained both (a) solvent-exposed and (b) buried regions. The heavy atoms (i.e., non-hydrogen atoms) were used to calculate drug RMSDs from the trajectories for both regions individually. Runs 1, 2, and 3 are colored red, green, and blue, respectively. MarvinSketch was used for drawing and displaying the 2D chemical structure, MarvinSketch 18.18.0, ChemAxon (<https://www.chemaxon.com>). Molecular graphics were generated using PyMOL and MOE.

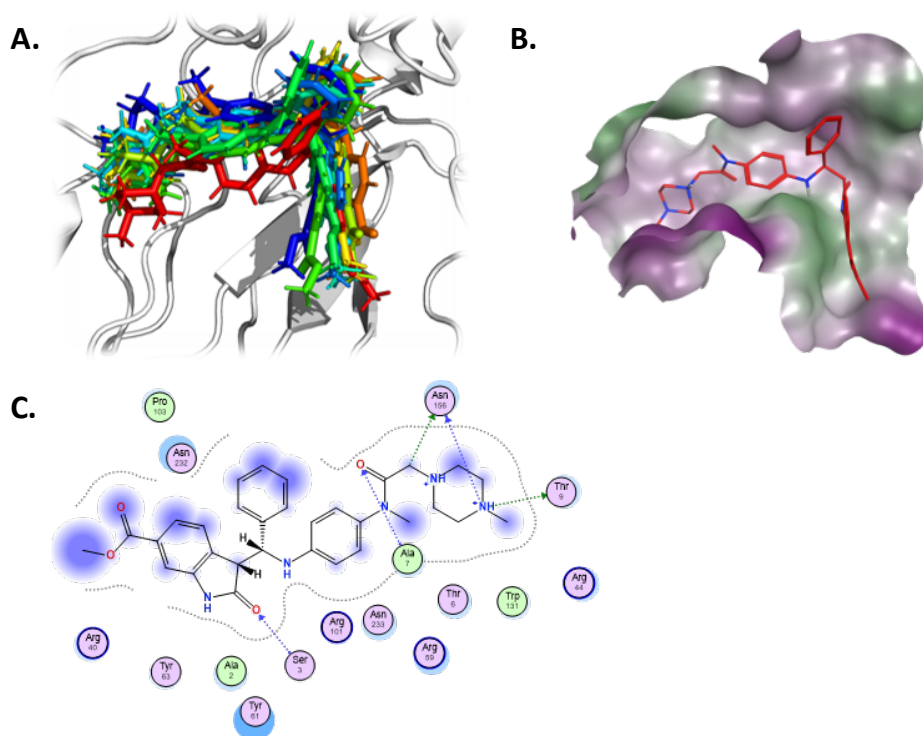


Fig.S12. Nintedanib interacts with the RNA-binding domain of the nucleocapsid protein. (A) Representative frames from run 1 are shown in 10 ns increments (blue → green → red). (B) Hydrophobic surface representation of the putative allosteric site to which nintedanib binds was determined using MOE, where green and purple denote hydrophobicity and hydrophilicity, respectively. (C.) A 2D interaction map of nintedanib with the system with final frame of first MD simulation replicate. Molecular graphics were generated using PyMOL and MOE.

Table S1. Homology Model Statistics and Modeling Data

<u>Protein or Domain</u>	<u>Species</u>	<u>E-value</u>	<u>Template PDB ID</u>	<u>% ID</u>	<u>Region</u>	<u>RMSD to Mean</u>	<u>Cα RMSD to Mean</u>	<u>Contact Energy</u>	<u>Packing Score</u>	<u>GB/VI</u>	<u>U</u>	<u>E_{sol}</u>	<u>E_{ele}</u>	<u>E_{vdw}</u>	<u>E_{bond}</u>	<u>Atom Clashes</u>	<u>BB Bond Outliers</u>	<u>BB Angle Outliers</u>	<u>BB Torsion Outliers</u>	<u>Rotamer Outliers</u>
NSP1	SARS	4.603e-60,	2GDT	86%	2-116	0.1601	0.00000	-106.62647	2.1425035	-32994	-8644	-1796	-9102	14	444	0	0	4	4	0
NSP3a	SARS	2.74E-48	2GRI	79%	17-112	0.0000	0.00000	-80.180763	2.2122338	-26454	-6145	-2470	-6852	217	491	9	0	1	2	0
NSP3 SUD	SARS	1.91E-126	2W2G	75%	1-264	0.0776	0.03018	-465.59048	2.2028725	-143896	-38098	-6419	-40012	-204	2117	8	3	20	13	0
NSP3 NAB	SARS	2.98E-49	2K87	82%	2-92	0.1494	0.00000	-61.182095	2.1671281	-22871	-5925	-1592	-6277	-19	371	0	0	2	4	0
NSP 4	Mouse Hepatitis A59	2.47E-23	3VCB	60%	5-94	0.4073	0.24649	-62.182476	2.1781962	-22020	-5164	-1031	-6574	292	1119	23	7	10	7	0
NSP13 (dimer)	SARS	0	6JYT	100%	3-603	0.0183131	0.007030345	-873.005	2.2819099	-468183	-112055	-18909	-119831	2262	5514	68	12	30	65	5
NSP14	SARS	0	5C8S	95%	1-527	0.286933	0.2587229	-809.2572	2.1527419	-383837	-91606	-19052	-99075	2099	5370	97	0	17	10	5
E Protein (Pentamer)	SARS	1.01E-23	5X29	89%	20-81	0.8426	0.76003	-118.68491	2.1943591	-1364	-245	-562	-513	22	245	115	4	7	19	3
Nucleocapsid CTD	SARS	4.65E-66	2CJR	96%	11-128	0.0000	0.00000	-258.84213	2.177073	-163763	-40916	-6551	-43357	435	2006	44	19	9	14	1

Table S4. List of 105 hits from the *in vitro* assay against SARS-CoV-2 infection in Huh-7 cells with potency $\leq 2 \mu\text{M}$ with methods as described in ¹⁰.

Drug Name	DrugBank ID	Drug Name	DrugBank ID	Drug Name	DrugBank ID
Acebutolol	DB01193	Ethionamide	DB00609	Nintedanib	DB09079
Acetazolamide	DB00819	Ethosuximide	DB00593	Olmесartan	DB00275
Acetohexamide	DB00414	Ethotoin	DB00754	Omeprazole	DB00338
Alfuzosin	DB00346	Ezetimibe	DB00973	Ondansetron	DB00904
Allopurinol	DB00437	Fedratinib	DB12500	Ospemifene	DB04938
Alvimopan	DB06274	Fludarabine	DB01073	Oxybutynin	DB01062
Aminolevulinic acid	DB00855	Fludrocortisone	DB00687	Pamidronic acid	DB00282
Amiodarone	DB01118	Gemcitabine	DB00441	Phentolamine	DB00692
Amprenavir	DB00701	Gilteritinib	DB12141	Phenytoin	DB00252
Apraclonidine	DB00964	Glasdegib	DB11978	Procainamide	DB01035
Avanafil	DB06237	Guanethidine	DB01170	Pyridoxine	DB00165
Bosutinib	DB06616	Hydroxychloroquine	DB01611	Quinestrol	DB04575
Calcipotriol	DB02300	Ingenol mebutate	DB05013	Raloxifene	DB00481
Capsaicin	DB06774	Ivabradine	DB09083	Remdesivir	DB14761
Carbinoxamine	DB00748	Ketorolac	DB00465	Rivastigmine	DB00989
Carmustine	DB00262	Lapatinib	DB01259	Rocuronium	DB00728
Cefotaxime	DB00493	Levamisole	DB00848	Rofecoxib	DB00533
Cefoxitin	DB01331	Levofloxacin	DB01137	Sevoflurane	DB01236
Chloroxine	DB01243	Levomefolic acid	DB11256	Sildenafil	DB00203
Chlorthalidone	DB00310	Lomitapide	DB08827	Simvastatin	DB00641
Cidofovir	DB00369	Loperamide	DB00836	Tafamidis	DB11644
Cilostazol	DB01166	Loratadine	DB00455	Tamoxifen	DB00675
Clofazimine	DB00845	Meclizine	DB00737	Terconazole	DB00251
Dasatinib	DB01254	Mepivacaine	DB00961	Tetrabenazine	DB04844
Dexmedetomidine	DB00633	Meropenem	DB00760	Thalidomide	DB01041
Dienogest	DB09123	Mestranol	DB01357	Thioridazine	DB00679
Dipivefrin	DB00449	Metformin	DB00331	Tioguanine	DB00352
Domperidone	DB01184	Metoclopramide	DB01233	Trimethadione	DB00347
Dorzolamide	DB00869	Metoprolol	DB00264	Ulipristal	DB08867
Duvelisib	DB11952	Naratriptan	DB00952	Valproic acid	DB00313
Edaravone	DB12243	Nebivolol	DB04861	Vardenafil	DB00862
Eflornithine	DB06243	Nevirapine	DB00238	Verapamil	DB00661
Eliglustat	DB09039	Niclosamide	DB06803	Vilazodone	DB06684
Entecavir	DB00442	Nifedipine	DB01115	Zanamivir	DB00558
Eslicarbazepine acetate	DB09119	Nilutamide	DB00665	Zonisamide	DB00909

Table S5. Reference drugs identified using FQ score.

Drug Name	DrugBank ID	Molecular Weight	Drug Type	Targets	Z Score
Alvimopan	DB06274	424.53	Peripherally acting μ opioid receptor antagonist	NSP1	2.25
Aminolevulinic Acid	DB00855	131.13	Photochemotherapy for actinic keratosis	NSP3-PLP, Site 3	2.86
Amprenavir	DB00701	505.63	HIV protease inhibitor	Spike Protein, Site 6	2.47
				NSP15, Site 1	2.23
				NSP5, Site 2	2.11
				NSP5, Site 1	2.11
				NSP15, Site 3	2.09
Avanafil	DB06237	484.95	Phosphodiesterase-5 inhibitor for erectile dysfunction	NSP15, Site 4	2.43
				NSP15, Site 2	2.24
				NSP3-PLP, Site 2	2.21
Calcipotriol	DB02300	412.60	Treatment of plaque psoriasis	Spike Protein, Site 8	2.21
Capsaicin	DB06774	305.41	Topical analgesic	Spike Protein, Site 7	2.33
Dipivefrin	DB00449	351.44	Prodrug of adrenaline	NSP7,8,12, Site 3	2.60
Domperidone	DB01184	425.91	Dopamine receptor antagonist	NSP3-ADRP	2.53
				NSP3-PLP, Site 4	2.30
				NSP3-SUD	2.16
Eliglustat	DB09039	404.55	Glucosylceramide synthase inhibitor	NSP7,8,12, Site 3	3.03
				NSP5, Site 3	2.44
				NSP14, Site 1	2.27

				Spike Protein, Site 7	2.12
Levomefolic Acid	DB11256	459.46	Food additive for folate deficiency	Spike Protein, Site 1	2.06
Metoprolol	DB00264	267.36	β 1 adrenergic receptor antagonist	NSP7,8,12, Site 3	2.54
Nintedanib	DB09079	539.62	Kinase inhibitor	Nucleocapsid, Site 4	2.35
				Envelope Protein, Site 1	2.26
Olmesartan	DB00275	446.50	Angiotensin II receptor antagonist	Spike Protein, Site 2	2.40
Ospemifene	DB04938	378.89	Selective estrogen receptor modulator	Nucleocapsid, Site 3	2.03
Oxybutynin	DB01062	357.49	Relief of overactive bladder	Nucleocapsid, Site 3	2.32
				Spike Protein, Site 7	2.01
Tamoxifen	DB00675	371.51	Selective estrogen receptor modulator	NSP14, Site 5	2.35
				NSP3-NSP3a	2.28
Terconazole	DB00251	532.46	14-alpha-demethylase inhibitor in fungi	NSP3-ADRP	2.52
				NSP3-PLP, Site 1	2.11
				Spike Protein, Site 2	2.01

Table S6. Number of Ions used in MD Simulations.

Protein	Drug	Na ⁺ Ions	Cl ⁻ Ions
NSP14	Tamoxifen	0	9
NSP15	Avanafil	162	0
NSP1	Alvimopan	6	0
NSP3-ADRP	Domperidone	0	0
NSP3-ADRP	Terconazole	0	0
NSP3-PLP	Aminolevulinic Acid	3	0
NSP7,8,12	Dipivefrin	12	0
NSP7,8,12	Eliglustat	12	0
NSP7,8,12	Metoprolol	12	0
Nucleocapsid	Ospemifene	0	84
Nucleocapsid	Oxybutynin	0	85
Nucleocapsid	Nintedanib	0	85
Spike	Levomefolic Acid	59	0
Spike	Olmesartan	59	0
Spike	Amprenavir	57	0
Spike	Capsaicin	57	0
Spike	Calcipotriol	57	0

3. Supplemental References

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