

Figure S1

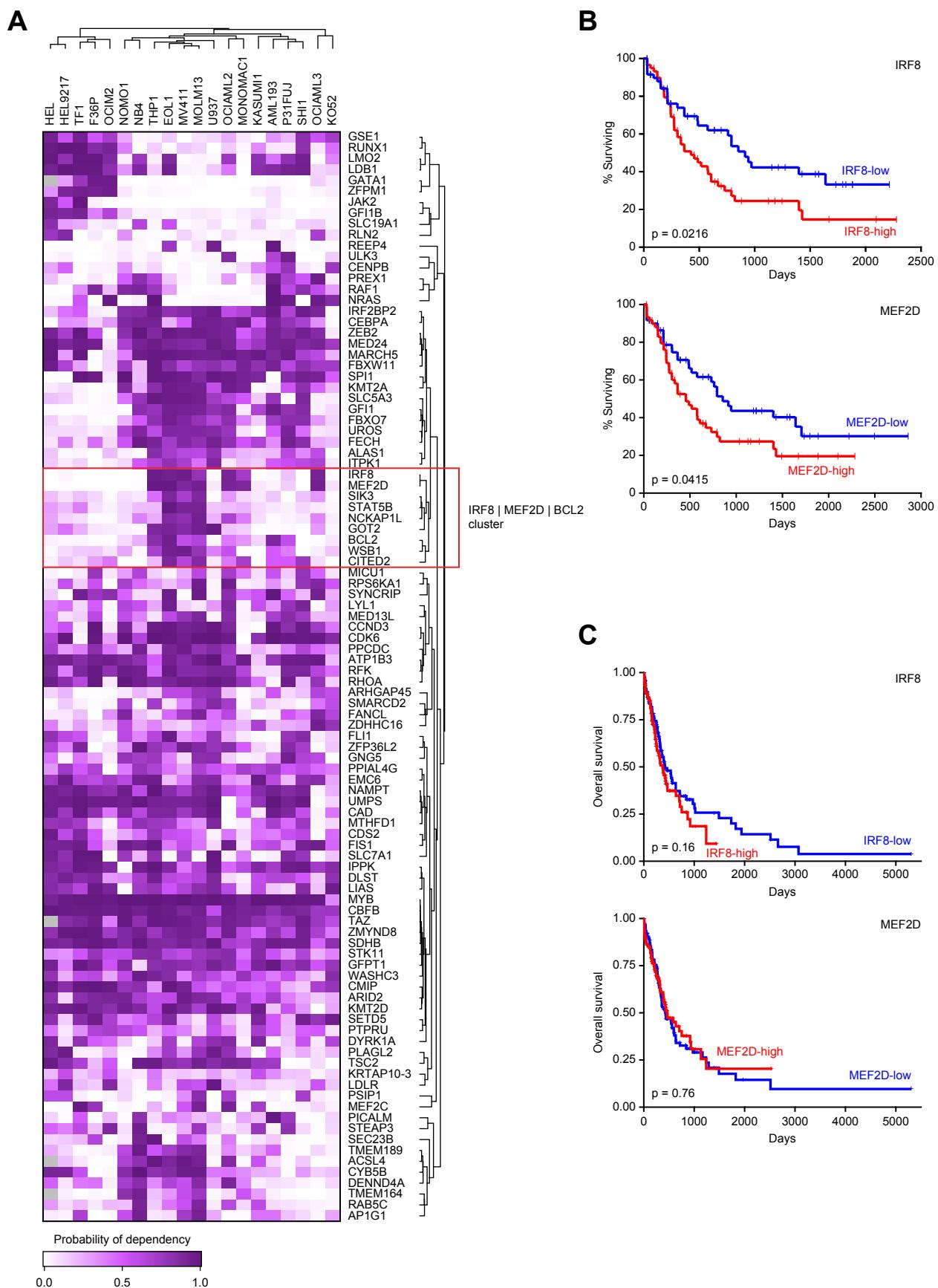


Figure S1, related to Figure 1. Selective AML gene dependencies

- A, A heatmap of 225 selective AML dependencies reflecting probability of dependency inferred from the skewed LRT scores (Methods). Rows and columns are hierarchically clustered on Pearson correlation with complete linkage.
- B, Kaplan-Meyer plots of overall survival in TCGA AML patients (Network 2013) with MEF2D and IRF8 expression above the 60th percentile (red line) and below the 40th percentile (blue line).
- C, Kaplan-Meyer plots of overall survival in BeatAML patients (Tyner et al. 2018) with MEF2D and IRF8 expression above the 60th percentile (red line) and below the 40th percentile (blue line).

Figure S2

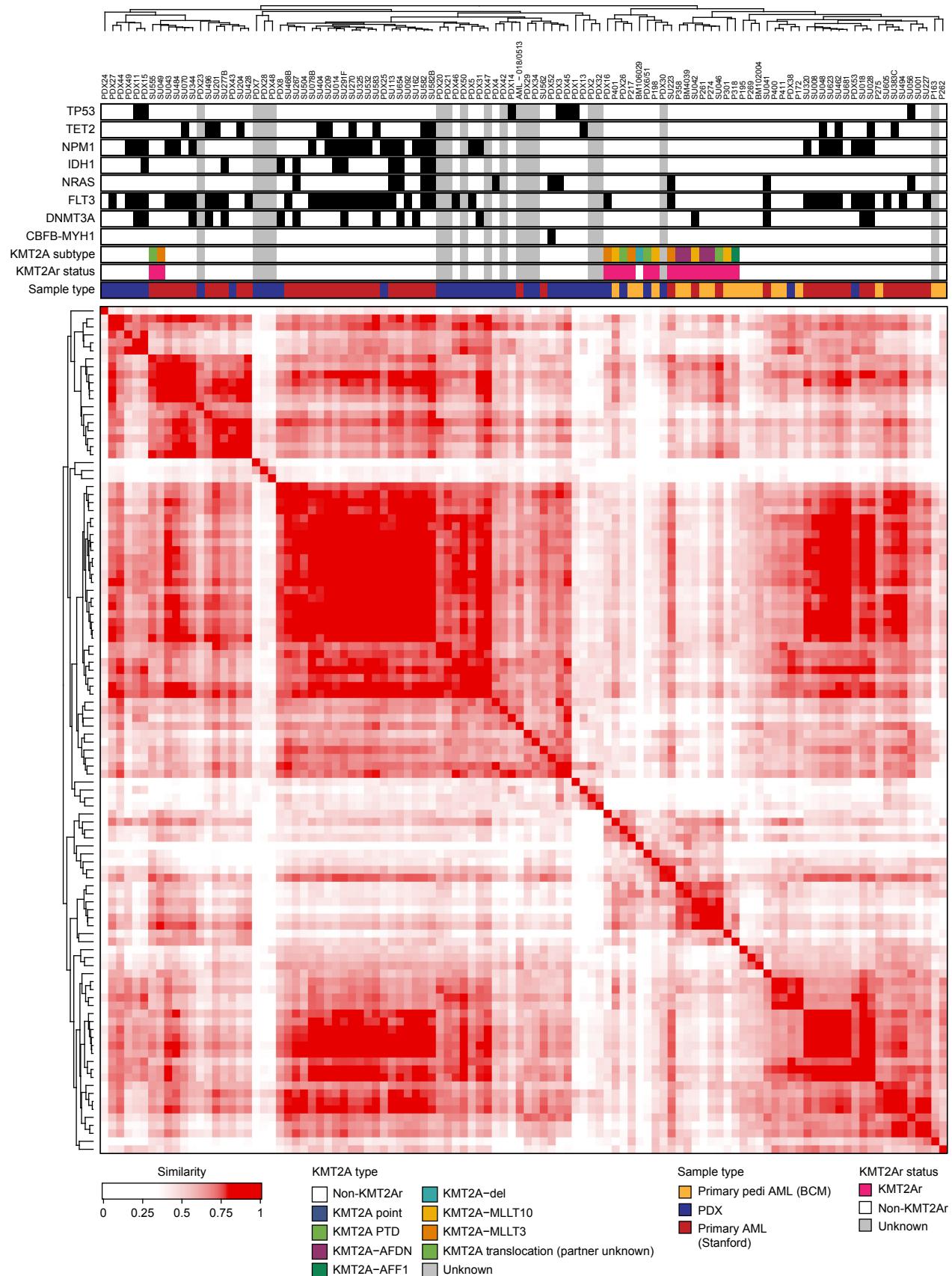


Figure S2, related to Figure 2. A superenhancer-based classification of AML: primary samples and PDX models. The heatmap is a similarity matrix based on AML classification in Figure 2B. Primary and PDX samples are hierarchically clustered with complete linkage using Pearson correlation of the scores of 4798 superenhancers recurrent in at least 2 samples. The bars at the top reflect presence of coding mutations (grey color designates unknown) and sample type.

Figure S3

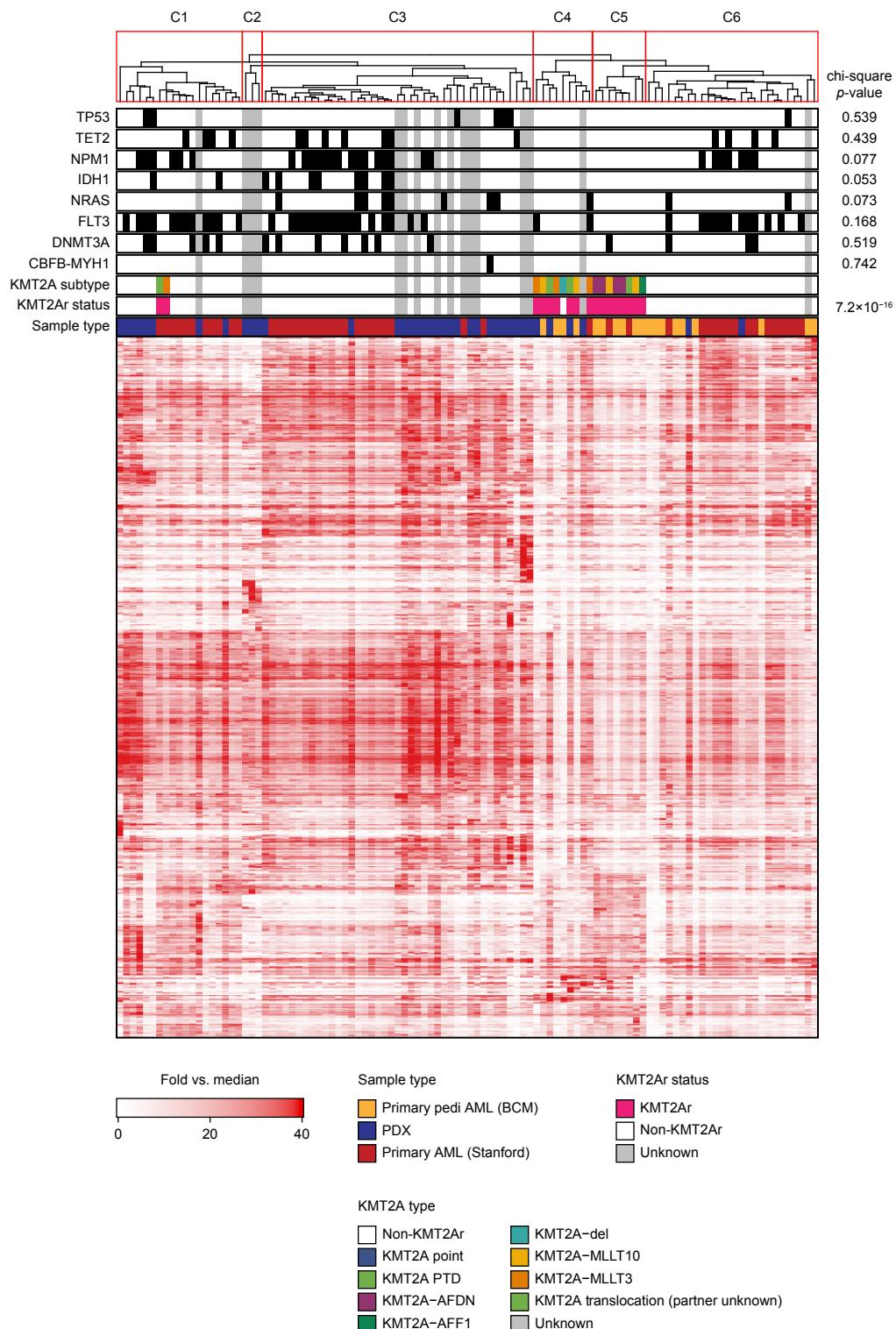


Figure S3, related to Figure 2. A superenhancer-based classification of AML: primary samples and PDX models. The heatmap is a signal matrix (a full version of the AML classification in Figure 2B). Primary and PDX samples are hierarchically clustered with complete linkage using Pearson correlation of the scores of 4798 superenhancers recurrent in at least 2 samples, forming 6 distinct clusters. The bars at the top reflect presence of coding mutations (grey color designates unknown) and sample type. The side plot reflects *p*-values of non-random distribution of somatic mutations across the 6 clusters, calculated by the one-sided chi-square test, demonstrating that KMT2A rearrangements are the only mutation that shows significant enrichment in any cluster ($p<0.05$).

Figure S4

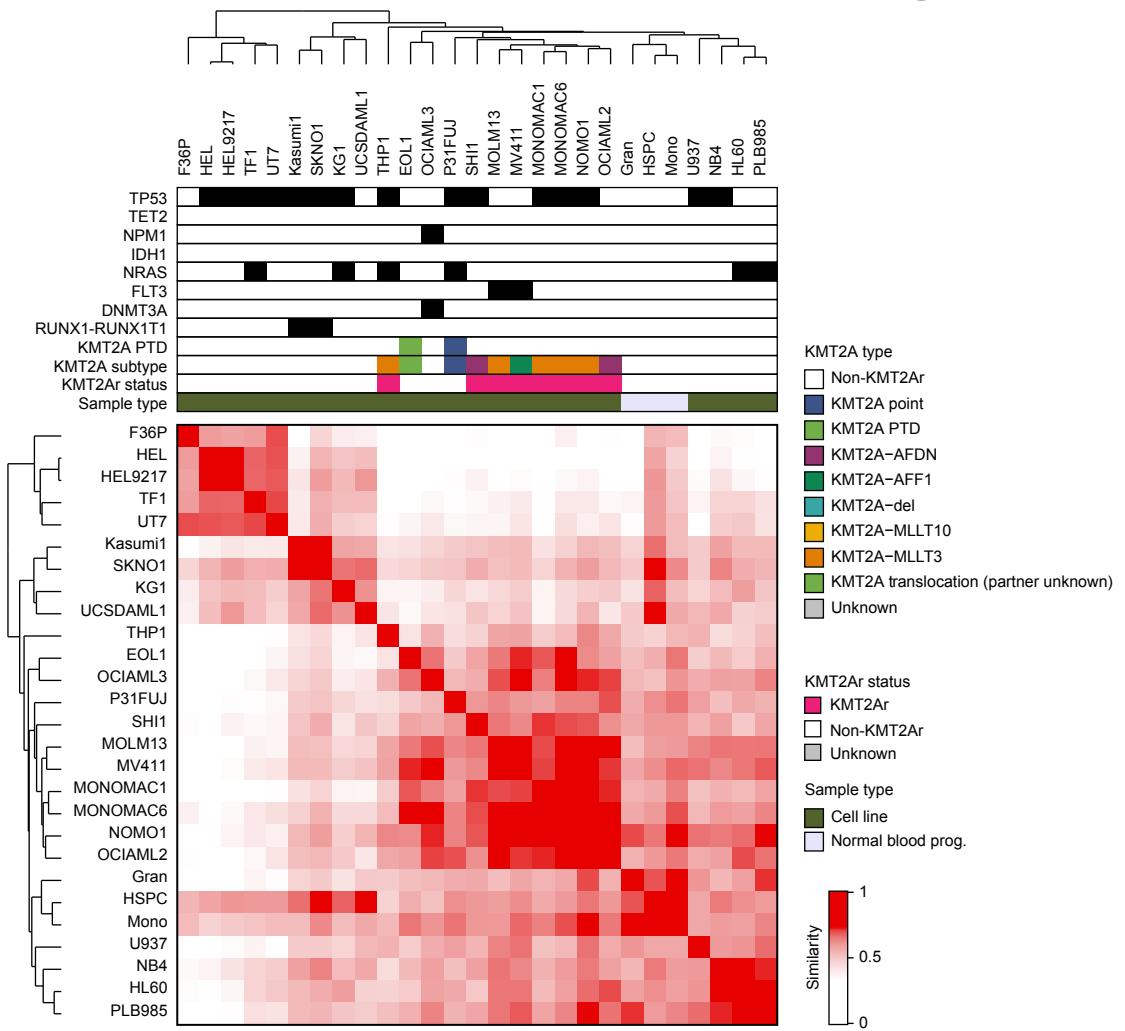
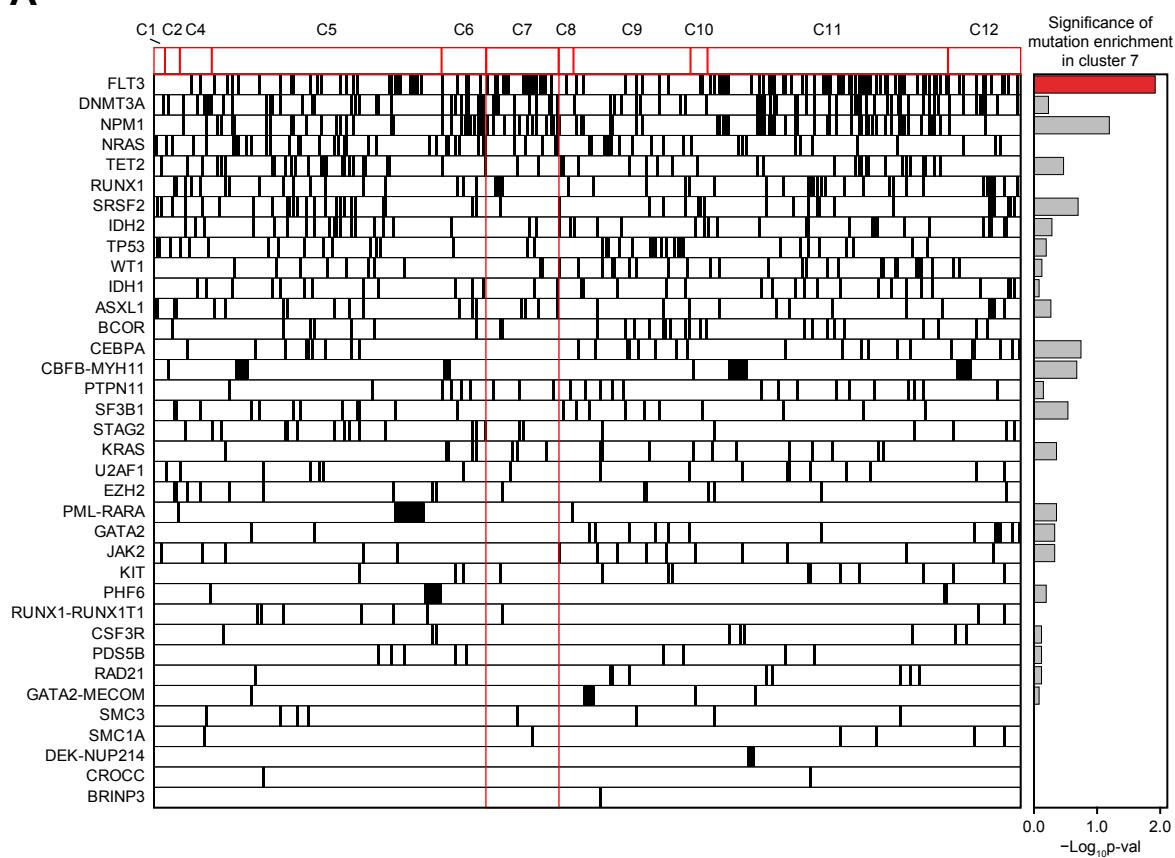


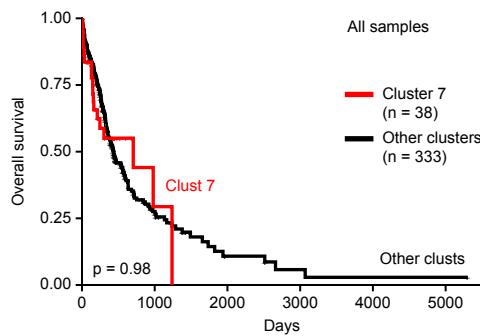
Figure S4, related to Figure 2. A superenhancer-based classification of AML: cell lines. The heatmap is a similarity matrix of the cell lines clustered with complete linkage using Pearson correlation of the scores of 4798 superenhancers recurrent in at least 2 samples. The heatmap reflects pairwise Pearson correlation of superenhancer scores. The bars at the top reflect presence of coding mutations and sample type.

Figure S5

A



B



C

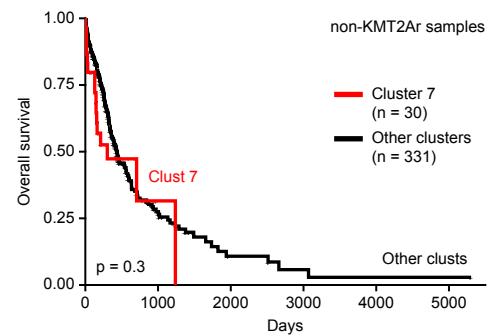


Figure S5, related to Figures 2 and 3. Mutational landscape and survival profile of KMT2Ar-like AML.

- A, Enrichment of somatic mutations in KMT2Ar-like AMLs from the BeatAML cluster 7 (defined by mRNA expression of core regulatory TFs; Figure 3A). The bar plot demonstrates *p*-values calculated by chi-square test of mutation distribution between KMT2Ar-negative samples in cluster 7 (*i.e.* KMT2Ar-like) versus KMT2Ar-negative samples in all other clusters. Only *FLT3* mutations are significantly over-represented in cluster 7, with an enrichment value of 1.7-fold compared to randomly expected distribution. The 38 most common point mutations and translocations from the BeatAML study were included in this analysis and are visualized in the order of decreasing overall frequency. Patients with KMT2Ar AML were excluded from the analysis.
- B, Kaplan-Meyer plot of overall survival of the BeatAML patients (Tyner et al. 2018), demonstrating no significant difference in survival of patients from cluster 7, including KMT2Ar and KMT2Ar-like AML, compared to other clusters.
- C, Kaplan-Meyer plot of overall survival in BeatAML patients (Tyner et al. 2018), demonstrating no significant difference in survival of KMT2Ar-like AML patients from cluster 7 compared to other clusters. KMT2Ar AMLs were excluded from the analysis.

Figure S6

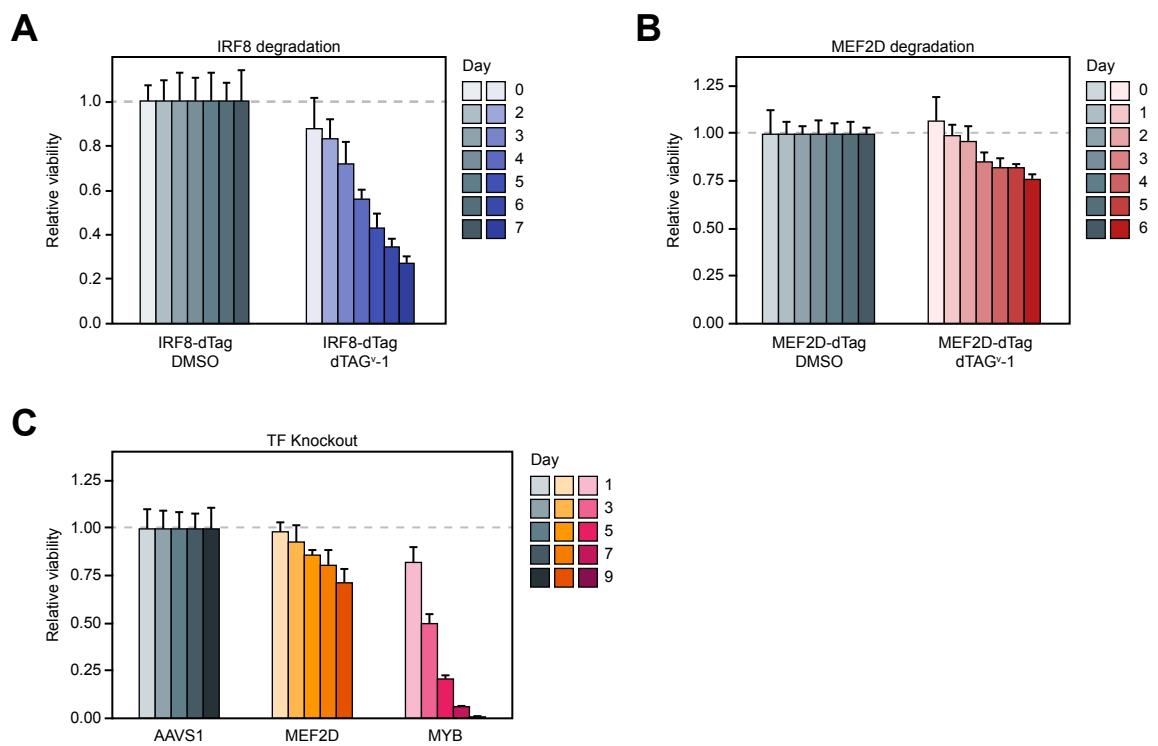


Figure S6, related to Figures 5-7. Application of targeted TF degradation to elucidate direct transcriptional effects of MEF2D and IRF8.

- A, Targeted degradation of IRF8 results in loss of cell viability. Cells carrying a fusion-based IRF8 degron were incubated with DMSO vs. dTAG^V-1 and cell viability was followed by quantification of ATP pools using a luciferin-based assay.
- B, Targeted degradation of MEF2D results in loss of cell viability. The experiment was carried out as in (B).
- C, CRISPR/Cas9 knockout of MEF2D results in a loss of cell viability that is quantitatively similar to the effects of MEF2D degradation. MYB knockout was used as positive control.

Figure S7

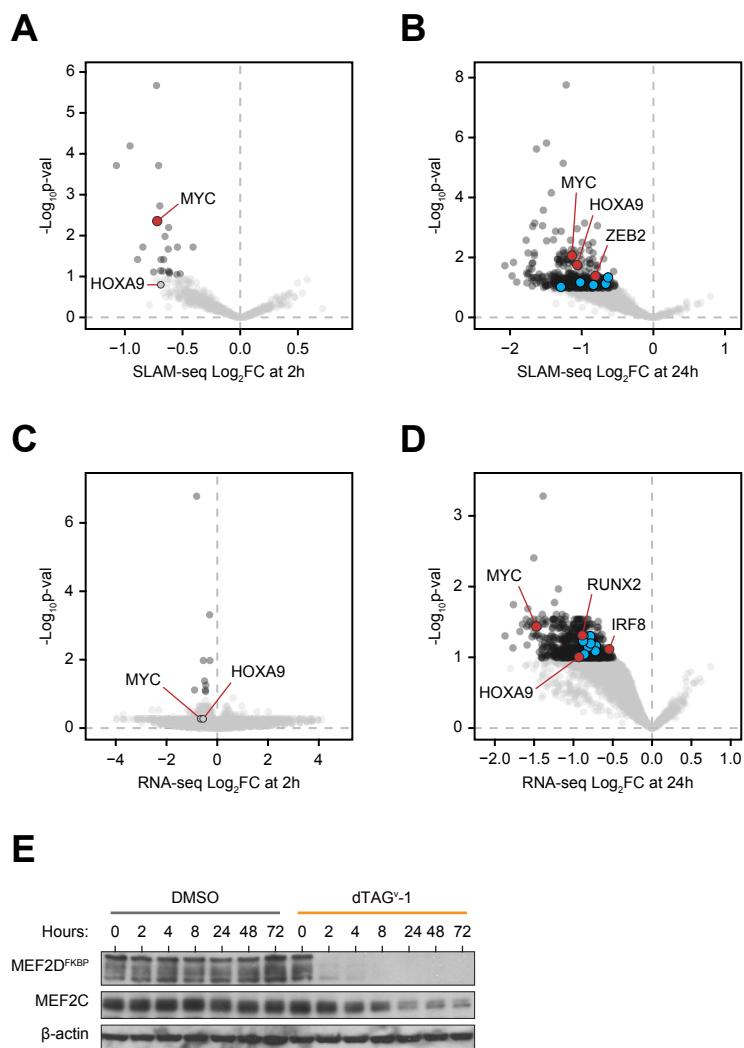


Figure S7, related to Figure 5. Volcano plots of SLAM-seq and mRNA-seq following MEF2D degradation, and regulation of MEF2C by MEF2D. Core regulatory TFs are highlighted in blue.

- A, SLAM-seq 2 hours after MEF2D degradation.
- B, SLAM-seq 24 hours after MEF2D degradation.
- C, mRNA-seq 2 hours after MEF2D degradation.
- D, mRNA-seq 24 hours after MEF2D degradation.
- E, Western blot showing reduced MEF2C levels after prolonged MEF2D degradation.

Figure S8

A

Gene ID			2 hrs SLAM-seq				24 hrs SLAM-seq				2 hrs RNA-seq				24 hrs RNA-seq			
	MV411 dep. probability		Pan-lethal	SE in MV411		log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	
	MV411	drop out score		TF	TF													
FUT4	0.009	-9E-04	FALSE	1	-0.725	2.6E-09	2.1E-06	-0.933	0.002	0.043	-0.452	2.9E-04	0.232	-0.757	0.004	0.089		
ZFP36	0.008	0.012	FALSE	1	-0.955	1.6E-07	6.4E-05	-1.630	3.2E-09	2.4E-06	-0.659	0.038	0.534	-1.477	0.025	0.124		
BHLHE40	0.001	0.141	FALSE	1	-0.707	8.3E-07	1.9E-04	-1.035	0.006	0.062	-0.500	0.018	0.534	-1.019	0.002	0.072		
PTGER4	0.001	0.143	FALSE	1	-1.073	9.5E-07	1.9E-04	-1.128	0.009	0.069	-0.944	0.003	0.446	-1.086	0.003	0.082		
KLF10	0.002	0.101	FALSE	1	-0.697	1.2E-05	0.002	-0.697	0.048	0.143	-0.545	0.009	0.521	-0.757	0.073	0.167		
MYC	1.000	-1.935	TRUE	1	-0.720	3.3E-05	0.004	-1.136	1.4E-04	0.009	-0.655	0.075	0.534	-1.470	2.7E-04	0.037		
F3	0.141	-0.259	FALSE		-0.620	5.5E-05	0.006	-0.828	0.010	0.070	-0.541	0.064	0.534	-0.953	0.002	0.076		
TNFAIP3	0.003	0.093	FALSE		-0.652	1.0E-04	0.010	-0.681	0.093	0.209	-0.520	0.006	0.503	-1.051	4.9E-04	0.046		
TXNIP	0.438	-0.449	FALSE		-0.544	2.4E-04	0.019	-1.039	6.0E-04	0.019	-0.468	0.058	0.534	-0.960	0.001	0.058		
PMAIP1	4E-04	0.252	FALSE	1	-0.844	2.3E-04	0.019	-0.962	0.003	0.046	-0.785	8.5E-04	0.299	-1.277	0.002	0.065		
PIM1	0.001	0.138	FALSE	1	-0.409	2.6E-04	0.019	-0.813	0.003	0.047	-0.455	0.008	0.521	-0.895	0.001	0.055		
IER2	0.003	0.072	FALSE	1	-0.624	3.2E-04	0.021	-1.219	8.0E-12	1.8E-08	-0.459	4.7E-05	0.086	-1.275	5.6E-04	0.047		
ID2	0.204	-0.309	FALSE	1	-0.890	6.1E-04	0.038	-1.545	3.5E-06	8.4E-04	-0.905	0.024	0.534	-1.212	0.007	0.097		
TP53RK	0.908	-0.843	TRUE	1	-0.663	6.6E-04	0.038	-0.880	0.017	0.090	-0.478	1.1E-04	0.164	-0.799	0.002	0.072		
KLF6	5E-04	0.222	FALSE	1	-0.687	7.1E-04	0.039	-1.656	2.4E-06	6.9E-04	-0.590	0.035	0.534	-1.293	6.7E-04	0.049		
KLF11	0.109	-0.228	FALSE	1	-0.691	1.4E-03	0.072	-1.325	0.003	0.046	-0.706	0.006	0.498	-0.975	8.4E-04	0.052		
CDKN1B	0.006	0.035	FALSE	1	-0.671	1.6E-03	0.074	-1.308	2.1E-04	0.011	-0.751	0.062	0.534	-1.478	6.3E-04	0.049		
CDC42EP3	0.012	-0.023	FALSE	1	-0.622	1.6E-03	0.074	-0.980	0.002	0.041	-0.584	0.008	0.521	-1.041	5.3E-04	0.047		
ZBTB33	0.032	-0.103	FALSE	1	-0.751	1.8E-03	0.078	-1.280	0.009	0.069	-0.637	0.041	0.534	-0.945	0.147	0.255		
RANBP6	0.038	-0.120	FALSE		-0.612	1.9E-03	0.079	-0.990	0.027	0.113	-0.477	0.011	0.534	-0.705	0.035	0.129		
CPEB2	2E-04	0.293	FALSE	1	-0.615	2.2E-03	0.085	-1.259	5.8E-05	0.005	-0.890	0.002	0.376	-1.474	1.7E-04	0.033		
SATB2	0.063	-0.168	FALSE	1	-0.519	2.3E-03	0.086	-1.119	0.007	0.062	-0.369	2.9E-04	0.232	-0.766	0.007	0.099		
DUSP6	0.005	0.037	FALSE	1	-0.550	2.5E-03	0.090	-0.914	0.005	0.056	-0.695	0.055	0.534	-1.520	1.5E-04	0.033		

B

Gene ID			2 hrs SLAM-seq				24 hrs SLAM-seq				2 hrs RNA-seq				24 hrs RNA-seq			
	MV411 dep. probability		Pan-lethal	SE in MV411		log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	
	MV411	drop out score		TF	TF													
MYC	1.000	-1.935	TRUE	1	-0.720	3.3E-05	0.004	-1.136	1.4E-04	0.009	-0.655	0.075	0.534	-1.470	2.7E-04	0.037		
LMO2	0.013	-0.032	FALSE	1	-0.446	0.006	0.138	-0.770	0.037	0.128	-0.171	0.001	0.334	-0.479	0.017	0.116		
ZEB2	0.931	-0.914	FALSE	1	-0.425	0.008	0.152	-0.809	0.002	0.041	-0.322	0.010	0.527	-0.877	0.001	0.059		
HOXA9	0.305	-0.374	FALSE	1	-0.677	0.011	0.172	-1.063	5.5E-04	0.018	-0.539	0.043	0.534	-0.930	0.008	0.099		
MYB	0.999	-1.468	FALSE	1	-0.356	0.047	0.307	-1.020	0.008	0.068	-0.440	0.056	0.534	-0.782	0.001	0.064		
IRF8	0.946	-0.984	FALSE	1	-0.279	0.091	0.376	-0.837	0.014	0.083	-0.019	0.803	0.929	-0.546	0.003	0.077		
RUNX1	0.167	-0.281	FALSE	1	-0.195	0.152	0.480	-0.671	0.041	0.134	-0.487	0.004	0.450	-0.861	0.005	0.091		
E2F3	0.200	-0.306	FALSE	1	-0.234	0.176	0.499	-0.498	0.098	0.213	-0.444	0.001	0.299	-0.840	0.002	0.065		
MEIS1	0.376	-0.414	FALSE	1	-0.274	0.184	0.513	-0.865	0.037	0.128	-0.184	0.222	0.574	-0.757	0.010	0.106		
ETV6	1E-04	0.378	FALSE	1	-0.160	0.296	0.614	-0.569	0.084	0.197	-0.029	0.767	0.916	-0.563	0.011	0.108		
GFI1	0.916	-0.864	FALSE	1	-0.123	0.288	0.614	-0.665	0.011	0.076	-0.172	0.036	0.534	-0.714	0.002	0.069		
RUNX2	0.132	-0.250	FALSE	1	-0.151	0.312	0.614	-0.810	0.027	0.114	-0.393	0.005	0.492	-0.887	6.4E-04	0.049		
PLAGL2	0.316	-0.380	FALSE	1	-0.149	0.383	0.665	-0.444	0.094	0.209	-0.161	0.009	0.521	-0.720	0.003	0.082		
SP1	0.864	-0.764	FALSE	1	-0.156	0.395	0.672	-0.653	0.038	0.130	-0.127	0.164	0.544	-0.624	0.024	0.123		
MEF2C	0.523	-0.496	FALSE	1	-0.205	0.428	0.693	-0.835	0.037	0.130	-0.369	0.045	0.534	-0.750	0.020	0.120		
CEBPA	0.485	-0.475	FALSE	1	-0.127	0.484	0.718	-0.293	0.137	0.257	0.103	0.377	0.687	-0.389	0.073	0.167		
RXRA	0.582	-0.530	FALSE	1	0.099	0.555	0.761	-0.371	0.248	0.366	0.104	0.249	0.597	-0.300	0.225	0.344		
ZMYND8	0.920	-0.875	FALSE	1	-0.059	0.738	0.884	-0.629	0.041	0.134	-0.058	0.754	0.915	-0.786	7.6E-04	0.050		
MEF2D	0.877	-0.784	FALSE	1	-0.018	0.913	0.962	-0.632	0.003	0.046	0.021	0.823	0.939	-0.444	0.060	0.153		
MAX	0.395	-0.425	TRUE	1	0.179	0.460		-0.708	0.074	0.183	-0.026	0.738	0.906	-0.471	0.033	0.128		
FLI1	0.318	-0.381	FALSE	1	-0.167	0.537		-1.290	0.019	0.096	-0.289	0.060	0.534	-0.571	0.047	0.139		
SP11	0.943	-0.967	FALSE	1	-0.182	0.603		-0.038	0.938	0.951	-0.236	0.050	0.534	-0.286	0.198	0.314		
STAT5B	0.898	-0.820	FALSE	1	0.572	0.143		-0.287	0.661		0.110	0.261	0.604	-0.306	0.114	0.215		
SREBF1	0.898	-0.820	FALSE	1	0.657	0.280		-0.133	0.813		0.243	0.180	0.549	-0.131	0.561	0.667		
TFAP4	0.941	-0.959	FALSE	1	0.089	0.833		-0.636	0.473		-0.064	0.587	0.824	-0.624	0.018	0.116		
ZNF281	0.074	-0.185	FALSE	1	-0.160	0.638		-0.746	0.099	0.214	-0.557	0.146	0.538	-1.042	0.021	0.121		
LYL1	0.142	-0.260	FALSE	1	0.401	0.437		0.018	0.966	0.974	0.243	0.081	0.534	-0.197	0.425	0.543		
FOSL2	0.400	-0.428	FALSE	1														

Figure S8, related to Figure 5. A table summary of SLAM-seq and RNA-seq data following MEF2D degradation

- A, Genes demonstrating a statistically significant change in transcription rate (SLAM-seq adjusted *p*-value <0.1) at 2 hours following MEF2D degradation.
- B, Changes in core regulatory TF transcription after MEF2D degradation.

Figure S9

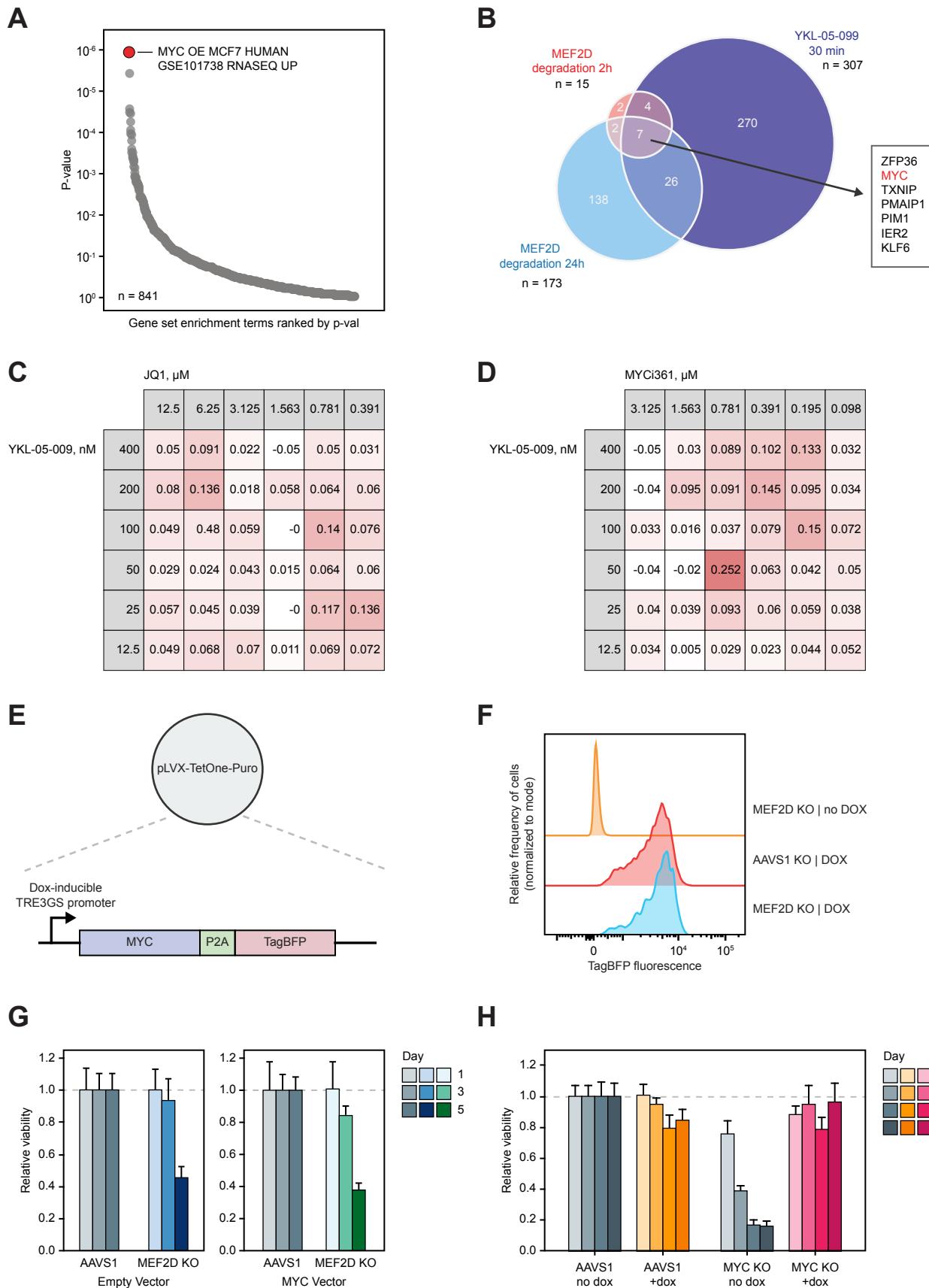


Figure S9, related to Figure 5. A functional interaction between MYC and MEF2D

- A, Gene set enrichment analysis of the genes showing significant changes in transcription rates after MEF2D degradation using Ehrichr, filtered to display only human-derived data (Kuleshov et al. 2016).
- B, Intersection of gene sets demonstrating significant changes in transcription (adjusted *p*-value <0.05) measured by SLAM-seq after MEF2D degradation and indirect MEF2C/D inhibition by YKL-05-099 treatment.
- C, Excess over bliss synergy matrix demonstrating synergy between SIK/MEF2 inhibitor YKL-05-099 and bromodomain inhibitor JQ1. Red color demonstrates synergy.
- D, Excess over bliss synergy matrix demonstrating synergy between SIK/MEF2 inhibitor YKL-05-099 and MYC inhibitor MYCi361. Red color demonstrates synergy.
- E, Cloning strategy for a doxycycline-inducible lentiviral vector expressing MYC-P2A-TagBFP.
- F, Induction of MYC expression demonstrated by TagBFP fluorescence after addition of doxycycline.
- G, Forced exogenous expression of MYC fails to rescue the cell viability loss following MEF2D degradation.
- H, As a positive control, forced exogenous expression of MYC completely rescues MYC knockout by CRISPR/Cas9. Exogenous MYC expression was induced by doxycycline.

Figure S10

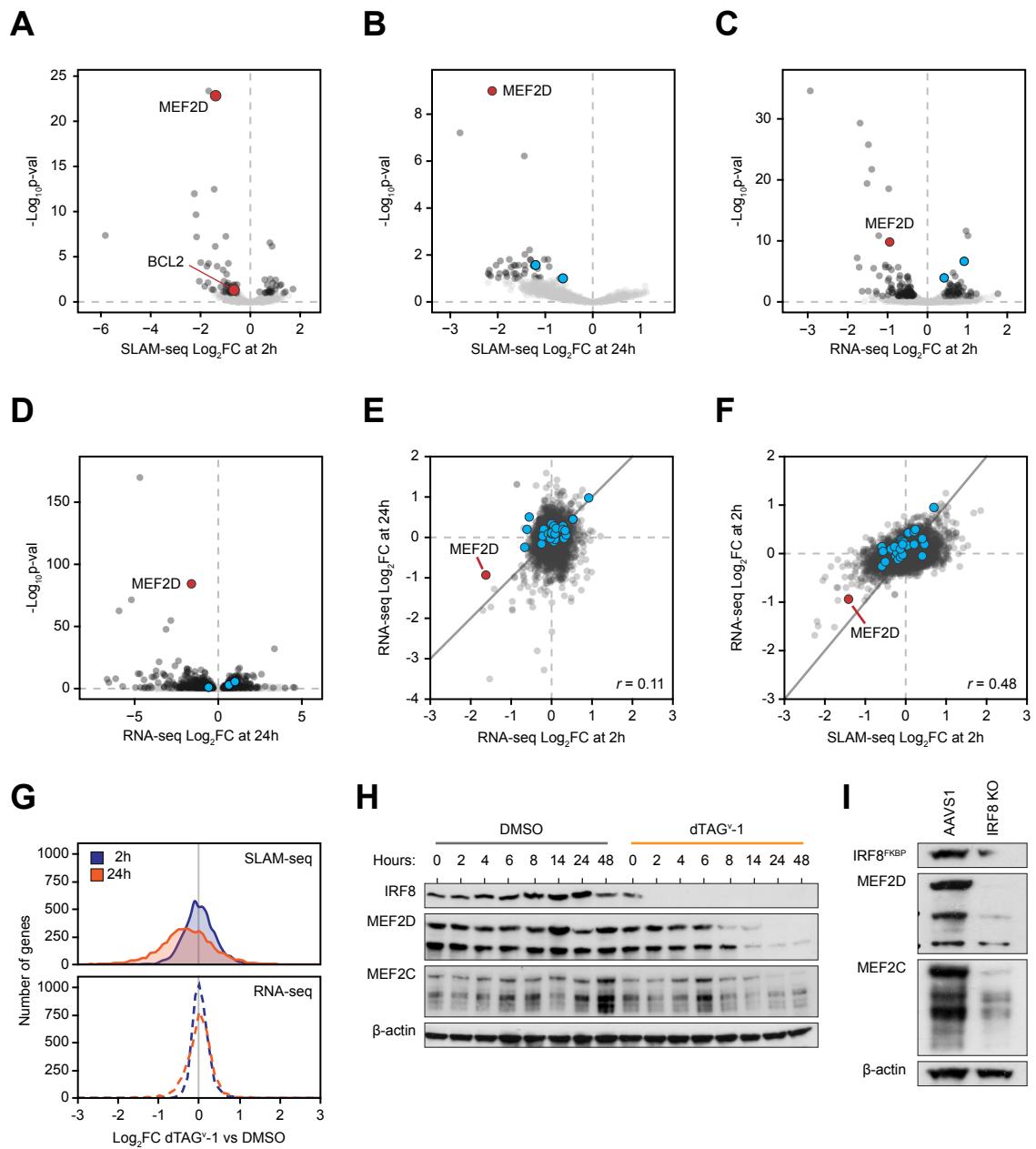


Figure S10, related to Figure 7. Targeted degradation of IRF8

- A, Volcano plot of SLAM-seq 2 hours after IRF8 degradation.
- B, Volcano plot of SLAM-seq 24 hours after IRF8 degradation. Core regulatory TFs are highlighted in blue.
- C, Volcano plot of RNA-seq 2 hours after IRF8 degradation. Core regulatory TFs are highlighted in blue.
- D, Volcano plot of RNA-seq 24 hours after IRF8 degradation. Core regulatory TFs are highlighted in blue.
- E, A cross-plot of genome-wide changes in mRNA pools measured after 2 vs. 24 hours of IRF8 degradation demonstrates a poor correlation between early and late transcriptional response.
- F, A cross-plot of genome-wide changes in mRNA pools (mRNA-seq) vs. transcription rates (SLAM-seq) measured after 2 hours of IRF8 degradation.
- G, A distribution plot of genome-wide changes in nascent transcription rates (SLAM-seq) and mRNA pools (RNA-seq) after 2 and 24 hours of IRF8 degradation.
- H, Western blot showing reduced MEF2C and MEF2D levels after IRF8 degradation.
- I, Western blot showing reduced MEF2C and MEF2D levels 24 hours after IRF8 knockout.

Figure S11

Gene ID			2 hrs SLAM-seq			24 hrs SLAM-seq			2 hrs RNA-seq			24 hrs RNA-seq		
	MV411 dep. probability	MV411 drop out score	log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pval	padj	log ₂ FC	pval	padj
CXorf21	0.027	-0.089	-1.672	9.9E-28	4.4E-24	-2.795	6.0E-11	6.6E-08	-0.971	1.4E-22	2.9E-19	-1.824	1.8E-04	0.009
MEF2D	0.877	-0.784	-1.393	6.7E-27	1.5E-23	-2.118	4.6E-13	1.0E-09	-0.946	1.2E-13	1.5E-10	-1.603	6.8E-89	5.0E-85
FAM107B	4E-04	0.237	-1.453	2.2E-16	3.2E-13	-1.241	0.002	0.113	-0.690	8.4E-06	0.003	-0.771	0.028	0.296
CCR2	0.268	-0.351	-2.247	8.7E-16	9.6E-13	-3.651	0.001		-1.684	8.2E-34	5.0E-30	-1.842	2.0E-13	6.3E-11
DCANP1	0.187	-0.296	-2.179	2.2E-13	1.9E-10	-2.188	0.003		-1.394	6.2E-26	1.9E-22	-1.295	2.7E-10	4.8E-08
KCNA3	0.006	0.033	-5.798	5.2E-11	3.8E-08	-5.960	1.8E-06		-2.926	2.1E-39	2.6E-35	-5.905	3.3E-67	1.2E-63
CBX6	0.007	0.017	-0.989	7.7E-11	4.9E-08	-1.325	6.2E-05	0.015	-0.378	0.002	0.145	-0.971	0.005	0.111
TIFAB	0.007	0.016	-2.156	1.0E-10	5.6E-08	-4.186	0.003		-1.478	4.2E-30	1.7E-26	-1.755	1.6E-19	1.6E-16
LDLR	0.579	-0.528	0.770	5.0E-10	2.5E-07	-0.116	0.617	0.859	0.957	1.2E-15	2.1E-12	0.132	0.351	0.800
MALAT1			0.856	1.4E-09	6.0E-07	-0.220	0.432	0.760	0.562	0.018	0.404	-0.296	0.477	0.855
CCR1	0.019	-0.061	-1.410	1.6E-09	6.3E-07	-1.636	2.3E-04	0.028	-0.902	7.6E-09	4.9E-06	-1.001	0.001	0.042
CPM	0.007	0.020	-1.991	1.0E-07	3.8E-05	-1.815	1.6E-02		-0.494	1.3E-05	3.4E-03	-1.606	7.2E-16	4.1E-13
IKZF1	0.434	-0.447	-0.889	1.4E-07	4.8E-05	-0.977	0.008	0.175	-0.532	3.9E-04	0.045	-0.301	0.261	0.735
SERPINB8	0.006	0.029	-1.678	1.6E-07	4.9E-05	-1.501	0.045		-0.693	1.2E-04	0.019	-1.033	0.026	0.286
POU2F2	8E-05	0.393	-1.745	3.6E-07	1.0E-04	-2.154	0.001	0.076	-1.510	1.6E-23	3.9E-20	-3.523	5.7E-26	9.2E-23
ABHD15	0.069	-0.178	-1.336	3.6E-07	1.0E-04	-1.321	0.043		-0.494	2.4E-07	1.1E-04	-0.628	2.5E-05	0.002
IL6R	0.021	-0.067	-1.098	9.6E-07	2.5E-04	-0.493	0.352	0.717	-0.517	1.4E-07	7.0E-05	-0.457	0.056	0.407
TMEM127	0.698	-0.605	-0.955	3.0E-06	7.4E-04	-1.121	0.029	0.314	-0.345	8.0E-04	0.077	-0.140	0.360	0.804
TRIB1	0.005	0.045	0.783	1.1E-05	0.003	-0.084	0.703	0.897	0.687	1.1E-07	5.7E-05	-0.075	0.750	0.947
SCD	0.919	-0.871	0.566	1.4E-05	0.003	-0.344	0.289	0.665	0.040	0.691	0.912	0.256	0.119	0.561
HMGCS1	0.965	-1.096	1.003	1.5E-05	0.003	-0.891	0.033	0.335	0.784	1.1E-05	0.003	-0.378	0.074	0.462
OPN3	0.002	0.128	-2.046	2.3E-05	0.004	-2.730	0.034		-0.561	1.9E-04	0.027	-0.488	0.103	0.531
ADGRG5	0.016	-0.044	-1.060	2.3E-05	0.004	-1.347	0.104		-0.579	2.6E-04	0.034	-0.288	0.008	0.148
CSF1R	0.229	-0.326	-1.702	3.5E-05	0.006	-1.519	0.109		-0.705	3.1E-07	1.3E-04	-0.881	0.009	0.158
MSMO1	0.002	0.110	0.887	4.0E-05	0.007	-1.452	0.008	0.187	1.004	9.2E-15	1.3E-11	0.107	0.278	0.748
TGFBR1	0.003	0.078	-0.899	4.2E-05	0.007	-1.727	3.1E-04	0.032	-0.332	0.079	0.649	-0.718	0.060	0.420
P2RY2	0.006	0.025	-0.538	6.6E-05	0.010	-0.101	0.723	0.904	-0.429	1.6E-04	0.024	0.068	0.643	0.916
DDIT3	0.160	-0.275	1.216	6.5E-05	0.010	-0.146	0.678	0.884	0.330	0.028	0.476	-0.153	0.452	0.845
ZBTB33	0.032	-0.103	-0.944	7.6E-05	0.011	-1.126	0.004	0.149	-0.902	3.4E-05	0.008	-1.022	0.005	0.112
B3GNT7	0.004	0.056	-0.684	7.6E-05	0.011	-0.416	0.270	0.646	-0.634	2.7E-06	9.7E-04	-0.229	0.008	0.140
CALR	0.040	-0.125	0.663	7.8E-05	0.011	0.583	0.096	0.477	0.093	0.447	0.820	0.623	2.6E-04	0.011
DUSP7	0.004	0.056	-0.633	8.4E-05	0.012	-0.518	0.326	0.694	-0.138	0.222	0.721	-0.656	0.058	0.417
INHBA	0.012	-0.023	-0.939	9.7E-05	0.013	-1.763	5.9E-05	0.015	-0.277	0.083	0.650	-1.321	2.1E-09	3.2E-07
JUN	0.374	-0.413	1.026	1.1E-04	0.013	0.314	0.272	0.646	0.765	2.4E-05	0.005	-0.152	0.584	0.895
284454			1.170	1.0E-04	0.013	0.681	0.023	0.298	0.639	0.042	0.548	0.039	0.871	0.979
SH2B3	0.002	0.107	-0.791	1.3E-04	0.016	-1.384	0.124	0.503	-0.461	3.6E-04	0.042	-0.374	0.274	0.746
LYZ	0.041	-0.126	-1.829	1.4E-04	0.017	-0.611	0.503		-0.275	0.044	0.556	0.546	0.135	0.588
TIFA	0.003	0.093	-1.191	1.6E-04	0.019	-1.692	0.012	0.210	-0.653	3.6E-04	0.042	-1.095	0.003	0.076
ZNF616	4E-04	0.249	-0.983	1.7E-04	0.019	-0.824	0.450		-0.213	0.223	0.721	0.232	0.363	0.806
RNF41	0.193	-0.301	-0.678	1.8E-04	0.020	-0.898	0.050	0.368	-0.183	0.074	0.644	-0.223	0.028	0.293
EGR1	0.007	0.017	0.837	2.9E-04	0.031	-0.124	0.578	0.845	0.606	8.9E-05	0.017	-0.342	0.240	0.715
HELQ	0.005	0.039	-0.792	3.1E-04	0.032	-0.613	0.173	0.564	-0.351	0.012	0.344	-0.180	0.562	0.887
BIN3-IT1			1.691	3.2E-04	0.033	0.252	0.652	0.872	0.828	0.025	0.453	-0.098	0.817	0.965
MBP	0.002	0.112	-0.650	3.5E-04	0.035	-0.755	0.010	0.205	-0.385	0.015	0.383	-0.184	0.377	0.813
ZNF765	3E-04	0.288	-1.544	3.6E-04	0.035	-0.895	0.492		-0.265	0.250	0.736	0.067	0.889	0.982
KLF2	0.002	0.121	0.989	3.7E-04	0.035	0.307	0.239	0.625	0.833	6.8E-05	0.014	-0.585	0.023	0.266
HES6	0.010	-0.009	0.760	3.9E-04	0.037	0.523	0.199	0.580	0.281	0.187	0.703	0.065	0.845	0.972
RHOU	0.030	-0.099	-1.481	4.3E-04	0.039	0.229	0.771		-0.475	0.003	0.188	0.131	0.627	0.911
TMED10	0.483	-0.473	-0.780	4.7E-04	0.042	-0.838	0.120	0.497	-0.254	0.093	0.653	-0.215	0.439	0.839
BCL2	0.576	-0.526	-0.649	5.4E-04	0.048	-1.205	0.005	0.159	-0.340	0.038	0.534	-0.713	3.2E-07	3.4E-05
FLVCR2	0.034	-0.110	-0.988	5.7E-04	0.048	-1.404	0.012	0.211	-0.418	1.2E-04	0.019	-0.426	9.7E-05	0.005
CRKL	0.902	-0.830	-0.611	5.6E-04	0.048	-0.039	0.967	0.986	-0.155	0.240	0.732	0.192	0.614	0.906
HNRNPA3	0.138	-0.256	0.733	6.3E-04	0.053	0.223	0.478	0.791	-0.119	0.464	0.825	-0.133	0.604	0.902
LRRC8C	0.041	-0.128	-0.817	6.8E-04	0.055	-2.041	1.8E-04	0.027	-0.366	0.060	0.611	-1.075	0.015	0.208

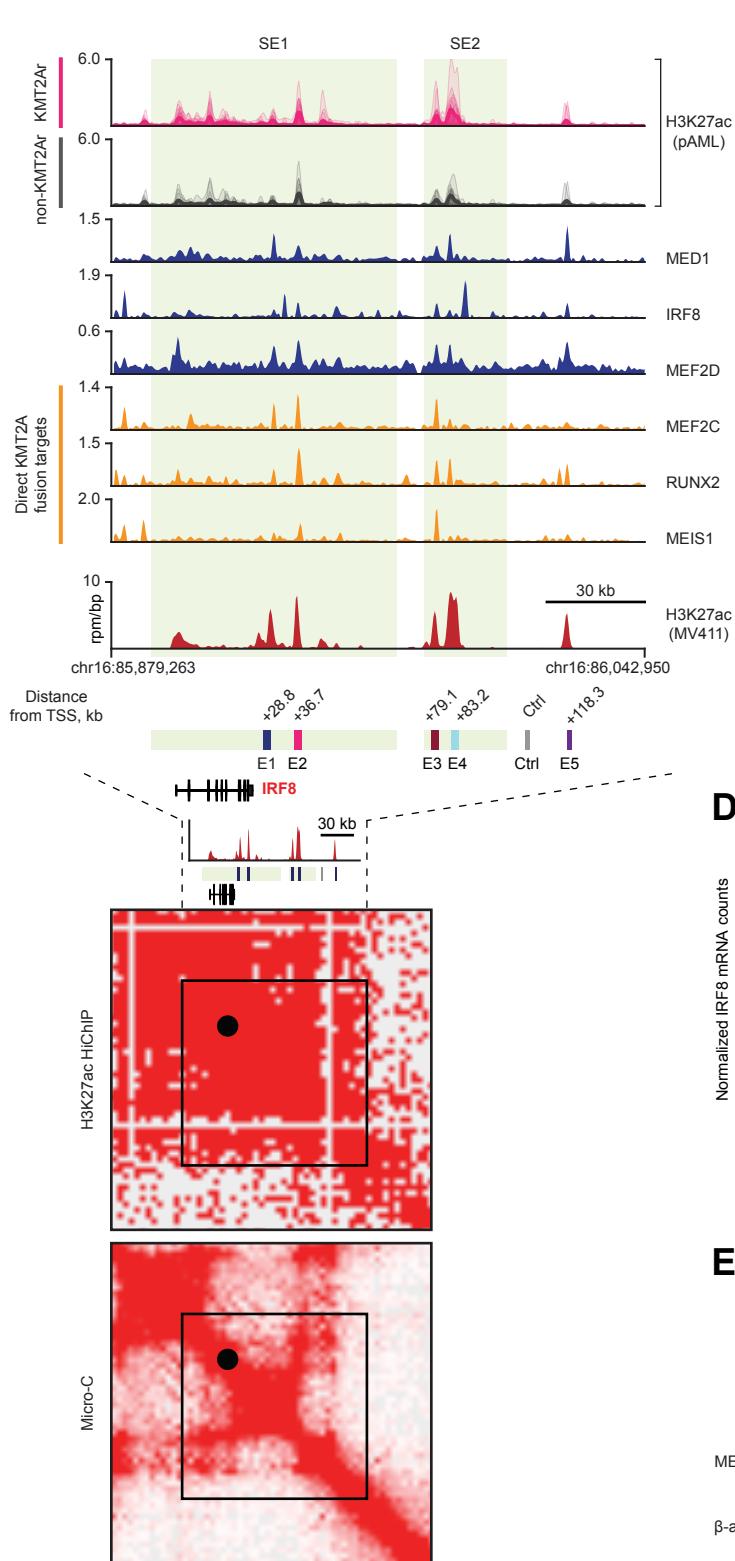
Figure S11 cont.

Gene ID	MV411 dep. probability	MV411 drop out score	2 hrs SLAM-seq			24 hrs SLAM-seq			2 hrs RNA-seq			24 hrs RNA-seq		
			log ₂ FC	pvalue	padj	log ₂ FC	pvalue	padj	log ₂ FC	pval	padj	log ₂ FC	pval	padj
LINC00599			0.849	7.3E-04	0.059	0.435	0.169	0.562	0.725	0.212	0.715	0.685	0.010	0.167
NLRP3	0.002	0.130	-1.080	8.1E-04	0.063	-1.668	1.8E-04	0.027	-0.688	7.2E-04	0.071	-0.959	1.4E-08	2.0E-06
ZNF189	5E-04	0.229	-0.858	8.2E-04	0.063	-1.239	0.036	0.347	-0.349	0.056	0.603	0.110	0.787	0.960
GLUL	0.002	0.122	0.357	9.0E-04	0.066	0.521	0.058	0.383	0.141	0.191	0.708	1.087	1.1E-05	8.1E-04
FADS1	0.006	0.034	0.710	9.1E-04	0.066	-0.743	0.145	0.531	-0.093	0.419	0.809	0.232	0.022	0.261
MIR9-3HG			0.780	9.1E-04	0.066	-0.337	0.355	0.720	0.125	0.467	0.826	0.078	0.607	0.903
GORASP1	0.091	-0.207	1.392	8.9E-04	0.066	0.727	0.366		0.303	0.040	0.542	0.998	2.1E-11	4.9E-09
ZNF766	0.001	0.215	-0.591	9.6E-04	0.068	-0.855	0.020	0.270	0.040	0.735	0.925	0.120	0.454	0.846
TMEM70	0.087	-0.203	-0.996	0.001	0.072	-1.093	0.029	0.312	-0.213	0.248	0.735	-0.138	0.544	0.880
GNA13	0.003	0.073	-0.838	0.001	0.072	-1.282	0.011	0.208	-0.540	0.008	0.291	-0.629	0.242	0.717
KCNQ1OT1			0.770	0.001	0.073	0.088	0.823	0.946	0.808	0.003	0.193	-0.002	0.994	0.999
RGS16	0.003	0.080	0.657	0.001	0.078	-0.996	0.005	0.149	0.694	0.001	0.100	-0.649	0.123	0.570
INSIG1	0.005	0.046	0.584	0.001	0.079	-0.431	0.227	0.609	0.705	0.000	0.004	0.100	0.727	0.940
RCSD1	0.060	-0.164	-0.778	0.001	0.080	-0.990	0.027	0.310	-0.279	0.018	0.405	-0.022	0.923	0.987
HNRNPA2B1	0.314	-0.379	0.585	0.001	0.086	0.488	0.189	0.571	-0.034	0.775	0.938	0.109	0.420	0.832
CXCL8	0.028	-0.094	0.819	0.001	0.086	-0.134	0.722	0.904	0.898	1.6E-05	0.004	0.062	0.902	0.984
DMXL2	0.091	-0.207	1.322	0.001	0.090	0.189	0.723	0.904	-0.009	0.952	0.990	0.547	0.222	0.699
BNIP2	0.060	-0.164	-0.792	0.002	0.093	-0.811	0.025	0.298	-0.523	0.015	0.388	-0.325	0.350	0.799
RAB11FIP1	3E-04	0.279	-0.560	0.002	0.097	-0.832	0.002	0.113	-0.352	4.7E-04	0.052	-0.210	0.221	0.697

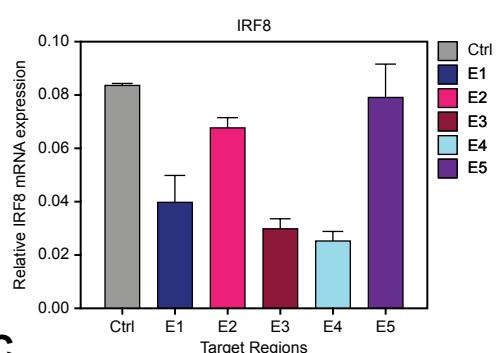
Figure S11, related to Figure 7. A table summary of SLAM-seq and RNA-seq data following IRF8 degradation. Genes demonstrating a statistically significant change in transcription rate (SLAM-seq adjusted p-value <0.1) at 2 hours following IRF8 degradation are presented.

Figure S12

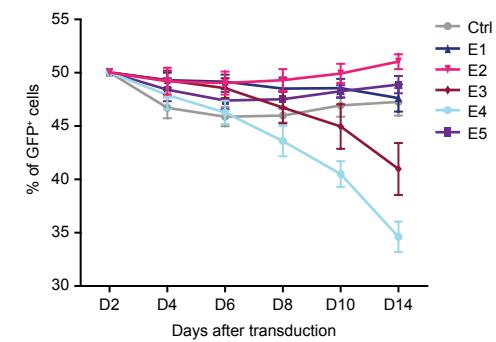
A



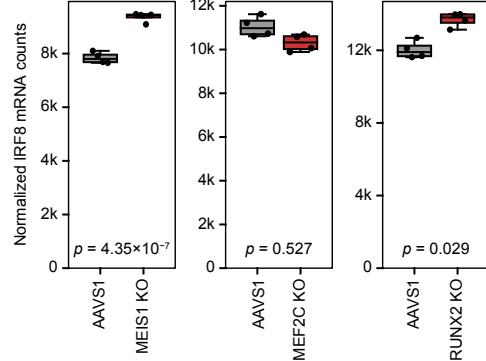
B



C



D



E

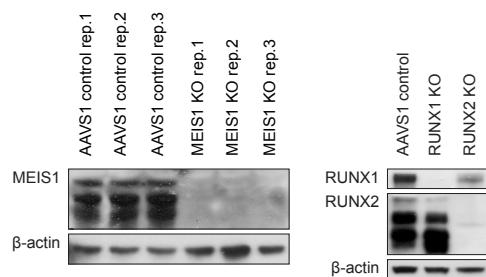


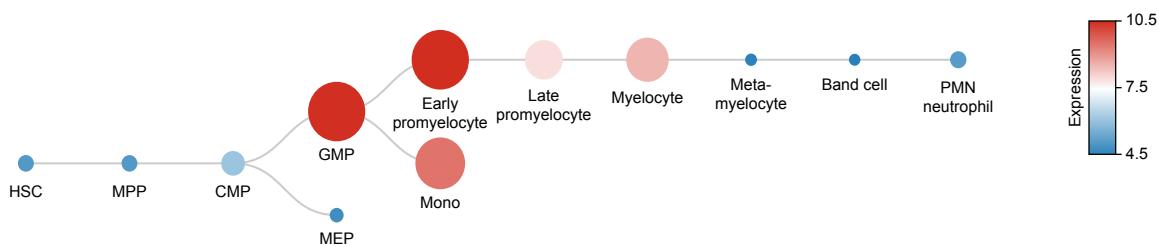
Figure S12, related to figure 7. Structure of the IRF8 locus and mechanism of its activation in KMT2A-rearranged AML.

- A, H3K27 acetylation, TF binding and 2D structure at the IRF8 locus. The top two tracks are H3K27ac metatracks representing primary AMLs. *Red*, tracks from pediatric patients with KMT2Ar AML ($n=10$); *grey*, tracks from the pediatric patients with non-KMT2Ar AML ($n=9$). Each metatrack is a collection of semi-transparent area plots representing individual samples and the average profile is represented by a thick line. The rest of the ChIP-seq tracks represent TF binding and H3K27 acetylation profiles in the MV411 cell line. Orange tracks represent TFs known to be the direct targets of KMT2A fusion oncoproteins and hypothesized to play a role in inducing IRF8 expression. Shaded areas represent 2 superenhancers associated with the IRF8 locus (SE1 and SE2). E1-E5 represent the constituent enhancers selected for the dCas9-KRAB-MeCP2 enhancer interference experiment. Superenhancer-promoter loops are demonstrated by the H3K27ac HiChIP and Micro-C 2D contact maps obtained in MV411 cells.
- B, MV411 cells stably expressing dCas9-KRAB-MECP2 were transduced with lentiviral vectors encoding gRNAs targeting the E1-E5 enhancers highlighted in panel A. Expression of IRF8 was measured by RT-qPCR.
- C, Viability of MV411 cells stably expressing dCas9-KRAB-MECP2 and transduced with GFP-expressing lentiviral vectors encoding gRNAs targeting the indicated enhancers was measured as fraction of GFP-positive cells.
- D, Knockouts of the 3 core regulatory TFs directly activated by KMT2A fusion oncoproteins do not decrease IRF8 expression. Expression of IRF8 was measured by RNA-seq 72 hours following electroporation of MV411 cells with pre-assembled Cas9/sgRNA complexes targeting the indicated TF genes. A guide RNA targeting the AAVS1 “safe harbor” locus was used as a negative control. P-values were calculated by BH-adjusted Wald test.

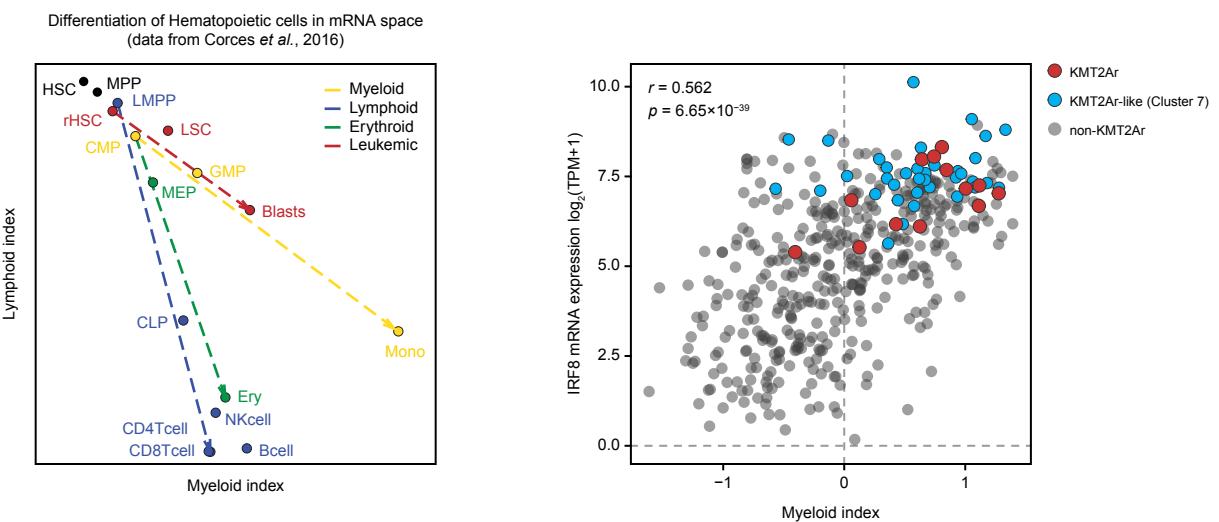
E, Validation of gene knockouts by Western blotting. MV411 cells were electroporated with pre-assembled Cas9/sgRNA complexes targeting the indicated genes and protein depletion was verified by Western blot 72 hours post electroporation. Paralog-specific knockout of RUNX2 is shown. Western blot visualizing the MEF2C knockout is found in Figure 6H.

Figure S13

A



B



C

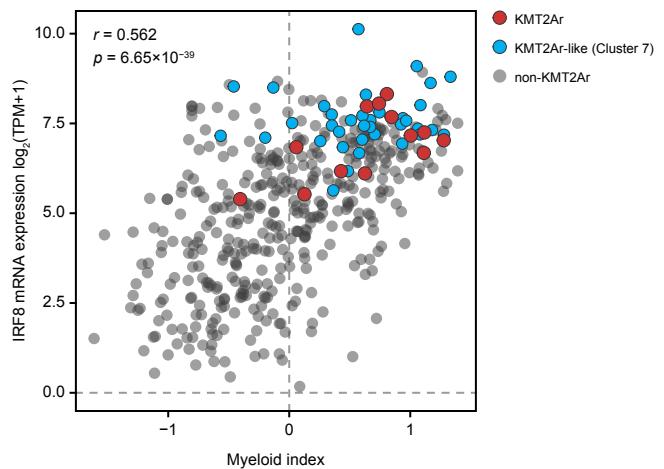


Figure S13, related to figure 7. Expression of IRF8 correlates with the myeloid differentiation state.

- A, Expression of IRF8 during normal human hematopoiesis. The plot was generated using the Bloodspot database with primary data from GSE42519 (Bagger et al. 2016; Rapin et al. 2014).
- B, Normal progenitors and leukemic cells are plotted according to their indices of myeloid and lymphoid development using primary data from (Corces et al. 2016). A myeloid differentiation index, computed from the mRNA expression of 19 cell type-specific markers (Methods), correctly reproduced both the normal myeloid trajectory and the functional AML hierarchy.
- C, Expression of IRF8 correlates with the myeloid differentiation index in the BeatAML dataset. KMT2A-rearranged and KMT2A-like leukemia samples are color-coded as shown.

Supplementary Figure References

- Bagger FO, Sasivarevic D, Sohi SH, Laursen LG, Pundhir S, Sønderby CK, Winther O, Rapin N, Porse BT. 2016. BloodSpot: a database of gene expression profiles and transcriptional programs for healthy and malignant haematopoiesis. *Nucleic Acids Res* 44: D917-24.
- Corces MR, Buenrostro JD, Wu B, Greenside PG, Chan SM, Koenig JL, Snyder MP, Pritchard JK, Kundaje A, Greenleaf WJ, et al. 2016. Lineage-specific and single-cell chromatin accessibility charts human hematopoiesis and leukemia evolution. *Nat Genet* 48: 1193–1203.
- Kuleshov MV, Jones MR, Rouillard AD, Fernandez NF, Duan Q, Wang Z, Koplev S, Jenkins SL, Jagodnik KM, Lachmann A, et al. 2016. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic acids research* 44: W90-7.
- Network CGAR. 2013. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. *New Engl J Medicine* 368: 2059–2074.
- Rapin N, Bagger FO, Jendholm J, Mora-Jensen H, Krogh A, Kohlmann A, Thiede C, Borregaard N, Bullinger L, Winther O, et al. 2014. Comparing cancer vs normal gene expression profiles identifies new disease entities and common transcriptional programs in AML patients. *Blood* 123: 894–904.
- Tyner JW, Tognon CE, Bottomly D, Wilmot B, Kurtz SE, Savage SL, Long N, Schultz AR, Traer E, Abel M, et al. 2018. Functional genomic landscape of acute myeloid leukaemia. *Nature* 562: 526–531.