

High-throughput Evaluation of Epilepsy-associated *KCNQ2* Variants Reveals Functional and Pharmacological Heterogeneity

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SUPPLEMENTARY MATERIAL

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- Fig. S2 Comparison of automated and manual patch clamp recording of *KCNQ2/KCNQ3*
- Fig. S3 Whole-cell currents from literature *KCNQ2* variants (homozygous state)
- Fig. S4 Whole-cell currents from literature *KCNQ2* variants (heterozygous state)
- Fig. S5 Manual and automated patch clamp analyses of *KCNQ2* variants (heterozygous state)
- Fig. S6 Whole-cell currents of *KCNQ2* population variants (homozygous state)
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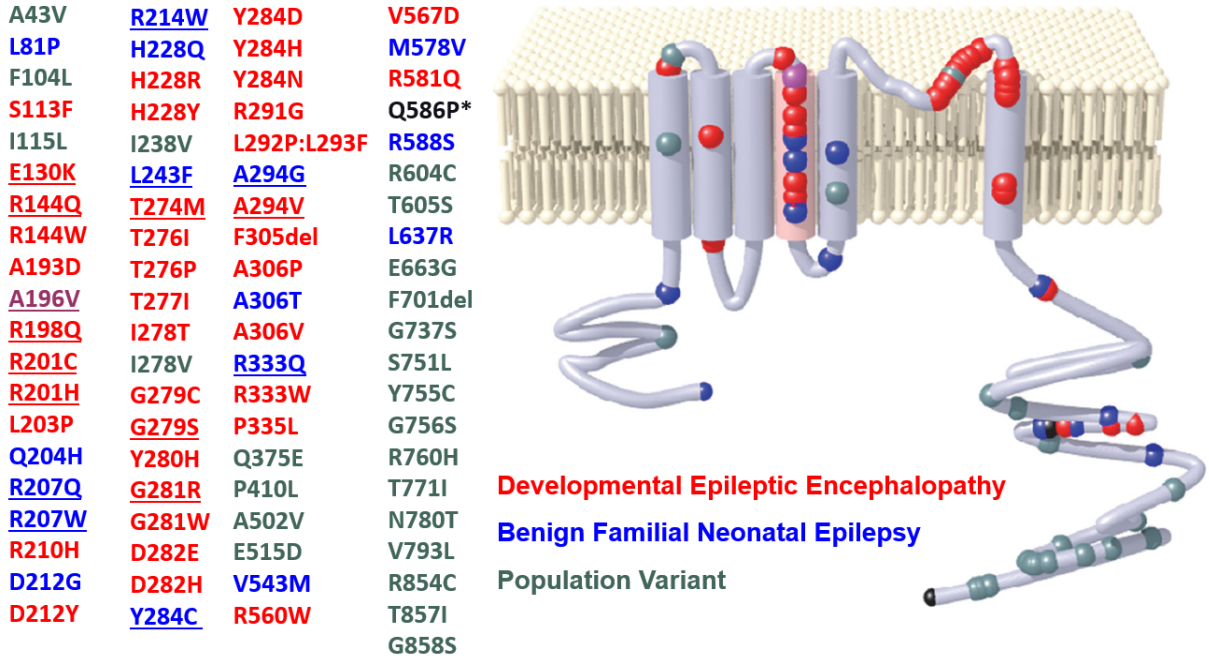


Figure S1. KCNQ2 variants analyzed in this study. Location and classification of the 81 KCNQ2 variants analyzed in this study. BFNE-associated variants are shown as blue dots, DEE-associated variants as red dots, the purple dot represents a variant associated with both BFNE and DEE, and population variants are denoted as green dots. Variant Q586P (marked by *) is associated with unknown phenotype category. Literature variants are underlined.

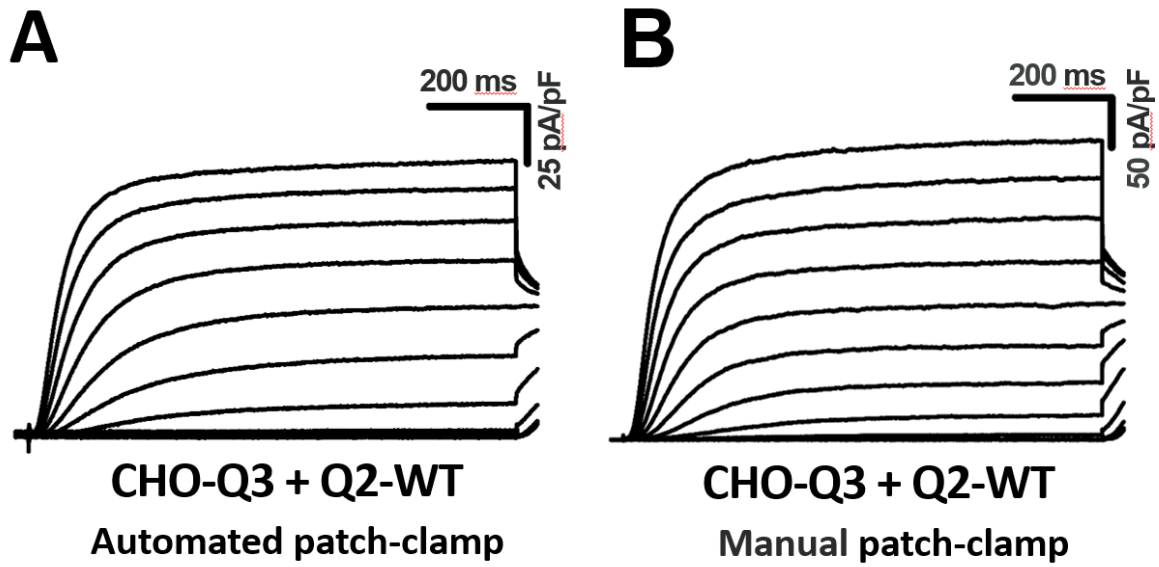


Figure S2. Comparison of automated and manual patch clamp recording of KCNQ2/KCNQ3. Whole cell current density recorded from CHO-Q3 cells electroporated with wild type KCNQ2 (Q2-WT) using either automated (**A**) or manual (**B**) patch clamp.

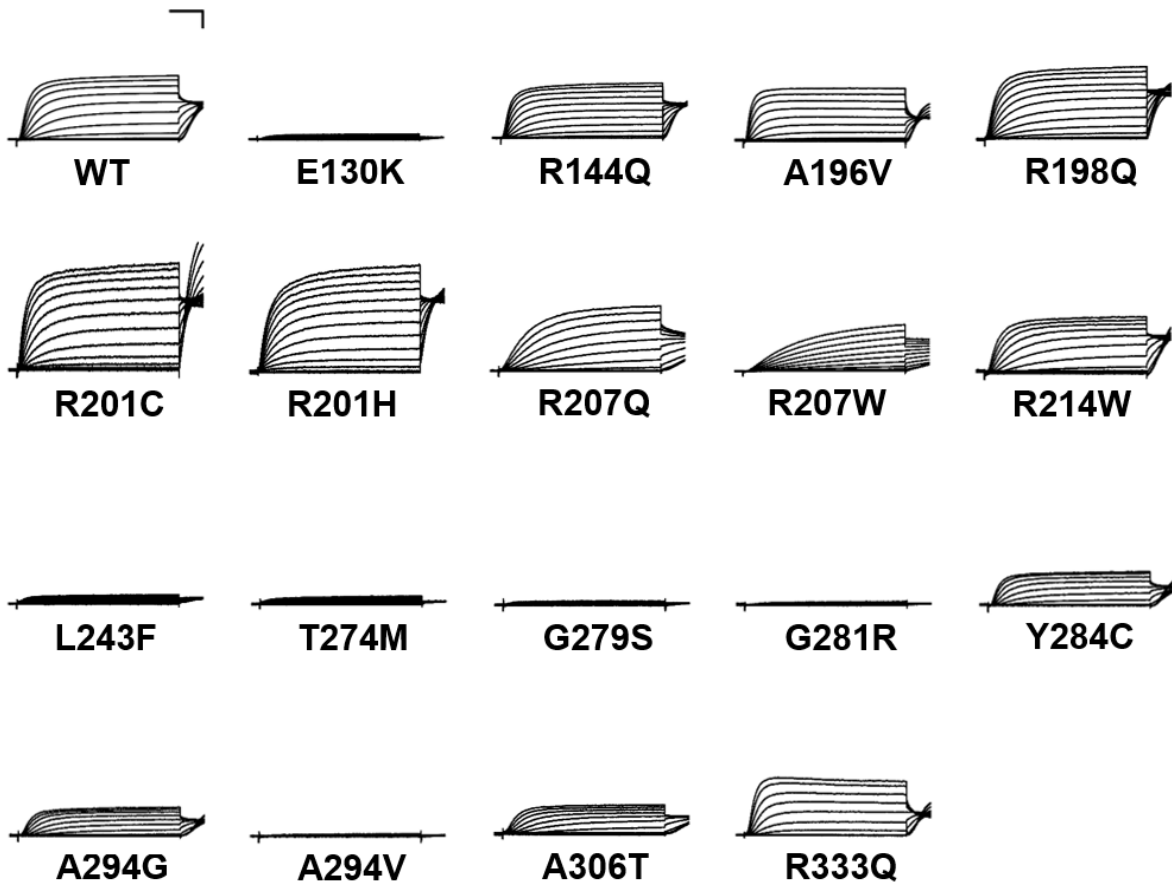


Figure S3. Whole-cell currents from literature KCNQ2 variants expressed as homozygous channels. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells electroporated with KCNQ2 variants from the literature set and normalized to wild type channel peak current recorded in parallel. For variant R201C, whole-cell currents were recorded from CHO-K1 cells co-electroporated with KCNQ3-WT plus KCNQ2-variant. Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

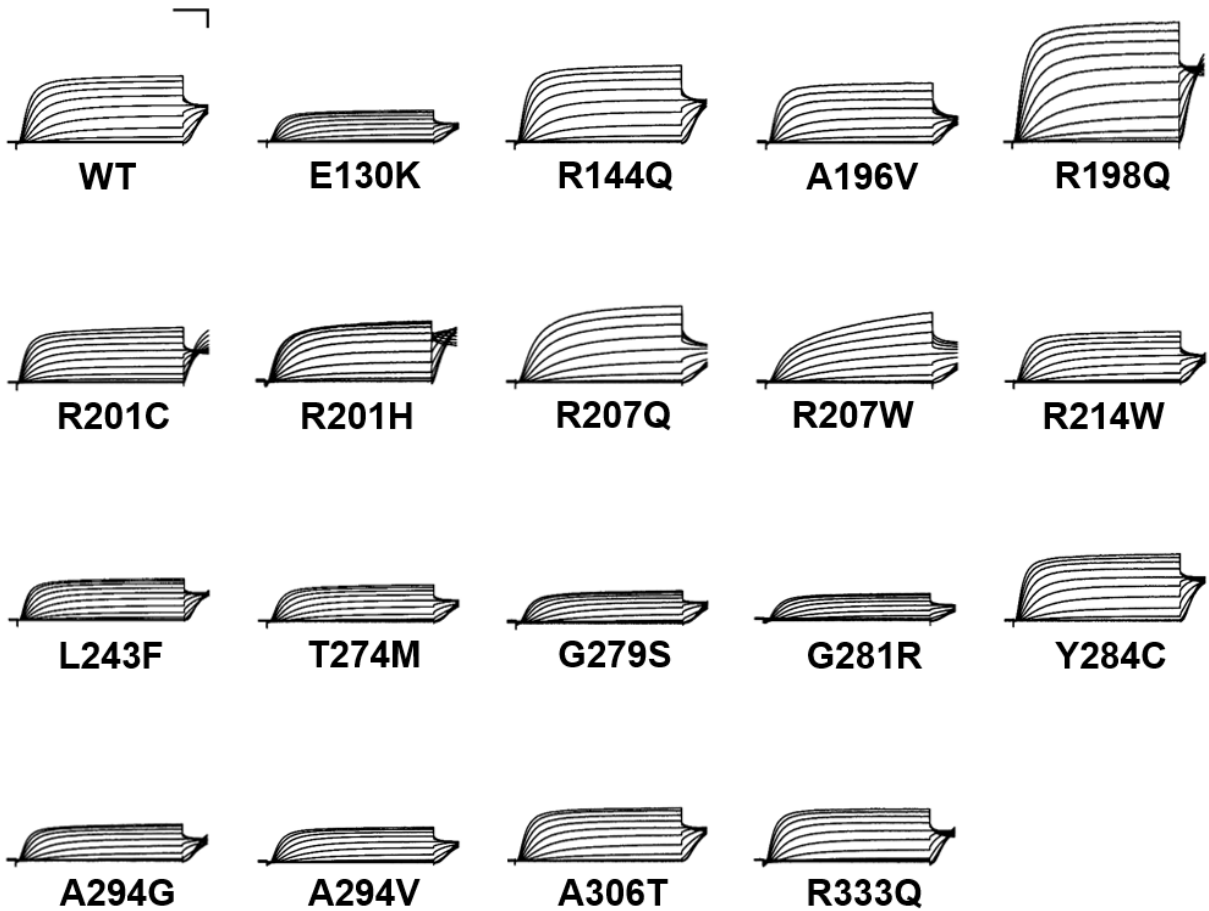


Figure S4. Whole-cell currents from literature KCNQ2 variants expressed as heterozygous channels. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells co-electroporated with wild type plus variant KCNQ2 cDNA from the literature set and normalized to wild type channel peak current recorded in parallel. Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

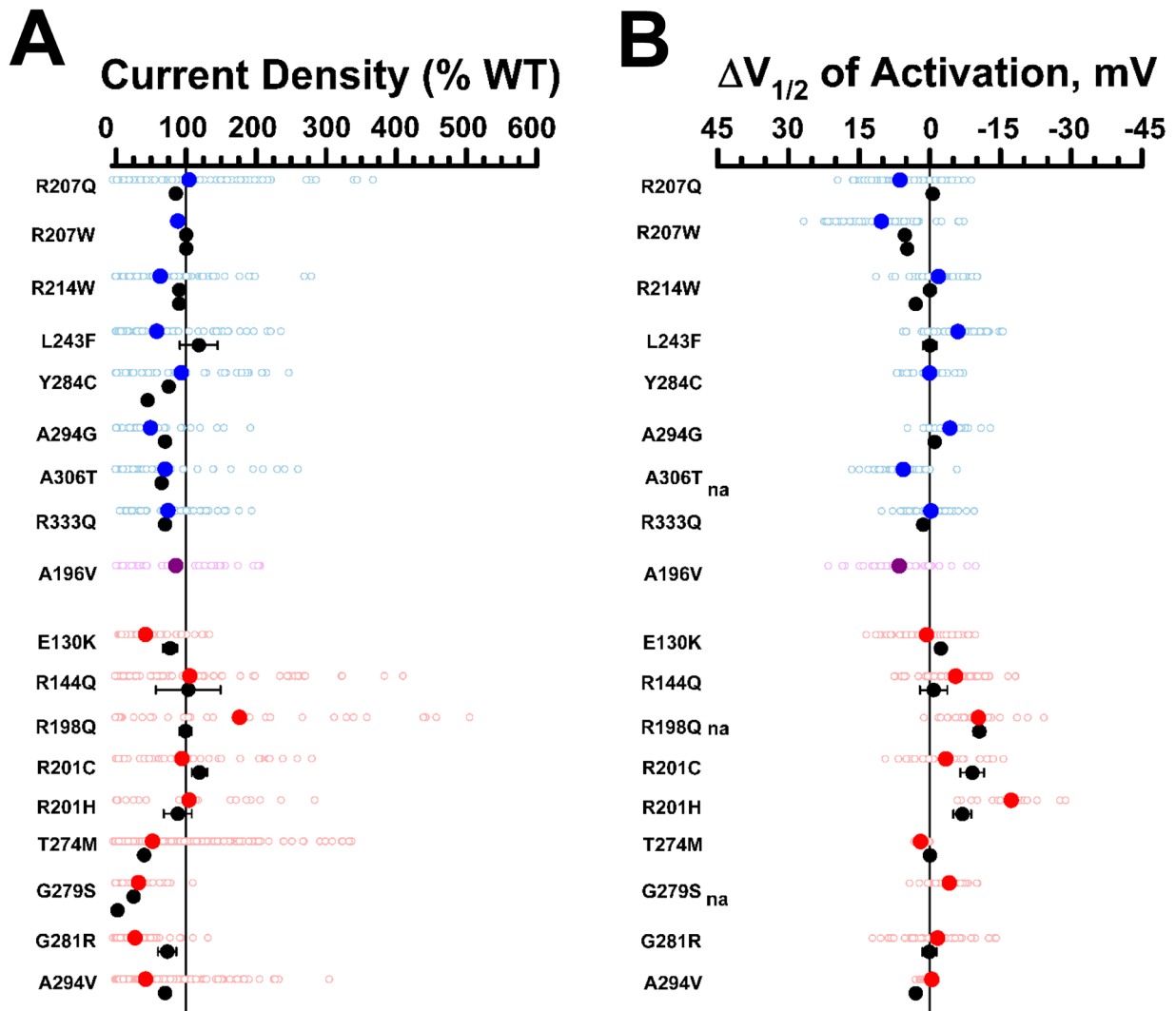


Figure S5. Manual and automated voltage clamp analyses of KCNQ2 variants expressed in the heterozygous state yield similar biophysical properties. **A.** Average whole-cell currents recorded at +40 mV from CHO-Q3 cells co-expressing variant + wild type KCNQ2 and normalized to WT channel peak current that was measured in parallel. **B.** Change (Δ) in voltage-dependence of activation $V_{1/2}$ determined for heterozygous KCNQ2 variants relative to the WT channel $\Delta V_{1/2}$ measured in parallel. Black symbols represent mean \pm SEM voltage-clamp data from literature reported variants (error bars are smaller than data symbol in some cases), while automated patch clamp results are shown as blue for BFNE, red for DEE, or purple symbols for BFNE/DEE pathogenic variants. All experimental data are presented as open circles with filled circles representing mean values. na = not available in the literature.

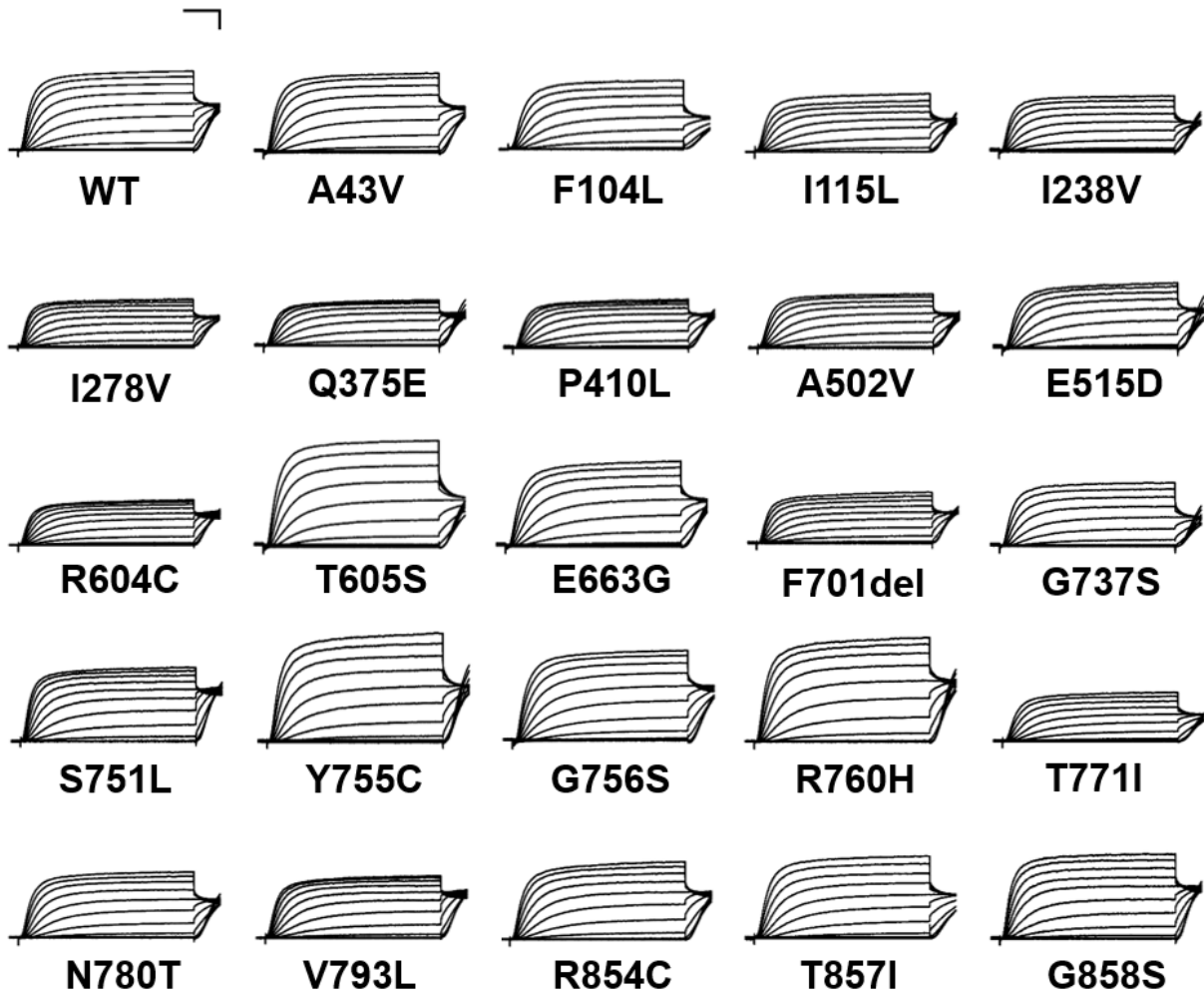


Figure S6. Average whole-cell currents recorded from CHO-Q3 cells electroporated with population KCNQ2 variants. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells electroporated with rare population KCNQ2 variants and normalized to wild type channel peak current measured in parallel. Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

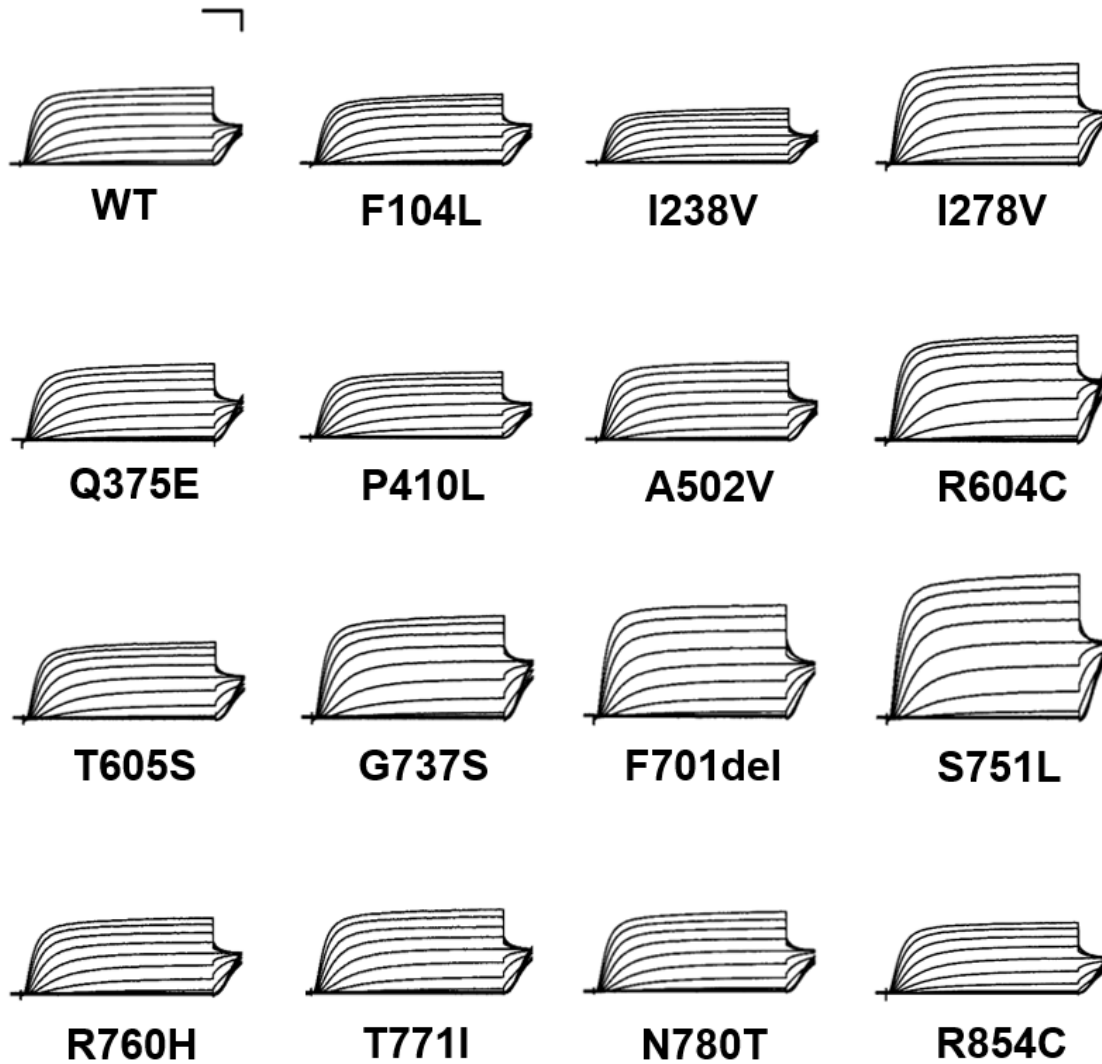


Figure S7. Average whole-cell currents recorded from CHO-Q3 cells co-electroporated with selected population variants plus wild type KCNQ2. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells co-electroporated with rare population variants plus wild type KCNQ2 and normalized to wild type channel peak current recorded in parallel. Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

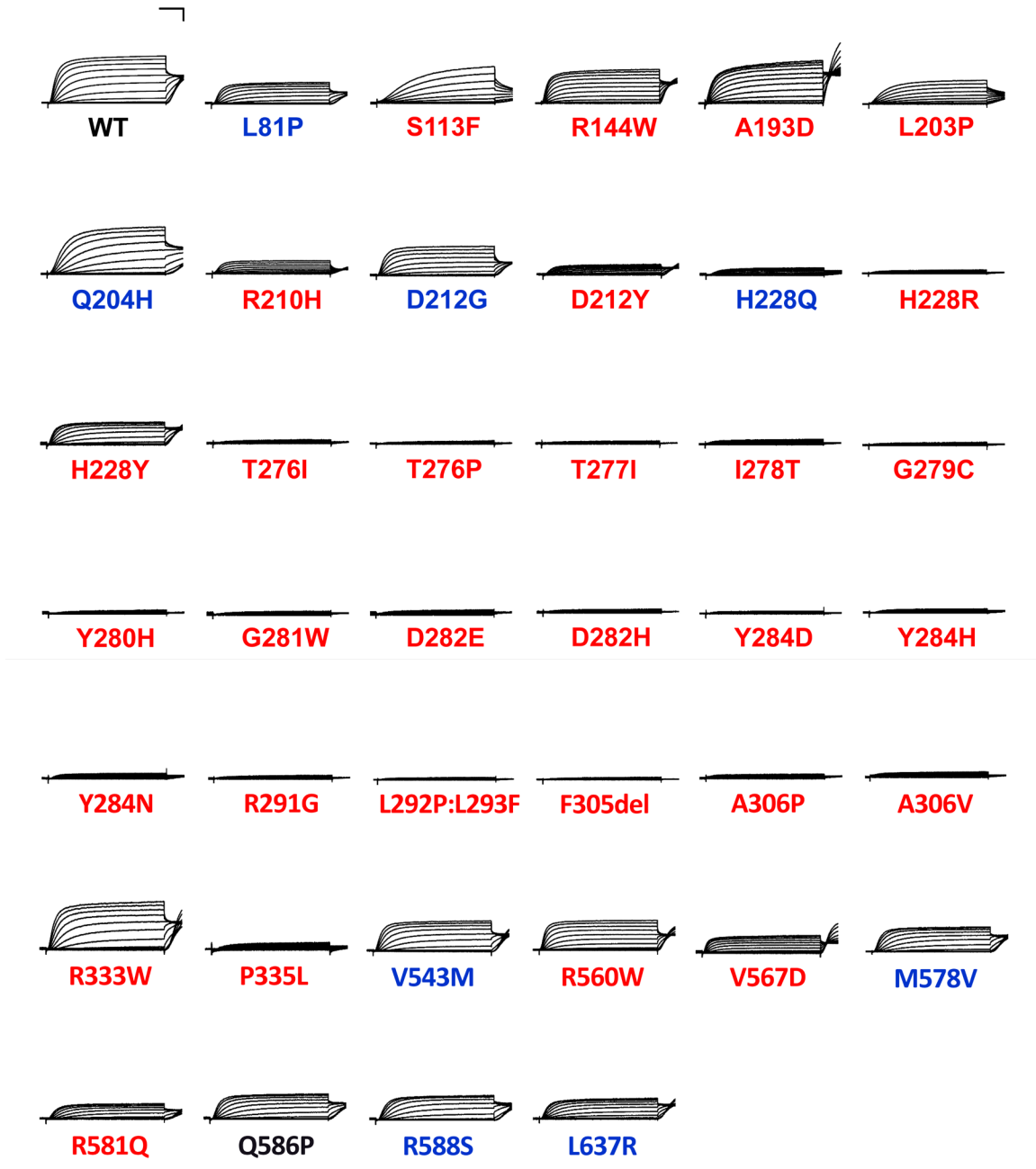


Figure S8. Whole-cell currents from epilepsy-associated KCNQ2 variants expressed as homozygous channels. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells electroporated with epilepsy-associated KCNQ2 variants and normalized to wild type channel peak current recorded in parallel. Variant labels: **Blue** = BFNE-associated; **Red** = DEE-associated; **Black** = unknown phenotype category (Q586P). For A193D and P335L, whole-cell currents were recorded from CHO-K1 cells co-electroporated with KCNQ3-WT plus KCNQ2-variant. Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

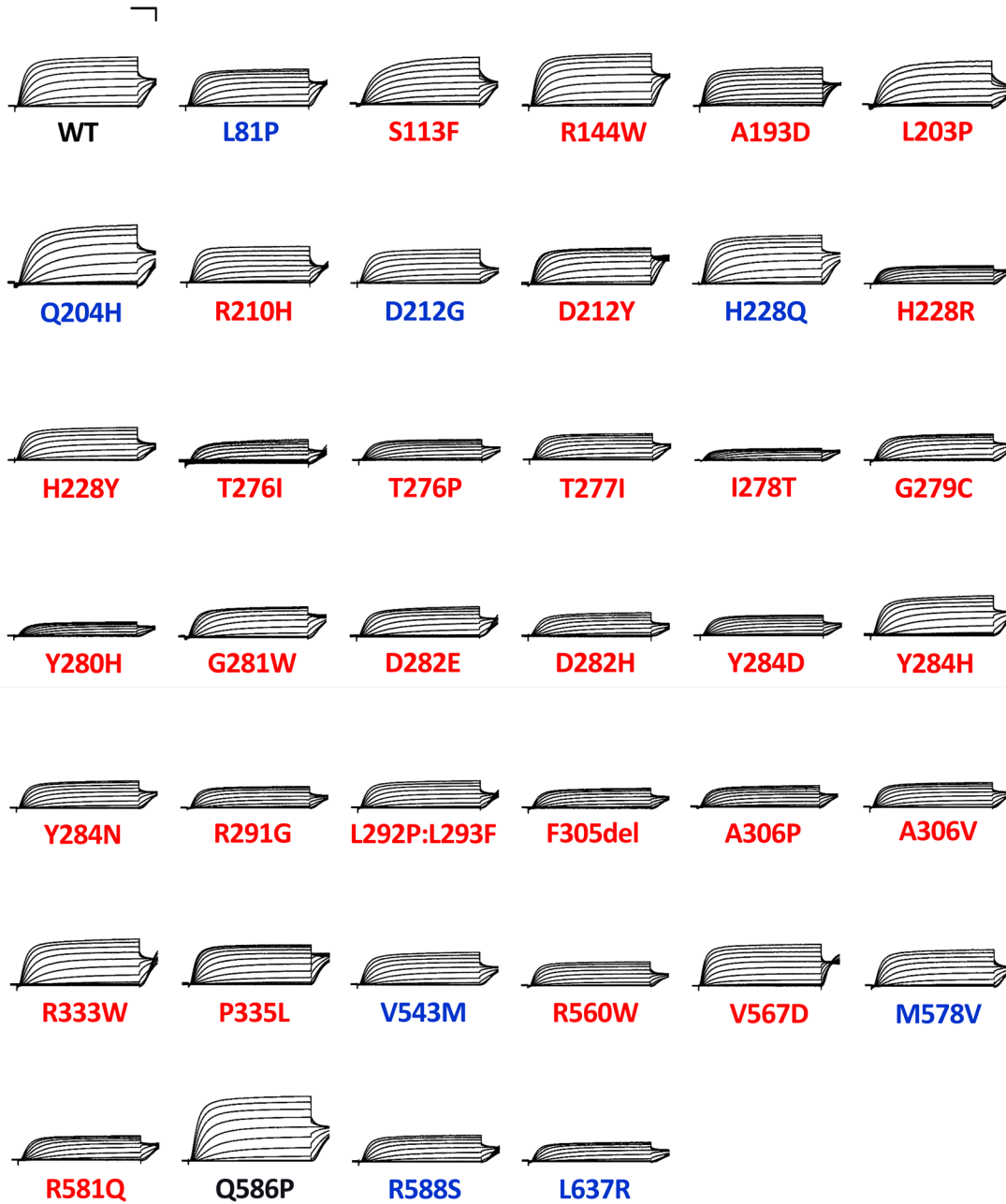


Figure S9. Average whole-cell currents recorded from CHO-Q3 cells co-electroporated with epilepsy-associated variants plus wild type KCNQ2. Average XE-991-sensitive whole-cell currents recorded by automated patch clamp from CHO-Q3 cells co-electroporated with epilepsy-associated KCNQ2 variants plus WT KCNQ2 and normalized to wild type channel peak current recorded in parallel. Variant labels: **Blue** = BFNE-associated; **Red** = DEE-associated; **Black** = unknown phenotype category (Q586P). Scale bars are 200 ms (horizontal) and 25% of WT channel current density (vertical).

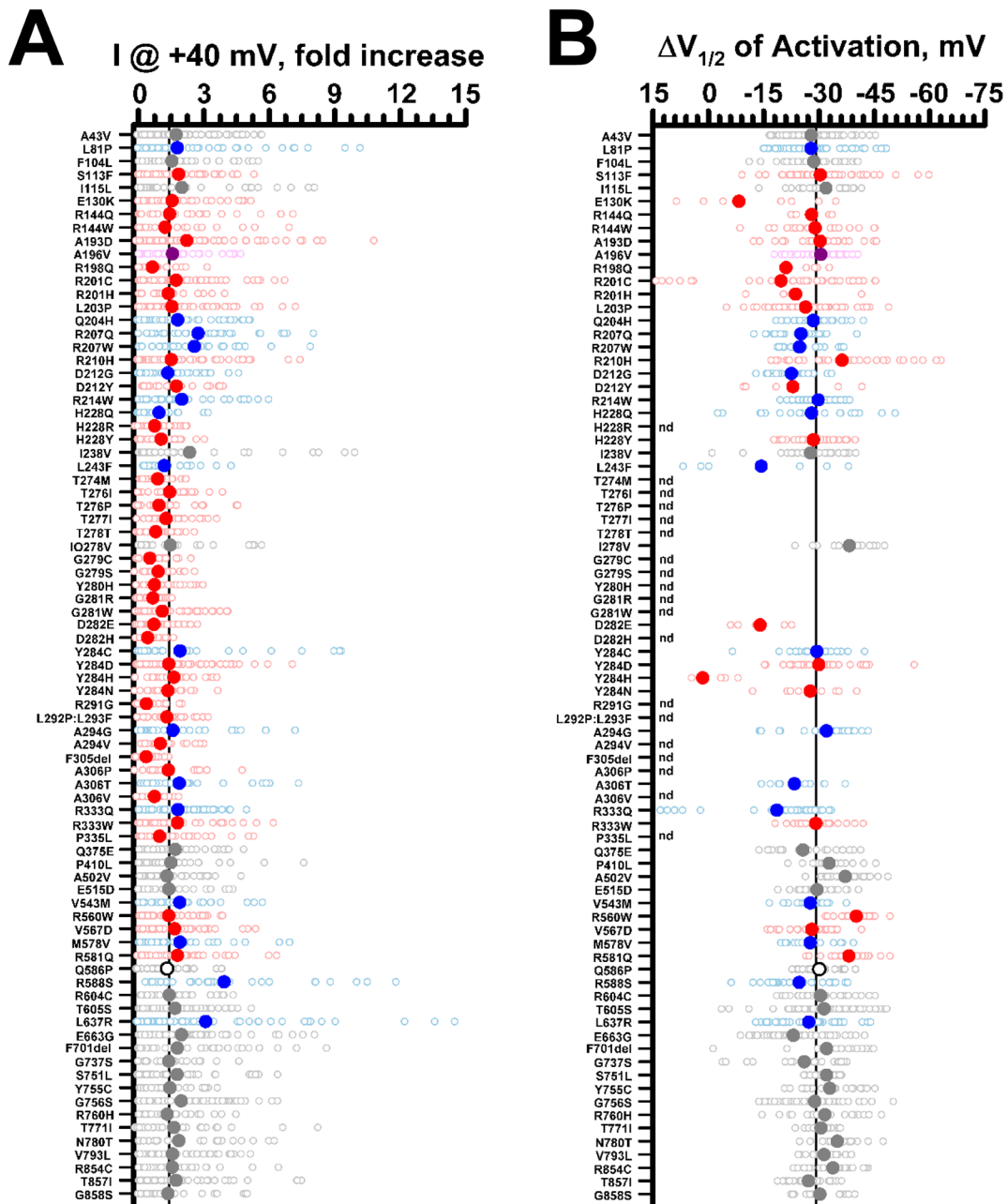


Figure S10. Retigabine effects on whole-cell currents recorded from KCNQ2 variants expressed in the homozygous state. A) Ratio of whole-cell currents recorded at +40 mV after exposure to 10 μ M retigabine and divided by the current measured under control conditions (n = 15-86). **B)** Change in voltage-dependence of activation $V_{1/2}$ determined for whole-cell currents after exposure to 10 μ M retigabine (n = 5-74). Dashed lines indicate average effect of retigabine on current amplitude and change in voltage-dependence of activation $V_{1/2}$ in the wild type channel. All experimental data are presented as open circles with larger filled circles representing mean values for BFNE (blue), DEE (red), BFNE/DEE (purple), unclear phenotype (white) or population (grey) variants. For complete list of results, see **Supplemental Table 6**.

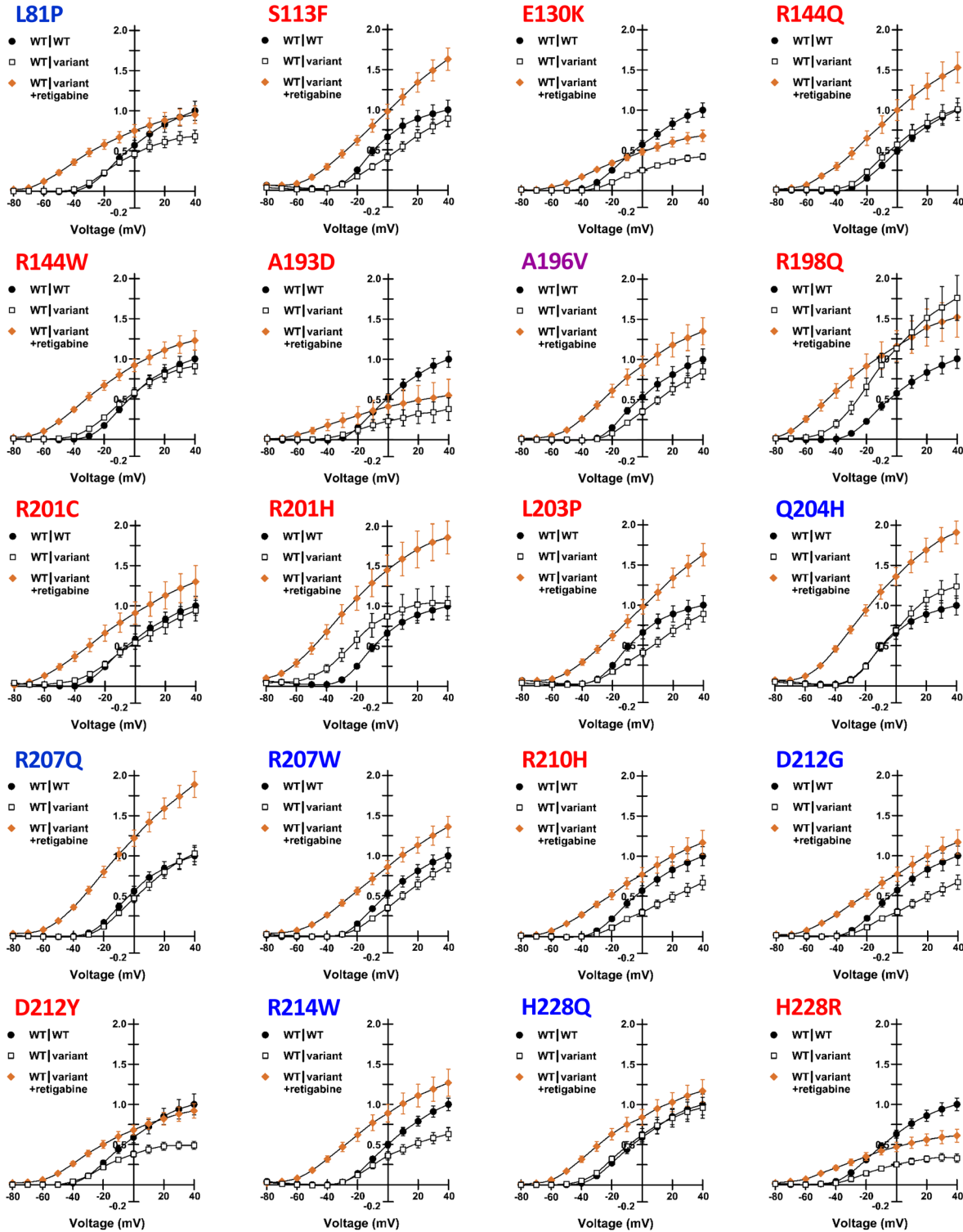


Figure S11. Retigabine effects on whole-cell currents recorded from epilepsy-associated KCNQ2 variants expressed in the heterozygous state. Normalized current-voltage relationships for each variant expressed in the heterozygous state recorded in the absence of retigabine (WT|variant, open squares) compared with heterozygous variants recorded in the absence of retigabine (WT|variant +retigabine, orange filled diamonds). Currents were first normalized to cell capacitance, then re-normalized to the peak current for WT channels recorded in parallel (WT|WT, filled circles). Variant labels: **Blue** = BFNE-associated; **Red** = DEE-associated; **Purple** = BFNE/DEE; **Black** = unknown phenotype category. Complete data sets are presented in **Table S7**.

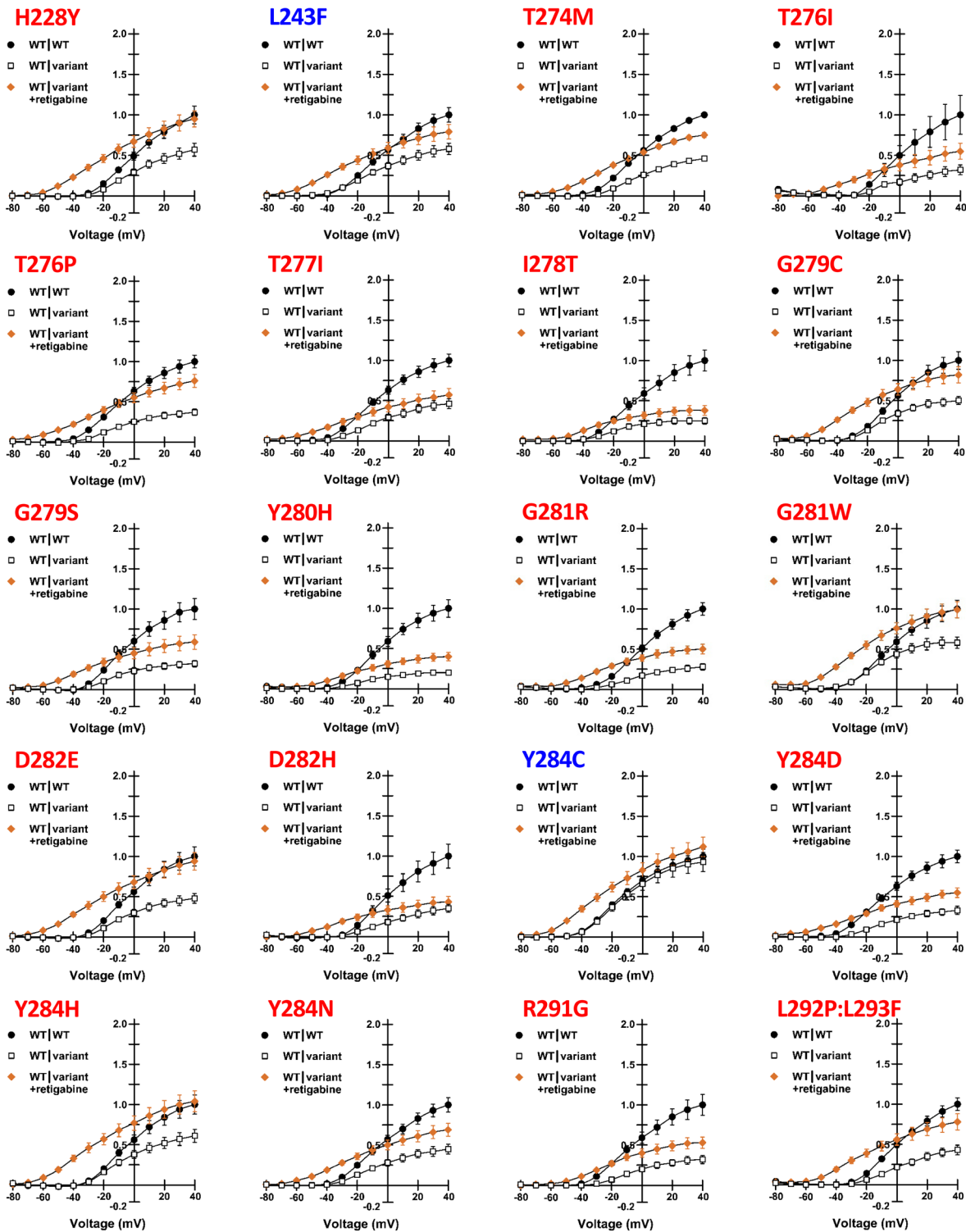


Figure S11 - continued. Retigabine effects on whole-cell currents recorded from epilepsy-associated KCNQ2 variants expressed in the heterozygous state. Normalized current-voltage relationships for each variant expressed in the heterozygous state recorded in the absence of retigabine (WT|variant, open squares) compared with heterozygous variants recorded in the absence of retigabine (WT|variant +retigabine, orange filled diamonds). Currents were first normalized to cell capacitance, then re-normalized to the peak current for WT channels recorded in parallel (WT|WT, filled circles). Variant labels: **Blue** = BFNE-associated; **Red** = DEE-associated; **Purple** = BFNE/DEE; **Black** = unknown phenotype category. Complete data sets are presented in Table S7.

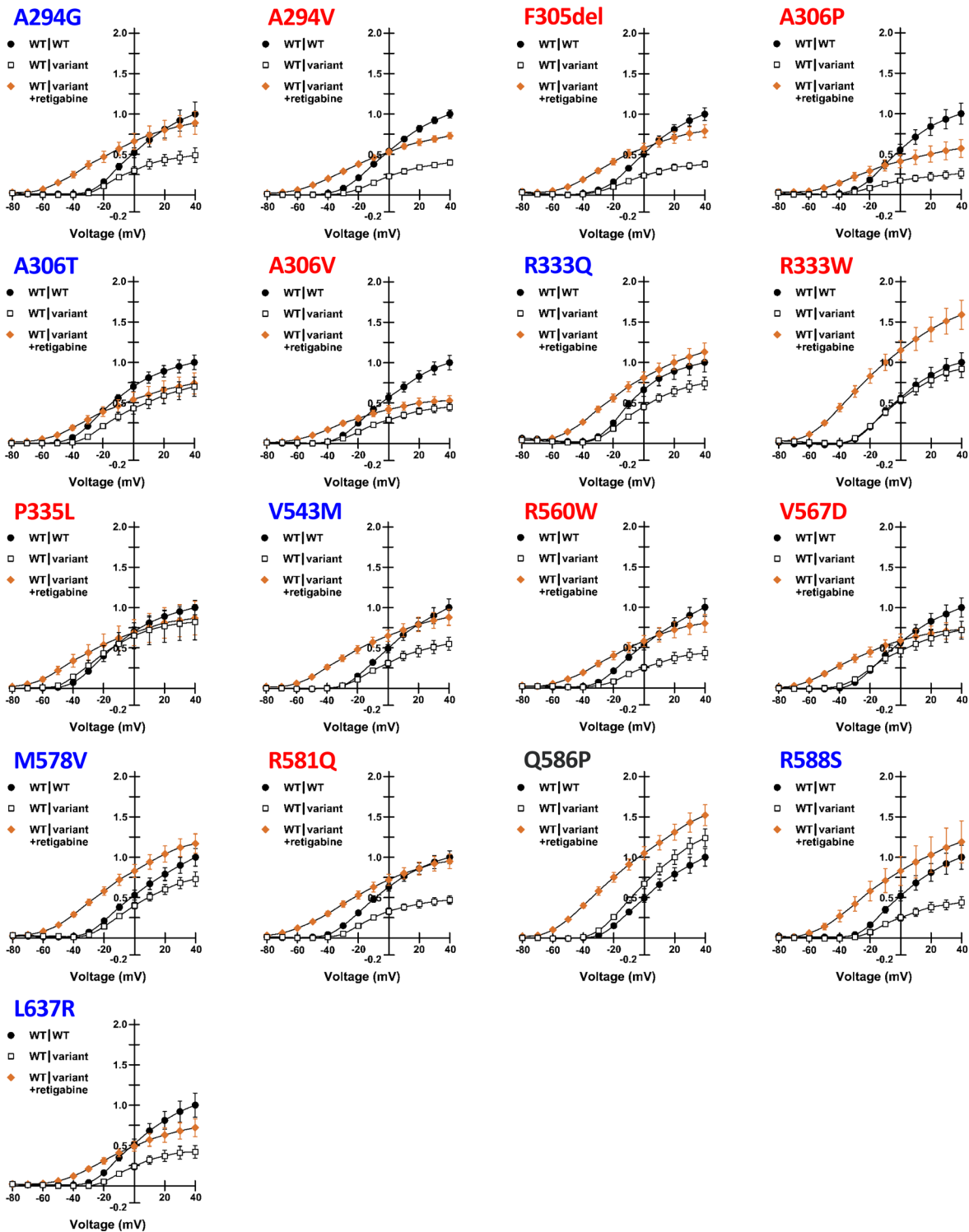


Figure S11 - continued. Retigabine effects on whole-cell currents recorded from epilepsy-associated KCNQ2 variants expressed in the heterozygous state. Normalized current-voltage relationships for each variant expressed in the heterozygous state recorded in the absence of retigabine (WT|variant, open squares) compared with heterozygous variants recorded in the absence of retigabine (WT|variant +retigabine, orange filled diamonds). Currents were first normalized to cell capacitance, then re-normalized to the peak current for WT channels recorded in parallel (WT|WT, filled circles). Variant labels: **Blue** = BFNE-associated; **Red** = DEE-associated; **Purple** = BFNE/DEE; **Black** = unknown phenotype category. Complete data sets are presented in **Table S7**.

Table S1 – KCNQ2 variant information

Nucleotide	Amino Acid	Channel Domain	Phenotype	MAF (gnomAD)	ClinVar	PubMed ID
c.128C>T	p.Ala43Val	N-term	PV	0.000176	LB/VUS	
c.242T>C	p.Leu81Pro	N-term	BFNE	0	N/A	29215089
c.312C>G	p.Phe104Leu	TMD: S1	PV	0.000008	N/A	
c.338C>T	p.Ser113Phe	TMD: S1-S2-Link	DEE	0	VUS/LP	29655203
c.343A>C	p.Ile115Leu	TMD: S1-S2-Link	PV	0.000016	N/A	
c.388G>A	p.Glu130Lys	TMD: S2	DEE	0	PATH	27535030
c.431G>A	p.Arg144Gln	TMD: S2-S3-Link	DEE	0	PATH/LP	23934111
c.430C>T	p.Arg144Trp	TMD: S2-S3-Link	DEE	0	PATH/LP	28628100; 28867141
c.578C>A	p.Ala193Asp	TMD: S2-S3-Link	DEE	0	PATH	27602407
c.587C>T	p.Ala196Val	TMD: S4	DEE	0	PATH	17475800
c.593G>A	p.Arg198Gln	TMD: S4	DEE	0	PATH	27861786
c.601C>T	p.Arg201Cys	TMD: S4	DEE	0	PATH/VUS	24107868
c.602G>A	p.Arg201His	TMD: S4	DEE	0	PATH	23708187
c.608T>C	p.Leu203Pro	TMD: S4	DEE	0	PATH	26007637
c.612G>T	p.Gln204His	TMD: S4	BFNE	0	LP	27602407
c.620G>A	p.Arg207Gln	TMD: S4	DEE	0	PATH/LP	17872363
c.619C>T	p.Arg207Trp	TMD: S4	BFNE	0	PATH	11572947
c.629G>A	p.Arg210His	TMD: S4	DEE	0	PATH	24107868
c.635A>G	p.Asp212Gly	TMD: S4	BFNE	0	N/A	19344764
c.634G>T	p.Asp212Tyr	TMD: S4	DEE	0	PATH	28817111
c.640C>T	p.Arg214Trp	TMD: S4	BFNE	0	PATH/LP	11175290; 29056246
c.684C>A	p.His228Gln	TMD: S4-S5-Link	BFNE	0	VUS	14534157
c.683A>G	p.His228Arg	TMD: S4-S5-Link	DEE	0	LP	
c.682C>T	p.His228Tyr	TMD: S4-S5-Link	BFNE	0	Not Provided	28837158
c.712A>G	p.Ile238Val	TMD: S5	PV	0.000008	VUS	
c.727C>T	p.Leu243Phe	TMD: S5	BFNE	0	PATH	14534157
c.821C>T	p.Thr274Met	TMD: P-loop	DEE	0	PATH	22275249
c.827C>T	p.Thr276Ile	TMD: P-loop	DEE	0	PATH	24463883
c.826A>C	p.Thr276Pro	TMD: P-loop	DEE	0	N/A	29720203
c.830C>T	p.Thr277Ile	TMD: P-loop	DEE	0	N/A	26544041
c.833T>C	p.Ile278Thr	TMD: P-loop	DEE	0	LP	30109124
c.832A>G	p.Ile278Val	TMD: P-loop	PV	0.000008	N/A	
c.835G>T	p.Gly279Cys	TMD: P-loop	DEE	0	PATH	25959266
c.836G>A	p.Gly279Ser	TMD: P-loop	DEE	0	N/A	27734276
c.838T>C	p.Tyr280His	TMD: P-loop	DEE	0	PATH	27779742
c.841G>A	p.Gly281Arg	TMD: P-loop	DEE	0	LP	24107868
c.841G>T	p.Gly281Trp	TMD: P-loop	DEE	0	PATH	25880994
c.846C>A	p.Asp282Glu	TMD: P-loop	DEE	0	N/A	28133863
c.844G>C	p.Asp282His	TMD: P-loop	DEE	0	VUS/LP	29655203
c.851A>G	p.Tyr284Cys	TMD: P-loop	BFNE	0	PATH	9425895
c.850T>G	p.Tyr284Asp	TMD: P-loop	DEE	0	PATH	27535030
c.850T>C	p.Tyr284His	TMD: P-loop	DEE	0	N/A	29588952
c.850T>A	p.Tyr284Asn	TMD: P-loop	DEE	0	N/A	
c.871A>G	p.Arg291Gly	TMD: P-loop	DEE	0	N/A	27779742
c.[875T>C:877C>T]	p.Leu292Pro:Leu293Phe	TMD: S6	DEE	0	LP, VUS	

Table S1 – (continued) KCNQ2 variant information

Nucleotide	Amino Acid	Channel Domain	Phenotype	MAF (gnomAD)	ClinVar	PubMed ID
c.881C>G	p.Ala294Gly	TMD: S6	BFNE	0	PATH	17129708
c.881C>T	p.Ala294Val	TMD: S6	DEE	0	PATH	17129708
c.913_915delTTC	p.Phe305del	TMD: S6	DEE	0	N/A	28554332; 28728838; 18640800
c.916G>C	p.Ala306Pro	TMD: S6	DEE	0	PATH	29655203
c.916G>A	p.Ala306Thr	TMD: S6	DEE	0	PATH	9425895; 26138355
c.917C>T	p.Ala306Val	TMD: S6	DEE	0	PATH	31152295
c.998G>A	p.Arg333Gln	C-term	BFNE	0.000004	PATH/LP	29215089; 14534157
c.997C>T	p.Arg333Trp	C-term	DEE	0	PATH	16039833
c.1004C>T	p.Pro335Leu	C-term	DEE	0	PATH/LP	28867141
c.1123C>G	p.Gln375Glu	C-term	DEE	0.000018	N/A	
c.1229C>T	p.Pro410Leu	C-term	PV	0.000043	VUS	
c.1505C>T	p.Ala502Val	C-term	PV	0.000036	VUS	
c.1545G>C	p.Glu515Asp	C-term	PV	0.002517	B/LB/VUS	19380078
c.1627G>A	p.Val543Met	C-term	BFNE	0.000004	VUS/LP	28399683
c.1678C>T	p.Arg560Trp	C-term	DEE	0	PATH/LP	22275249
c.1700T>A	p.Val567Asp	C-term	DEE	0	LP	27888506
c.1732A>G	p.Met578Val	C-term	BFNE	0	PATH/LP	25982755
c.1742G>A	p.Arg581Gln	C-term	DEE	0	PATH/LP	27864847
c.1757A>C	p.Gln586Pro	C-term	DEE	0	VUS	
c.1764A>T	p.Arg588Ser	C-term	BFNE	0	PATH	25982755
c.1810C>T	p.Arg604Cys	C-term	PV	0.000008	VUS	
c.1814C>G	p.Thr605Ser	C-term	PV	0.000056	VUS/LB	
c.1910T>G	p.Leu637Arg	C-term	BFNE	0	PATH	25982755
c.1988A>G	p.Glu663Gly	C-term	PV	0.000047	N/A	
c.2101_2103delTCT	p.Phe701del	C-term	PV	0.000009	N/A	
c.2209G>A	p.Gly737Ser	C-term	PV	0.000016	VUS	
c.2252C>T	p.Ser751Leu	C-term	PV	0.000065	VUS	
c.2264A>G	p.Tyr755Cys	C-term	PV	0.002953	B/LB	
c.2266G>A	p.Gly756Ser	C-term	PV	0.000264	LB	
c.2279G>A	p.Arg760His	C-term	PV	0.000059	VUS	
c.2312C>T	p.Thr771Ile	C-term	PV	0.000047	VUS	
c.2339A>C	p.Asn780Thr	C-term	PV	0.609194	B	
c.2377G>C	p.Val793Leu	C-term	PV	0.000025	VUS	
c.2560C>T	p.Arg854Cys	C-term	PV	0.000226	B/LB	
c.2570C>T	p.Thr857Ile	C-term	PV	0.000019	N/A	
c.2572G>A	p.Gly858Ser	C-term	PV	0.000030	VUS	

Table S2. Sequence of mutagenic primers used to generate KCNQ2 variants.

Nucleotide change	Amino acid change	Forward Primer	Reverse Primer
c.128C>T	p.Ala43Val	GCTGATCGTCGGCTCCGAGCCCCCAAG	CGGAGCCGACGATCAGCAGCGCCCCGTCC
c.242T>C	p.Leu81Pro	GCAGAATTTCCCTACAACGTGCTGGAGCGGCC	TGTAGGGGAAATTCCTGAGCTTCGGGTAGAAGG
c.312C>G	p.Phe104Leu	CTGGTTTTGCTCTGCCTCGTGTCTGTGTTTTT	AGGCAGGACAAAACCAGGAGAACCGTAGGCGTG
c.338C>T	p.Ser113Phe	GTGTTTTTCACCATCAAGGAGTATGAGAAGAGCTCG	TTGATGGTGA AAAACACAGACAGCAGGCGAGG
c.343A>C	p.Ile115Leu	TTCCACCCTCAAGGAGTATGAGAAGAGCTCGGAGG	ACTCCTTGAGGGTGGAAAAACAGACAGCAGGAGG
c.388G>A	p.Glu130Lys	CCTGA AAAATCGTGACTATCGTGGTGTGGCGTG	TAGTCACGATTTTCAGGATGTAGAGGGCCCCCTC
c.430C>T	p.Arg144Trp	GTACTTCGTGTGGATCTGGGCCGAGGCTGC	AGATCCACACGAACTCCACGCCAAACACC
c.431G>A	p.Arg144Gln	TACTTCGTGCAGATCTGGGCCGAGGCTGCTG	CAGATCTGCACGAACTCCACGCCAAACAC
c.578C>A	p.Ala193Asp	CGTCTTTGACACATCTGCGCTCCGGAGCCT	CAGATGTGTCAAAGACGTTGCCCTGGGAGCC
c.587C>T	p.Ala196Val	GCCACATCTGTGCTCCGAGCCTGCGCTTCC	CGGAGCACAGATGTGGCAAAGACGTTGCCCTG
c.593G>A	p.Arg198Gln	TCTGCGCTCCAGAGCCTGCGCTTCTGCAGAT	CGCAGGCTCTGGAGCGCAGATGTGGCAAAGAC
c.601C>T	p.Arg201Cys	GCCTGTGCTTCTGCAGATTCTGCGGATGATC	CTGCAGGAAGCACAGGCTCCGGAGCGCAGATGT
c.602G>A	p.Arg201His	CCTGCACTTCTGCAGATTCTGCGGATGATCC	TCTGCAGGAAGTGCAGGCTCCGGAGCGCAGATGT
c.608T>C	p.Leu203Pro	CTTCCCGCAGATTCTGCGGATGATCCGCATG	GCAGAATCTCGGGAAGCGCAGGCTCCGGAGC
c.612G>T	p.Gln204His	CCTGCATATTCTGCGGATGATCCGCATGGACC	TCCGCAGAATATGCAGGAAGCGCAGGCTCCGG
c.620G>A	p.Arg207Gln	AGATTTCTGCAGATGATCCGCATGGACCGCGG	GATCATCTGCAGAATCTGCAGGAAGCGCAGG
c.619C>T	p.Arg207Trp	CAGATTCTGTGGATGATCCGCATGGACCGGC	ATCATCCACAGAATCTGCAGGAAGCGCAGG
c.629G>A	p.Arg210His	GCCGATGATCCACATGGACCGCGGGGAGGC	CCATGTGGATCATCCCGAGAATCTGCAGGAAG
c.635A>G	p.Asp212Gly	GATCCGCATGGCGCCGGCGGGAGGCACCTG	GCCCGCCATCGGATCATCCCGAGAATCTGC
c.634G>T	p.Asp212Tyr	TGATCCGCATGTACCGCGGGGAGGCACCTG	CCGGTACATCGGATCATCCCGAGAATCTGC
c.640C>T	p.Arg214Trp	TGGACCGGTGGGGAGGCACCTGGAAGCTGC	GCCTCCCAACCGGTCCATCGGATCATCCG
c.684C>A	p.His228Gln	TATGCCCAAGCAAGGAGCTGGTCACTGCCTG	TCCTTGCTTGGGCATAGACCACAGAGCCAG
c.683A>G	p.His228Arg	CTATGCCCGCAGCAAGGAGCTGGTCACTGCC	CCTTGCTGCGGCATAGACCACAGAGCCAG
c.682C>T	p.His228Tyr	CTATGCCACAGCAAGGAGCTGGTCACTGCC	CCTTGCTGTAGGCATAGACCACAGAGCCAGC
c.712A>G	p.Ile238Val	TGGTACGCTGGCTTCTTTGTCTCATCTGCG	AGGAAGCCGACGTACCAGGCAGTGACCAGCTCC
c.727C>T	p.Leu243Phe	TTCTTTGTTCATCTGGCCTCGTTCTGCTGGTG	AGGATGAAACAAAGGAAGCCGATGTACCAGGCAG
c.821C>T	p.Thr274Met	GGTGGGCTGATCATGCTGACCAACA	TGGTGGTCAGCATGATCAGGCCCAACC
c.827C>T	p.Thr276Ile	CGCTGATCACCATTGGCTACGGGGACAAGTAC	GCCAATGGTGTATCAGCGTGATCAGGCCCAACC
c.826A>C	p.Thr276Pro	CACGCTGCCACCATTGGCTACGGGGACAAG	CAATGGTGGGACGCGTGATCAGGCCCAACA
c.830C>T	p.Thr277Ile	TGACCACTATTGGCTACGGGGACAAGTACCC	GTAGCCAATGTATGGTCAGCGTGATCAGGCC
c.833T>C	p.Ile278Thr	GACCACCACTGGCTACGGGGACAAGTACCCC	CGTAGCCAATGGTGGTCAGCGTGATCAGGCC
c.832A>G	p.Ile278Val	GACCACCACTGGCTACGGGGACAAGTACCCC	CGTAGCCAATGGTGGTCAGCGTGATCAGGCC
c.835G>T	p.Gly279Cys	ACCACCATTGCTACGGGGACAAGTACCCC	CCGTAGCAATGGTGGTCAGCGTGATCAGGCC
c.836G>A	p.Gly279Ser	ACCACCATTGCTACGGGGACAAGTACCCC	CCGTAGCAATGGTGGTCAGCGTGATCAGGCC
c.838T>C	p.Tyr280His	ACCATTGGCCACGGGGACAAGTACCCCAGAC	TCCCCGTGGCAATGGTGGTCAGCGTGATCAG
c.841G>A	p.Gly281Arg	ATTGGCTACAGGGACAAGTACCCCAGACCTG	TTGTCCCTGTAGCAATGGTGGTCAGCGTGATC
c.841G>T	p.Gly281Trp	ATTGGCTACAGGGACAAGTACCCCAGACCTG	TTGTCCCTGTAGCAATGGTGGTCAGCGTGATC
c.846C>A	p.Asp282Glu	TACGGGGA AAGTACCCCAGACTGGAACGG	GGGTACTTTCCCGTAGCCAATGGTGGTCAG
c.844G>C	p.Asp282His	CTACGGGCAACAAGTACCCCAGACTGGAACG	GGTACTTTGCCCCGTAGCCAATGGTGGTCAGC
c.851A>G	p.Tyr284Cys	GGGACAAGTCCCCAGACTGGAACGGCAG	CTGGGGCACTTTGCCCGTAGCCAATGGTGG
c.850T>G	p.Tyr284Asp	GGGACAAGGACCCCAGACTGGAACGGCA	TGGGGGTCTTGTCCCCGTAGCCAATGGTGG
c.850T>C	p.Tyr284His	GGGACAAGCACCCCAGACTGGAACGGCA	TGGGGGTCTTGTCCCCGTAGCCAATGGTGG

Table S2 - continued. Sequence of mutagenic primers used to generate KCNQ2 variants.

Nucleotide change	Amino acid change	Forward Primer	Reverse Primer
c.850T>A	p.Tyr284Asn	GGGGACAAG <u>A</u> ACCCCCAGACCTGGAACGGCA	TGGGGG <u>T</u> CTTGTCCCGTAGCCAATGGTGG
c.871A>G	p.Arg291Gly	ACCTGGAACGG <u>G</u> GGCTCCTTGCGG	CCGCAAGGAGCC <u>G</u> CCGTTCCAGGT
c.[875T>C:877C>T]	p.Leu292Pro:Leu293Phe	CAGGC <u>C</u> TTTGGCGGAACCTTCACCCATCG	TTGCCGCAAA <u>A</u> GGGCTGCCGTTCCAGGTCTGGG
c.881C>T	p.Ala294Val	CTCCTTG <u>T</u> GGCAACCTTCACCTCATCGGTG	AAGGTTGCC <u>A</u> CAAGGAGCCTGCCGTTCCAGG
c.881C>G	p.Ala294Gly	CTCCTTG <u>G</u> GGCAACCTTCACCTCATCGGTG	AAGGTTGCC <u>C</u> CAAGGAGCCTGCCGTTCCAGG
c.913_915delITC	p.Phe305del	TGTCTCCTTC...GCGCTGCTGCAGGCATCTTG	CAGCGC...GAAGGAGACACCGATGAGGGTGAAG
c.916G>C	p.Ala306Pro	CTCCTTCTTC <u>C</u> CGTGCCTGCAGGCATCTTGG	GCAGCG <u>G</u> AAGAAGGAGACACCGATGAGGGTG
c.916G>A	p.Ala306Thr	CTCCTTCTTC <u>A</u> CGTGCCTGCAGGCATCTTGG	GCAGCG <u>T</u> AAGAAGGAGACACCGATGAGGGTG
c.917C>T	p.Ala306Val	CCCTTCTCG <u>T</u> GCTGCCTGCAGGCATCTTGGG	AGGCAGC <u>A</u> CGAAGAAGGAGACACCGATGAGGG
c.998G>A	p.Arg333Gln	GAGAAGAGG <u>C</u> AAGACCCGGCAGCAGGCCTGAT	GGGTT <u>C</u> IGCTCTTCTCAAAGTGCTTCTGCCTG
c.997C>T	p.Arg333Trp	TTGAGAAGAGG <u>T</u> GGAACCCGGCAGCAGGCCTG	GTTCC <u>A</u> CTCTTCTCAAAGTGCTTCTGCCTGTG
c.1004C>T	p.Pro335Leu	AGGCGGAACCT <u>G</u> GGCAGCAGGCCTGATCCAGTC	GCTGCC <u>A</u> GGTTCCGCCTTCTCAAAGTGCTTC
c.1123C>G	p.Gln375Glu	TACAGTT <u>C</u> GAAACTCAAACCTACGGGCTCTCC	GTTTGAGTTT <u>C</u> GAACTGTACATGGGCAGGGTG
c.1229C>T	p.Pro410Leu	AGGACCCCT <u>G</u> CCGCGAGCCGTCTCCAAGCC	CTCCGGC <u>G</u> GGGGGCTCCTTCTCAAAGCGGAG
c.1505C>T	p.Ala502Val	GTGCCG <u>T</u> GACGGCAGAAGCAAGCAAGC	CTGCCG <u>T</u> GAC <u>C</u> GGCACCCCTTGATGCGGAAAGC
c.1545G>C	p.Glu515Asp	CGGAGAC <u>G</u> ACATTGTGGATGACAAGAGCTGCC	CCACAATG <u>C</u> TCTCCGGGAGGCTTCTCTTCTG
c.1627G>A	p.Val543Met	CAGAGC <u>C</u> ATGTGTGTCATGCGGTTCTGGTGTG	TGACACAC <u>A</u> GGCTCTGATGCTGACTTTGAGGGC
c.1678C>T	p.Arg560Trp	GGAGAGCCT <u>G</u> GGCCCTACGACGTGATGGACG	AGGGCC <u>A</u> CAGGCTCTCTTGAACCTCCGCTTG
c.1700T>A	p.Val567Asp	ATGACG <u>C</u> ATCGAGCAGTACTCAGCCGGC	TGCTCGAT <u>G</u> CTGCATCAGTCGTAGGGCC
c.1732A>G	p.Met578Val	TGGAC <u>G</u> TGCTGCCGAATTAAAGACCTGCAG	TCGGGACAGC <u>A</u> GTCAGGTGGCCGGCTGAGTA
c.1742G>A	p.Arg581Gln	TCCC <u>A</u> AATTAAGAGCCTCGAGTCCAGAGTGGAC	AGGCTCTTAAT <u>T</u> TGGGACAGCATGTCCAGGTGGC
c.1757A>C	p.Gln586Pro	AGAGCCTG <u>C</u> CTCCAGAGTGACAGATCGTGG	TCTGGAC <u>G</u> CAGGCTCTTAATTCGGGACAGCATG
c.1764A>T	p.Arg588Ser	TGCAGTCCAG <u>T</u> GTGACAGATCGTGGGGCGG	GTCCAC <u>A</u> CTGGACTGCAGGCTCTTAATTCGGG
c.1810C>T	p.Arg604Cys	GGACAAGGACT <u>G</u> CACCAAGGGCCCGCCGAG	CCTTGGT <u>G</u> CTCTGTCCGTGATCGCTGG
c.1814C>G	p.Thr605Ser	AAGGACCGCAG <u>C</u> CAAGGGCCCGCCGAGGC	CCCTTGT <u>G</u> CGGCTCTTCCGTGATCGC
c.1910T>G	p.Leu637Arg	AGAAGC <u>G</u> GGACTTCTGGTGAATATCTACATGCAGC	CAGGAAGTCC <u>G</u> CTTCTTCCATGGACAAGACCTGC
c.1988A>G	p.Glu663Gly	GGGGCCAAAG <u>G</u> GCCGGAGCCGGCCGCGCC	TCCGGCCTT <u>T</u> GGCCCCAAGTAGGCTCCG
c.2101_2103delITC	p.Phe701del	CCAGAAGA <u>A</u> C...TCGGCGCCCCGGCCGCG	CGCCGA...GTTCTTGGCCCGTGAGCTG
c.2209G>A	p.Gly737Ser	GGACCAC <u>A</u> GTCTCCCTGGTGCATCCCG	CCAGGGAG <u>C</u> TGTGTCCCCACGGGGGAG
c.2252C>T	p.Ser751L eu	CGAGCGG <u>T</u> GTGCTGCCCTACGGCGGG	CGGACAGC <u>A</u> ACCCTCGTGGCGAGGGGG
c.2264A>G	p.Tyr755Cys	TGTCCGCT <u>C</u> GGCGGGGGCAACCGCGC	CCCCCGC <u>A</u> GCGGACAGCGACCGCTCG
c.2266G>A	p.Gly756Ser	TCCGCCTAC <u>A</u> GGCGGGGGCAACCGCGCCA	CCCCCGC <u>T</u> GTAGGCGGACAGCGACCGCTC
c.2279G>A	p.Arg760His	GGCAACC <u>A</u> CGCCAGCATGGAGTTCTGCG	ATGCTGGC <u>G</u> TGGTTCGCCCGCGGTAGGC
c.2312C>T	p.Thr771Ile	CAGGAGGACAT <u>C</u> CCGGGCTGCAGGCCCC	CCCCGG <u>A</u> TGTCTCTGCGCGAGAACTC
c.2339A>C	p.Asn780Thr	AGGGGAC <u>C</u> CTGCGGGACAGCGACACGTC	GTCCCGCAGG <u>G</u> TCCCTCGGGGGCCCTGC
c.2377G>C	p.Val793Leu	TCCCGTCC <u>T</u> TGGACCACGAGGAGCTGGAGC	GTGGTCCAG <u>G</u> GACGGGATGGAGATGGACGTG
c.2560C>T	p.Arg854Cys	CCCCGCCA <u>T</u> GCTCGGCCACCGCGGAGG	GGCCGAGC <u>A</u> TGGCGGGGGCCCGCACGG
c.2570C>T	p.Thr857Ile	CGGCCA <u>T</u> CGGCGAGGGTCCCTTTGGTGA	ACCCTCGCC <u>A</u> TGGCCGAGCGTGGCGGG
c.2572G>A	p.Gly858Ser	GCCACC <u>A</u> GCGAGGGTCCCTTTGGTGACG	GGACCTCGC <u>T</u> GTTGGCCGAGCGTGGCGG

Table S3. Manual patch clamp and high throughput functional results.

See Excel file:

<https://digitalhub.northwestern.edu/files/151bf778-d6be-49a9-b4b7-6f75ba9c0e2e>

Table S4. Functional properties of CHO-Q3 cells electroporated with homozygous variant KCNQ2 cDNA recorded under control conditions.

See Excel file:

<https://digitalhub.northwestern.edu/files/5e63c462-56c3-4cf5-9468-f4e2e20af714>

Table S5. Functional properties of CHO-Q3 cells co-electroporated with heterozygous variant plus wild type KCNQ2 cDNA recorded under control conditions.

See Excel file:

<https://digitalhub.northwestern.edu/files/33bd2d57-21b4-4d10-aa59-07e1d8551ada>

Table S6 Functional properties of CHO-Q3 cells electroporated with homozygous variant KCNQ2 cDNA recorded following exposure to retigabine.

See Excel file:

<https://digitalhub.northwestern.edu/files/ffb6d08c-4af6-41f9-b319-26b593987e01>

Table S7. Functional properties of CHO-Q3 cells co-electroporated with heterozygous variant plus wild type KCNQ2 cDNA recorded following exposure to retigabine.

See Excel file:

<https://digitalhub.northwestern.edu/files/b2c17941-0e0e-4255-b0fd-cf34f4628ea6>