Supplementary Appendix

This appendix has been provided by the authors to give readers additional information about their work.

Supplement to: Hsu C, Yang W, Parikh RV, et al. Race, genetic ancestry, and estimating kidney function in CKD. N Engl J Med 2021;385:1750-60. DOI: 10.1056/NEJMoa2103753

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Table S1. Baseline CRIC study visit measurement details.

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Table S2. Baseline characteristics of development and validation datasets.

Table S3. Root mean squared errors and precision of estimated GFRs for all serum creatinine (SCr)-based and Cystatin C models reported in manuscript Tables 2 and 4.

Abbreviations: CI: confidence interval; IQR, interquartile range; iGFR, ¹²⁵I-iothalamate glomerular filtration rate; eGFR, estimated glomerular filtration rate Models derived on a development subset of 844 (67%) participants and performance of estimated GFR reported on a validation set of 404 (33%) participants. All 95% confidence intervals correspond to the 2.5th and 97.5th percentile values from 1000 bootstrapped samples of the validation set. Root mean square error (RMSE) is calculated in the validation set on the same scale as the measured and estimated GFR.

Table S4. 10-fold cross validation metrics for manuscript Tables 2 and 4 using the full study sample (N=1248) instead of split-sample development and validation.

Abbreviations: SCr, serum creatinine; CI: confidence interval; iGFR, ¹²⁵I-iothalamate glomerular filtration rate; eGFR, estimated glomerular filtration rate; P30, percent of estimated GFR within 30% of iothalamate GFR; P10, percent of estimated GFR within 10% of iothalamate GFR

Models metrics reported using predictions from the combined validation folds from 10-fold cross validation, corresponding to the full study sample size (N=1248).

Table S5. Model performance metrics corresponding to manuscript Tables 2 and 4 using models with interaction terms between self-reported race or African ancestry and serum creatinine and cystatin C.

Abbreviations: SCr, serum creatinine; CI: confidence interval; iGFR, ¹²⁵I-iothalamate glomerular filtration rate; eGFR, estimated glomerular filtration rate; P30, percent of estimated GFR within 30% of iothalamate GFR; P10, percent of estimated GFR within 10% of iothalamate GFR Models derived on a development subset of 844 (67%) participants and performance of estimated GFR reported on a validation set of 404 (33%) participants. All 95% confidence intervals correspond to the 2.5th and 97.5th percentile values from 1000 bootstrapped samples of the validation set.

† In model for urine creatinine, both urine creatinine and iGFR are natural log-transformed.

‡Models for CrCl/iGFR and CrCl-iGFR are only adjusted for age and sex and not iGFR.

Abbreviations: BIA, bioelectric impedance analysis; BSA, body surface area; CrCl, creatinine clearance; iGFR, ¹²⁵I-iothalamate glomerular filtration rate.

Figure S1. Assembly of study sample from the Chronic Renal Insufficiency Cohort (CRIC) study.

Figure S2. Conceptual approach to evaluating associations of self-reported race, genetic ancestry, serum creatinine, serum cystatin C and measured glomerular filtration rate in adults with chronic kidney disease.

1 Is it possible to estimate GFR just as well using genetically-defined ancestry instead of self-reported race in adults with mild-to-moderate chronic kidney disease?

2 Are genetic ancestry or self-reported Black race independently associated with components of creatinine production, secretion or excretion that contribute to variations in SCr levels independent of GFR? Can these variables be used to attenuate the race or ancestry coefficient in GFR estimating equations?

3 Are genetic ancestry or self-reported Black race necessary for GFR estimation when using serum cystatin C, and is a cystatin C-only equation without race or ancestry similar in accuracy to a serum creatinine-based equation that includes race or ancestry?

Figure S3. Distribution of % genetic ancestry by self-reported Black race (top figure) vs. non-Black race (bottom figure) in the study population.

Figure S4. Measured GFR vs. estimated GFR in the validation set using different estimating equations.

Each dot represents one individual in the validation set. The solid lines are the linear fit between the measured GFR and estimated GFR stratified by self-reported race. A deviation from the dotted diagonal line indicates bias associated the GFR estimating equation.

Supplementary Methods. Methods for CRIC Ancestry estimation.

1 A general admixture model

To begin with, we introduce a general admixture model. Consider a genotype dataset $G = \{g_{ij}\}\$ with genotypes at J single nucleotide polymorphisms (SNPs) from I unrelated individuals. These individuals are drawn from an admixed population with contributions from K postulated ancestral populations. Population k contributes a fraction q_{ik} of individual i's genome. The effect allele at the SNP j has frequency f_{kj} in population k for $k = 1, \ldots, K$ and g_{ij} is the observed effect allele counts of SNP j of individual i. Here we only consider the bi-allelic genetic variants so the g_{ij} will take values from $\{0,1,2\}$. Both the q_{ik} and the f_{kj} are unknown.

The observed dataset can be modeled by a mixture model with parameters $\{q_{ik}\}\$ and $\{f_{kj}\}\$. Under the assumption that the genotypes of individuals are formed by the random union of gametes, g_{ij} is modeled by the binomial distribution $Binom(2, \sum_k q_{ik} f_{kj})$ with

$$
P(g_{ij} = c) = \binom{2}{c} \left[\sum_{k} q_{ik} f_{kj} \right]^c \left[\sum_{k} q_{ik} (1 - f_{kj}) \right]^{2 - c}, \quad c = 0, 1, 2. \tag{1}
$$

Therefore for independent individuals and genetic variants in linkage equilibrium, the loglikelihood of the entire sample ${g_{ij}}$ (up to an additive constant) is

$$
L(Q, F) = \sum_{i} \sum_{j} \left\{ g_{ij} \ln[\sum_{k} q_{ik} f_{kj}] + (2 - g_{ij}) \ln[\sum_{k} q_{ik}^{t} (1 - f_{kj})] \right\}
$$
(2)

where
$$
0 \le f_{kj} \le 1
$$
, $\sum_{k=1}^{K} q_{ik} = 1$. (3)

The parameter matrices $Q = \{q_{ik}\}\$ and $F = \{f_{kj}\}\$ have dimensions $I \times K$ and $K \times J$. An efficient algorithm for estimating the parameters is implemented by the software Admixture [Alexander and Lange, 2011].

$\overline{2}$ **Projection Analysis**

A number of large genome-wide datasets of human populations such as 1000 Genomes Project [Consortium et al., 2015] are now publicly available. Since these large datasets summarize worldwide human population structure, we use them as reference panels in combination with the study sample to estimate individual ancestry of study sample. This function is implemented by the projection command in Admixture Shringarpure et al. 2016 .

Specifically, we first do the admixture analysis on the 1000 Genomes Project data and estimate effect allele frequency $\hat{F} = \{\hat{f}_{kj}\}\$ for each learned clusters $k = 1, \dots, K$ based on (2). Since the population ancestry information of each learned cluster is known from the reference data, \hat{F} can be viewed as the learned population structure. Then with $\hat{F} = \{\hat{f}_{ki}\}\$ estimated from reference data and a set of CRIC study genotype data $\{g_{ij}^{(s)}\}$, we estimate the individual ancestry of CRIC data by maximizing following function,

$$
L(Q^{(s)}; \hat{F}, \{g_{ij}^{(s)}\}) = \sum_{i} \sum_{j} \left\{ g_{ij}^{(s)} \ln[\sum_{k} q_{ik}^{(s)} \hat{f}_{kj}] + (2 - g_{ij}^{(s)}) \ln[\sum_{k} q_{ik}^{(s)} (1 - \hat{f}_{kj})] \right\}
$$
(4)

where $Q^{(s)} = \{q_{ik}^{(s)}\}$ are the ancestry proportion parameters for each CRIC participants and $\sum_{k=1}^{K} q_{ik}^{(s)} = 1$. The optimized function (4) has almost the same form as (2) except that \hat{F} is fixed here.

The projection approach has many advantages when a good reference dataset is available, as pointed by Shringarpure et al. [2016]. First, when a new dataset is strongly unbalanced in its distribution of populations, the accuracy of ancestry inference may be affected by the unbalance Shringarpure and Xing, 2014, while the projection method avoids this problem. Besides, the meaning of each cluster in the study samples is suggested from the reference panel. Finally, it can be applied to estimate individual ancestry for a set of related individuals in the study samples without excluding related samples.

$\boldsymbol{3}$ **Implement Details**

Pre-processing: For the CRIC study data, we first removed SNPs and individuals that did not meet standard quality-control criteria: SNPs with missing rates > 0.05 , minor allele frequency < 0.01 , with no founder genotypes observed were excluded as well as the individuals with missing rates > 0.1 . In addition, we only kept the genotype data of bi-allelic variants on the chromosome 1-22. SNPs were further pruned for linkage disequilibrium (LD) with a window size of 50 SNPs, a step size of 10 SNPs, and a R^2 threshold of 0.1 using PLINK 1.9. Besides, we removed SNPs that did not intersect between the two datasets and palindromic $(A/T, G/C)$ SNPs. Finally there are 102317 variants, 3635 individuals left for the target data and 2503 individuals for the 1KGP reference data.

Then, the following steps were taken to estimate the individual ancestry of CRIC data:

- From the 1000 Genome Project data, get $\hat{F} = \{\hat{f}_{kj}\}$ given pre-determined $K = 5$.
- For the CRIC study data, estimate individual ancestry $\hat{Q}^{(s)}$ based on the estimated $\hat{F} = \{\hat{f}_{kj}\}\$ and pre-determined K according to (4).
- The meaning of $\hat{Q}^{(s)}$ is acquired from the known ancestry information of reference data.

A brief discussion on the choice of K : based on previous studies, it is reasonable to choose $K = 5$ as our final input parameter, corresponding to the 5 super-population in reference data, African, American, European, East Asian and South Asian. Besides, this choice is also verified by comparing the log-likelihood value of fitted function (4) with different value of K. In Figure 1, it clearly shows that $K = 5$ is a sensible modeling choice for CRIC data.

Figure 1: Log-likelihood value of fitted model for CRIC data using K from 2 to 8. For each value of K, first we get estimated \hat{F} from the reference data and then estimate individual ancestry $\hat{Q}^{(s)}$ of CRIC study samples based on the log-likelihood function (4). An elbow in the curve can be seen at $K = 5$ and for $K > 5$, the log-likelihood values are similar to each other. Therefore, $K = 5$ was decided to be an appropriate number of population for the CRIC data.

References for CRIC Ancestry estimation

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