

Supplementary Information

Peripheral blood CD4⁺CCR6⁺ compartment differentiates HIV-1 infected or seropositive elite controllers from long-term successfully treated individuals

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Supplementary Table 1: Treatment regimen

Treatment regimen, N (%)	ART
3TC/DTG	1 (1.8)
ABC/3TC/DRV/ritonavir	3 (5.5)
ABC/3TC/DTG	20 (37)
ABC/3TC/EFV	5 (9.3)
ABC/3TC/NVP	2 (3.7)
ABC/3TC/RPV	4 (7.4)
CAB/RPV*	1 (1.8)
DRV/COB/DTG	1 (1.8)
RAL/3TC/EFV	1 (1.8)
TAF/FTC/DTG	5 (9.3)
TAF/FTC/EFV	1 (1.8)
TDF/FTC/DTG	2 (3.7)
TDF/FTC/EFV	3 (5.5)
TDF/TFC/EVG/Cob	1 (1.8)
TDF/FTC/RPV	4 (7.4)

*3TC, Lamivudine; ABC, Abacavir; *CAB/RPV, Cabotegravir given together with rilpivirine intramuscularly as long-acting drugs; COB, Cobicistat; DRV, Darunavir; r, ritonavir (boosterdose); DTG, Dolutegravir; EFV, Efavirenz; EVG, Elvitegravir; FTC, Emtricitabine; N, Number; NVP, Nevirapine; RAL, Raltegravir; RPV, Rilpivirine; TDF, Tenofovir Disoproxil; TAF, Tenofovir Alafenamide*

Supplementary Table 2: Patient characteristics of the longitudinal data

Parameter	T1	T2	P-values
<i>N</i>	10	10	NA
Gender, Female, <i>N</i> (%)	2 (20)	2 (20)	NA
At sampling			
Age in years, mean (SD)	52 (7.93)	55 (7.93)	0.4087 [#]
CD4 count (cells/ μ L); median (IQR)	510 (460-557.5)	510 (370-700)	0.9847*
CD8 count (cells/ μ L); median (IQR)	500 (425-640)	430 (345-655)	0.7039*
CD4:CD8 ratio; median (IQR)	1.084 (0.7811-1.31)	1.214 (0.8712-1.479)	0.5490*
Years on treatment; median (IQR)	19 (16.75-23.25)	22 (19.75-26.25)	0.1152*
Viral Load; Log ₁₀ copies/mL (IQR)	0 (0-0)	0 (0-0)	0.4737*

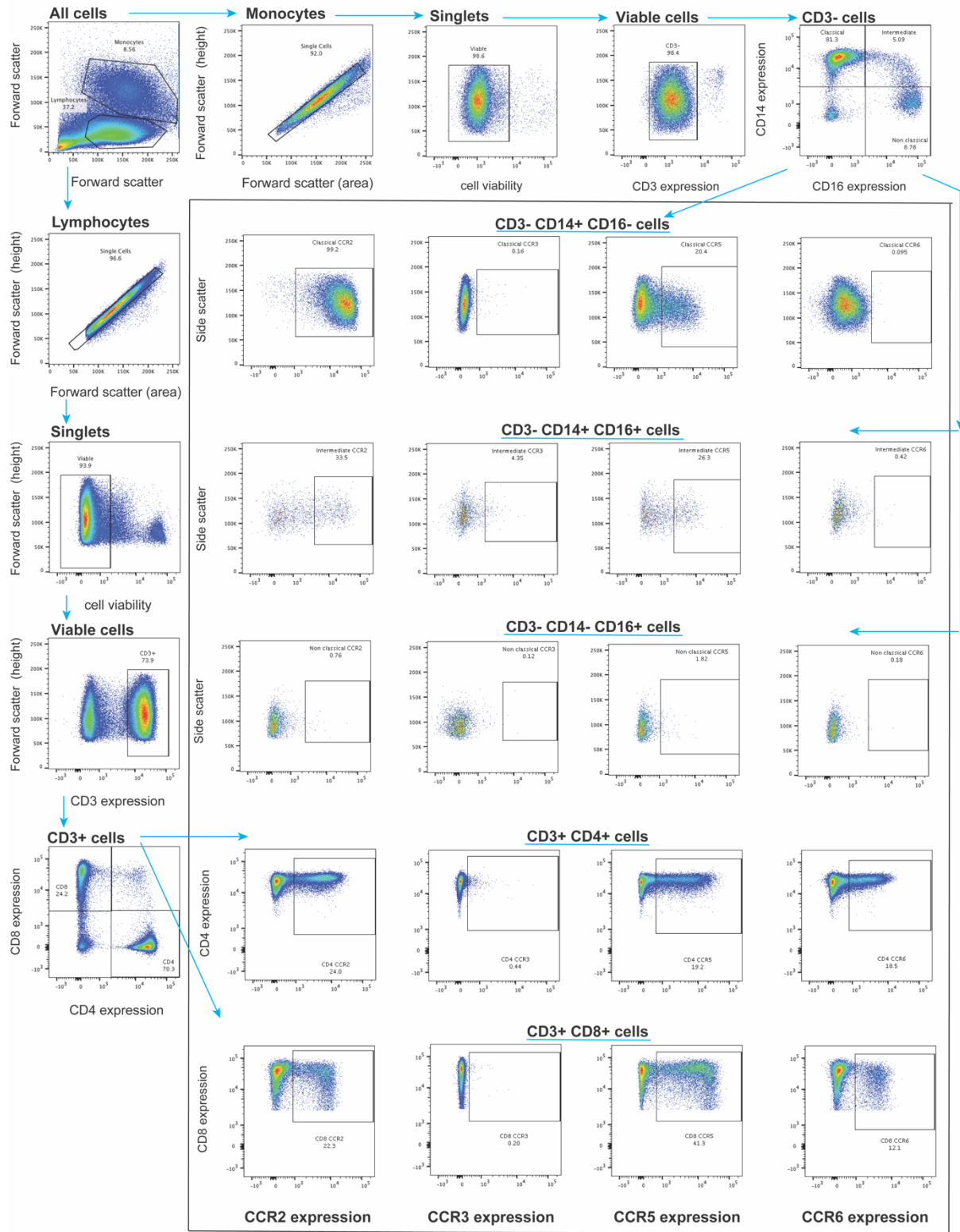
IQR, Interquartile Range; *N*, Number; *NA*, Not Applicable; *SD*, Standard Deviation; *Mann-Whitney U-test; [#]Unpaired t-test

Supplementary Table 3: Primers and probes used for quantification of HIV-1 DNA

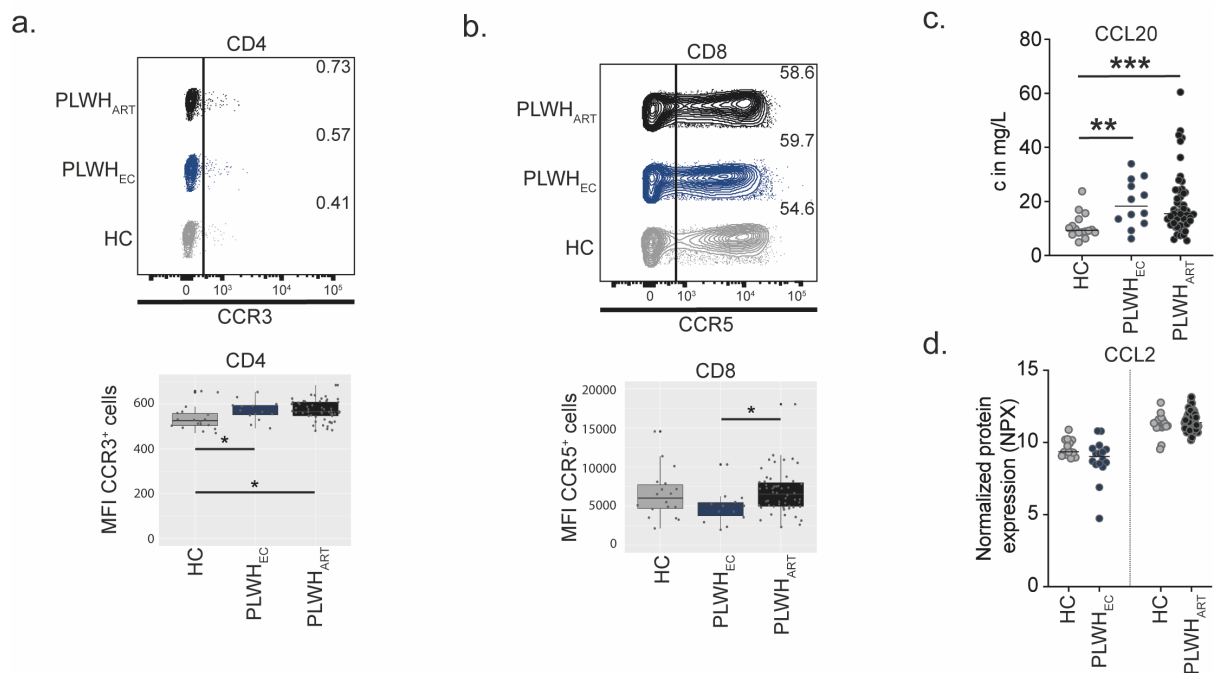
Total HIV DNA quantification	
Beta globin probe	HEX-ATCCACGTTACCTTGCCCCACA-TAM
Beta globin forward	AGGGCCTCACCACCAACTT
Beta globin reverse	GCACCTGACTCCTGAGGAGAA
HIV (HXB2) probe	FAM-AAGTAGTGTGTGCCCGTCTG-MGBEQ
HIV (HXB2) forward	GCCTCAATAAAGCTTGCCTTGA
HIV (HXB2) reverse	GGCGCCACTGCTAGAGATTTT
FAM coupled probes + primers	
RPP30 3' (ctrl) probe	/56-FAM/AGAGAGCAA/ZEN/CTTCTTCAAGGGCCCC/3IABkFQ/
RPP30 3' forward	GTGTGAGTCAATCACTAGACAGAA
RPP30 3' reverse	AAACTGCAACAACATCATAGAGC
2-LTRcircle probe	/56-FAM/ACACTACTT/ZEN/GAAGCACTCAAGGCAAGCTTT/3IABkFQ/
2-LTRcircle forward	AACTAGGGAACCCACTGCTTAAG
2-LTRcircle reverse	TCCACAGATCAAGGATATCTTGTC
Pol probe	/56-FAM/AAGCCAGGA/ZEN/ATGGATGGCC/3IABkFQ/
Pol forward	GCACTTTAAATTTCCCATTAGTCCTA
Pol reverse	CAAATTTCTACTAATGCTTTTATTTTTTC
Psi (Ψ) probe	/56-FAM/TTTTGGCGT/ZEN/ACTCACCAGT/3IABkFQ/
Psi (Ψ) forward	CAGGACTCGGCTTGCTGAAG
Psi (Ψ) reverse	GCACCCATCTCTCTCCTTCTAGC
HEX coupled probes + primers	
RPP30 (ctrl) probe	/5HEX/CTGACCTGA/ZEN/AGGCTCT/3IABkFQ/
RPP30 forward	GATTTGGACCTGCGAGCG
RPP30 reverse	GCGGCTGTCTCCACAAGT
5'LTR probe	/5HEX/AAGTAGTGT/ZEN/GTGCCCGTCTG/3IABkFQ/
5'LTR forward	GCCTCAATAAAGCTTGCCTTGA
5'LTR reverse	GGCGCCACTGCTAGAGATTTT
Env7781 probe	/5HEX/CCTTGGGTT/ZEN/CTTGGGA/3IABkFQ/
Env7781 forward	AGTGGTGCAGAGAGAAAAAAGAGC
Env7781 reverse	GTCTGGCCTGTACCGTCAGC
Uncoupled probes	
Env 7781 hypermutation probe	CCTTAGGTTCTTAGGAGC/3IABkFQ/

Supplementary Table 4: TMT11plex labelling distribution for CCR6⁺ and CCR6⁻ samples

B1		B2		B3	
ART01 +	126	ART03 +	127N	ART05 +	131N
ART01 -	127N	ART03 -	128C	ART05 -	130N
ART02 +	127C	ART04 +	129N	ART06 +	129N
ART02 -	128N	ART04 -	130C	ART06 -	128N
EC01 +	128C	EC03 +	131N	EC05 +	127N
EC01 -	129N	EC03 -	130N	EC05 -	126
EC02 +	129C	EC04 +	129C	EC06 +	130C
EC02 -	130N	EC04 -	128N	EC06 -	127C
HC02 +	130C	HC01 +	127C	HC03 +	128C
HC02 -	131N	HC01 -	126	HC03 -	129C
Norm	131C	Norm	131C	Norm	131C

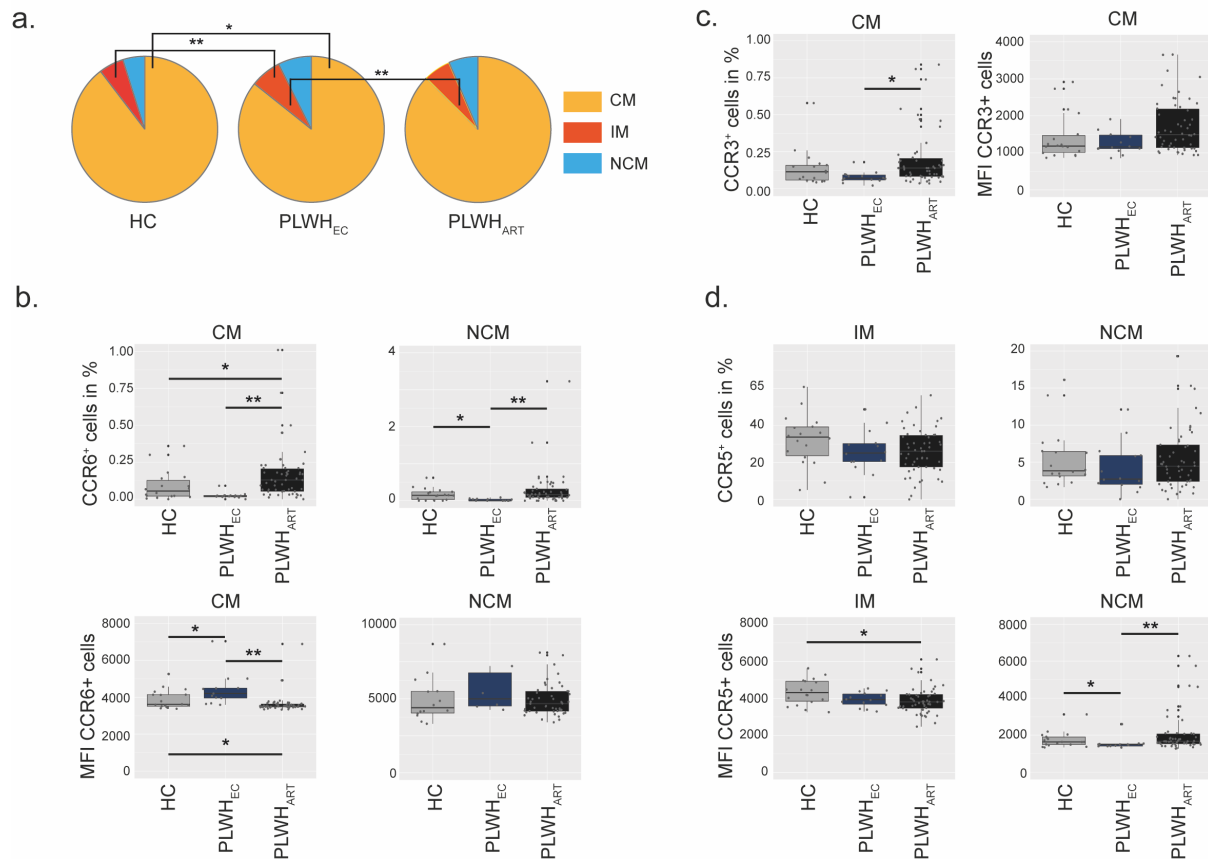


Supplementary Fig. 1: Gating strategy for CCR2, CCR3, CCR5, and CCR6 flow cytometry detection



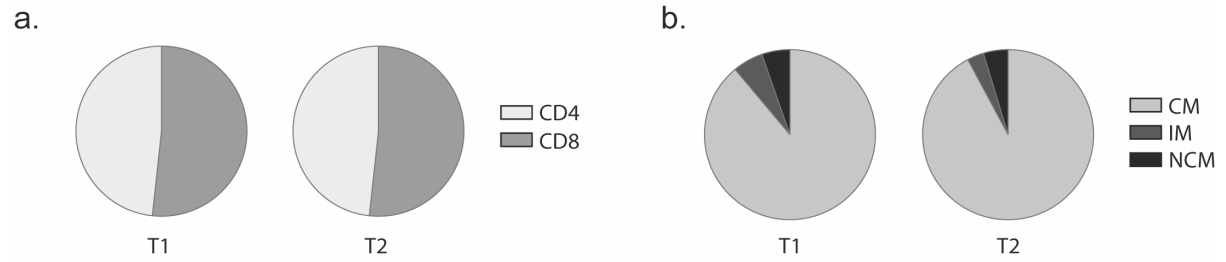
Supplementary Fig. 2: Chemokine receptor expression on lymphocytic cell populations.

Flow cytometry detection in lymphocytic cell populations in PLWH_{EC} ($n=14$), PLWH_{ART} ($n=54$), and HC ($n=18$). **a** Expression of CCR3 in CD4⁺ T cells. The contour plot is a median representative sample of % of cells expressing CCR3, and the boxplot shows the median fluorescence intensity (MFI). **b** Expression of CCR5 in CD8⁺ T cells. The contour plot is a median representative sample of % of cells expressing CCR5, and the boxplot shows the median MFI. **c** Chemokine detection of CCL20 in plasma of PLWH_{EC} ($n=12$), PLWH_{ART} ($n=49$), and HC ($n=16$). **d** Normalized expression profile of CCL2 plasma levels in PLWH_{EC} ($n=14$), PLWH_{ART} ($n=51$), and HC ($n=17$). Chemokine detection by ELISA was performed in technical duplicates. Statistical significance was determined using two-tailed Mann-Whitney U-test (significance level $p < 0.05$, with $* < 0.05$, $** < 0.001$) and represented with median and 95% CI.



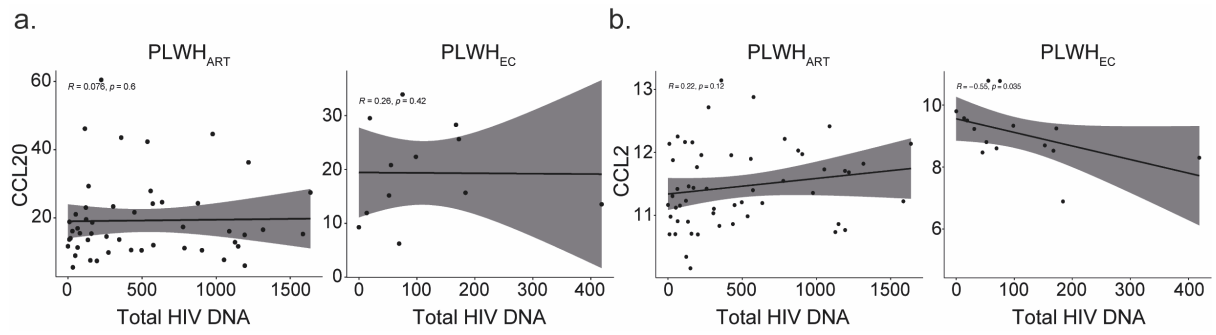
Supplementary Fig. 3: Chemokine receptor expression on monocytic cell population.

Flow cytometry detection in monocytic cell populations (classical monocytes (CM); CD14⁺CD16⁻, intermediate monocytes (IM); CD14⁺CD16⁺, and non-classical monocytes (NCM); CD14⁻CD16⁺) in PLWH_{EC} ($n=14$), PLWH_{ART} ($n=54$), and HC ($n=18$). **a** Frequency distribution of CM, IM, and NCM. **b** Expression of CCR6 in CM and NCM cells. The upper row shows % of cells expressing the receptor and the bottom row shows the median fluorescence intensity (MFI). **c** Expression of CCR3 in CM. The left boxplot shows % of cells expressing the receptor and the right boxplot shows the MFI. **d** Expression of CCR5 in IM and NCM. The upper row shows % of cells expressing the receptor and the bottom row shows the MFI. Statistical significance was determined using two-tailed Mann-Whitney U-test (significance level $p < 0.05$, with * < 0.05 , ** < 0.001) and represented as pie charts or with median using 95% CI.



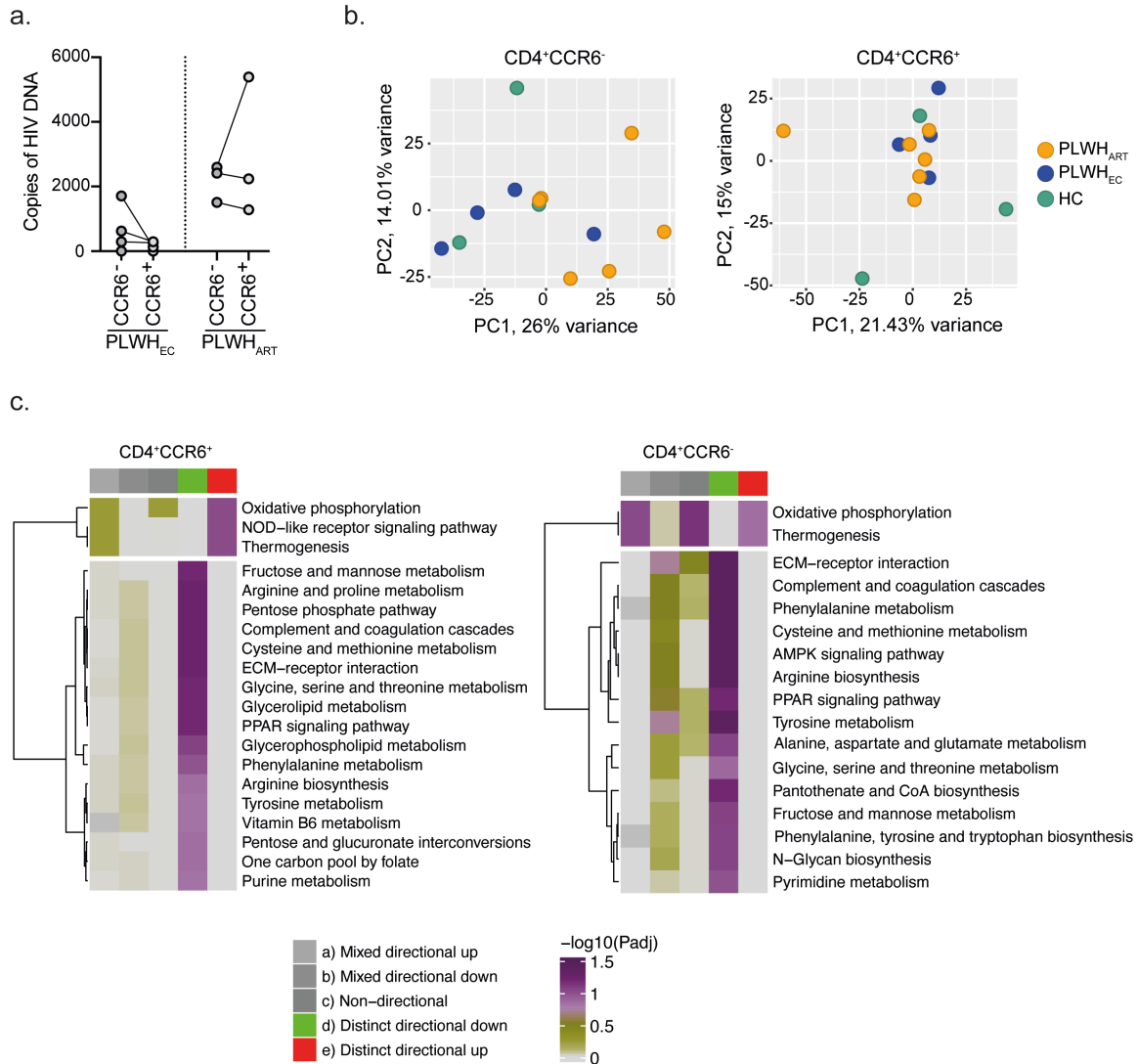
Supplementary Fig. 4: Frequency of cell populations in longitudinal data of PLWH_{ART}.

Flow cytometry analysis of longitudinal dataset of HIV-1 infected individuals (timepoint 1 (T1), $n=10$ and timepoint 2 (T2), $n=10$). **a, b** Frequency of lymphocytic cell populations ($CD4^+$ and $CD8^+$) (**a**) and monocyte cell populations (classical monocytes (CM); $CD14^+CD16^-$, intermediate monocytes (IM); $CD14^+CD16^+$, and non-classical monocytes (NCM); $CD14^-CD16^+$) (**b**) in T1 and T2.



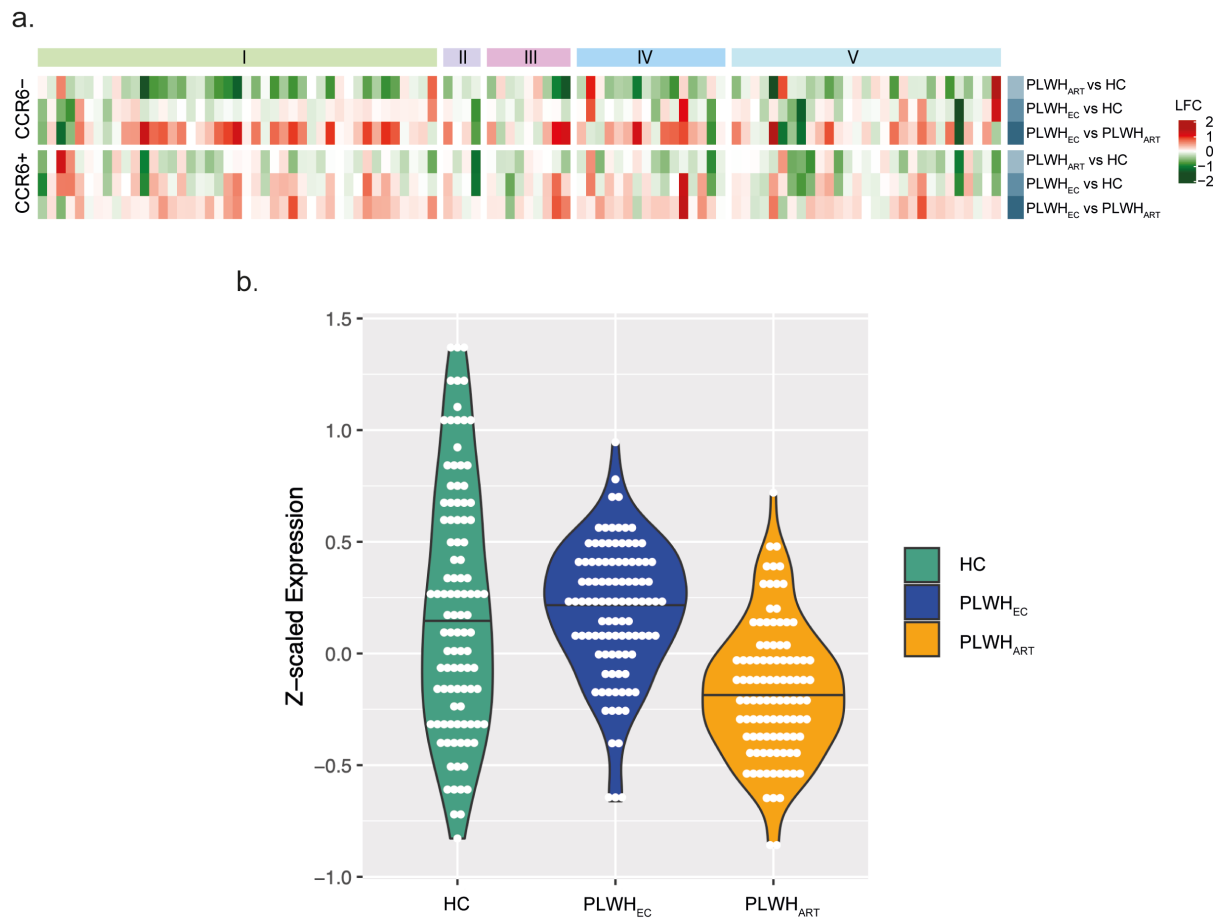
Supplementary Fig. 5: Correlation of chemokines CCL20 and CCL2 to total HIV-1 DNA.

a, b Correlation between total HIV-1 DNA and plasma levels of CCL20 (**a**) and CCL2 (**b**) in PLWH_{ART} and PLWH_{EC}. Statistical significance was tested using Spearman's correlation.



Supplementary Fig. 6: Analysis of CD4⁺CCR6⁺ and CD4⁺CCR6⁻ cell populations in PLWH_{EC} compared to PLWH_{ART}.

a HIV-1 DNA quantified in CD4⁺CCR6⁺ and CD4⁺CCR6⁻ cells from PLWH_{EC} ($n=5$) and PLWH_{ART} ($n=3$). Data is represented as paired datapoints where line is drawn within between the two populations in the same patient. The quantification was performed in technical duplicates. **b** Two-dimensional visualization of PC1 and PC2 of principal components analysis using CD4⁺CCR6⁻ and CD4⁺CCR6⁺ samples. **c** Heatmap representation of significant pathways found differentially regulated in CD4⁺CCR6⁺ PLWH_{EC} compared to CD4⁺CCR6⁺ PLWH_{ART} and in CD4⁺CCR6⁻ PLWH_{EC} compared to CD4⁺CCR6⁻ PLWH_{ART} using KEGG 2021. Color gradient is corresponding to the negative log scaled adjusted p-values. Each column represents p-values of various directionality classes, calculated for the pathways. Non-directional p-values are calculated based on gene-level statistics regardless of the direction of expression. Mixed directional up and mixed directional down p-values are calculated using the subset of the gene statistics that are up-regulated and down-regulated, respectively. Distinct directional up and distinct directional down p-values are calculated from gene statistics with expression direction.



Supplementary Fig. 7: Abundance of OXPPOS proteins detected from the proteomics analysis of CD4⁺CCR6⁺ and CD4⁺CCR6⁻ cells.

a Heatmap visualization of regulation of OXPPOS pathway proteins detected in the proteomics experiment. Color gradient represents log₂ scaled fold change values. Column annotation denotes each of the five complexes of OXPPOS pathway. Row annotation denotes each of the pair-wise analysis performed. **b** Cumulative Z-scale expression of the OXPPOS genes.