nature portfolio

Corresponding author(s):	Jianquan Liu
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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	nfirmed
	x	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	x	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	×	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
X		A description of all covariates tested
	x	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	×	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	×	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x		Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
		Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection has been described in the Methods and Supplementary Information. No software has been used for data collection

Software used are listed as follows: Guppy (v3.2.2), Trimmomatic (v0.36), Jellyfish (v2.2.9), NextDenovo (v2.0), SmartDenovo (v1.0.0), Pilon (v1.22), bowtie2 (v2.3.2), HiC-Pro (v3.1.0), LACHESIS (v1), RepeatMasker (v4.0.7), RepeatModeler (v1.0.10), PASA pipeline (v2.1.0), Augustus (v3.2.3), HISAT2 (v2.1.0), TBLASTN (v2.2.31), GeneWise (v2.4.1), EVidenceModeler (v1.1.1), BLAST+ (v2.2.31), InterProScan (v5.25-64.0), Blast2GO (v4.1), BUSCO (v3), MCScanX (v1), Circos (v0.69), OrthoFinder (v2.3.8), PRANK (v170427), RAXML (v8.2.12), LAST (v963), R (v4.1.2), ASTRAL (v5.6.3), DendroPy (v4.2.0), BWA (v0.7.17), SAMtools (v1.5), GATK (v3.8), HyDe (v0.4.3), and Dsuit (v0.4).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Data analysis

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

All the sequencing data have been deposited in the National Genomics Data Center (https://ngdc.cncb.ac.cn). The sequencing data for the de novo genome of Carpinus viminea (including the ONT long reads, Illumina reads of WGS, Hi-C reads, and transcriptomes) have been deposited under the accession number PRJCA005724. The Hi-C sequencing data for Ostrya rehderiana have been deposited under the accession number PRJCA005717. The resequencing data for all

individuals have been deposited under the accession numbers PRICA003130 and PRICA005842. The genome assemblies and annotations of C. viminea and O. rehderiana have been uploaded to figshare (https://doi.org/10.6084/m9.figshare.14988777). The SwissProt and TrEMBL databases used in this study are available from https://www.uniprot.org. KEGG PATHWAY database can be available from https://www.kegg.jp. There is no restriction on data availability. Source data are provided with this paper.

Field-specific reporting					
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	w that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection. Behavioural & social sciences * Ecological, evolutionary & environmental sciences				
Life sciences For a reference copy of the docum	■ Behavioural & social sciences ■ Ecological, evolutionary & environmental sciences ent with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf				
To a reference copy of the docum	ent with an sections, see <u>nature.com/documents/in-reporting-summary nat.pur</u>				
Ecological, e	volutionary & environmental sciences study design				
All studies must disclose or	these points even when the disclosure is negative.				
Study description	We generated a chromosome-level reference genome of Carpinus viminea of sect. Carpinus and re-sequenced genomes of 47 individuals from the genera Carpinus, Ostrya, and Ostryopsis (as outgroup) in Betulaceae. Our integrated analyses together suggested that sect. Distegocarpus with three species originated through HHS during the early divergence between Carpinus and Ostrya.				
Research sample	Species names, population codes, sample locations, and sample sizes for whole genome resequencing of individuals are described in Supplementary Data 2. All were adult individuals collected in the wild. Within each population, sampling was performed randomly to ensure representative samples.				
Sampling strategy	We collected a wild plant of Carpinus viminea for de novo genome sequencing and a wild plant of Ostrya rehderiana for Hi-C sequencing. We collected 47 individuals (including a total of 21 species) from three different lineages (and outgroup) for population genomic re-sequencing: 10 individuals (from 10 species) of sect. Carpinus, 27 individuals (from 3 species) of sect. Distegocarpus, 7 individuals (from 7 species) of Ostrya, and 3 individuals (from 1 species) of Ostryopsis as outgroup. Except for the outgroup, all the sampled individuals were selected from different populations (one individual per population). The samples of sect. Carpinus and Ostrya covered all the major species of these two lineages. The samples of sect. Distegocarpus covered all 3 species in this lineage and almost the whole of their distributions in the wild.				
Data collection	Samples were collected in the wild by C.C. Sampling sites were positioned by GPS and recorded in a laptop. DNA was prepared by Z.W. Genome sequencing data were generated by different types of sequencers.				
Timing and spatial scale	Spatial scale of sampling for the study is described in Supplementary Data 2 (spanning USA, Slovenia, Russia, and China). All samples were collected between 2016-2020.				
Data exclusions	No data were excluded.				
Reproducibility	All sequencing for genome assembly were more than 100 fold coverage. For each of the 47 re-sequenced samples, an average of more than 25 fold coverage clean bases were sequenced. All phylogenetic analyses were performed with 100 bootstraps.				
Randomization	ation Samples were randomized for collection.				
Blinding	Blinding is not applicable in our study because it does not involve subjects which receive different treatments.				
Did the study involve field	d work? Yes X No				
	r specific materials, systems and methods				
	authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, evant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.				
Materials & experime	ental systems Methods				
n/a Involved in the study	<u> </u>				
Antibodies Fukaryotis sell lines	ChIP-seq				
Eukaryotic cell lines Palaeontology and a	yotic cell lines X Flow cytometry ontology and archaeology MRI-based neuroimaging				
	entology and archaeology MRI-based neuroimaging lals and other organisms				
Clinical data	Clinical data				
Dual use research o	f concern				