Supporting Information

The pattern of apolipoprotein A-I lysine carbamylation reflects its lipidation

state and the chemical environment within human atherosclerotic aorta

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GB subset
(n=117)
60.7(55.6-66.5)
45.6
131.0(120.0-144.0)
5.5(5.2-5.9)
38.8(32.0-48.0)
105.0(87.0-123.0)
171.4(151.9-194.3)
97.5(77.3-140.8)
80.4(70.3-91.4)
1.74(0.86-4.57)

 Table S1. Demographics and laboratory values of the clinical cohort subset (GeneBank, GB)

 used in the analysis of protein bound HCit content in plasma.

Clinical and laboratory characteristics of GB subjects (n=117) for whom protein-bound HCit was quantified in plasma proteins. Subjects included were confirmed to be without known cardiovascular or metabolic diseases, and with normal renal function, as described under Experimental Procedures. Plasma was analyzed for protein bound HCit content using stable isotope dilution LC/MS/MS methods as described. Values shown are median (interquartile range) for continuous variables, and percent for dichotomous variables.

Abbreviations: Systolic blood pressure (SBP), hemoglobinA1C (HgbA1C), high density lipoprotein cholesterol (HDLc), low density lipoprotein cholesterol (LDLc), estimated glomerular filtration rate (eGFR), C-reactive protein (CRP)

Table S2. Age and gender of subjects from whom human atherosclerotic aortic samples were used in recovery of apoA-I for proteomic analysis.

Human aorta sample	Age	Gender	Sequence coverage %
1	79	Male	93
2	76	Female	93
3	74	Male	93
4	81	Male	90
5	76	Male	90
6	75	Female	93
7	73	Female	92
8	68	Female	90
9	82	Female	90
10	71	Male	90

Human aortic tissue was obtained as discarded material at time of organ removal from transplant recipients. Atherosclerotic donor tissues were from subjects (n=10) of average age 75yr +/- 4yr. Both males and females were analyzed. ApoA-I was immunoprecipitated from aortic tissue homogenates using a cocktail of anti-apoA-I antibodies using methods shown to result in quantitative recovery of total apoA-I from samples, as described under Experimental Procedures. Recovered apoA-I protein was subjected to proteolysis with GluC and mass spectrometry was performed as described under Experimental Procedures. Proteome Discoverer 2.3 was used to identify peptide sequences and % coverage.

 Table S3. Peptide search parameters for Proteome Discoverer.

Protein Database: ApoA1_His.fasta
- Enzyme Name: GluC
- Maximum Missed Cleavage Sites: 5
Tolerances:
- Precursor Mass Tolerance: 10 ppm
- Fragment Mass Tolerance: 0.6 Da
- Max. Identical Dynamic Modifications Per Peptide: 3
- Max. Dynamic Modifications Per Peptide: 4
Fixed Value PSM Validator:
Maximum Delta Cn: 0.05
Maximum Rank: 0
Protein FDR Validator:
Target FDR (Strict): 0.01
Target FDR (Relaxed): 0.05
Dynamic Modification:
- Oxidation / +15.995 Da (M, W)
- Carbamylation / +43.006 Da (K)
- Acetylation / +42.001 Da (K)

Peak lists were generated with Proteome Discoverer 2.3 (Thermo Fischer Scientific, Waltham, MA) using the parameters listed. The resulting Unified Search Files (*.srf) were searched against the Uniprot FASTA of human apoA-I. Modifications used for the searches included, oxidized methionine (variable), oxidized tryptophan (variable), carbamylated lysine (variable, mass addition 43 Da) and acetylated lysine (variable). GluC peptides with a maximum of 5 missed cleavage sites were allowed in the database searches. Monoisotopic precursor ions were searched with a tolerance of 10 ppm with 0.6 Da for the fragment ions.

Table S4. Abundance and frequency of detection of reference peptide candidates used in the proteomic analysis.

Ref	erence	Sample									
pep can	tide didate	rHDL+M	-0	rHDL+CN	0-	apoA-I+M	PO	apoA-I+C	CNO ⁻	Human pla	que
		RPA	DFRP	RPA	DFRP	RPA	DFR	RPA	DFRP	RPA	DFRP
		(%)	(%)	(%)	(%)	(%)	P (%)	(%)	(%)	(%)	(%)
Clu	ster 1 of re	ference p	eptide ca	ndidates							
1A	213DLRQ GLLPVL E ²²³	99±0.2	100	66±12	100	82±0.5	100	87±1	100	98±0.4	100
1B	²⁰⁶ KAKP ALEDLR QGLLPV LE ²²³	1±0.2	100	22±6	100	14±0.9	100	11±1	100	2±0.4	100
1C	¹⁹⁹ HLSTL SEKAKP ALEDLR QGLLPV LE ²²³	<0.001	100	12±6	100	4±0.4	100	2±0.1	100	<0.1±.01	80
Clu	ster 2 of re	ference p	eptide ca	ndidates							
2A	⁷⁹ TEGLR QE ⁸⁵	100±0	100	100±0	100	4±0.3	100	6±0.5	67	79±15	100
2B	⁸¹ GLRQE MSKDLE ⁹¹	<0.001	100	<0.001	100	96±0.3	100	94±0.5	100	9±5.6	100
2C	⁷⁹ TEGLR QEMSK D ⁸⁹	<0.001	100	<0.001	100	<0.001	100	<0.001	100	12±13	100
Clu	ster 3 of re	ference p	eptide ca	ndidates							
3А	¹⁸⁴ NGGA RLAE ¹⁹¹	<0.1±0.1	100	0.3±0.1	100	11±4.6	100	35±1.3	67	68±16	100
3B	¹⁸⁰ ALKE NGGARL AE ¹⁹¹	99±0.1	100	99±0.1	100	89±4.6	100	65±1.2	67	32±16	100
Clu	Cluster 4 of reference peptide candidates										
4A	⁶³ QLGPV TQE ⁷⁰	24±30	100	26±24	67	62±3.5	100	21±2	100	99±0.1	100

4B	⁶³ QLGPV TQEFW DNLEKE 78	<0.001	100	17±15	100	1.0±0.1	100	<0.001	100	<0.1±0.1	70
4C	³⁵ GSALG KQLNLK LLDNWD SVTSTF SKLREQ LGPVTQ EFWDNL EKE ⁷⁸	76±30	100	65±11	100	37±3.5	100	79±2	100	0.5±0.6	90
Clu	ster 5 of ref	ference p	eptide ca	ndidates							
5A	⁷¹ FWDNL E ⁷⁶	-	-	-	-	1±0.1	100	1±0.1	100	-	-
5B	⁷¹ FWDNL EKE ⁷⁸	-	-	-	-	56±0.4	100	55±0.9	100	-	-
5C	⁷¹ FWDNL EKETE ⁸⁰	-	-	-	-	43±0.5	100	44±1	100	-	-

To select optimal apoA-I peptides that might serve as an internal reference peptide for relative quantification of carbamyllysine harboring apoA-I peptides in samples, we used the following criteria: 1) the reference peptide candidate doesn't contain lysine residues and if possible, methionine, histidine, or other nucleophilic or oxidizable residues; 2) the reference peptide candidate should be readily detectable in all sample replicates; 3) the reference peptide candidate should be readily detectable in all samples from all analyses groups (i.e. rHDL + MPO; rHDL + CNO⁻; apoA-I + MPO; apoA-I + CNO⁻; and Human plaque).

RPA (%) = average reference peptide candidate abundance \pm standard deviation = 100 X (peak area of reference peptide candidates / (sum of peak areas of reference peptide candidates in the same peptide cluster, (e.g. peptides 1A, 1B, 1C from reference peptide candidates cluster 1)).

DFRP (%) = detection frequency of reference peptide candidate = $100 \times (number of times reference peptide candidate was observed in replicate samples / total of replicate samples). The DFRP is listed in bold face if the reference peptide was not seen in all replicates and therefore its priority for use was reduced.$

(-) indicates reference peptide candidate not used in analysis.

We observed similar qualitative results, whether using three different single reference peptides, vs. a weighted average of the top three performing reference peptides (e.g. see Figure S3, and S4). However, based on the data analyses shown above, we also noted that only reference peptide D₂₁₃-E₂₂₃ showed optimal characteristics (i.e. it was the only reference peptide observed in all *in vitro* and *in vivo* samples). We therefore elected to use this internal reference peptide as the denominator for all apoA-I quantifications.

Peptide Sequence	m/z	Charge	Δm (ppm)	Xcorr Sequest Score
GSALGKQLNLK45LLDNWDSVTSTFSKLRE	791.67	4	-0.2	5.89
V <mark>K</mark> 94AKVQPYLDDFQKKWQEE	808.08	3	-1.58	3.04
MELYRQK118VEPLRAELQE	725.71	3	0.08	3.22
LYRQ <mark>K</mark> 118VEPLRAE	515.62	3	-0.32	2.91
LYRQ <mark>K₁₁₈VEPLRAELQE</mark>	639.01	3	0.03	3.4
AL <mark>K₁₈₂ENGGARLAE</mark>	636.33	2	-0.5	3.14
SFK226VSFLSALEEYTKKLNTQ	792.75	3	1.1	4.95
SFK226VSFLSALEE	700.35	2	0.09	3.98
YTK <mark>K</mark> 239LNTQ	519.78	2	0.19	2.44
GSALGK40QLNLK45LLDNWDSVTSTFSK59 LREQLGPVTQE	1004.78	4	0.44	5.14
Reference	peptide can	didates		
²¹³ DLRQGLLPVLE ²²³	626.86	2	-0.85	2.84
⁷⁹ TEGLRQE ⁸⁵	416.71	2	0.32	2.82
¹⁸⁴ NGGARLAE ¹⁹¹	394.20	2	-0.86	3.03

Table S5. *In vitro* CNO⁻ modified rHDL/apoA-I peptides containing carbamylated lysine residues detected in the proteomic analyses

LC-MS/MS analysis of GluC digests from *in vitro* modified rHDL (CNO⁻) preparations yielded identification of \geq 90% of apoA-I protein sequence (Table S2). Peptide sequence, m/z, Δ m, charge and Xcorr produced by Proteome Discoverer for the *in vitro* CNO⁻ modified rHDL/apoA-I peptides containing carbamylated lysine residues included in the proteomic analysis are listed above.

Carbamylated lysine residue within the peptide sequence is shown in red.

m/z = the ratio of the mass (Da) and the charge (a.u.) of the digested peptide.

 Δm = mass difference (ppm) between measured peptide and its theoretical mass.

Charge (positive ion mode) = peptide charge (a.u.).

Table S6. *In vitro* MPO modified rHDL/apoA-I peptides containing carbamylated lysine residues detected in the proteomic analysis.

Peptide Sequence	m/z	Charge	Δm (ppm)	Xcorr Sequest Score
LATVYVDVL <mark>K</mark> 23DSGRDYVSQFE	816.40	3	-2.07	4.82
GSALG <mark>K</mark> 40QLNLKLLDNWDSVTSTFSKL RE	791.67	4	-0.2	6.38
VK94AKVQPYLDDFQKKWQEE	808.08	3	-1.58	3.7
LYRQ <mark>K118</mark> VEPLRAE	515.62	3	3.83	2.88
YHA <mark>K₁₉₅ATEHLSTLSEKAKP</mark>	524.77	4	-0.02	1.78
SF <mark>K₂₂₆</mark> VSFLSALEE	700.35	2	-2.53	4.43
YT <mark>K</mark> 238KLNTQ	519.78	2	-0.99	2.71
Reference	e peptide ca	ndidates		
²¹³ DLRQGLLPVLE ²²³	626.86	2	-0.85	3.03
⁷⁹ TEGLRQE ⁸⁵	416.71	2	0.32	2.48
¹⁸⁴ NGGARLAE ¹⁹¹	394.20	2	-0.86	2.95

LC-MS/MS analysis of GluC digests from *in vitro* modified rHDL (MPO) preparations yielded identification of \geq 90% of apoA-I protein sequence. Peptide sequence, m/z, Δ m, charge and Xcorr produced by Proteome Discoverer are listed above.

Carbamylated lysine residue is shown in red.

m/z = the ratio of the mass (Da) and the charge (a.u.) of the digested peptide.

 Δm = mass difference (ppm) between measured peptide and its theoretical mass.

Charge (positive ion mode) = peptide charge (a.u.).

Peptide Sequence	m/z	Charge	Δm (ppm)	Xcorr Sequest Score
LATVYVDVL <mark>K</mark> 23DSGRDYVSQFE	816.40	3	-2.07	3.05
GSALGK40QLNLKLLDNWDSVTSTFSKLREQ				
LGPVTQE	1004.78	4	-0.59	5.07
V <mark>K</mark> 94AKVQPYLDDFQKKWQEE	808.08	3	-1.58	4.58
LYRQ <mark>K118</mark> VEPLRAELQE	639.01	3	-1.02	3.87
LYRQK118VEPLRAE	515.62	3	3.83	3.06
YHA <mark>K</mark> 195ATEHLSTLSE	543.93	3	-2.22	1.44
SFK226VSFLSALEE	700.35	2	-2.53	3.58
VKA <mark>K</mark> 96VQPYLDDFQ <mark>K106K107</mark> WQEE	836.76	3	8.17	1.22
GSALGK40QLNLKLLDNWDSVTSTFSK59				
LRE	1055.22	3	-1.37	7.01
Reference p	eptide cand	idates		
²¹³ DLRQGLLPVLE ²²³	626.86	2	-0.85	2.85
⁶³ QLGPVTQE ⁷⁰	436.22	2	-1.78	2.5
⁷¹ FWDNLE ⁷⁸	823.36	1	-0.86	1.87

Table S7. *In vitro* CNO⁻ modified lipid poor apoA-I peptides containing carbamylated lysine residues detected in the proteomic analysis.

LC-MS/MS analysis of GluC digests from *in vitro* CNO⁻ modified lipid poor apoA-I preparations yielded identification of \ge 90% of apoA-I protein sequence. Peptide sequence, m/z, Δ m, charge and Xcorr produced by Proteome Discoverer are listed above.

Carbamylated lysine residue is shown in red.

m/z = the ratio of the mass (Da) and the charge (a.u.) of the digested peptide.

 Δm = mass difference (ppm) between measured peptide and its theoretical mass.

Charge (positive ion mode) = peptide charge (a.u.).

Peptide Sequence	m/z	charge	Δm (ppm)	Xcorr Sequest Score		
GSALG <mark>K</mark> 40QLNLKLLDNWDSVTSTFSKLRE	791.67	4	0.03	6.31		
GSALG K40QLNLKLLDNWDSVTSTFSKLREQLGPVTQE	1004.78	4	-0.59	4.83		
LYRQ <mark>K118</mark> VEPLRAE	515.62	3	-3.17	2.88		
YHA <mark>K195</mark> ATEHLSTLSE	543.93	3	-1.65	1.24		
SGRDYVSQFEGSALGK40QLNLK45LLDNWD	1047.18	3	2.67	1.48		
Reference peptide candidates						

Table S8. *In vitro* MPO modified lipid poor apoA-I peptides containing carbamylated lysine residues detected in the proteomic analysis.

LC-MS/MS analysis of GluC digests from *in vitro* modified (MPO) lipid poor apoA-I preparations yielded identification of \ge 90% of apoA-I protein sequence. Peptide sequence, m/z, Δ m, charge and Xcorr produced by Proteome Discoverer are listed above.

626.86

436.22

823.36

-0.85

-1.78

-0.86

2.94

2.51

1.85

2

2

1

Carbamylated lysine residue is shown in red.

m/z = the ratio of the mass (Da) and the charge (a.u.) of the digested peptide.

 Δm = mass difference (ppm) between measured peptide and its theoretical mass.

Charge (positive ion mode) = peptide charge (a.u.).

²¹³DLRQGLLPVLE²²³

⁶³QLGPVTQE⁷⁰

⁷¹FWDNLE⁷⁸

Peptide Sequence	m/z	Charge	Δm (ppm)
DEPPQSPWDRVK12D	537.92	3	-2.65
LATVYVDVLK23D	639.85	2	-1.41
VLK23DSGRDYVSQFE	843.41	2	-4
VYVDVLK23DSGRDYVSQFE	1081.5	2	1.15
LATVYVDVLK23DSGRDYVSQFE	816.41	3	0.1
GSALGK40QLN	465.75	2	-2.0
GSALG <mark>K₄₀</mark> QLNLK	586.34	2	0.64
ALGK40QLNLKLLDNWDSVTSTFSKLRE	766.41	4	-5.44
SALGK40QLNLKLLDNWDSVTSTFSKLRE	1036.2	3	-3.33
GSALG <mark>K₄₀</mark> QLNLKLLD	504.96	3	-2.19
GSALGKQLNLK45LLDNWDSVTS	1151.6	2	-0.89
LNLK45LLDNWDSVTSTFSKLRE	841.45	3	0.63
QFEGSALGKQLNLK45LLDNWDSVTSTFSKLRE	714.38	5	1.16
WDSVTSTFS <mark>K</mark> ₅9LRE	799.89	2	-1.97
NWDSVTSTFS <mark>K₅</mark> 9LRE	856.91	2	-0.96
LKLLDNWDSVTSTFSK59LRE	765.74	3	1.03
NLKLLDNWDSVTSTFSK59LRE	803.75	3	-3.54
GSALGKQLNLKLLDNWDSVTSTFSK59L	960.18	3	0.20
FWDNLE <mark>K77</mark> ETE	677.3	2	-1.16
GLRQEMS <mark>K</mark> 88DLEE	747.35	2	-0.78
TEGLRQEMSK88DLEE	862.39	2	-0.71
V <mark>K</mark> 94AKVQPYLD	602.34	2	-2.07
V <mark>K</mark> 94AKVQPYLDD	659.85	2	-2.05
VKAK ₉₆ VQPYLD	602.34	2	0.97
FQKK107WQEE	583.28	2	-1.36
YRQ <mark>K</mark> 118VEPLRAE	477.93	3	-1.53
LYRQK118VEPLRAE	515.62	3	-2.45
MELYRQK118VEPLRAE	602.32	3	-1.84
MELYRQK118VEPLRAELQE	725.71	3	-3.2
LQEK140LSPLGEE	643.33	2	-0.60
K140LSPLGEEMRD	667.32	2	-1.82
ALK182ENGGARLAE	636.33	2	-7.60
KAK ₂₀₈ PALE	400.24	2	-3.08
SFK226VSFL	435.74	2	-1.13
SFK226VSFLSA	514.77	2	0.54

Table S9. Human atherosclerotic aorta apoA-I peptides containing carbamylated lysine residues detected by the proteomic analysis.

Peptide Sequence (Table S9 Continued)	m/z	charge	Δm (ppm)
SFK226VSFLSAL	571.32	2	2.23
K ₂₂₆ VSFLSALEE	583.31	2	-1.17
SFK226VSFLSALE	635.84	2	-1.64
FK226VSFLSALEE	656.84	2	-0.99
SFK226VSFLSALEE	700.36	2	-0.61
YTK <mark>K</mark> 239LNT	455.75	2	0.53
YTKK ₂₃₉ LNTQ	519.78	2	-1.81
K40QLNLK45LLDNWDSVTSTFSKLRE	706.12	4	-2.25
SALGK40QLNLK45LLDNWDSVTSTFSKLRE	788.17	4	-0.5
GSALG <mark>K</mark> 40QLNL <mark>K</mark> 45LLDNWDSVTSTF	850.78	3	-0.71
GSALG <mark>K40</mark> QLNL <mark>K</mark> 45LLDNWDSVTSTFS <mark>K</mark> 59LRE	791.68	4	1.81
VK94AK96VQPYLDDFQK106K107WQEE	808.08	3	-0.90
EV <mark>K</mark> 94AK96VQPYLDDFQ K106K107WQEE	851.1	3	-0.69
Reference peptide candi	dates		
⁶³ QLGPVTQE ⁷⁰	436.21	2	1.06
²¹³ DLRQGLLPVLE ²²³	626.86	2	-0.33
⁷⁹ TEGLRQE ⁸⁵	416.72	2	-1

LC MS/MS analysis of GluC peptide digests generated from apoA-I isolated from human atherosclerotic aorta preparations yielded coverage of \geq 90% of apoA-I protein sequence. Peptide sequence, m/z, Δ m and charge are listed in the table above.

Carbamylated lysine residue is shown in red

m/z = the ratio of the mass (Da) and the charge (a.u.) of the digested peptide

 Δm = mass difference (ppm) between measured peptide and its theoretical mass

Charge (positive ion mode) = peptide charge (a.u.)

	Hu	Human subject aorta carbamylated apoA-I peptide Xcorr Values (n=10)					les			
	1	2	3	4	5	6	7	8	9	10
Peptide Sequence										
DEPPQSPWDRVK12D	2.16	2.38	2.26	2.33	2.65	2.26	2.38	2.32	2.93	2.11
LATVYVDVL <mark>K</mark> 23D	-	-	-	-	-	2.39	-	-	-	-
VLK23DSGRDYVSQFE	-	2.46	-	-	3.18	2.01	-	2.26	2.25	2.63
VYVDVLK23DSGRDYVSQFE	-	-	2.46	-	-	-	-	-	-	-
LATVYVDVL <mark>K</mark> 23DSGRDYVSQ FE	3.71	3.31	-	3.31	3.72	4.04	3.57	3.65	3.4	3.73
GSALG <mark>K₄₀</mark> QLN	2.71	2.15	-	2.61	2.74	2.27	2.64	2.72	2.9	-
GSALG <mark>K₄₀</mark> QLNLK	-	2.62	-	-	-	•	-	-	-	-
ALGK40QLNLKLLDNWDSVTS TFSKLRE	4.41	-	-	-	-	3.54	4.29	-	4.88	5.02
SALGK40QLNLKLLDNWDSVT STFSKLRE	4.22	-	4.8	-	4.98	-	-	-	-	-
GSALG <mark>K₄₀QLNLKLLD</mark>	2.85	3.11	-	-	-	2.64	2.69	2.03	2.56	-
GSALGKQLNL <mark>K</mark> 45LLDNWDSV TS	-	-	-	-	-	-	2.85	-	3.26	3.31
LNL <mark>K</mark> 45LLDNWDSVTSTFSKL RE	-	3.52	-	-	-	-	-	-	-	-
QFEGSALGKQLNLK45LLDNW DSVTSTFSKLRE	-	-	-	-	-	-	3	-	-	•
WDSVTSTFSK59LRE	2.16	-	3.05	2.78	2.79	3.58	2.97	-	-	-
NWDSVTSTFSK ₅₉ LRE	4.27	3.48	3.61	4.38	4.59	3.56	4.39	3.71	3.7	3.94
LKLLDNWDSVTSTFSK59LRE	4.16	4.32	4.91	4.3	4.29	3.76	3.86	4.21	4.07	4.11
NLKLLDNWDSVTSTFS <mark>K</mark> 59LR E	-	3.19	-	-	-	-	-	2.93	-	•
GSALGKQLNLKLLDNWDSVT STFS <mark>K</mark> 59L	-	3.2	-	-	-	-	-	-	-	-
FWDNLEK77ETE	-	-	-	2.54	2.18	2.13	2.44	-	-	2.4
GLRQEMSK88DLEE	-	2.5	-	2.61	2.29	-	2.59	2.57	2.63	-
TEGLRQEMSK88DLEE	-	-	-	-	-	2.81	-	2.23	3.1	3.05
V <mark>K</mark> 94AKVQPYLD	-	-	-	-	-	2.13	-	-	-	-
VK94AKVQPYLDD	-	-	2.44	2.33	2.13	-	2.11	-	-	2.41
VKAK96VQPYLD	-	-	-	2.44	-	-	-	-	-	-
FQKK ₁₀₇ WQEE	-	-	2.05	-	-	-	-	-	-	-
YRQ <mark>K118</mark> VEPLRAE	-	2.69	-	-	2.63	-	-	-	-	-
LYRQ <mark>K118</mark> VEPLRAE	2.98	-	2.69	3.54	2.93	2.84	3.38	2.68	3.06	2.72
MELYRQK ₁₁₈ VEPLRAE	-	-	2.89	3.22	3.7	3.12	-	-	2.78	2.5
MELYRQ <mark>K118</mark> VEPLRAELQE	-	-	-	-	-	2.52	-	-	-	-
LQE <mark>K140</mark> LSPLGEE	-	-	-	-	-	-	-	-	2.33	-

 Table S10. Xcorr Sequest scores for human atherosclerotic lesion apoA-I peptides containing carbamylated lysine residues detected during proteomic analyses.

Peptide Sequence (Table S10 Continued)	1	2	3	4	5	6	7	8	9	10
K ₁₄₀ LSPLGEEMRD	2.12	-	-	-	-	2.49	2.22	-	2.01	-
ALK182ENGGARLAE	1.9	2.3	2.1		2.5	2.7	1.2			2.7
KAK208PALE	2.2	2.1	-	-	-	-	2	-	2.05	-
SFK226VSFL	-	2.2	2.13	-	-	-	-	-	-	-
SFK226VSFLSA	-	-	2.48	-	-	-	-	-	-	-
SFK226VSFLSAL	-	2.16	-	-	-	-	-	-	-	-
K ₂₂₆ VSFLSALEE	-	-	-	-	-	-	-	-	-	2.15
SFK226VSFLSALE	-	2.66	2.36	-	-	2.61	2.8	-	-	-
FK226VSFLSALEE	-	-	-	-	-	-	2.09	-	-	-
SFK226VSFLSALEE	4.58	3.9	4.1	4.33	4.51	4.51	3.85	3.49	3.87	3.52
YTK <mark>K</mark> 239LNT	2.04	-	-	-	-	-	-	-	-	-
YTK <mark>K</mark> 239LNTQ	-	2.6	2.22	2.48	2.46	2.45	2.46	2.44	2.61	2.31
K40QLNLK45LLDNWDSVTSTF SKLRE	5.68	4.4	4.89	-	5.84	4.65	-	-	-	3.32
SALGK40QLNLK45LLDNWDSV TSTFSKLRE	-	-	-	-	-	3.72	3.92	-	-	-
GSALG <mark>K</mark> 40QLNL <mark>K</mark> 45LLDNWDS VTSTF	-	-	-	-	2.99	-	-	-	-	-
GSALGK40QLNLK45LLDNWDS VTSTFSK59LRE	5.85	5.91	6.11	5.26	-	5.04	-	5.64	5.74	5.42
VK94AK96VQPYLDDFQK106K10 7WQEE	4.39	4.74	3.51	-	4.08	-	3.91	4.42	-	4.09
EVK94AK96VQPYLDDFQ K106K107WQEE	-	-	3.85	2.75	2.65	2.84	3.33	-	2.69	-
Reference peptide candidates										
²¹³ DLRQGLLPVLE ²²³	2.55	2.48	2.75	2.53	2.45	2.66	2.49	2.43	2.59	2.44
⁷⁹ TEGLRQE ⁸⁵	2.18	2.05	2.50	2.10	2.23	2.08	2.27	2.15	2.02	2.39
⁶³ QLGPVTQE ⁷⁰	2.25	2.30	2.48	2.36	2.22	2.46	2.27	2.66	2.47	2.62

LC MS/MS analysis of GluC digests generated from apoA-I isolated from human atherosclerotic lesions yielded identification of \geq 90% of the protein sequence. Peptide sequence and the distribution of Xcorr values produced by Proteome Discoverer are listed above.

Carbamylated lysine residue is shown in red

Xcorr (Proteome Discoverer) = cross-correlation value from the search.

(-)= Not detected.

Table S11. Kinetic and binding affinity data for lipid poor apoA-I binding to MPO determined by Surface Plasmon Resonance spectroscopy.

k _{on} (1/Ms)	k _{off} (1/s)	K _d (M)
5.02 X 10 ⁴	3.77 X 10 ⁻³	7.51 X 10 ⁻⁸

To determine whether MPO specifically interacts with lipid-poor apoA-I we performed Surface Plasmon Resonance (SPR) spectroscopy measurements under physiological salt and pH conditions. MPO was immobilized on a CM5 sensor chip. Samples of apoA-I concentrations ranging from 0.5 nM to ~3000 nM were prepared in binding buffer (10 mM PBS, pH 7.4) and flowed over the sensor chip at a flow rate of 20 μ L/min. The association rate constant (kon), dissociation rate constant (koff), and apparent dissociation constant (Kd) calculated from the kinetic/binding data are listed above. The results indicate that apoA-I binds tightly to MPO with Kd in the nanomolar range.

kon is the kinetic rate constant for apoA-I/MPO binding.

koff is the kinetic rate constant for apoA-I/MPO dissociation.

K_d is the dissociation constant of the complex MPO/apoA-I.

Table S12 .PRIDE accessible raw file names for in-vitro modified apoA-I and apoA-I isolated from human lesion samples used in the proteomics analysis. Files can be accessed using the data set identifier PXD027881.

	•		
	Sample Name	Sample Description	File Name
1	HDL_MPO	in vitro Modified rHDL by MPO system	HDL_MPO.raw
2	HDL_MPO2	in vitro Modified rHDL by MPO system	HDL_MPO2.raw
3	HDL_MPO3	in vitro Modified rHDL by MPO system	HDL_MPO3.raw
4	HDL_OCN	in vitro Modified rHDL by CNO- system	HDL_OCN.raw
5	HDL_OCN2	in vitro Modified rHDL by CNO- system	HDL_OCN2.raw
6	HDL_OCN3	in vitro Modified rHDL by CNO- system	HDL_OCN3.raw
7	LP_MPO	in vitro Modified LP apoA-I by MPO system	LP_MPO.raw
8	LP_MPO2	in vitro Modified LP apoA-I by MPO system	LP_MPO2.raw
9	LP_MPO3	in vitro Modified LP apoA-I by MPO system	LP_MPO3.raw
10	LP_OCN	in vitro Modified LP apoA-I by CNO- system	LP_OCN.raw
11	LP_OCN2	in vitro Modified LP apoA-I by CNO- system	LP_OCN2.raw
12	LP_OCN3	in vitro Modified LP apoA-I by CNO- system	LP_OCN3.raw

Table S12A. ApoA-I samples used in the proteomics analysis shown in Fig. 3 and Fig. 4

Table S12B. ApoA-I samples used in the in solution proteomic analysis shown in Fig. 5

	Sample Name	Sample Description	File Name
1	Lum_19may2101	apoA-I isolated from human subject 1	Lum_19may2101.raw
2	Lum_19may2102	apoA-I isolated from human subject 2	Lum_19may2102.raw
3	Lum_19may2103	apoA-I isolated from human subject 3	Lum_19may2103.raw
4	Lum_19may2104	apoA-I isolated from human subject 4	Lum_19may2104.raw
5	Lum_19may2105	apoA-I isolated from human subject 5	Lum_19may2105.raw
6	Lum_19may2106	apoA-I isolated from human subject 6	Lum_19may2106.raw
7	Lum_19may2107	apoA-I isolated from human subject 7	Lum_19may2107.raw
8	Lum_19may2108	apoA-I isolated from human subject 8	Lum_19may2108.raw
9	Lum_19may2109	apoA-I isolated from human subject 9	Lum_19may2109.raw
10	Lum_19may2110	apoA-I isolated from human subject 10	Lum_19may2110.raw

Table S12C. ApoA-I samples used in the in gel proteomic analysis shown in Fig. 6

	Sample Name	Sample Description	File Name
1	Lum_21nov1701	apoA-I Dimer isolated from human subject 1	Lum_21nov1701.raw
2	Lum_21nov1702	apoA-I Dimer isolated from human subject 2	Lum_21nov1702.raw
3	Lum_21nov1703	apoA-I Dimer isolated from human subject 3	Lum_21nov1703.raw
4	Lum_21nov1704	apoA-I Dimer isolated from human subject 4	Lum_21nov1704.raw
5	Lum_21nov1705	apoA-I Dimer isolated from human subject 5	Lum_21nov1705.raw
6	Lum_21nov1706	apoA-I Dimer isolated from human subject 6	Lum_21nov1706.raw
7	Lum_21nov1707	apoA-I Dimer isolated from human subject 7	Lum_21nov1707.raw
8	Lum_21nov1708	apoA-I Dimer isolated from human subject 8	Lum_21nov1708.raw
9	Lum_21nov1709	apoA-I Dimer isolated from human subject 9	Lum_21nov1709.raw
10	Lum_21nov1711-re	apoA-I Dimer isolated from human subject 10	Lum_21nov1711-re.raw

11	Lum_21nov1713	apoA-I Monomer isolated from human subject 1	Lum_21nov1713.raw
12	Lum_21nov1714	apoA-I Monomer isolated from human subject 2	Lum_21nov1714.raw
13	Lum_21nov1715	apoA-I Monomer isolated from human subject 3	Lum_21nov1715.raw
14	Lum_21nov1716	apoA-I Monomer isolated from human subject 4	Lum_21nov1716.raw
15	Lum_21nov1717	apoA-I Monomer isolated from human subject 5	Lum_21nov1717.raw
16	Lum_21nov1718	apoA-I Monomer isolated from human subject 6	Lum_21nov1718.raw
17	Lum_21nov1719	apoA-I Monomer isolated from human subject 7	Lum_21nov1719.raw
18	Lum_21nov1720	apoA-I Monomer isolated from human subject 8	Lum_21nov1720.raw
19	Lum_21nov1721	apoA-I Monomer isolated from human subject 9	Lum_21nov1721.raw
20	Lum_21nov1722	apoA-I Monomer isolated from human subject 10	Lum_21nov1722.raw



Figure S1. Characterization of reconstituted HDL particles. Reconstituted HDL (rHDL) particles were prepared using a modified cholate dialysis method at a molar ratio of 100:10:1, POPC: Cholesterol: apoA-I. Left: native-PAGE gel analysis of rHDL containing purified recombinant human apoA-I. Reconstituted HDL was purified by gel filtration and loaded (10 µg of protein) on a 4-20% gradient gel. The rHDL particle has a diameter of approximately 10 nm, consistent with preparations used in previous studies. Right: Purified 96-Å diameter POPC/Cholesterol rHDL-containing human apoA-I was cross-linked with BS3 cross-linker for 5 min at 37 °C and then quenched by the addition of Tris, pH 7.4 and samples fractionated on 12% SDS-PAGE gel. Left lane shows molecular mass standards and right lane shows bands for crosslinked rHDL.



Figure S2. Western blot analysis of lipid poor apoA-I crosslinked with MPO. To further confirm the result of the Surface Plasmon Resonance (SPR) spectroscopy experiment, we crosslinked lipid poor apoA-I with MPO using the DSSO crosslinker as described in Experimental Procedures. The indicated samples were fractionated on a 12% reducing SDS-PAGE gel and proteins transferred to PVDF membranes for western blot analysis. A. Western blot analysis using anti-MPO antibody in the presence or absence of DSSO are shown in the gel. Lane 1: Marker, Lane 2: apoA-I only, Lane 3: MPO only, Lane 4: apoA-I+DSSO, Lane 5: MPO+DSSO, and Lane 6: MPO+apoA-I+DSSO. Samples were probed either with anti-MPO antibody or anti-apoA-I antibody (10G) to detect the presence of MPO or apoA-I in the sample, respectively.



Figure S3. Quantification of carbamylated lysine residues in lipid poor apoA-I modified *in vitro* by **the MPO or CNO⁻ system.** The top, middle, and bottom panels show the peak areas of carbamylated lysine containing peptides divided by the peak area average of 3 distinct reference peptide candidates (multiplied by 10⁴). Reference peptide candidates D₂₁₃-E₂₂₃, F₇₁-E₇₈ and Q₆₃-E₇₀ were used for quantitation in this analysis. Note that the overall patterns observed are highly aligned, regardless of which reference peptide was used.



Figure S4. Quantification of site-specific carbamylation of apoA-I recovered from human atherosclerotic aorta. A-C. Each panel displays the peak area of peptides containing modified lysine residues divided by the peak area of a distinct reference peptide candidates (multiplied by 10^4). Reference peptide candidates D₂₁₃-E₂₂₃, T₇₉-E₈₅ and Q₆₃-E₇₀ were used for quantitation in this analysis. Note that the overall patterns observed are highly aligned, regardless of which reference peptide was used.



Figure S5. Lysine carbamylation sites identified by the proteomic analysis of rHDL and lipid poor apoA-I modified *in vitro* by CNO⁻. A) Lysine residues carbamylated by the CNO⁻ system in rHDL are mapped onto the Double Belt model of rHDL. B) Lysine residues carbamylated by the CNO⁻ system in rHDL are mapped onto the Double Super Helix model of rHDL. C) Lysine residues carbamylated by the CNO⁻ system in lipid poor apoA-I are mapped onto a model of lipid poor apoA-I. Dark blue spheres depict lysine residues that were carbamylated to the greatest extent, while the light blue spheres depict lysine residues that are carbamylated to a lesser extent in the *in vitro* model systems. White spheres are additional sites that were observed to be carbamylated in apoA-I recovered from human atherosclerotic aorta.



#1	b+	b²+	Seq.	у+	У ²⁺	#2
1	116.03422	58.52075	D			13
2	245.07681	123.04204	E	1496.71283	748.86005	12
3	342.12958	171.56843	Р	1367.67024	684.33876	11
4	439.18234	220.09481	Р	1270.61748	635.81238	10
5	567.24092	284.12410	Q	1173.56471	587.28599	9
6	654.27295	327.64011	S	1045.50613	523.25671	8
7	751.32571	376.16649	Р	958.47411	479.74069	7
8	937.40502	469.20615	W	861.42134	431.21431	6
9	1052.43197	526.71962	D	675.34203	338.17465	5
10	1208.53308	604.77018	R	560.31509	280.66118	4
11	1307.60149	654.30438	V	404.21398	202.61063	3
12	1478.70227	739.85477	K-Carbamyl	305.14556	153.07642	2
13			D	134.04478	67.52603	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S6: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide DEPPQSPWDRVK₁₂(carbamyl)D. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs. within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top -CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples. ND, not detected.



#1	b⁺	b²+	Seq.	у+	У ²⁺	#2
1	114.09134	57.54931	L			11
2	185.12845	93.06787	A	1165.60993	583.30860	10
3	286.17613	143.59170	Т	1094.57282	547.79005	9
4	385.24455	193.12591	V	993.52514	497.26621	8
5	548.30788	274.65758	Y	894.45672	447.73200	7
6	647.37629	324.19178	V	731.39340	366.20034	6
7	762.40323	381.70525	D	632.32498	316.66613	5
8	861.47165	431.23946	V	517.29804	259.15266	4
9	974.55571	487.78149	L	418.22963	209.61845	3
10	1145.65649	573.33188	K-Carbamyl	305.14556	153.07642	2
11			D	134.04478	67.52603	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S7: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide LATVYVDVLK₂₃(carbamyl)D. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top -CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the *indicated in vitro* carbamylated apoA-I samples. ND, not detected.



#1	b+	b²+	Seq.	у*	У ²⁺	#2
1	58.02874	29.51801	G			9
2	145.06077	73.03402	S	873.47886	437.24307	8
3	216.09788	108.55258	A	786.44683	393.72705	7
4	329.18195	165.09461	L	715.40971	358.20850	6
5	386.20341	193.60534	G	602.32565	301.66646	5
6	557.30419	279.15573	K-Carbamyl	545.30419	273.15573	4
7	685.36276	343.18502	Q	374.20341	187.60534	3
8	798.44683	399.72705	L	246.14483	123.57605	2
9			N	133.06077	67.03402	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	yes

Figure S8: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide GSALGK₄₀(carbamyl)QLN. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	y+	У ²⁺	#2
1	115.05020	58.02874	N			14
2	301.12952	151.06840	W	1598.78091	799.89409	13
3	416.15646	208.58187	D	1412.70160	706.85444	12
4	503.18849	252.09788	S	1297.67466	649.34097	11
5	602.25690	301.63209	V	1210.64263	605.82495	10
6	703.30458	352.15593	Т	1111.57421	556.29075	9
7	790.33661	395.67194	S	1010.52654	505.76691	8
8	891.38429	446.19578	Т	923.49451	462.25089	7
9	1038.45270	519.72999	F	822.44683	411.72705	6
10	1125.48473	563.24600	S	675.37841	338.19285	5
11	1296.58551	648.79639	K- Carbamyl	588.34639	294.67683	4
12	1409.66957	705.33842	L	417.24561	209.12644	3
13	1565.77068	783.38898	R	304.16155	152.58441	2
14			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S9: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide NWDSVTSTFSK₅₉(carbamyl)LRE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top -CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	y+	y ²⁺	#2
1	148.07569	74.54148	F			10
2	334.15500	167.58114	W	1206.52732	603.76730	9
3	449.18195	225.09461	D	1020.44801	510.72764	8
4	563.22487	282.11608	N	905.42107	453.21417	7
5	676.30894	338.65811	L	791.37814	396.19271	6
6	805.35153	403.17940	E	678.29408	339.65068	5
7	976.45231	488.72979	K-Carbamyl	549.25148	275.12938	4
8	1105.49490	553.25109	E	378.15071	189.57899	3
9	1206.54258	603.77493	Т	249.10811	125.05769	2
10			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S10: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide FWDNLEK₇₇(carbamyl)ETE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b⁺		b ²⁺	Seq.	y⁺	y ²⁺	#2
1	58.02874	2	9.51801	G			12
2	171.11280	8	6.06004	L	1436.66858	718.83793	11
3	327.21392	16	4.11060	R	1323.58452	662.29590	10
4	455.27249	22	8.13988	Q	1167.48341	584.24534	9
5	584.31509	29	2.66118	E	1039.42483	520.21605	8
6	731.35049	36	6.17888	M-Oxidation	910.38224	455.69476	7
7	818.38251	40	9.69490	S	763.34684	382.17706	6
8	989.48329	49	5.24528	K-Carbamyl	676.31481	338.66104	5
9	1104.51023	55	2.75876	D	505.21403	253.11066	4
10	1217.59430	60	9.30079	L	390.18709	195.59718	3
11	1346.63689	67	3.82208	E	277.10303	139.05515	2
12				E	148.06043	74.53386	1
				Sample:	Observed in:		
			Hur	man plaque	yes		
			In-Vit	ro rHDL OCN	ND		
			In-Vit	ro rHDL MPO	ND		
			In-Vitro	LP apoA-I OCN	ND		
			In-Vitro	I P apoA-I MPO	ND		

Figure S11: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide GLRQEMSK₃₈(carbamyl)DLEE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	у+	y ²⁺	#2
1	100.07569	50.54148	V			11
2	271.17647	136.09187	K-Carbamyl	1219.63173	610.31950	10
3	342.21358	171.61043	А	1048.53095	524.76911	9
4	470.30854	235.65791	K	977.49384	489.25056	8
5	569.37696	285.19212	V	849.39887	425.20308	7
6	697.43554	349.22141	Q	750.33046	375.66887	6
7	794.48830	397.74779	Р	622.27188	311.63958	5
8	957.55163	479.27945	Y	525.21912	263.11320	4
9	1070.63569	535.82148	L	362.15579	181.58153	3
10	1185.66263	593.33496	D	249.07173	125.03950	2
11			D	134.04478	67.52603	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S12: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide VK₉₄(carbamyl)AKVQPYLDD. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	y+	У ²⁺	#2
1	100.07569	50.54148	V			10
2	228.17065	114.58897	К	1104.60479	552.80603	9
3	299.20777	150.10752	A	976.50982	488.75855	8
4	470.30854	235.65791	K-Carbamyl	905.47271	453.23999	7
5	569.37696	285.19212	V	734.37193	367.68960	6
6	697.43554	349.22141	Q	635.30352	318.15540	5
7	794.48830	397.74779	Р	507.24494	254.12611	4
8	957.55163	479.27945	Y	410.19218	205.59973	3
9	1070.63569	535.82148	L	247.12885	124.06806	2
10			D	134.04478	67.52603	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S13: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide VKAK₉₆(carbamyl)VQPYLD. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	y+	y²+	#2
1	148.07569	74.54148	F			8
2	276.13427	138.57077	Q	1018.49524	509.75126	7
3	404.22923	202.61825	К	890.43666	445.72197	6
4	575.33001	288.16864	K-Carbamyl	762.34169	381.67449	5
5	761.40932	381.20830	W	591.24092	296.12410	4
6	889.46790	445.23759	Q	405.16160	203.08444	3
7	1018.51049	509.75888	E	277.10303	139.05515	2
8			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S14: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide FQKK₁₀₇(carbamyl)WQEE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b⁺	b²+	Seq.	у+	У ²⁺	#2
1	114.09134	57.54931	L			12
2	277.15467	139.08097	Y	1431.77029	716.38878	11
3	433.25578	217.13153	R	1268.70696	634.85712	10
4	561.31436	281.16082	Q	1112.60585	556.80656	9
5	732.41513	366.71121	K-Carbamyl	984.54727	492.77727	8
6	831.48355	416.24541	V	813.44649	407.22689	7
7	960.52614	480.76671	E	714.37808	357.69268	6
8	1057.57891	529.29309	Р	585.33549	293.17138	5
9	1170.66297	585.83512	L	488.28272	244.64500	4
10	1326.76408	663.88568	R	375.19866	188.10297	3
11	1397.80119	699.40424	A	219.09755	110.05241	2
12			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S15: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide LYRQK₁₁₈**(carbamyl)VEPLRAE.** Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples. LQEK₁₄₀(+43)LSPLGEE



#1	b+	b²+	Seq.	у⁺	У ²⁺	#2
1	114.09134	57.54931	L			11
2	242.14992	121.57860	Q	1172.57936	586.79332	10
3	371.19251	186.09989	E	1044.52078	522.76403	9
4	542.29329	271.65028	K-Carbamyl	915.47819	458.24273	8
5	655.37735	328.19231	L	744.37741	372.69234	7
6	742.40938	371.70833	S	631.29335	316.15031	6
7	839.46214	420.23471	Р	544.26132	272.63430	5
8	952.54621	476.77674	L	447.20855	224.10792	4
9	1009.56767	505.28747	G	334.12449	167.56588	3
10	1138.61026	569.80877	E	277.10303	139.05515	2
11			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S16: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide LQEK₁₄₀(carbamyl)LSPLGEE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	у +	У ²⁺	#2
1	72.04439	36.52583	A			12
2	185.12845	93.06787	L	1200.63313	600.82020	11
3	356.22923	178.61825	K-Carbamyl	1087.54906	544.27817	10
4	485.27182	243.13955	E	916.44828	458.72778	9
5	599.31475	300.16101	N	787.40569	394.20648	8
6	656.33622	328.67175	G	673.36276	337.18502	7
7	713.35768	357.18248	G	616.34130	308.67429	6
8	784.39479	392.70103	A	559.31984	280.16356	5
9	940.49590	470.75159	R	488.28272	244.64500	4
10	1053.57997	527.29362	L	332.18161	166.59444	3
11	1124.61708	562.81218	A	219.09755	110.05241	2
12			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S17: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide ALK₁₈₂(carbamyl)ENGGARLAE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



					-					
#1	b+	b ²	2+	b ³⁺	Se	q.	У+	y ²⁺	У ³⁺	#2
1	164.07061	82.5	3894	55.36172	Y	,				14
2	301.12952	151.0	6840	101.04802	Н		1466.72340	733.86534	489.57932	13
3	372.16663	186.5	8695	124.72706	A		1329.66448	665.33588	443.89301	12
4	543.26741	272.1	3734	181.76065	K-Cart	bamyl	1258.62737	629.81732	420.21397	11
5	614.30452	307.6	5590	205.43969	A		1087.52659	544.26694	363.18038	10
6	715.35220	358.1	7974	239.12225	Т		1016.48948	508.74838	339.50134	9
7	844.39479	422.7	0103	282.13645	E		915.44180	458.22454	305.81879	8
8	981.45370	491.2	3049	327.82275	Н		786.39921	393.70324	262.80459	7
9	1094.53777	547.7	7252	365.51744	L		649.34030	325.17379	217.11828	6
10	1181.56980	591.2	8854	394.52812	S		536.25623	268.63175	179.42360	5
11	1282.61748	641.8	1238	428.21068	Т		449.22420	225.11574	150.41292	4
12	1395.70154	698.3	5441	465.90536	L		348.17653	174.59190	116.73036	3
13	1482.73357	741.8	7042	494.91604	S		235.09246	118.04987	79.03567	2
14					E		148.06043	74.53386	50.02500	1
	•	•		Sample:		Obs	served in:	•		
	Human plaque			ND						
	In-Vitro rHDL OCN NE		ND							
			In	-Vitro rHDL M	IPO		yes			
			In-V	itro LP apoA-l	OCN		yes			

Figure S18: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide YHAK₁₉₅(carbamyl)ATEHLSTLSE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.

yes

In-Vitro LP apoA-I MPO



#1	b+	b²+	Seq.	у+	У ²⁺	#2
1	129.10224	65.05476	К			7
2	200.13935	100.57331	А	671.37227	336.18977	6
3	371.24013	186.12370	K-Carbamyl	600.33515	300.67121	5
4	468.29289	234.65009	Р	429.23438	215.12083	4
5	539.33001	270.16864	А	332.18161	166.59444	3
6	652.41407	326.71067	L	261.14450	131.07589	2
7			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S19: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide KAK₂₀₈(carbamyl)PALE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	у+	У ²⁺	#2
1	88.03930	44.52329	S			12
2	235.10772	118.05750	F	1312.67834	656.84281	11
3	406.20850	203.60789	K-Carbamyl	1165.60993	583.30860	10
4	505.27691	253.14209	V	994.50915	497.75821	9
5	592.30894	296.65811	S	895.44074	448.22401	8
6	739.37735	370.19231	F	808.40871	404.70799	7
7	852.46142	426.73435	L	661.34030	331.17379	6
8	939.49344	470.25036	S	548.25623	274.63175	5
9	1010.53056	505.76892	A	461.22420	231.11574	4
10	1123.61462	562.31095	L	390.18709	195.59718	3
11	1252.65721	626.83225	E	277.10303	139.05515	2
12			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	ND

Figure S20: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide SFK₂₂₆(carbamyl)VSFLSALEE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	y+	У ²⁺	#2
1	164.07061	82.53894	Y			8
2	265.11828	133.06278	Т	875.49451	438.25089	7
3	436.21906	218.61317	K-Carbamyl	774.44683	387.72705	6
4	564.31402	282.66065	К	603.34605	302.17666	5
5	677.39809	339.20268	L	475.25109	238.12918	4
6	791.44101	396.22415	N	362.16702	181.58715	3
7	892.48869	446.74798	Т	248.12410	124.56569	2
8			Q	147.07642	74.04185	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S21: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide YTK₂₃₈(carbamyl)KLNTQ. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	у+	У ²⁺	#2
1	164.07061	82.53894	Y			8
2	265.11828	133.06278	Т	875.49451	438.25089	7
3	393.21325	197.11026	К	774.44683	387.72705	6
4	564.31402	282.66065	K-Carbamyl	646.35187	323.67957	5
5	677.39809	339.20268	L	475.25109	238.12918	4
6	791.44101	396.22415	N	362.16702	181.58715	3
7	892.48869	446.74798	Т	248.12410	124.56569	2
8			Q	147.07642	74.04185	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S22: Annotated MS/MS spectrum and table of fragmentation ions for carbamylated apoA-I peptide YTKK₂₃₉ (carbamyl)LNTQ. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I peptide sequence harboring a carbamyllysine residue. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the carbamyllysine (K+43)-harboring apoA-I peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b⁺	b²+	Seq.	у⁺	У ²⁺	#2
1	116.03422	58.52075	D			11
2	229.11828	115.06278	L	1137.69902	569.35315	10
3	385.21939	193.11334	R	1024.61496	512.81112	9
4	513.27797	257.14262	Q	868.51384	434.76056	8
5	570.29944	285.65336	G	740.45527	370.73127	7
6	683.38350	342.19539	L	683.43380	342.22054	6
7	796.46756	398.73742	L	570.34974	285.67851	5
8	893.52033	447.26380	Р	457.26568	229.13648	4
9	992.58874	496.79801	V	360.21291	180.61009	3
10	1105.67281	553.34004	L	261.14450	131.07589	2
11			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	yes

Figure S23: Annotated MS/MS spectrum and table of fragmentation ions for apoA-I reference peptide DLRQGLLPVLE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I reference peptide sequence. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the apoA-I reference peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	b²+	Seq.	у+	У ²⁺	#2
1	129.06585	65.03657	Q			8
2	242.14992	121.57860	L	743.39340	372.20034	7
3	299.17138	150.08933	G	630.30933	315.65830	6
4	396.22415	198.61571	Р	573.28787	287.14757	5
5	495.29256	248.14992	V	476.23510	238.62119	4
6	596.34024	298.67376	Т	377.16669	189.08698	3
7	724.39882	362.70305	Q	276.11901	138.56314	2
8			E	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	yes
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	yes

Figure S24: Annotated MS/MS spectrum and table of fragmentation ions for apoA-I reference peptide QLGPVTQE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I reference peptide sequence. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the apoA-I reference peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b⁺	b²+	Seq.	у+	У ²⁺	#2
1	102.05496	51.53112	Т			7
2	231.09755	116.05241	E	731.36824	366.18776	6
3	288.11901	144.56314	G	602.32565	301.66646	5
4	401.20308	201.10518	L	545.30419	273.15573	4
5	557.30419	279.15573	R	432.22012	216.61370	3
6	685.36276	343.18502	Q	276.11901	138.56314	2
7			Е	148.06043	74.53386	1

Sample:	Observed in:
Human plaque	ND
In-Vitro rHDL OCN	yes
In-Vitro rHDL MPO	yes
In-Vitro LP apoA-I OCN	ND
In-Vitro LP apoA-I MPO	ND

Figure S25: Annotated MS/MS spectrum and table of fragmentation ions for apoA-I reference peptide TEGLRQE. Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I reference peptide sequence. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the apoA-I reference peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.



#1	b+	Seq.	у+	#2
1	148.07569	F		6
2	334.15500	W	676.29368	5
3	449.18195	D	490.21437	4
4	563.22487	N	375.18743	3
5	676.30894	L	261.14450	2
6		E	148.06043	1

Sample:	Observed in:
Human plaque	ND
In-Vitro rHDL OCN	ND
In-Vitro rHDL MPO	ND
In-Vitro LP apoA-I OCN	yes
In-Vitro LP apoA-I MPO	yes

Figure S26: Annotated MS/MS spectrum and table of fragmentation ions for apoA-I reference **peptide FWDNLE.** Human apoA-I was either recovered via immunoprecipitation from homogenized human atherosclerotic aorta (Human plaque), or recombinant human apoA-I (lipid poor (LP) vs within rHDL) was exposed to the indicated *in vitro* carbamylation systems. ApoA-I was then subjected to proteolysis and proteomics analyses as described under Experimental Procedures. Top - CID spectrum for the indicated apoA-I reference peptide sequence. Middle – Fragmentation table of peptide ions. Bottom – Table stating whether the apoA-I reference peptide was observed either in apoA-I recovered from human plaque or within the indicated *in vitro* carbamylated apoA-I samples.