

Figure S1| Characterization of RBCEVs, mouse lung epithelial cells and α-EGFR-VHH binding affinity

(A) Western blot of ALIX, TSG101, GPA, HBA, β -actin and GAPDH in RBCs and RBCEVs. (B) Single-EV flow cytometry analysis of GPA in RBCEVs with a gating strategy to obtain a distinct population of RBCEVs from the background noise. (C) Flow cytometry analysis of EpCAM expression in mouse lung epithelial cells and α -EGFR-VHH binding affinity to multiple cells.



Figure S2| The effects of immRNA-loaded RBCEVs on cancer cells and non-malignant cells

(A-D) qPCR analysis of *DDX58* and its downstream effectors relative to *GAPDH* in MDA-MB-468 (A), MDA-MB-231 (B), MCF10A (C) and mouse lung epithelial cells (D) treated with 0.1 μ g/ μ L unloaded or NC-RNA-loaded, immRNA-loaded RBCEVs for 24 hours (n = 3-4, RNA loaded using REG1) ND, not detected. (E) Representative flow cytometric plots of ANXV/PI staining in 4T1, CA1a, H358, MDA-MB-468 (MB468), MDA-MB-231 (MB231), MCF10A and mouse lung epithelial (mLE) cells treated with 0.1 μ g/ μ L unloaded RBCEVs, NC RNA-loaded RBCEVs and immRNA-loaded RBCEVs for 72 hours. (F) Average percentage of ANXV⁺PI⁺ population in MB468, MB231, MCF10A and mLE cells treated with RBCEVs as in (E) (n = 3). All bar graphs represent mean ± SEM. **P* < 0.05, ***P* < 0.01 and *****P* < 0.0001 determined by Student's two-tailed *t*-test.



Figure S3| The effects of 3p-125b-ASO-loaded RBCEVs on cancer cells and non-malignant cells

(A) Secondary structure of 3p-125b-ASO. (B-D) qPCR analysis of *DDX58* and its downstream effectors relative to *GAPDH* in MDA-MB-231 (B), MCF10A (C) and mouse lung epithelial cells (D) treated with 0.1 μ g/ μ L unloaded or NC-RNA-loaded, 125b-ASO-loaded and 3p-125b-ASO-loaded RBCEVs for 24 hours (n = 3-4, RNA loaded using REG1) ND, not detected. (E) Representative flow cytometric plots of ANXV/PI staining in 4T1, CA1a, H358, MDA-MB-231 (MB231), MCF10A and mouse lung epithelial (mLE) cells treated with 0.1 μ g/ μ L unloaded or NC RNA-loaded, 125b-ASO-loaded and 3p-125b-ASO-loaded RBCEVs for 72 hours. (F) Average percentage of ANXV⁺PI⁺ population in MB231, MCF10A and mLE cells treated with RBCEVs as in (E) (n = 3). (G) Representative flow cytometric plots of ANXV/PI staining in 4T1 cells and CA1a cells after a treatment with combined 3p-125-ASO- and immRNA-loaded RBCEVs for 72 hours. All bar graphs represent mean ± SEM. **P* < 0.05, ***P* < 0.01, ****P* < 0.001 and *****P* < 0.001 determined by Student's two-tailed *t*-test.



Figure S4| Therapeutic efficacy of immRNA-loaded RBCEVs in mammary tumors

(A) Flow cytometry analysis of immune cells in 4T1 mammary tumors treated with immRNA-loaded RBCEVs as in Figure 4. Cells were gated based on FSC-A and SSC-A to exclude the debris and dead cells. The single cells were further gated based on FSC-W and FSC-H to exclude doublets and aggregates. The live cells were gated from singlet population base on Sytox[™] Blue negative. Subsequently, the fluorescence of phenotypic markers for immune cells were gated based on the fluorescent channels and subjected to viSNE analysis. (B) Survival of mice with orthotopic 4T1 tumors treated with 2.5 mg/kg NC-RNA-RBCEVs (i.t.), 2.5 mg/kg immRNA-RBCEVs (i.t.), 2 mg/kg anti-PD-L1 antibody (i.p.) as well as 2.5 mg/kg immRNA-RBCEVs (i.t.) in combination with 2 mg/kg anti-PD-L1 antibody (i.p.) (n = 2-4 mice). (C) Tumor growth of mice treated as in (B) (n = 2-4 mice). (D) ELISA quantification of IFNβ in the sera of mice bearing 4T1 tumors treated with immRNA-loaded RBCEVs as in Figure 4. (F) Volume of MDA-MB-468 tumors injected intratumorally with 2.5 mg/kg RBCEVs containing immRNA or NC RNA every three days (n = 3-4 mice). (G) qPCR analysis of the RIG-I pathway gene expression relative to *GAPDH* in untreated and treated MDA-MB-468 tumors (n = 3-4 mice). All bar graphs represent mean ± SEM. ***P* < 0.01, ****P* < 0.001 and *****P* < 0.0001 determined by Student's two-tailed *t*-test.



Figure S5| Flow cytometry analysis of immune cells in 4T1 mammary tumors treated with 3p-125b-ASO-loaded RBCEVs Representative viSNE plots of immune cells in 4T1 mammary tumors treated with 3p-125b-ASO-loaded RBCEVs.



Figure S6| EGFR-VHH-RBCEVs target 4T1-hEGFR cells *in vitro* and 4T1-hEGFR lung metastatic tumors *in vivo* (A) qPCR analysis of the RIG-I pathway gene expression relative to *Gapdh* in 4T1-hEGFR cells and parental 4T1 cells treated with 0.1 μ g/ μ L uncoated, CtrI-VHH-coated and EGFR-VHH-coated RBCEVs containing immRNA (n = 3). All bar graphs represent mean ± SEM. ***P* < 0.01 and ****P* < 0.001 determined by Student's two-tailed *t*-test. (B) Representative viSNE plots of biodistribution of EGFR-VHH-coated CFSE-RBCEVs in respective cell type in the lungs with 4T1-hEGFR metastatic tumors.



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Untreated
NC RNA-EVs
immRNA-EVs
Ctrl-VHH-immRNA-EVs
EGFR-VHH-immRNA-EVs

Figure S7| Flow cytometry analysis of immune cells in the lungs with 4T1 metastatic tumors treated with EGFR-VHH-immRNA-RBCEVs

(A) Representative viSNE plots of respective cell type in the lungs with 4T1 metastatic tumors treated with EGFR-VHH-immRNA-RBCEVs. (B) qPCR analysis of cytokine gene expression relative to *Gapdh* in the lungs of mice with 4T1-hEGFR metastatic tumors (n = 4 mice). All bar graphs represent mean \pm SEM. **P* < 0.05, ***P* < 0.01, ****P* < 0.001 and *****P* < 0.0001 determined by Student's two-tailed *t*-test.

Human Genes F/R Mouse F AGGTCGGTGTGAACGGATTTG GGAGCGAGATCCCTCCAAAAT GAPDH TGTAGACCATGTAGTTGAGGTCA R GGCTGTTGTCATACTTCTCATGG F GAG AGT CAC GGG ACC CAC T GCCATTACACTGTGCTTGGAGA DDX58 CGG TCT TAG CAT CTC CAA CG CCAGTTGCAATATCCTCCACCA R F TGATGCACTATTCCAAGAACTAAC GAGCAACTTCTTTCAACCACAG MDA5 R TCTGTGAGACGAGTTAGCCAAG CACTTCCTTCTGCCAAACTTG F ACAGCCAAGACATCCTTCGT CACAAAGAAGTGTCCTGCTTGGT RSAD2 R AAAAGTTGATCTTCTCCAAACCA AAGCGCATATATTCATCCAGAAT F CTGCCTCACAGCTAGTGACC AGGAGACAGATGGAGACACA MAVS R CCGGCGCTGGAGATTATTG CAGAACTGGGCAGTACCC AACCTCACCTACAGGGCGGACTT F CTCTCCTGTTGTGCTTCTCC С **IFNB** TCCCACGTCAATCTTTCCTCTTGC R GTCAAAGTTCATCCTGTCCTTG т F CGGAAAGAAGTGTTGCGGTT ACCAGCCGTGGACCAAGAG IRF3 TTTTCCTGGGAGTGAGGCAG R TACCAAGGCCCTGAGGCAC F AGGGCGTTTTATCTTGCG TGGTCCTGGTGAAGCTGGAA IRF7 GATGTCGTCATAGAGGCTGTTG R TGGAGCCCAGCATTTTCTCT G F TACAGGCTGGAGTGTGCTGAGA TAGCCAACATGTCCTCACAGAC ISG56 R CTCCACTTTCAGAGCCTTCGCA TCTTCTACCACTGGTTTCATGC F ATCATCGGCATTTTGAACGAGG IL4 R GCAGCTCCATGAGAACACTA F CTCTGTTGACAAGCAATGAGACG IL5 R TCTTCAGTATGTCTAGCCCCTG F GCCACGGCACAGTCATTGA IFNG R TGCTGATGGCCTGATTGTCTT F GTGCTCCTTGTCAACAGCG IL2 R GGGGAGTTTCAGGTTCCTGTA F CTTCAATACGTCAGACATTCGGG TGFB1 R GTAACGCCAGGAATTGTTGCTA F GCAACTGTTCCTGAACTCAACT IL1B R ATCTTTTGGGGTCCGTCAACT F GACTCTTGCGTCAACTTCAAGG IL18 R CAGGCTGTCTTTTGTCAACGA F AGCCTTATCGGAAATGATCCAGT IL10 R GGCCTTGTAGACACCTTGGT F TCTATACCACTTCACAAGTCGGA IL6 R **TCTATACCACTTCACAAGTCGGA** F TGGTTTGCCATCGTTTTGCTG IL12B R ACAGGTGAGGTTCACTGTTTCT

Table S1| List of primers

TNF	F	CCTGTAGCCCACGTCGTAG
	R	GGGAGTAGACAAGGTACAACCC
IFNA4	F	CTGGTAATGATGAGCTACTACTG
		G
	R	CCTTCTCCAAGGGGAATCCAA
IFNA11	F	GGTCCTGGCACAAATGAGGA
	R	TCCAAGCAGCAGATGAGTCC
IFNA12	F	AAGACTGAGTGAGAAGGAGTGAG
	R	GAGATGCCAGAATTTGAGCAGTG