





Supplemental Table 4. Basal Serum Tryptase, Tryptase Copy Number Variants and Mast Cell Numbers by Participants

Number of Participants	Sex	Year of Birth	BST (ng/mL)	Tryptase Alpha Copy Number	Tryptase Beta Copy Number	Mast Cells/HPF
1.	М	2001	14.0	2	3	30
2.	F	1974	15.0	2	3	27
3.	F	1959	23.0	2	3	102
4.	F	1956	13.0	3	2	54
5 .	F	1975	12.0	3	2	39
6.	F	1964	9.9	3	2	42
7.	F	1981	22.0	2	3	72
8.	F	1995	10.5	3	2	60
9.	F	1965	18.0	2	3	40
10.	F	1944	31.0	2	3	63
11.	F	1975	20.6	2	3	35
12.	F	1946	14.6	2	3	25
13.	F	1983	13.4	3	2	35
14.	F	1993	9.2	3	2	47
15.	M	2001	10.8	2	3	32
16.	M	1941	26.5	2	3	56
17.	M	2004	15.0	2	3	45
18.	F	1981	21.0	2	3	45
19.	F	1980	13.0	2	3	21
20.	M	2002	16.0	2	3	45
21.	F	1982	8.1	3	2	23

Supplemental Table 1. Summary of $H\alpha T$ -Associated Clinical Features

Characteristics, N = 21 individuals				
Mean age, years \pm SD	41 ± 19.4			
Female sex, N (%)	16 (76)			
Male sex, N (%)	5 (24)			
Clinical GI Symptoms				
GERD, N (%)	4/21 (19.05)			
Abdominal Pain, N (%)	19/21 (90.48)			
Diarrhea, N (%)	13/21 (61.90)			
Constipation, N (%)	9/21 (42.86)			
Food Sensitivity, N (%)	5/21 (23.81)			
GI Dysmotility, N (%)	4/21 (14.29)			
Nocturnal awakening due to abdominal pain or diarrhea, N (%)	21/21 (100)			
Laboratory Findings, Median (IQR)				
BST ng/mL	14 (10.9 - 20.8)			
GI small intestinal MCs*	40 (33.5 – 52.0)			

Abbreviations: GERD, gastroesophageal reflux disease; GI, gastrointestinal; IQR, interquartile

range; BST, basal serum tryptase; MCs, mast cells.

Autoimmune Disorders, N (%) – 12/21 (57.14)

*Refer to Figure 2A.

Supplemental Table 2. Antibody and Metal List

Metal	General Panel Clone		Source	
147	CD45RO	UCHL1	Harvard Core	
151	CD2	TS1/8	Fluidigm	
153	CD45RA	HI100	Harvard Core	
154	CD33	WM53	Harvard Core	
165	Mast-cell_tryptase	AA1	Harvard Core, custom conj.	
175	IgD	IA6-2	Harvard Core	
191	DNA1	N/A	Fluidigm	
193	DNA2	N/A	Fluidigm	
148	CD28	CD28.2	Harvard Core	
152	CD14	M5E2	Harvard Core	
141	c kit	1 04D2	Harvard Core, Self Conj.	
142	CD19	HIB19	Harvard Core	
143	HLA-DR	L243	Fluidigm	
144	CD64	10.1	Harvard Core	
145	CD16	3G8	Harvard Core	
149	CD25	2A3	Fluidigm	
155	CD27	L128	Fluidigm	
158	CD3	UCHT1	Harvard Core	
159	CD11c	Bu15	Harvard Core	
164	CD161	HP-3G10	Harvard Core	
166	CD24	ML5	Harvard Core	
168	CCR6	G034E3	Harvard Core	
170	CCR7	G043H7	Harvard Core	
172	IgM	MHM-88	Harvard Core	
174	CD4	OKT4	Harvard Core	
161	CD203c	NP4D6	Harvard Core, self conj.	
146	CD8a	RPA-T8	Harvard Core	
150	FCeR1	AER-37 (CRA-1)	Fluidigm	
162	CD56	HCD56	Harvard Core	
163	CXCR3	G025H7	Fluidigm	
171	CD127	A019D5	Harvard Core	
173	CD335	9E2	Harvard Core, self conj.	
115	CD34	581	Harvard Core, self conj.	
89	CD45	HI30	Fluidigm	

Supplemental Table 3. GI Significance of Antigen Production (IgG) in Individuals with H α T, CD, and Non-H α T

Antibody	Presence in HaT patients	Presence in CD patients	Presence in non- HaT patients	Gastrointestinal Reference
Liver/Kidney Microsomal Type I (LKM1)	10/21	5/20	5/19	Autoimmune hepatitis. Targets the enzyme, CYP2D6, found primarily in liver cells.
Gliadin	9/21	4/20	4/19	Gliadin is protein component of gluten. May indicate gluten-sensitive enteropathies. Gliadin alters barrier properties of intestinal epithelial cells.
SP100	8/21	5/20	4/19	Associated with primary biliary cirrhosis. Targets SP100 nuclear antigen.
Intrinsic Factor (IF)	7/21	8/20	6/19	IF protein is produced by parietal cells in the stomach. Presence of antibodies often leads to autoimmune induced anemia secondary to inability to absorb vitamin cobalamin.
Scl-70/Topoisomerase I	6/21	8/20	6/19	Mainly in systemic scleroderma and found in patients with connective tissue disease.
Tissue Transglutaminase (TTG)	6/21	9/20	5/19	Autoantigen of Celiac Disease. Marker for intestinal barrier defects.
Lipopolysaccharide (LPS)	5/21	6/20	5/19	Contributes to systemic inflammation and impacts the intestinal barrier. Elevated in Inflammatory Bowel Disease.
Amyloid	5/21	8/20	8/19	Demonstrated in intestinal epithelial cells, can impact absorption and secretion. Commonly studied in CNS and Alzheimer's Disease.
Collagen IV	2/21	5/20	4/19	Collagen IV is a major constituent of basement membranes. Useful in detecting loss of parts of basement membrane in carcinomas.

Supplementary Figure legends

Supplemental Figure 1. Terminal ileum T cells phenotype differs between H α T and quiescent CD. tSNE plots of automated clustering by FlowSOM for leukocytes in H α T and CD (All) (Panel A). CD4 (clusters: 1,4,6), CD8 (clusters: 9-14, 16-20), and DNT (clusters: 2,3,5,7,8). Cluster identity is determined by marker expression of the heatmap in Panel B. Panel B. Heatmap of indicated antibody expression in the clusters and the associated phenotype of the cells in each cluster. Panel C. tSNE maps of patients with CD and individuals with H α T that are associated with Panel A.

Supplemental Figure 2. Terminal ileum B cells phenotype reveal expanded CD27 memory B cells in H α T. tSNE plots of automated clustering by FlowSOM for leukocytes in H α T and CD (All) (Panel A). Memory B cells (clusters: 2-6, 9 and 13), IgM B cells (clusters: 14-19). Panel B. Heatmap of indicated antibody expression in the clusters and the associated phenotype of the cells in each cluster. Panel C. tSNE maps of patients with CD and individuals with H α T that are associated with Panel A.

Supplemental Figure 3. Panel A. Principal Component Analysis (PCA) array of IgM and IgG in non-H α T, H α T, and quiescent CD individuals. Panel B. Heatmap indicating antibody expression in non-H α T, H α T, and quiescent CD. Note: Race and ethnicity were accounted for in the analysis, and removing these variables had no effect on the result.