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2	Human serum triggers antibiotic tolerance in Staphylococcus aureus
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Supplementary Fig. 1. S. aureus CFU counts do not change during incubation in human serum. Log₁₀
CFU ml⁻¹ during incubation of TSB-grown S. aureus in human serum for 22 h. Graph represents the
geometric mean ± geometric standard deviation of three independent experiments. Data were
analysed by one-way ANOVA and no statistically significant differences were observed (log₁₀ CFU ml⁻¹
at time points vs at 0 h). Source data are provided as a Source Data file.

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55 Supplementary Fig. 2. Incubation of S. aureus in serum but not TSB or PBS for 16 h results in 56 tolerance towards daptomycin. TSB-grown S. aureus cultures were incubated for 16 h in TSB, PBS or serum, adjusted to 2 x 10⁸ CFU ml⁻¹ where necessary and resuspended in fresh serum containing 80 57 58 μ g ml⁻¹ daptomycin. Log₁₀ CFU ml⁻¹ was measured over 6 h. Graph represents the geometric mean ± 59 geometric standard deviation of three independent repeats and data were analysed by two-way ANOVA with Tukey's post-hoc test (serum-adapted vs PBS/TSB-adapted at each time point). * P = 60 0.0016 (TSB 2 h), 0.0065 (TSB 4 h), 0.022 (TSB 6 h), 0.0292 (PBS 2 h), 0.0075 (PBS 4 h), 0.0023 (PBS 6 61 62 h). Source data are provided as a Source Data file. 63



Supplementary Fig. 3. Incubation of S. aureus in serum results in tolerance towards antimicrobials from various classes. Log₁₀ CFU ml⁻¹ of TSB-grown and serum-adapted cultures of *S. aureus* USA300 WT after/throughout a 6 h incubation in serum with 128 μ g ml⁻¹ nisin (**a**), 160 μ g ml⁻¹ gramicidin (**b**), μ g ml⁻¹ vancomycin (c), 160 μ g ml⁻¹ nitrofurantoin (d) or 40 μ g ml⁻¹ gentamicin (e). Graphs represent the geometric mean ± geometric standard deviation of three (a, b, d, e) or two (c) independent replicates. Graphs in **a** – **b** were analysed by two-way ANOVA with Sidak's *post-hoc test* (* for **a**, P < 0.0001. For **b**, P < 0.0001. Graphs in **c** – **e** were analysed by two-way ANOVA with Tukey's post-hoc test; TSB-grown compared to serum-adapted at each time-point. (* for c, P = 0.0103. For d, P = 0.0197 (4 h), 0.0257 (6 h). For e, P = 0.0122 (4 h), 0.0491 (6h)). In e, some data points fell below the limit of detection of 100 CFU ml⁻¹. Source data are provided as a Source Data file.

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Supplementary Fig. 4. TCS-deficient mutants survive as well as WT in human serum. Log₁₀ CFU ml⁻¹ of *S. aureus* JE2 WT and mutants defective for the sensor kinase components of TCS after 16 h incubation in human serum. Data represent the geometric mean ± geometric standard deviation of three independent experiments. Data were analysed by one-way ANOVA and no statistically significant differences were observed demonstrating there were no differences in survival in serum between the WT and any of the mutants. Source data are provided as a Source Data file.



Supplementary Fig. 5. Complementation of the vraS::Tn and graS::Tn mutants restores tolerance to WT levels. Log₁₀ CFU ml⁻¹ of TSB-grown and serum-adapted cultures of *S. aureus* JE2 WT, graS::Tn and graS::Tn complemented with pCN34 or PgraXRS (a) and WT, vraS::Tn and vraS::Tn complemented with empty pCN34 or PvraUTSR (b) over 6 h incubation in serum with 80 µg ml⁻¹ daptomycin. Graphs represent the geometric mean ± geometric standard deviation of three independent experiments. * for a, P = 0.0008 (2 h graS::Tn pEmpty Serum), 0.0019 (2 h graS::Tn Serum), 0.0279 (4 h graS::Tn Serum). For **b**, P = 0.002 (2 h vraS::Tn pEmpty Serum), 0.0002 (6 h vraS::Tn pEmpty Serum), 0.0198 (6 h vraS::Tn Serum) determined by two-way ANOVA with Dunnett's post-hoc test, serum-adapted WT compared to serum-adapted mutants at each time-point. Some data points fell below the limit of detection of 100 CFU ml⁻¹. Source data are provided as a Source Data file.



127 Supplementary Fig. 6. Colistin activates GraRS signalling and triggers daptomycin tolerance. TSBgrown cultures of S. aureus JE2 WT (solid lines) and the graS::Tn mutant (dashed lines) containing 128 129 PdltA-gfp were exposed to various concentrations of colistin $(2.5 - 20 \,\mu g \,m^{-1})$ in RPMI 1640 (a). GFP 130 fluorescence (RFU) and OD₆₀₀ were measured every 15 min for 12 h. Fluorescence values were divided by OD₆₀₀ measurements to normalise for changes in cell density. S. aureus JE2 WT and graS::Tn mutant 131 132 were each incubated for 16 h in RPMI 1640 only or RPMI 1640 supplemented with indicated concentrations of colistin before determination of CFU counts prior to (0 h) and after (6 h) exposure 133 134 to 80 μ g ml⁻¹ daptomycin (**b**). Graph in **a** represents the mean of three independent experiments and 135 error bars have been omitted for clarity. Graph in **b** represents the geometric mean ± geometric standard deviation of three independent experiments. *, $P = 0.0001 (2.5 \ \mu g \ ml^{-1})$, < 0.0001 (5 $\ \mu g \ ml^{-1})$, 136 137 < 0.0001 (10 μ g ml⁻¹), < 0.0001 (20 μ g ml⁻¹) determined by two-way ANOVA with Dunnett's *post-hoc* 138 test, RPMI 1640 and colistin compared to RPMI 1640 only. RFU, relative fluorescent units; OD₆₀₀, optical density at 600 nm. Source data are provided as a Source Data file. 139

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Supplementary Fig. 7. Exposure of *S. aureus* to verteporfin does not affect bacterial survival in serum. Log₁₀ CFU ml⁻¹ of *S. aureus* JE2 WT grown in TSB or after 16 h incubation in human serum with indicated concentrations of verteporfin. Data represent the geometric mean ± geometric standard deviation of three independent experiments. Data were analysed by two-way ANOVA and no statistically significant differences were observed between bacteria incubated in serum ± verteporfin confirming that verteporfin does not compromise *S. aureus* survival in serum in the absence of daptomycin. Source data are provided as a Source Data file.



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Time (h)

-o-- Water

> 100 kDa

Undialvsed serum

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0

2

4 Time (h)

164 Supplementary Fig. 8. A 0.5 - 5 kDa fraction of serum is necessary and sufficient to trigger 165 daptomycin tolerance. Serum was fractionated by dialysis against PBS using membranes with various molecular weight cut-offs. Next, CFU ml⁻¹ were determined for *S. aureus* JE2 WT which had been 166 incubated for 16 h in serum fractions, or in PBS only or unfractionated serum, then exposed to 80 µg 167 ml⁻¹ daptomycin in fresh human serum over 6 h (a). Log₁₀ CFU ml⁻¹ of *S. aureus* JE2 WT which had been 168 incubated for 16 h in < 5 kDa or > 5 kDa serum fractions, or water only or undialysed serum, then 169 exposed to 80 µg ml⁻¹ daptomycin in fresh human serum over 6 h (**b**). Graphs represent the geometric 170 171 mean ± geometric standard deviation of three independent experiments and were analysed by two-172 way ANOVA with Dunnett's post-hoc test. (* for a, P = 0.0373 (>100 kDa 2 h), 0.0016 (>100 kDa 4 h), 173 0.0003 (>100 kDa 6 h), 0.0106 (>5 kDa 2 h), 0.0037 (>5 kDa 4 h), 0.0053 (>5 kDa 6 h), 0.0008 (PBS 2 h), 0.0033 (PBS 4 h), 0.0121 (PBS 6 h). For b, P = 0.0233 (>5 kDa 2 h), 0.0009 (>5 kDa 4 h), 0.0002 (>5 kDa 174 6 h), 0.0206 (Water 4 h), 0.0002 (Water 6 h) comparing undialysed serum vs serum 175 176 fractions/PBS/water). Source data are provided as a Source Data file.



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Supplementary Fig. 9. Human neutrophil peptide-1, dermcidin and platelet AMPs do not activate GraRS signalling or induce daptomycin tolerance. TSB-grown cultures of S. aureus JE2 WT (solid lines) and the graS::Tn mutant (dashed lines) containing PdltA-qfp were exposed to a range of concentrations of hNP-1 (a), DCD (b) or activated platelet supernatant (c) in RPMI 1640 and GFP fluorescence (RFU) and OD₆₀₀ were measured every 15 min for 16 h. Fluorescence values were divided by OD₆₀₀ measurements to normalise for changes in cell density. S. aureus JE2 WT and graS::Tn mutant were pre-incubated in RPMI 1640 containing hNP-1 (d), DCD (e) or activated platelet supernatant (f) for 16 h before CFU ml⁻¹ were determine before (0 h) and after (6 h) exposure to 80 µg ml⁻¹ daptomycin. Graphs in $\mathbf{a} - \mathbf{c}$ represent the mean of at least two independent experiments with error bars omitted for clarity. Graphs in d - f represent the geometric mean \pm geometric standard deviation of three independent experiments. Data in $\mathbf{d} - \mathbf{f}$ were analysed by two-way ANOVA. No statistically significant differences were observed (RPMI 1640 + AMPs compared to RPMI 1640 alone). Source data are provided as a Source Data file.



Supplementary Fig. 10. Exposure of *S. aureus* to LL-37 does not affect bacterial survival. Log₁₀ CFU ml⁻¹ of *S. aureus* JE2 WT after 16 h incubation in RPMI 1640 with indicated concentrations of LL-37. Data represent the geometric mean ± geometric standard deviation of three independent experiments. Data were analysed by two-way ANOVA and no statistically significant differences were observed (RPMI 1640 + LL-37 compared to RPMI 1640 alone). Source data are provided as a Source Data file.



Supplementary Fig. 11. Exposure of *S. aureus* to anti-hNP-1 or anti-LL-37 antibodies did not affect bacterial survival. Log₁₀ CFU ml⁻¹ of *S. aureus* JE2 WT after incubation for 16 h in human serum supplemented with indicated concentrations of antibodies against hNP-1 or LL-37. Data represent the geometric mean ± geometric standard deviation of three independent experiments. Data were analysed by two-way ANOVA and no statistically significant differences were observed. Source data are provided as a Source Data file.

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Supplementary Fig. 12. Complementation of *∆dltD* mutant with WT *dltD* restores tolerance to WT levels. Log₁₀ CFU ml⁻¹ of TSB-grown and serum-adapted cultures of WT, the $\Delta dltD$ mutant or $\Delta dltD$ mutant complemented with either pCN34 or PdltD after a 16 h incubation in serum (0 h) and after exposure to daptomycin in serum (6 h). Graph represents the geometric mean ± geometric standard deviation of three independent experiments (one data point fell below the limit of detection of 100 CFU ml⁻¹). Data were analysed by two-way ANOVA with Dunnett's *post-hoc* test (* P < 0.0001 ($\Delta dltD$ serum 6 h), < 0.0001 (pEmpty serum 6 h) comparing serum-adapted WT with serum-adapted mutants). Source data are provided as a Source Data file.

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266 Supplementary Fig. 13. The $\Delta dltD$ mutant incorporates less HADA than WT during serum adaptation.

The fluorescence of individual TSB-grown and serum-adapted WT and $\Delta dltD$ cells was quantified. Graph represents the fluorescence of 50 cells per biological replicate (150 cells in total) with the mean

of the three replicates indicated. Each biological replicate is depicted in a different colour. Data were analysed by Kruskal Wallis test (* P = < 0.0001 (WT TSB vs Serum), < 0.0001 (WT Serum vs $\Delta dltD$

Serum), < 0.0001 ($\Delta dltD$ TSB vs $\Delta dltD$ Serum)). Source data are provided as a Source Data file.



Supplementary Fig. 14. Inhibition of peptidoglycan synthesis during serum adaptation did not affect bacterial viability. Log_{10} CFU ml⁻¹ over 6 h of TSB-grown *S. aureus* or cultures which had been incubated in serum supplemented, or not, with 64 µg ml⁻¹ fosfomycin for 16 h. Graph represents the geometric mean ± geometric standard deviation of three independent experiments. No statistically significant differences were observed (two-way ANOVA, log_{10} CFU ml⁻¹ at each time-point vs 0 h). Source data are provided as a Source Data file.





Supplementary Fig. 15. Complementation of the *cls2*::Tn mutant restores daptomycin tolerance. 309 310 Log₁₀ CFU ml⁻¹ of TSB-grown and serum-adapted cultures of S. aureus JE2 WT, cls2::Tn or cls2::Tn 311 complemented with empty pCN34 or Pcls2 before (0 h) or after (6 h) exposure to 80 µg ml⁻¹ daptomycin in serum. Graph represents the geometric mean ± geometric standard deviation of three 312 313 independent experiments (some data points fell below the limit of detection of 100 CFU ml⁻¹). Data were analysed by two-way ANOVA with Dunnett's post-hoc test (* P < 0.0001 (cls2::Tn serum 6 h), < 314 0.0001 (pEmpty serum 6 h) comparing serum-adapted WT with serum-adapted mutants). Source data 315 316 are provided as a Source Data file.