Figure S1. Related to Figure 1.



Figure S1. GLUT3 expression in Th17 cells and the phenotype of mice with T cell-specific ablation of GLUT3 (related to Fig. 1). (A and B) Analysis of subcellular GLUT3 expression in Jurkat T cells using immunofluorescence (A) or immunoblot (B) analyses. (C-J) GLUT3 expression in Th17 cells is coordinated by multiple pathways. (C) Slc2a3 (GLUT3) gene expression in naïve and differentiated Th17 cells of WT and *Ppp3r1*<sup>fl/fl</sup>*Cd4*<sup>Cre</sup> mice with conditional deletion of the calcineurin regulatory subunit B $\alpha$  in T cells; means ± SEM of 4 mice. (D) S/c2a3 (GLUT3) gene expression in naïve and differentiated Th17 cells of WT and CD28deficient (Cd28<sup>-/-</sup>) mice; means ± SEM of 3 mice. (E) Analysis of S/c2a3 (GLUT3) gene expression in naïve and differentiated Th17 cells of WT and Irf4<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice with conditional deletion of IRF4 in T cells; means ± SEM of 4 mice. (F) Slc2a3 (GLUT3) gene expression analysis in naïve and differentiated Th17 cells of WT and Stat3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice with conditional deletion of STAT3 in T cells; means ± SEM of 4 mice. (G) Analysis of SIc2a3 (GLUT3) gene expression Th17 cells differentiated under normoxic (21% O<sub>2</sub>) and hypoxic (2% O<sub>2</sub>) culture conditions for 3 days; means ± SEM of 3 mice. (H) Slc2a3 (GLUT3) gene expression analysis in naïve and differentiated Th17 cells of WT and *Hif1a<sup>fl/fl</sup>Cd4<sup>Cre</sup>* mice with conditional deletion of HIF-1a in T cells; means ± SEM of 4 mice. (I) In silico analysis of the SIc2a3 (GLUT3) gene locus in murine T cells. Chromatin accessibility was determined by genome-wide DNase I hypersensitivity analysis (Bevington et al., 2016) and ATAC-seq. (Mognol et al., 2017). Analysis of ChIP-seq datasets showed the binding of NFATc1 (Klein-Hessling et al. 2017) and NFATc2 (Martinez et al., 2015) to the promoter region and to an -16 kb upstream regulatory element, binding of STAT3 (Hirahara et al., 2015) and IRF4 (Man et al., 2013) to the -16 kb element, and binding of HIF-1 $\alpha$  (Ciofani et al., 2012) to an additional element located -22 kb upstream of the SIc2a3 transcription start site (TSS). (J) Upregulation of GLUT3 expression by antigen receptor ligation (TCR), co-stimulation (CD28) and cytokine signaling (IL-6/IL-23) is orchestrated by NFAT, IRF4, STAT3 and HIF-1  $\alpha$  in Th17 cells. (K-R) Analysis of mice with T cell-specific deletion of GLUT3 (*Slc2a3*<sup>fl/fl</sup>*Cd4*<sup>Cre</sup> mice). (K and L) Analysis of Slc2a3 (GLUT3) (K), Slc2a1 (GLUT1), Slc2a6 (GLUT6) and Slc2a8 (GLUT8) (L) gene expression in naïve and anti-CD3/CD28 stimulated CD4<sup>+</sup> T cells of WT and S/c2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice by gRT-PCR; means ± SEM of 4 mice. (M) Total cell numbers of thymus, spleen and lymph nodes (LNs) of 8-14 weeks old WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM of 11-13 mice. (N) Thymic development of T cells in WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice. Representative flow cytometric analyses of CD4<sup>-</sup>CD8<sup>-</sup> (DN), CD4<sup>+</sup>CD8<sup>+</sup> (DP) and single positive (SP) thymocytes of WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM of 11-13 mice. (O) Analysis of peripheral T cell subsets in WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice. Representative flow cytometric analyses of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in spleen and LNs of WT and *Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup>* mice; means ± SEM of 11-13 mice. (P and Q) Analysis of CD44<sup>-</sup>CD62L<sup>+</sup> (naïve), CD44<sup>+</sup>CD62L<sup>+</sup> (central memory) and CD44<sup>+</sup>CD62L<sup>-</sup> (effector) CD4<sup>+</sup> (P) and CD8<sup>+</sup> (Q) T cells of WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice by flow cytometry; means ± SEM of 12-13 mice. (R) Analysis of Foxp3<sup>+</sup> Treg cell frequency and total cell number in the thymus, spleen and LNs of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM of 11-13 mice. Statistical analyses in (C-H), (K-M), (O) and (R) by unpaired Student's t-tests. \*, p<0.05; \*\*, p<0.01, \*\*\*, p<0.001.

# Figure S2. Related to Figure 1.



Figure S2. GLUT3-deficient T cells show normal activation and proliferation but defective Th17 cell effector function (related to Fig. 1). (A and B) Analysis of glucose uptake by WT and GLUT3-deficient (SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup>) Th1 and Th17 cells using the fluorescent glucose analogue 2-NBDG (A) and tritiated [<sup>3</sup>H] 2-desoxy-glucose (2-DG) (B); means ± SEM of 2-4 mice. (C and D) Flow cytometric analysis of cell cycle entry (expression of CD25 and Ki-67) (C) and cell size (FSC-A) (D) of WT and GLUT3-deficient T cells after polarization for 3 days under Th1 and Th17 culture conditions. (E) Analysis of activation marker (CD69, CD25, CD44, Ki-67, PD-1, OX40, ICOS, CD62L) expression on WT and GLUT3-deficient T cells cultured for 3 days under Th1, Th17 and pTh17 culture conditions by flow cytometry; means ± SEM of 3-4 mice. (F) Proliferation analysis of WT and GLUT3-deficient Th1, Th17 and pTh17 cells by CFSE dilution over a course of 4 days in vitro; means ± SEM of 3 mice. (G and H) Analysis of viability (G) and cellular expansion (H) of WT and GLUT3-deficient T cells cultured for 3 days under Th1, Th17 and pTh17 culture conditions by flow cytometry; means ± SEM of 5-7 mice. (I) Flow cytometric analyses of T-bet and RORyt expression in WT and GLUT3deficient T cells cultured for 3 days under Th1 and pTh17 cell-polarizing conditions; means ± SEM of 6-10 mice. (J-L) Flow cytometric analysis of GM-CSF (K) and IL-2 (L) production by WT and GLUT3-deficient Th1, Th17 and pathogenic Th17 (pTh17) cells after re-stimulation with PMA/Iono for 5 h; means ± SEM of 8-17 mice. (M and N) Analysis of Foxp3 (M) and GATA3 (N) expression in WT and GLUT3-deficient T cells differentiated under iTreg and Th2-polarizing conditions, respectively; means ± SEM of 3-11 mice. (O-S) Ectopic expression of GLUT3 in T cells augments the pathogenicity of Th17 cells. (O) Measurement of glycolytic proton efflux rate (PER) of GLUT3 or empty vector (EV)-transduced and FACS-sorted Th17 cells on day 3 and 6 of culture using a Seahorse extracellular flux analyzer; means ± SEM of 3 experiments. (P and Q) Flow cytometric analysis of GM-CSF and IL-2 cytokine production of GLUT3 and EV-transduced Th1 and Th17 after re-stimulation with PMA/Iono for 5 h; means ± SEM of 4-8 mice. (R and S) Ectopic expression of GLUT3 augments EAE immunopathology. (R) Analysis of immune cell populations in the CNS after transfer of EV or GLUT3-transduced 2D2 Th17 cells. (S) Analysis of IL-17, GM-CSF and IL-2 expression in CNSinfiltrating 2D2 T cells transduced with GLUT3 or EV; means ± SEM of 4 mice per cohort. Statistical analyses in (A), (B), (D-I), (K-N), (Q) and (S) by unpaired Student's t-tests. \*, p<0.05, \*\*, p<0.01, \*\*\*, p<0.001, n.s., non-significant.

### Figure S3. Related to Figure 2.



Figure S3. GLUT3 controls Th17 cell-mediated immunity in vivo (related to Fig. 2). (A-K) T cell-specific GLUT3 deletion protects mice from experimental autoimmune encephalomyelitis (EAE). (A) Relative weight change of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice after immunization with MOG<sub>35-55</sub> peptide emulsified in CFA; means ± SEM of 9 mice per cohort. (B-D) Flow cytometric analysis of relative (B and C) and absolute numbers (D) of immune cell populations in the CNS of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice after immunization with MOG<sub>35-55</sub> peptide; means ± SEM of 6 mice per cohort. (E) Absolute CD4<sup>+</sup>T cell numbers in the spleen, inguinal lymph nodes (inLN) and CNS of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice after immunization with MOG<sub>35-55</sub> peptide; means ± SEM of 6 mice per cohort. (F and G) T-bet and RORyt transcription factor expression in encephalitic CD4<sup>+</sup>T cells in the spleen, inLNs and CNS of WT and  $S/c2a3^{fl/fl}Cd4^{Cre}$  mice after immunization with MOG<sub>35-55</sub> peptide; means ± SEM of 6 mice per cohort. (H-K) Cytokine production of encephalitic CD4<sup>+</sup>T cells. Frequencies of IFNγ (H) and IFNγ plus IL-17 (I) as well as total cell numbers of IL-17 (J) and GM-CSF-producing (K) CD4<sup>+</sup> T cells in the spleen, inLNs and CNS of WT and *Slc2a3*<sup>fl/fl</sup>*Cd4*<sup>Cre</sup> mice 20 days after MOG<sub>35-55</sub> immunization; means ± SEM of 5-7 mice. (L-P) GLUT3 controls Th17 cell effector function in response to C. rodentium infection. (L) Frequencies of IL-17, IFN $\gamma$  and TNF $\alpha$ -producing T cells in the spleen, mesenteric (m)LNs and colon of WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice 10 days after C. rodentium infection; means ± SEM of 3-5 mice. (M) Absolute cell numbers of IL-17-producing CD4<sup>+</sup> T cells in the spleen, mLNs and colon of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice 10 days after C. rodentium infection; means ± SEM of 3-5 mice. (N and O) Relative weight change (N) and bacterial load in the feces (O) of WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice after infection with C. rodentium; means ± SEM of 3-10 mice per cohort. (P) Analysis of IFN<sub>γ</sub> expression of all and LCMV-specific (NP<sub>309-328</sub> tetramer<sup>+</sup>) CD4<sup>+</sup> T cells in the spleen of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice 10 days after the infection with the Armstrong strain of LCMV; means ± SEM of 5-7 mice. Statistical analyses in (C-E), (G-M) and (P) by unpaired Student's t-tests. \*, p<0.05, \*\*, p<0.01, \*\*\*, p<0.001, n.s., non-significant.

Figure S4. Related to Figure 4.



- - 5 10 20 Glutamine (mM)

5 10 20 Pyruvate (mM)

- - 5 10 20 Lactate (mM)

Figure S4. Tracing of glucose-derived metabolites in GLUT3-deficient Th1 and Th17 cells (related to Fig. 4). (A) Experimental setup to trace <sup>13</sup>C-glucose-derived polar metabolites in WT and GLUT3-deficient (Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup>) Th1 and Th17 cells. (B) Volcano plot of differential metabolite concentrations between Th1 and Th17 cells, analysis is based on 4 biological replicates per T cell subset and genotype. (C and D) Metabolic tracing of glucose-derived <sup>13</sup>C-metabolites of the glycolytic pathway (C) and the tricarboxylic acid (TCA) cycle (D) in WT and GLUT3-deficient Th1 and Th17 cells; means ± SEM of 4 biological replicates. (E) Venn diagram analysis of > 2-fold differential metabolite concentrations (p < 0.05) comparing WT and GLUT3deficient Th1 with Th17 cells; red and blue arrows indicate higher and lower metabolite abundances, respectively. (F) Analysis of ATP concentrations in whole cell lysates (WCL) and isolated cytosolic and mitochondrial fractions of WT and GLUT3-deficient Th17 cells; means ± SEM of 2-4 mice. (G) Metabolite set enrichment analysis (MSEA) of differential metabolite concentrations (p < 0.05) between WT and GLUT3deficient Th1 cells. (H) Ratio of oxygen consumption rate (OCR) to extracellular acidification rate (ECAR) in Th1, Th17 and pathogenic (p)Th17 cells; means ± SEM of 6 experiments. (I-M) Effects of exogenous pyruvate (J), lactate (K), glutamine (L) and dimethyl- $\alpha$ -ketoglutarate (M) on the cytokine production of GLUT3-deficient Th17 cells. Flow cytometric analyses of IL-17 production of WT and Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> Th17 cells treated with different metabolites for 24 h before re-stimulation with PMA/lono; means ± SEM of 2- 11 mice. (N-R) Defective cytokine production is not due to impaired lipid metabolism in GLUT3- deficient Th17 cells. (N) Lipidomic analyses of sphingoid long-chain bases (dhSph, dihydrosphingosine; Sph, sphingosine; S1P, sphingosine 1-phosphate), sphingomyelins (SM), ceramides (Cer) and dihydroceramides (dhCer) in WT and GLUT3-deficient Th17 cells by LC/MS; means ± SEM of 3 mice. Effect of oleic (O) or palmitic acid (P) on the cytokine production of GLUT3-deficient Th17. Flow cytometric analyses of IL-17 production of WT and SIc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup> Th17 cells treated with oleic or palmitic acid for 24 h before re-stimulation with PMA/Iono for 5 h; means ± SEM of 2-11 mice. Neutral lipid content of WT and GLUT3-deficient Th17 cells after exogenous addition of oleic acid (Q) or palmitic acid (R) by flow cytometry using BODIPY labeling. Statistical analyses in (H), (J-M), (O) and (P) by unpaired Student's t-tests. \*\*, p<0.01; \*\*\*, p<0.001.

### Figure S5. Related to Figure 5



Figure S5. ACLY controls cytokine expression of Th17 cells (related to Fig. 5). (A and B) Inducible deletion of ACLY in Th17 cells using CRISPR/Cas9-mediated gene editing. (A) Transduction of Cas9expressing Th17 cells (isolated from Rosa26<sup>LSL-Cas9</sup>Cd4<sup>Cre</sup> mice) with retroviral vectors expressing control (ctrl.) or Acly-targeting gRNAs. Immunoblot analysis to confirm successful ACLY protein deletion 3 days after retroviral transduction. (B) Flow cytometric analysis of IFN<sub>Y</sub> and IL-17 production in Th1 and Th17 cells after Cas9-mediated ACLY deletion and stimulation with PMA/Iono for 5 h; means ± SEM of 8 mice. (C) Analysis of Acly gene expression in naïve and anti-CD3/CD28 activated WT and ACLY-deficient (Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup>) CD4<sup>+</sup> T cells by qRT-PCR; means ± SEM of 3 mice. (D-I) Phenotypic characterization of lymphocyte subsets in WT and Aclv<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice. (D) Total cell number of thymus, spleen and lymph nodes (LNs) in 7-12 weeks old WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM of 7 mice. (E) Thymic development of T cells in WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice. Representative flow cytometric analyses of CD4<sup>-</sup>CD8<sup>-</sup> (DN), CD4<sup>+</sup>CD8<sup>+</sup> (DP) and single positive (SP) thymocytes of WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice and summary of 7 mice; means ± SEM. (F) Analysis of peripheral T cell subsets in WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM. (G and H) Analysis of CD44<sup>-</sup>CD62L<sup>+</sup> (naïve), CD44<sup>+</sup>CD62L<sup>+</sup> (central memory) and CD44<sup>+</sup>CD62L<sup>-</sup> (effector) CD4<sup>+</sup> (G) and CD8<sup>+</sup> (H) T cells of WT and Aclv<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice by flow cytometry; means ± SEM of 7 mice. (I) Flow cytometric analysis of Foxp3<sup>+</sup> Treg cells in the thymus, spleen and LNs of WT and  $Acly^{fl/fl}Cd4^{Cre}$  mice; means ± SEM of 7 mice. (J) Analysis of cell cycle entry (expression of CD25 and Ki-67) of WT and ACLY-deficient CD4<sup>+</sup> T cells after polarization for 3 days under Th1 and Th17 culture conditions. (K) Activation marker expression (CD69, CD25, CD44, Ki-67. PD-1. OX40. ICOS) on WT and ACLY-deficient (Ac/v<sup>fl/fl</sup>Cd4<sup>Cre</sup>) T cells cultured for 3 days under Th1. Th17 and pathogenic Th17 (pTh17) cell-polarizing conditions by flow cytometry. Heatmap showing row-normalized geometric MFIs of the activation markers; means of 3 mice. (L) Representative proliferation analysis of WT and ACLY-deficient Th1 and Th17 cells by cell trace violet dilution over 4 days in vitro. (M) Representative intracellular analyses of T-bet and RORyt transcription factor expression in WT and ACLY-deficient Th1 and Th17 cells, respectively. (N) Flow cytometric analysis of GM-CSF and IL-2 production by WT and ACLYdeficient Th1, Th17 and pTh17 cells after re-stimulation with PMA/lono for 5 h; means ± SEM of 6-11 mice. (O) Representative histopathological examination of caudal spinal cord sections from WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice 16 days after MOG<sub>35-55</sub> immunization stained with Luxol fast blue (myelin) and Cresyl violet (nuclei). (P) Flow cytometric analysis of immune cell infiltration in the CNS of WT and Acly<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice 16 days after immunization with MOG<sub>35-55</sub>; means ± SEM of 3-6 mice per cohort. (Q and R) Flow cytometric analyses of GM-CSF expression in CD4<sup>+</sup> T cells (Q) and quantification of total cytokine-producing CD4<sup>+</sup> T cells (R) in the CNS of WT and Aclv<sup>fl/fl</sup>Cd4<sup>Cre</sup> mice; means ± SEM of 3-6 mice. Statistical analyses in (B), (C), (N) and (P-R) by unpaired Student's t-tests. \*, p<0.05; \*\*, p<0.01, \*\*\*, p<0.001.

# Figure S6. Related to Figure 6



Figure S6. GLUT3-dependent acetate metabolism controls histone acetylation in Th17 cells (related to Fig. 6). (A) Gene set enrichment analyses (GSEA) of WT versus GLUT3-deficient Th17 cells indicate dysregulated protein and histone acetylation processes. (B and C). Analysis of global histone 3 (H3) trimethylation (me<sup>3</sup>) at lysins K9 (B) and K27 (C) in WT and GLUT3-deficient (Slc2a3<sup>fl/fl</sup>Cd4<sup>Cre</sup>) Th17 cells using flow cytometry; means ± SEM of 5-6 mice. (D-G) Genome-wide analysis of H3 K9/14 acetylation in WT and GLUT3-deficient Th17 cells by chromatin immunoprecipitation followed by DNA-sequencing (ChIP-seq). (D) Mapping of differentially acetylated regions to CpG-rich elements (upper panel) and gene bodies (lower panel) using deepTools. (E-G) Analysis of H3 K9/14 acetylation at housekeeping (Actb, Pgk1, Hprt, B2m), activation marker (Mki67, Il2ra, Cd44) and transferrin receptor (Tfrc) gene loci in WT and GLUT3-deficient Th17 cells. Promoters (green shading), enhancers and other conserved noncoding regions (orange shading) were determined using ATAC-seq datasets of different Th17 cell populations (Qiu et al., 2020). (H) Flow cytometric analysis of CD71 protein expression (encoded by *Tfrc*) in WT and GLUT3-deficient Th17 cells; means  $\pm$  SEM of 3 mice. (I) Pathway enrichment analysis using differentially acetylated genes (p < 0.05) between WT and GLUT3- deficient Th17 cells; dot size and color represent enrichment and significance, respectively. Statistical analyses in (C) and (H) by unpaired Student's t-tests. \*, p<0.05; \*\*, p<0.01, \*\*\*, p<0.001.

## Figure S7. Related to Figure 7



Figure S7. Pharmacological inhibition of ACLY prevents Th17 cell-mediated autoimmunity (related to Fig. 7). (A) Representative proliferation analysis of WT Th17 cells treated with 2-Hydroxycitrate (2-HC) using cell trace violet. (B-G) Treatment of mice with 2-HC ameliorates EAE immunopathology. (B) Relative weight change of mice after immunization with MOG<sub>35-55</sub> peptide treated with or without 500 mg/kg 2-HC; means ± SEM of 4-6 mice per cohort. (C) Flow cytometric analysis of absolute immune cell numbers in the CNS of WT mice after immunization with MOG<sub>35-55</sub> peptide with or without 500 mg/kg 2-HC treatment; means ± SEM of 4-6 mice per cohort. (D and E) Quantification of CD4<sup>+</sup> (D) and cytokine-producing T cells (E) in the spleen, inguinal (in)LNs and CNS of WT mice after immunization with MOG<sub>35-55</sub> peptide with or without 500 mg/kg 2-HC treatment; means ± SEM of 4-6 mice per cohort. (F and G) Histopathological examination of caudal spinal cord sections of WT mice 18 days after MOG<sub>35-55</sub> peptide immunization (**F**) and *Rag1<sup>-/-</sup>* mice 14 days after transfer of 2D2 Th17 cells and MOG<sub>35-55</sub> peptide immunization (G). Samples were stained with Luxol fast blue (myelin) and Cresyl violet (nuclei); white arrows and asterisks indicate leukocytic infiltrates and areas of demyelination, respectively. (H-K) 2-HC inhibits cytokine production of human CD4<sup>+</sup> memory T cells. Treatment of human PBMCs with 5 mM 2-HC does not prevent cell cycle entry (nuclear Ki-67 expression) (H) and proliferation (cell trace violet dilution assay) (I) or promotes apoptosis (Annexin V staining) (J) in human CD4<sup>+</sup> T cells; means ± SEM of 5 different donors. (K) Flow cytometric analysis of GM-CSF and IL-2 expression in naïve, CD45RO<sup>+</sup> and CD45RO<sup>+</sup>CCR6<sup>+</sup> memory CD4<sup>+</sup>T cell subsets from healthy human donors after stimulation with PMA/lono for 5 h in presence or absence of 2-HC; n=6 donors. Statistical analyses in (E) and (K) by unpaired Student's t-tests. \*, p<0.05; \*\*, p<0.01, \*\*\*, p<0.001.

Name	gRNA-sequence	Oligo1	Oligo1
gAcly.1	GGTACGCCTCAC GCCCAAAGGGG	CACCGGGTACGCCTCACGCCCAAAG	AAACCTTTGGGCGTGAGGCGTACCC
gAcly.2	GGGTCCCACTCAT ACCTCGGAGG	CACCGGGGTCCCACTCATACCTCGG	AAACCCGAGGTATGAGTGGGACCCC
gAcly.3	CGAGTAAAATCGG TAAACTGAGG	CACCGCGAGTAAAATCGGTAAACTG	AAACCAGTTTACCGATTTTACTCGC
gAcly.4	GTCCGCTTACGAC AGCACCATGG	CACCGGTCCGCTTACGACAGCACCA	AAACTGGTGCTGTCGTAAGCGGACC

Table S1. Sequences of gRNAs for CRISPR/Cas9 genome editing (Related to STAR Methods).

#### Table S2. Mouse primer for qRT-PCR (Related to STAR Methods).

Gene name	Forward primer	Reverse primer	Source
mouse Acly	ACCCTTTCACTGGGGATCACA	GACAGGGATCAGGATTTCCTTG	This study
mouse Slc2a1	GAGACCAAAGCGTGGTGAGT	GAGTTCGGCTATAACACTGG	This study
mouse Slc2a3	ATCGTGGCATAGATCGGTTC	TCTCAGCAGCTCTCTGGGAT	This study
mouse 18S	CGGCGACGACCCATTCGAAC	GAATCGAACCCTGATTCCCCGT	This study
mouse <i>Hif1a</i>	AAACTTCAGACTCTTTGCTTCG	CGGCGAGAACGAGAAGAA	This study

#### Table S3. Primer for ChIP-qPCR (Related to STAR Methods).

Gene name	Forward primer	Reverse primer	Source
Mouse Actb prom	TGCAAAGAAGCTGTGCTCGC	GCCGTTCCGAAAGTTGCCTT	PMID: 17218320
mouse II17a prom	GCAGCAGCTTCAGATATGTCC	TGAGGTCAGCACAGAACCAC	PMID: 17218320
mouse II17f prom	GGGAATCAAAGGGGGGACCCTAA	AAAGCAGAACCCACACGCAGAG	PMID: 22244845
mouse II17 CNS2	ATGGGCCTCTCTTTCCACTGATG	GGAATTTGTGGTGGAAGGGAGTG	PMID: 22244845