## Supplemental Online Material

## **Optogenetic Stimulation of Midbrain Dopamine Neurons**

## **Produces Striatal Serotonin Release**

Merel Dagher,<sup>1†</sup> Katie A. Perrotta,<sup>2†</sup> Sara A. Erwin,<sup>1</sup> Ayaka Hachisuka,<sup>3</sup> Rahul Iyer,<sup>4</sup>

Sotiris C. Masmanidis,<sup>3,5,7</sup> Hongyan Yang,<sup>6</sup> and Anne M. Andrews<sup>1,2,5,6,7\*</sup>

<sup>1</sup>Molecular Toxicology Interdepartmental Program, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>2</sup>Department of Chemistry and Biochemistry, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>3</sup>Department of Neurobiology, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>4</sup>Department of Electrical Engineering, University of California, Los Angeles, Los Angeles, CA, 94720

<sup>5</sup>Neuroscience Interdepartmental Program, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>6</sup>Department of Psychiatry and Biobehavioral Sciences, Semel Institute for Neuroscience & Human Behavior, and Hatos Center for Neuropharmacology, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>7</sup>California Nanosystems Institute, University of California, Los Angeles, Los Angeles, CA 90095, United States

<sup>†</sup>These authors contributed equally to this work

\*To whom correspondence should be addressed: <u>aandrews@mednet.ucla.edu</u>



Figure S1: Head-fixed recording set-up. A. Schematic showing the locations of the head-bar implants (in blue and to scale) and the stimulation (stim) and recording (dSTR) site craniotomies relative to a mouse skull. B. Schematic of the head-bar plate holder (in gray and to scale). The head-bar plate holder was 30 mm long, 28 mm wide, and 1.3 mm thick. The mini-plates, which attach the holder to the head bars, were 9 mm long, 7 mm wide, and 0.65 mm thick, with a 10 mm gap between them. C. Schematic of the custom head-fixed tube used for fast microdialysis recordings with optical stimulation. The restraint tube (2" diameter), constructed of opaque (black) plexiglass, provided loose restraint to reduce spontaneous and stimulated physical movement, which can evoke movement-induced dopamine release artifacts in dorsal striatum.



Figure S2: Optical stimulation of midbrain dopamine neurons increases in striatal serotonin and 3-methoxytyramine. Representative chromatograms from a Chrimson-transfected mouse under basal conditions (gray), and in response to optical stimulation (orange) or high-K<sup>+</sup> perfusion (red). Both optical stimulation and high-K<sup>+</sup> perfusion induced increases in neurochemicals (peaks 2 and 3), in addition to dopamine (peak 1).
 Chromatogram of a standard containing 250 pM dopamine (peak 1) and serotonin (peak 3) is shown in black. Peaks 2 and 3 in the dialysate samples could not be definitively identified based on comparison with retention times in the standard chromatogram.



Figure S3: Normalized responses to optical stimulation. A. Time courses of %basal dialysate levels for DA (top, red), 3-MT (middle, pink), and serotonin (5-HT; bottom, blue) in mice expressing Chrimson (left) vs. mice transfected with a control protein (right). Optically induced overflow of dopamine, 3-MT, and serotonin were only detected in the Chrimson animals. B. The magnitudes of overflow are represented as areas under the curve percent (AUC (%)). Dialysate serotonin concentrations were below the detectable threshold in 2/11 Chrimson mice and 2/9 control mice. The yellow bars indicate optical stimulations (5 min).
 \*\*P<0.01 and \*\*\*P<0.001.</li>



Figure S4: RNAscope in situ hybridization controls in dorsal raphe. A. The RNAscope<sup>®</sup> Multiplex Fluorescent Assay as a 3-plex positive control. The RNA polymerase II subunit RPB1 (Polr2a, C1 channel), cyclophilin B (PPIB, C2 channel), and ubiquitin C (UBC, C3 channel) are mRNAs found in all mouse cells. Cell nuclei stained by DAPI are shown in blue. The overlay is shown on the right
B. The RNAscope<sup>®</sup> Multiplex Fluorescent Assay as a 3-plex negative control. A probe for DapB, an mRNA that codes for a reductase enzyme from *Bacillus subtilis*, was used in all three channels with each of the opal dyes to evaluate background staining.



Figure S5: Standard curves for dopamine, 3-methyltyramine 3-MT), and serotonin. Fourteen standards (0 nM, 0.008 nM, 0.016 nM, 0.032 nM, 0.063 nM, 0.125 nM, 0.250 nM, 0.500 nM, 0.625 nM, 1 nM, 1.25 nM, 2.5 nM, 5 nM, and 10 nM) were injected into the HPLC (20 μL volumes) to create standard curves. Insets are zoomed in on the lower concentrations ranging from 0-1 nM. Quadratic curve-fits were applied to A. dopamine, B. 3-MT, and C. serotonin standards. Each point represents *N*=3 replicates measured on different days. Error bars (standard errors of the means) are too small to be visualized in some cases.

## Table S1: Statistical summary

FIGURE	COMPARISON	TEST	RESULTS	SIGNIFICANT?
2C	Basal DA: control vs.	Unpaired two-tailed	t (18)=1.6; <i>P</i> >0.1	No
	Chrimson	t-test		
	AUC DA: control vs.	Unpaired two-tailed		
2D	Chrimson	t-test	t (18)=3.0; <i>P</i> <0.01	**
	DA: 5 mins pre- <i>vs.</i> 60			
3A	mins post-ESC	Paired two-tailed t-test	t (2)=0.92; <i>P</i> >0.4	No
	5HT: 5 mins pre- vs.	Paired two-tailed		
3A	60 mins post-ESC	t-test	t (2)=5.7; <i>P</i> <0.05	*
	DA: 5 mins pre vs. 60	Ratio paired two-tailed		
3C	mins post TOL	t-test	t (3) = 0.83; <i>P</i> >0.46	No
	3MT: 5 mins pre vs.	Ratio paired two-		alaala
30	60 mins post TOL	tailed t-test	t(3) = 9.6; P < 0.01	**
20	5HT: 5 mins pre vs. 60	Ratio paired two-tailed		N
36	mins post IUL	t-test	t(3) = 1.3; P > 0.29	NO
4.4	Basal 3M1: control vs.	t tost	+ (10)=0.27, 0>0.7	No
<b>4A</b>	Racal 5HT: control vs	I-lesi	l(10)=0.27; P>0.7	NO
4.4	Chrimson	t-test	t (18)-0 52· P>0 6	No
тл	AIIC 3MT: control vs	Unnaired two-tailed	t(10)=0.52, 1>0.0	NO
<b>4</b> C	Chrimson	t-test	t (18)=3.1: <i>P</i> <0.01	**
10	AUC 5HT: control vs.	Unpaired two-tailed		
4C	Chrimson	t-test	t (15)=4.4; <i>P</i> <0.001	***
	Basal DA: pre- <i>vs.</i>	Ratio paired two-		
6A	post-SCH	tailed t-test	t (3)=4.4; <i>P</i> <0.05	*
	Basal 3MT: pre- <i>vs.</i>	Ratio paired two-tailed		
6A	post-SCH	t-test	t (3) = 0.17; <i>P</i> >0.87	No
	Basal 5HT: pre- <i>vs.</i>	Ratio paired two-		
6A	post-SCH	tailed t-test	t (3) = 3.5; <i>P</i> <0.05	*
	AUC DA: pre- vs. post-	Ratio paired two-		
6C	SCH	tailed t-test	t (3) = 6.2; <i>P</i> <0.05	**
	AUC 3MT: pre- <i>vs.</i>	Ratio paired two-		
6C	post-SCH	tailed t-test	t (3) = 4.8; <i>P</i> <0.05	*
	AUC 5HT: pre- vs.	Ratio paired two-		ala.
6C	post-SCH	tailed t-test	t (3) = 4.5; <i>P</i> <0.05	*
70	AUC (%) DA: pre vs.	Ratio paired two-tailed	+ (2) 2 4 D 0 1	<b>Т</b>
/B	POSUSCH	l-lest	l(3) = 2.4; P < 0.1	Trend
70	AUC (%) 3MT: pre vs.	tailed t test	+ (2) = 2 7, D<0.0E	*
7 D	AUC (06) 5HT: prove	Patio paired two tailed	t(3) = 3.7; F < 0.05	-
7 <b>R</b>	nost SCH	t-test	t (3) – 0 41· P>0 71	No
	Basal DA: nre vs. nost	Ratio paired two-tailed	(0) = 0.71, 1 > 0.71	110
8A	ETC	t-test	$t(3) = 0.31 \cdot P > 0.78$	No
	Basal 3MT: pre vs. post	Ratio paired two-tailed		
8A	ETC	t-test	t (3) = 0.81: <i>P</i> >0.47	No
	Basal 5HT: pre vs. post	Ratio paired two-tailed		
8A	ETC	t-test	t (2) = 2.7; <i>P</i> >0.11	No

	AUC DA: pre vs. post	Ratio paired two-tailed		
8C	ETC	t-test	t (3) = 1.5; <i>P</i> <0.23	No
	AUC 3MT: pre vs. post	Ratio paired two-tailed		
8C	ETC	t-test	t (3) = 3.1; <i>P</i> <0.06	Trend
	AUC 5HT: pre vs. post	Ratio paired two-tailed		
8C	ETC	t-test	t (2) = 1.4; <i>P</i> >0.28	No
	AUC (%) DA: pre vs.	Ratio paired two-tailed		
9B	post ETC	t-test	t (3) = 2.6; <i>P</i> <0.08	Trend
		D		
	AUC (%) 3MT: pre vs.	Ratio paired two-		
9B	AUC (%) 3MT: pre vs. post ETC	Ratio paired two- tailed t-test	t (3) = 4.4; <i>P</i> <0.05	*
9B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs.	<b>tailed t-test</b> Ratio paired two-tailed	t (3) = 4.4; <i>P</i> <0.05	*
9B 9B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC	Ratio paired two- tailed t-test Ratio paired two-tailed t-test	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21	* No
9B 9B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC AUC (%) DA: control	Ratio paired two-tailed t-testRatio paired two-tailedt-testUnpaired two-tailed	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21	* No
9B 9B S3B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC AUC (%) DA: control vs. chrimson	Ratio paired two- tailed t-test Ratio paired two-tailed t-test Unpaired two-tailed t-test	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21 t (18) = 5.9; P<0.001	* No ***
9B 9B \$3B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC AUC (%) DA: control vs. chrimson AUC (%) 3MT: control	Ratio paired two- tailed t-test Ratio paired two-tailed t-test Unpaired two-tailed t-test Unpaired two-tailed	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21 t (18) = 5.9; P<0.001	* No ***
9B 9B S3B S3B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC AUC (%) DA: control vs. chrimson AUC (%) 3MT: control vs. chrimson	Ratio paired two- tailed t-test Ratio paired two-tailed t-test Unpaired two-tailed t-test Unpaired two-tailed t-test	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21 t (18) = 5.9; P<0.001 t (18) = 4.1; P<0.001	* No ***
9B 9B S3B S3B	AUC (%) 3M1: pre vs. post ETC AUC (%) 5HT: pre vs. post ETC AUC (%) DA: control vs. chrimson AUC (%) 3MT: control vs. chrimson AUC (%) 5HT: control	Ratio paired two- tailed t-test Ratio paired two-tailed t-test Unpaired two-tailed t-test Unpaired two-tailed t-test Unpaired two-tailed	t (3) = 4.4; P<0.05 t (2) = 1.8; P>0.21 t (18) = 5.9; P<0.001 t (18) = 4.1; P<0.001	* No *** ***

\*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001