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Supplemental information

**A dual-function phage regulator controls
the response of cohabiting phage elements
via regulation of the bacterial SOS response**

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C. Clustal W alignment of deduced amino acid sequences of AriS (LMRG_02920; *comK*-prophage, *L. monocytogenes* strain 10403S) and Lin2418 (*comK*-prophage, *L. innocua* strain CLIP 11262). Alignment length: 266 residues. Identical amino acids (30.45%) are shown in red and marked with asterisks, highly similar residues (24.81%) in green and marked with colons, and weakly similar residues (12.03%) in blue and marked with dots; different residues (32.71%) in black and unmarked. A tyrosine residue (Y99) of AriS that was predicted to be phosphorylated is shown in bold and underlined. The glutamine Q144 of AriS (in bold and yellow highlighted) is the last residue of the recombinant amino-terminal domain expressed by pPL2-*antA/B*. The basic sequence alignment was performed using the public server of the PRABI Rhone-Alpes Bioinformatics Center (France) running the Clustal W program (https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_clustalw.html), and then manually curated. **D.** Clustal W alignment of deduced amino acid sequences of AriS (LMRG_02920; *comK*-prophage, *L. monocytogenes* strain 10403S) and Lmo2324 (*comK*-prophage, *L. monocytogenes* strain EGD-e). Alignment length: 259 residues. Identical amino acids (83.40%) are shown in red and marked with asterisks, highly similar residues (7.34%) in green and marked with colons, and weakly similar residues (3.86%) in blue and marked with dots; different residues (5.41%) in black and unmarked. Tyrosine residues of AriS and Lmo2324 (Y99 and Y98, respectively) that were predicted to be phosphorylated are shown in bold and underlined. The glutamine Q144 of AriS (in bold and yellow highlighted) is the last residue of the recombinant amino-terminal domain expressed by pPL2-*antA/B*. The basic sequence alignment was performed using the public server of the PRABI Rhone-Alpes Bioinformatics Center (France) running the Clustal W program (https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_clustalw.html), and then manually curated. Related to Figure 4.

Figure S2

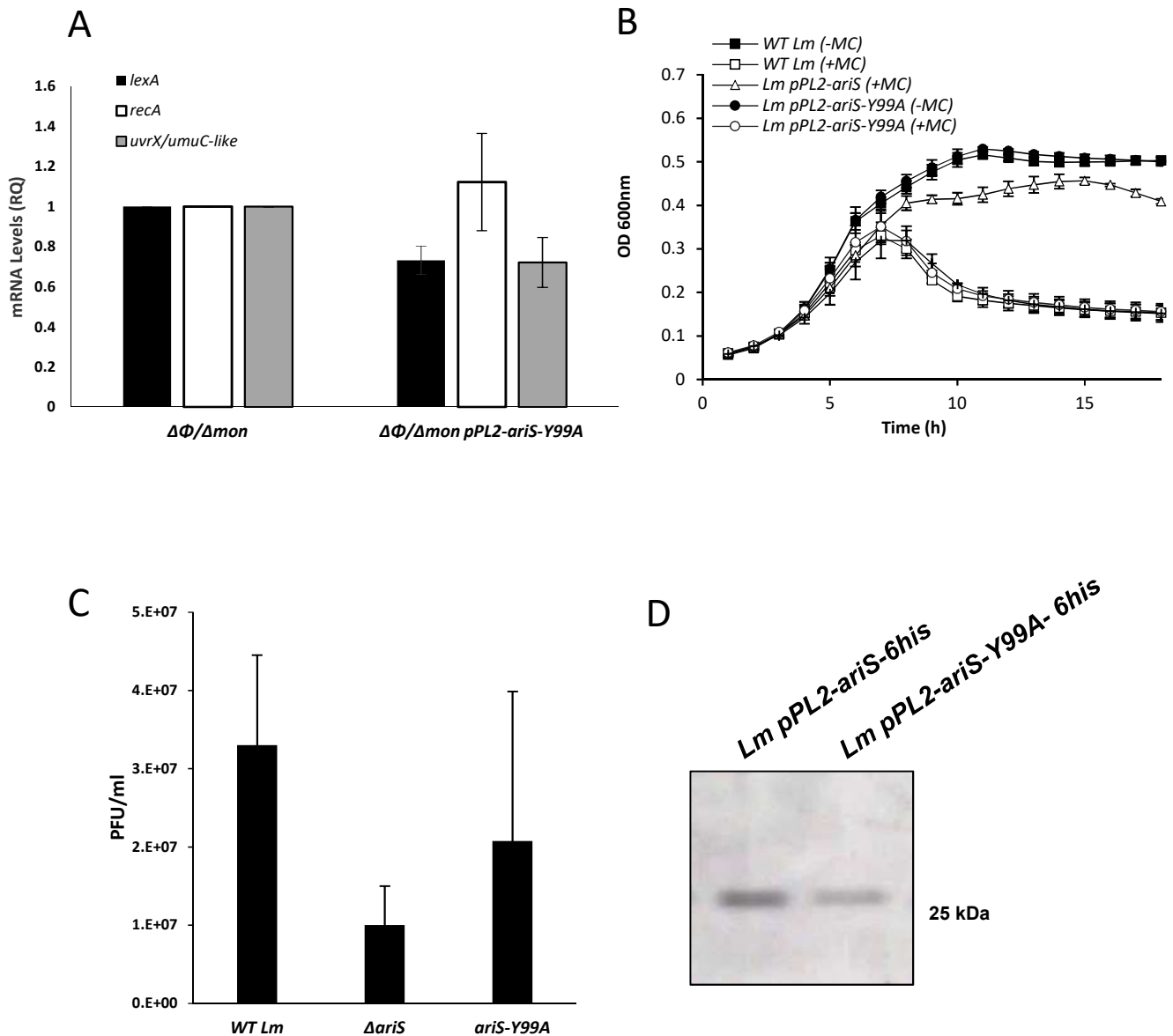


Figure S2. Substitution of AriS tyrosine 99 with alanine abolishes its function.

A. RT-qPCR analysis of representative SOS genes (*recA*, *lexA* and *uvrX*) in $\Delta\phi/\Delta mon$ and $\Delta\phi/\Delta mon$ bacteria expressing AriS-Y99A (using pPL2-*ariS*-Y99A). Indicated strains were grown in BHI medium with MC treatment, at 30 °C, for 45 min. mRNA levels are presented as relative quantity (RQ), relative to their levels in WT bacteria. The error bars represent the standard deviation of three independent experiments. **B.** Growth analysis of WT *Lm* with and without overexpression of AriS-Y99A (*Lm* pPL2-*ariS*-Y99A) in the presence (+) or absence (-) of MC, at 30 °C. Overexpression of *ariS* (*Lm* pPL2-*ariS*) was used as a control. The experiment was performed three times and the figure shows a representative result. The error bars represent the standard deviation of three independent experiments, and are sometimes hidden by the symbols. **C.** Virions obtained from MC-treated cultures (6 h after MC treatment) of WT *Lm*, a deletion mutant of the *ariS* gene ($\Delta ariS$) or *Lm* bacteria possessing the *ariS*-Y99A mutation in the prophage genome (i.e., in the *Lm* genome) were tested on an indicator strain for plaque formation (numerated as plaque-forming units, PFUs). The error bars represent the standard deviation of three independent experiments. **D.** Western blot analysis comparing the protein levels of His-tagged AriS and AriS-Y99A proteins expressed using the pPL2 plasmid. Bacteria were grown in BHI medium for 6 h. Equal amounts of total proteins were separated on 15% SDS-PAGE, blotted and probed with anti-His tag antibody. Related to Figure 7.