

# **Prospective multicentre head-to-head validation of host blood transcriptomic biomarkers for pulmonary tuberculosis by real-time PCR**

Simon C. Mendelsohn, Stanley Kimbung Mbandi, Andrew Fiore-Gartland, Adam Penn-Nicholson, Munyaradzi Musvosvi, Humphrey Mulenga, Michelle Fisher, Katie Hadley, Mzwandile Erasmus, Onke Nombida, Michèle Tameris, Gerhard Walzl, Kogieleum Naidoo, Gavin Churchyard, Mark Hatherill, and Thomas J. Scriba

## **SUPPLEMENTARY INFORMATION**

### **Contents**

Supplementary Notes 1	2
Supplementary Notes 2	5
Supplementary Figures	6
Supplementary References	21

## Supplementary Notes 1

### The CORTIS Study Team

*Centre for the AIDS Programme of Research in South Africa (CAPRISA), and MRC-CAPRISA HIV-TB Pathogenesis and Treatment Research Unit, Doris Duke Medical Research Institute, University of KwaZulu-Natal, Durban, South Africa.*

Bianca Bande  
Thilagavathy Chinappa  
Cara-Mia Corris  
Thobelani Cwele  
Celaphiwe Dlamini  
Dhineshree Govender  
Goodness Khanyisile Gumede  
Nonhlanhla Zanele Elsie Gwamanda  
Senzo Halti  
Razia Hassan-Moosa  
Senzo Ralph Hlathi  
Zandile Patrica Jali  
Lungile Khanyile  
Bhavna Maharaj  
Jabulisiwe Lethabo Maphanga  
Nonhle Bridgette Maphanga  
Siyabonga Mbatha  
Atika Moosa  
Nompumelelo Ngcobo  
Ntombozuko Gloria Ntanjana  
Sphelele Simo Nzimande  
Nesri Padayatchi  
Dirhona Ramjit  
Thandiwe Yvonne Shezi  
Zibuyile Phindile Penlee Sing  
Chandrapharbha Singh  
Philile Thembela  
Londiwe Zaca  
Mbali Ignatia Zulu

*Department of Molecular Microbiology, Washington University in St. Louis, St. Louis, MO, USA*  
Shabaana A. Khader

*Division of Medical Microbiology, Department of Pathology, University of Cape Town, Cape Town, South Africa*  
Slindile Mbhele  
Mark P. Nicol  
Judi van Heerden

*DST/NRF Centre of Excellence for Biomedical TB Research and SAMRC Centre for TB Research, Division of Molecular Biology and Human Genetics, Department of Biomedical Sciences, Faculty of Medicine and Health Sciences, Stellenbosch University, Cape Town, South Africa.*  
Petri Ahlers  
Roslyn Beukes  
Maria Didloff  
Marika Flinn  
Bernadine Fransman  
Andriëtte Hiemstra

Justine Khoury  
Belinda Kriel  
Jaftha Kruger  
Andre Loxton  
Elizna Maasdorp  
Stephanus T. Malherbe  
Onesisa Mpofo  
Mpho Mtlali  
Liesel Muller  
Bronwyn Smith  
Dorothy Solomons  
Kim Stanley  
Susanne Tonsing  
Khayaletu Toto  
Ayanda Tsamane  
Susanne Tönsing

*South African Tuberculosis Vaccine Initiative, Institute of Infectious Disease and Molecular Medicine, Division of Immunology, Department of Pathology, University of Cape Town, South Africa.*

Charmaine Abrahams  
Hadn Africa  
Denis Arendsen  
Nomfuneko Cynthia Batyi  
Nicole Bilek  
Natasja Botes  
Samentra Braaf  
Sivuyile Buhlungu  
Alida Carstens  
Balie Carstens  
Nompumelelo Cetywayo  
Yolundi Cloete  
Lorraine Coetzee  
Alessandro Companie  
Ilse Davids  
Marwou de Kock  
Bongani Diamond  
Palesa Dolo  
Margareth Erasmus  
Juanita Ferreira  
Christal Ferus  
Elizabeth Filander  
Hennie Geldenhuys  
Diann Gempies  
Yolande Gregg  
Rieyaat Hassiem  
Roxane Herling  
Yulandi Herselman  
Chris Hikuam  
Henry Issel  
Ruwyda Jansen  
Lungisa Jaxa  
Fabio Julies  
Fazlin Kafaar  
Masooda Kaskar  
Sophie Keffers  
Xoliswa Kelepu

Gloria Khomba  
 Sandra Kruger  
 Sunelza Lakay  
 Thelma Leopeng  
 Angelique Kany Kany Luabeya  
 Simbarashe Mabwe  
 Lauren Mactavie  
 Nomsitho Magawu  
 Lebohang Makhete  
 Lebohang Makhethe  
 Faheema Meyer  
 Miriam Moses  
 Boitumelo Mosito  
 Angelique Mouton  
 Julia Noble  
 Nambitha Nqakala  
 Fajwa Opperman  
 Christel Petersen  
 Patiswa Plaatjie  
 Abe Pretorius  
 Rodney Raphela  
 Frances Ratangee  
 Maigan Ratangee  
 Susan Rossouw  
 Elisma Schoeman  
 Constance Schreuder  
 Alison September  
 Cashwin September  
 Justin Shenje  
 Marcia Steyn  
 Sonia Stryers  
 Leticia Swanepoel  
 Anne Swarts  
 Asma Toefy  
 Petrus Tyambetyu  
 Habibullah Valley  
 Linda van der Merwe  
 Elma van Rooyen  
 Ashley Veldsman  
 Helen Veldtsman  
 Kelvin Vollenhoven  
 Elaine Zimri

*TB Modelling Group, TB Centre, Centre for  
 Mathematical Modelling of Infectious Diseases,  
 Department of Infectious Disease Epidemiology,  
 London School of Hygiene & Tropical Medicine,  
 London, United Kingdom.*

Tom Sumner  
 Richard G. White

*The Aurum Institute, Johannesburg, Gauteng, South  
 Africa.*

*Aurum Klerksdorp Site*  
 Tebogo Badimo  
 Kagiso Baepanye  
 Kesenogile Edna Baepanye  
 Tshepiso Baepanye  
 Ken Clarke  
 Marelize Collignon  
 Audrey Lebohang Dhlamini  
 Audrey Dlamini

Candice Eyre  
 Tebogo Feni  
 Moogo Fikizolo  
 Phinda Galane  
 Alia Gangat  
 Thelma Goliath  
 Craig Innes  
 Bonita Janse van Rensburg  
 Elba Janse van Rensburg  
 Olebogeng Jonkane  
 Boitumelo Sophy Kekana  
 Gomotsegang Virginia Khobedi  
 Marietjie King  
 Adrienne Kock  
 Ndlela Israel Kunene  
 Aneesa Lakhi  
 Nondumiso Langa  
 Hildah Ledwaba  
 Marilynn Lumphoko  
 Immaculate Mabasa  
 Tshegofatso Dorah Mabe  
 Nkosinathi Charles Mabuza  
 Molly Majola  
 Mantai Makhetha  
 Blossom Makhubalo  
 Mpho Makoanyane  
 Vernon Malay  
 Shirley Malefo-Grootboom  
 Juanita Market  
 Selvy Matshego  
 Lungile Mbata  
 Nontsikelelo Mbipa  
 John Mdlulu  
 Tsiamo Mmotsa  
 Karabo Moche  
 Sylvester Modipa  
 Joseph Panie Moloko  
 Kabelo Molosi  
 Samuel Mopati  
 Palesa Moswegu  
 Primrose Mothaga  
 Dorothy Muller  
 Grace Nchwe  
 Nhlamulo Ndlovu  
 Maryna Nel  
 Lindiwe Nhlangulela  
 Tanya Nielson  
 Bantubonke Bertrum Ntamo  
 Lawrence Ntoahae  
 Tedrius Ntshauba  
 Thandiwe Papalagae  
 Pedro Pinho  
 Pearl Nomsa Sanyaka  
 Sharfuddin Sayed  
 Letlhogonolo Seabela  
 Raesibe Agnes Pearl Selepe  
 Melissa Neo Senne  
 Moeti Serake  
 Naydene Slabbert  
 Constance Takavamanya  
 Marthinette Taljaard  
 Maria Thlapi

Mugwena Thompo  
Vincent Tshikovhi  
Lebogang Isaac Tswaile  
Amanda van Aswegen  
Marietjie Zietsman

*Aurum Rustenburg Site*

Laudicia Tshenolo Bontsi  
Obakeng Peter Booi  
Mari Cathrin Botha  
William Brumskine  
Salemeng Matseliso Carol  
Kgomotso Violet Chauke  
Mooketsi Theophilius Cwaile  
Isabella Johanna Davies  
Emilia De Klerk  
Blanchard Mbay Iyemosolo  
James Michael Jeleni  
Christian Mabika Kasongo  
Sebaetseng Jeanette Kekana  
Lucky Sipho Khoza  
Gloria Keitumetse Kolobe  
Lerato Julia Lekagane  
Sheiley Christina Lekotloane  
Ilze Jeanette Louw  
Sarah Teboso Lusale  
Perfect Tiisetso Maatjie  
Kamogelo Fortunate Mabena  
Johanna Thapelo Madikwe  
Octavia Mahkosazana Madikwe  
Rapontwana Letlhogonolo Maebana  
Malobisa Sylvester Magwasha  
Vutlhari-I-Vunhenha Fairlord Manzini  
Isholedi Samuel Maroele  
Omphile Petunia Masibi  
July Rocky Mathabanzini  
Tendamudzimu Ivan Mathode  
Ellen Ditaba Matsane  
Lungile Mbata

Nyasha Karen Mhandire  
Thembsiwe Miga  
Caroline Mkhokho  
Neo Hilda Mkwalase  
Nondzakazi Mnqonywa  
Brenda Matshidiso Modisaotsile  
Patricia Pakiso Mokgetsengoane  
Kegomoditswe Magdeline Molatlhegi  
Thuso Andrew Molefe  
Motlatsi Evelyn Molotsi  
Tebogo Edwin Montwedi  
Boikanyo Dinah Monyemangene  
Hellen Mokopi Mooketsi  
Tshplpfelo Mapula Mosito  
Ireen Lesebang Mosweu  
Banyana Olga Motlagomang  
Funeka Nomvula Mthembu  
Themba Phakathi  
Mapule Ozma Phatshwane  
Victor Kgothatso Rameetse  
Kelebogile Magdeline Segaetsho  
Ni Ni Sein  
Melissa Neo Senne  
Sifiso Cornelius Shezi  
Zona Sithetho  
Bongiwe Stofile  
Mando Mmakhora Thaba  
Nosisa Charity Thandeka  
Lethabo Collen Theko  
Dimakatso Sylvia Tsagae

*Vaccine and Infectious Disease Division, Fred  
Hutchinson Cancer Research Center, Seattle, WA, USA.*

Bhavesb Borate  
Eva Chung  
Michelle Chung  
Ellis Hughes  
Alicia Sato  
Steven Self

**The Cross-sectional TB Cohort Study Team**

*South African Tuberculosis Vaccine Initiative, Institute  
of Infectious Disease and Molecular Medicine,  
Division of Immunology, Department of Pathology,  
University of Cape Town, South Africa.*

Hadn Africa  
Janelle Botes  
Fatoumatta Darboe  
Elizabeth Filander  
Lebohang Makhetha  
Sindile Matiwane  
Melissa Murphy

Constance Schreuder  
Marcia Steyn  
Michele van Rooyen  
Noncedo Xoyana

*Desmond Tutu HIV Centre, and Institute of Infectious  
Disease and Molecular Medicine (IDM), University of  
Cape Town, Cape Town, South Africa*

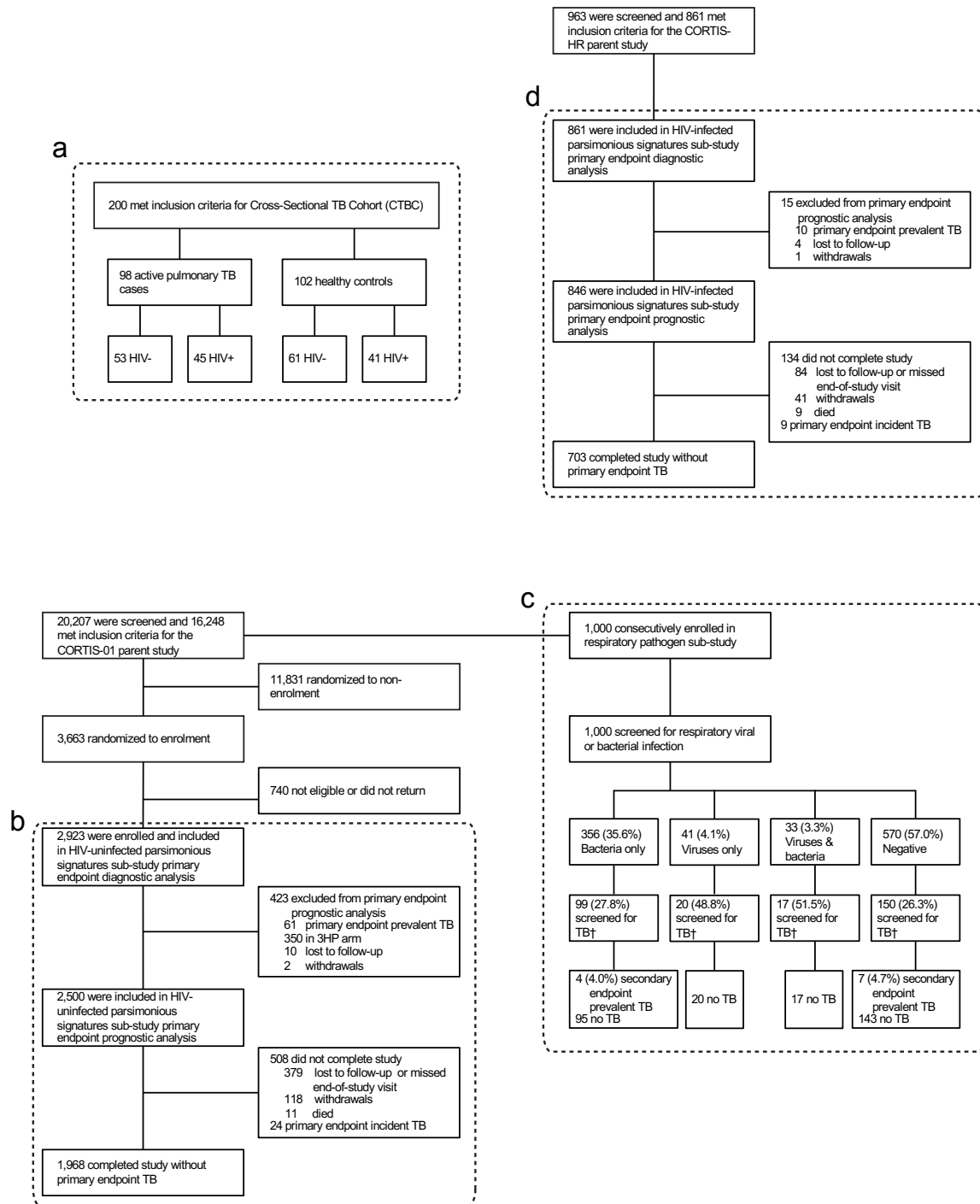
Carl Morrow  
Robin Wood

## Supplementary Notes 2

### *Signature failure rate*

Although different pass rates may have biased comparison of performance between signature, the primary aim of the study was to compare individual signature performance to the WHO Target Product Profile (TPP) criteria benchmarks rather than against each other. Hence samples with failed signatures were still included in analysis to increase statistical power. Failure rate is also an indicator of signature robustness; an important criterion when selecting biomarkers for further development and clinical translation. The high failure rate of the Suliman4 signature was attributed to the GAS6 primer-probe assay, which performed well in assay qualification (**Table S3 in Supplementary Data 1**) and validation in high yield and purity manually extracted PAXgene RNA samples. However, performance of the GAS6 assay was less robust in lower yield and quality RNA from automated robotic extraction in the CORTIS studies. Due to the pair-wise ensemble structure of three up- and three down-regulated genes, RISK6 score can still be calculated even if one or more transcript is not detected due to failed PCR amplification.<sup>1</sup> As a result, RISK6 was resilient to single failed PCR reactions, with highest signature pass rate. Only HIV-uninfected participants with successful measurement of the RISK11 score were enrolled in the CORTIS-01 study, whereas HIV-infected participants were enrolled irrespective of RISK11 score in CORTIS-HR. This effectively excluded participants in CORTIS-01 with very low yield or purity RNA samples, resulting in a lower signature failure rates in CORTIS-01 as compared to CORTIS-HR.

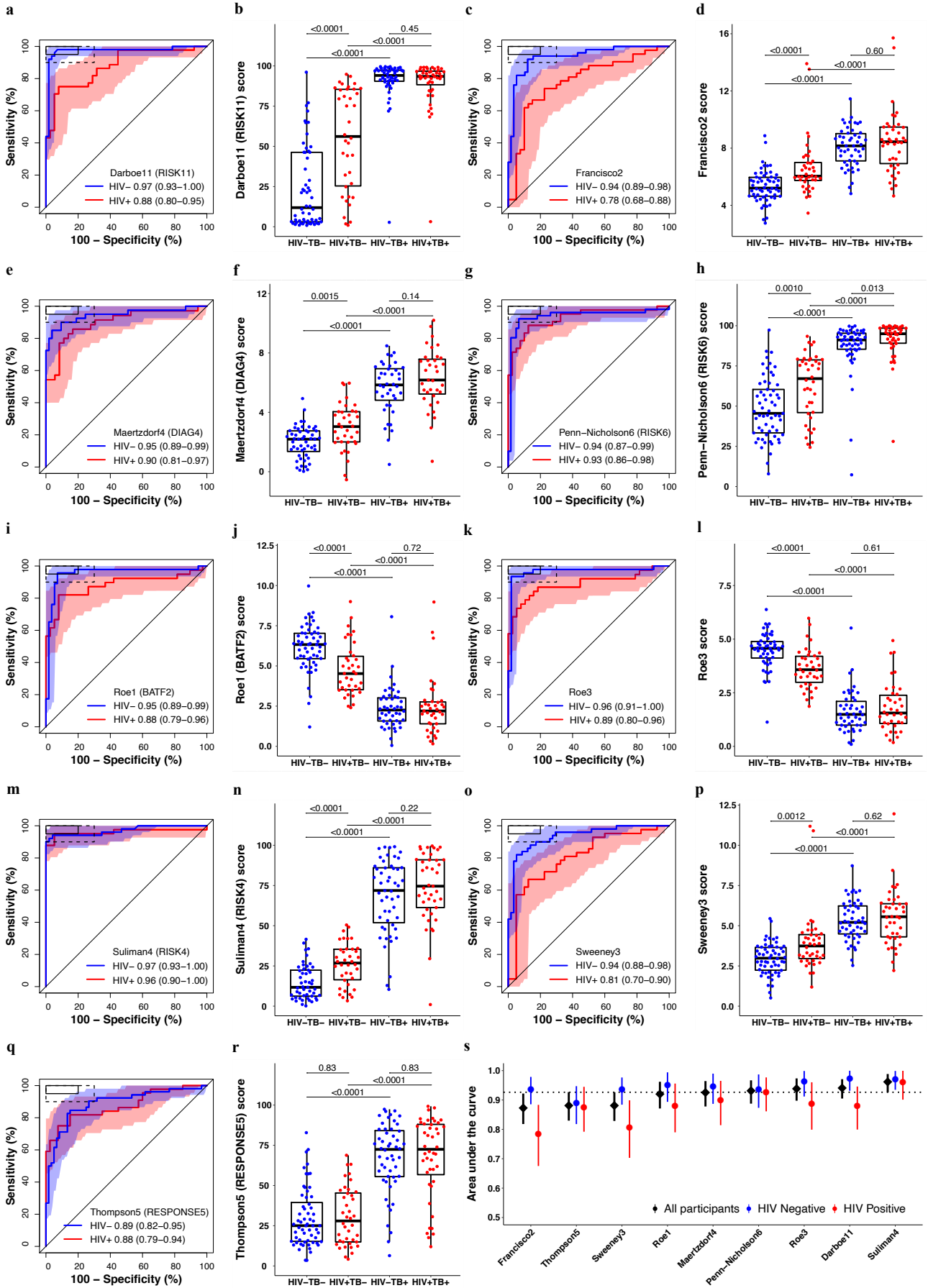
## Supplementary Figures



**Figure S1: Study flow diagram**

(a) HIV-infected and HIV-uninfected participants with and without TB were recruited into the Cross-Sectional TB Cohort (CTBC) case-control study. HIV-uninfected participants were recruited from the CORTIS-01 parent study and co-enrolled into the (b) HIV-uninfected parsimonious signatures sub-study and (c) respiratory pathobionts sub-study. HIV-infected participants were recruited from the CORTIS-HR parent study and co-enrolled into the (d) HIV-infected parsimonious signatures sub-study. For the CTBC study and respiratory pathobionts sub-study, TB disease was defined by a single sputum sample positive for *Mtb* on either Xpert MTB/RIF and/or liquid culture at enrolment. The coprimary endpoints in the CORTIS studies were baseline prevalent TB disease and incident disease through 15 months follow-up confirmed by a positive Xpert MTB/RIF, Xpert Ultra, or MGIT culture, on two or more separate sputum samples collected within any 30-day period. The secondary endpoint was microbiologically-confirmed TB disease on at least one sputum sample.

†Out of 1000 participants enrolled in the respiratory pathobionts cohort, only 286 (28.6%) participants were co-enrolled in the CORTIS-01 parent study and investigated for TB at enrolment; 11/286 (3.8%) had prevalent TB confirmed by *Mycobacterium tuberculosis* liquid culture and/or Xpert MTB/RIF.



**Figure S2: Parsimonious signatures diagnostic performance and signature score distributions in the Cross-sectional Tuberculosis Cohort (CTBC) study**

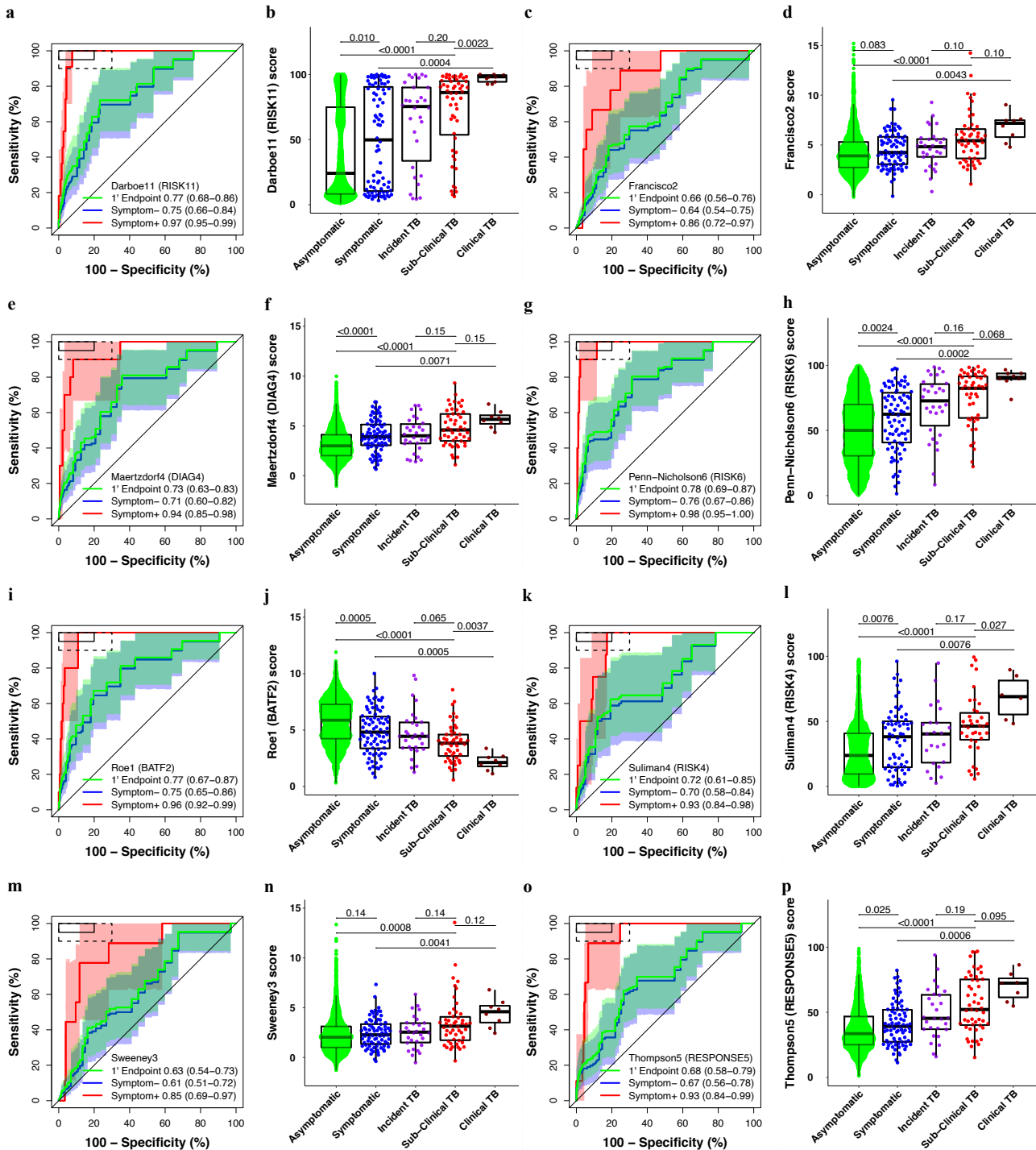
Paired receiver operating characteristic (ROC) curves and box-and-whisker plots for the (a-b) Darboe11 (RISK11), (c-d) Francisco2, (e-f) Maertzdorf4 (DIAG4), (g-h) Penn-Nicholson6 (RISK6), (i-j) Roe1 (BATF2), (k-l) Roe3, (m-n) Suliman4 (RISK4), (o-p) Sweeney3, and (Q-R) Thompson5 (RESPONSE5) signatures in the Cross-sectional Tuberculosis Cohort (CTBC) study.

The ROC curves depict diagnostic performance (area under the curve, AUC, with 95% CI) of the parsimonious signatures for diagnosing TB, stratified by HIV status. The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup>

The box-and-whisker plots depict signature score distribution by HIV (HIV+/HIV-) and prevalent TB disease (TB+/TB-) status. Each dot represents a participant. p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney *U* test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Boxes depict the IQR, the midline represents the median, and the whiskers indicate the  $IQR \pm (1.5 \times IQR)$ .

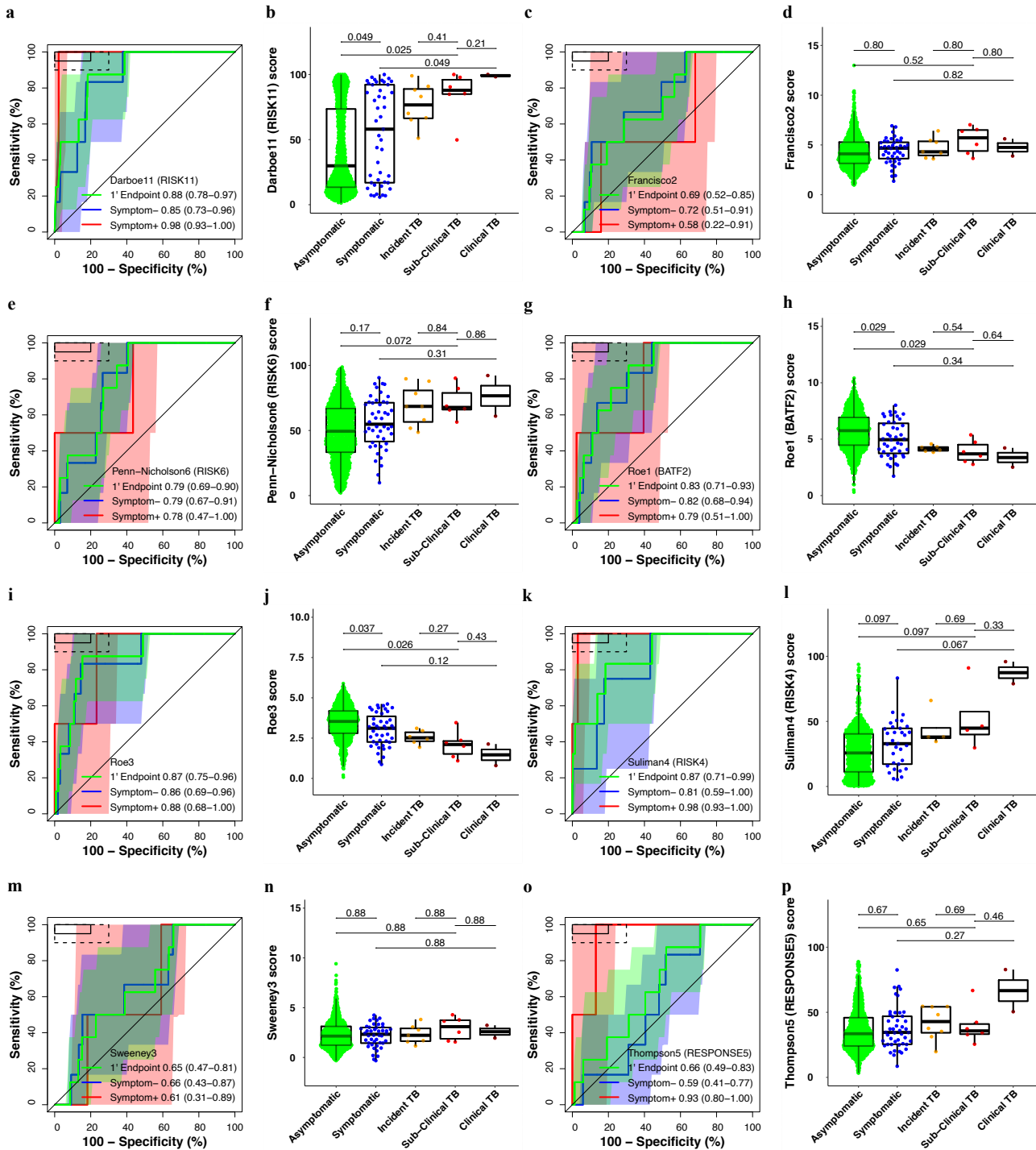
(S) Summary of signature diagnostic performance in order of AUC estimates in all participants. The diagnostic AUC estimates in HIV-infected and HIV-uninfected participant sub-groups are also shown. The midline indicates the AUC estimate, the error bars indicate the 95% CIs, and the black dotted line indicates the lower bound of the 95% CI for the best performing signature for all participants.



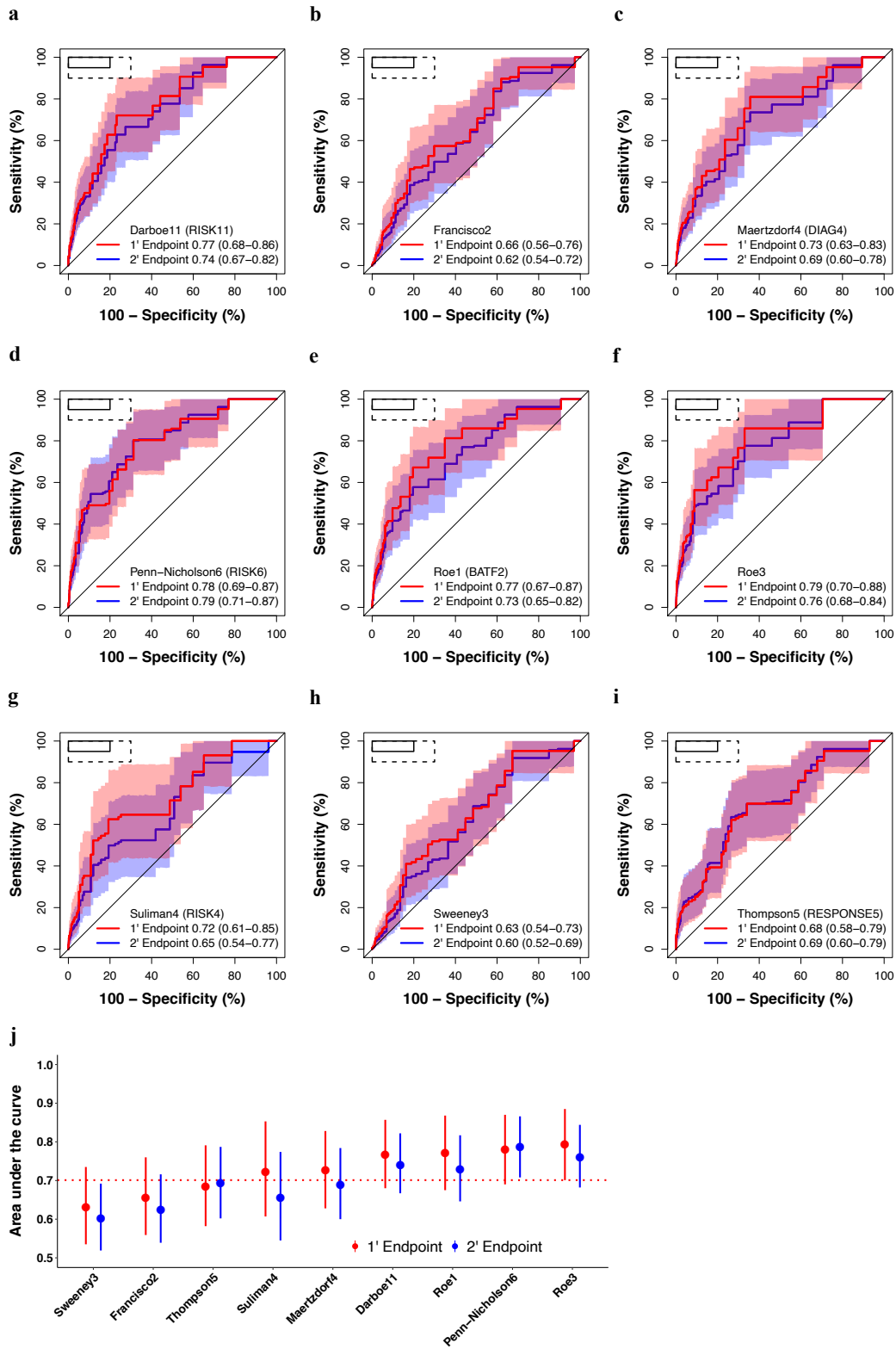


**Figure S3: Parsimonious signature diagnostic performance for prevalent TB and signature score distributions in people without HIV**

Paired receiver operating characteristic (ROC) curves and box-and-whisker plots for the (a-b) Darboe11 (RISK11), (c-d) Francisco2, (e-f) Maertzdorf4 (DIAG4), (g-h) Penn-Nicholson6 (RISK6), (i-j) Roe1 (BATF2), (k-l) Suliman4 (RISK4), (m-n) Sweeney3, and (o-p) Thompson5 (RESPONSE5) signatures in the CORTIS-01 study of people without HIV. The ROC curves depict diagnostic performance (area under the curve, AUC, with 95% CI) of the parsimonious signatures for the primary endpoint (1' Endpoint), i.e. TB diagnosed on two or more liquid culture-positive or Xpert MTB/RIF-positive sputum samples. The ROC curves show participants with symptomatic clinical prevalent TB versus symptomatic controls (Symptom+), and participants with asymptomatic, subclinical prevalent TB versus asymptomatic controls (Symptom-). The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup> The box-and-whisker plots depict signature score (measured at enrolment) distribution by symptom status (each dot represents a participant) in asymptomatic and symptomatic participants with no TB, participants who progressed to incident TB, and participants with prevalent subclinical (asymptomatic) and clinical (symptomatic) TB. Prevalent and incident TB comprised all primary endpoint cases. Symptoms were recorded at the time of enrolment for participants without TB and those with prevalent TB. p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney *U* test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Boxes depict the IQR, the midline represents the median, and the whiskers indicate the IQR  $\pm$  (1.5  $\times$  IQR).



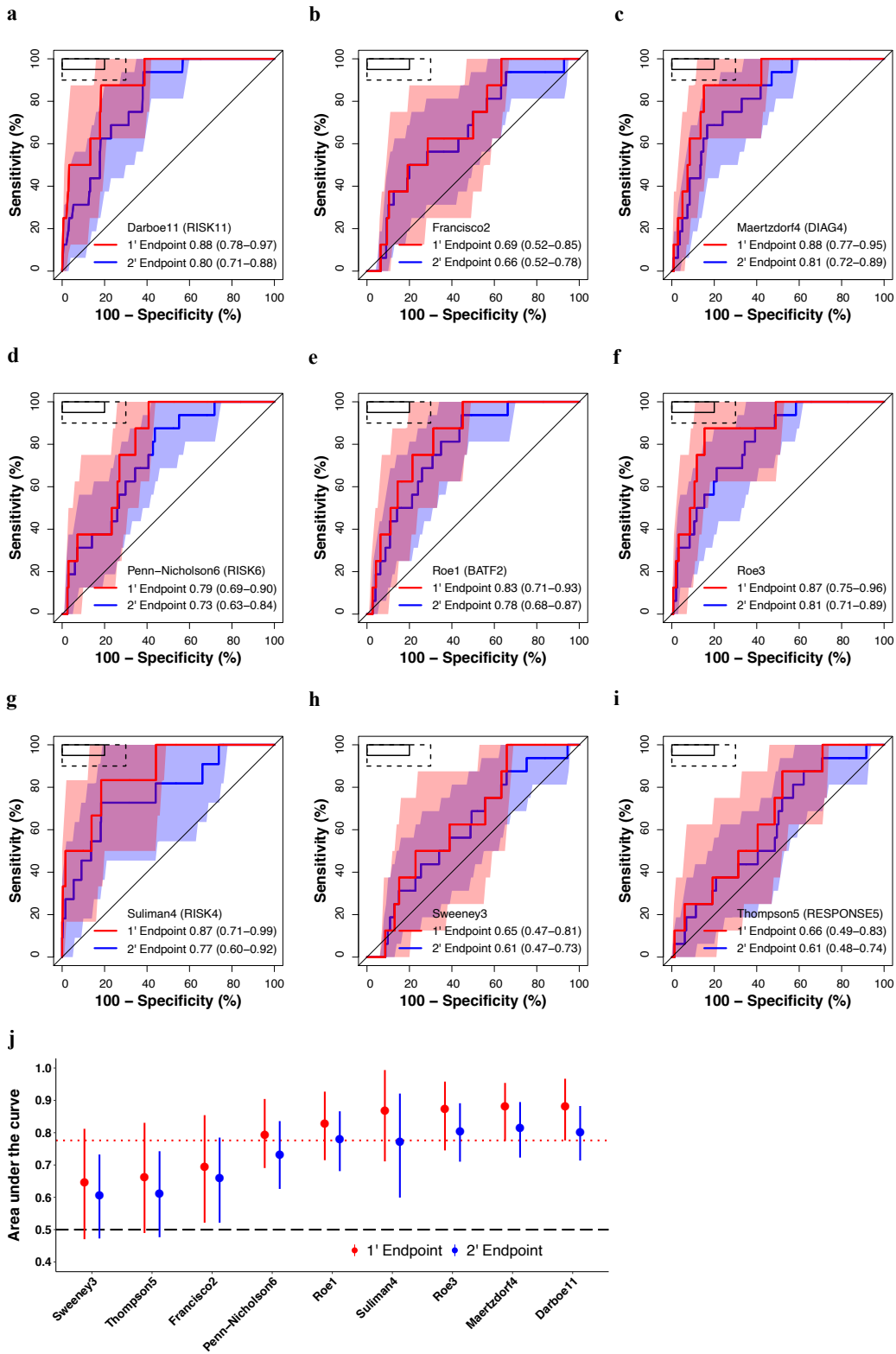
**Figure S4: Parsimonious signatures diagnostic performance for prevalent TB and signature score distributions in people living with HIV.** Paired receiver operating characteristic (ROC) curves and box-and-whisker plots for the (a-b) Darboe11 (RISK11), (c-d) Francisco2, (e-f) Penn–Nicholson6 (RISK6), (g-h) Roe1 (BATF2), (i-j) Roe3, (k-l) Suliman4 (RISK4), (m-n) Sweeney3, and (o-p) Thompson5 (RESPONSE5) signatures in the CORTIS-HR study of people living with HIV. The ROC curves depict diagnostic performance (area under the curve, AUC, with 95% CI) of the parsimonious signatures for TB diagnosed for the primary endpoint (1' Endpoint), i.e. TB diagnosed on two or more liquid culture-positive or Xpert MTB/RIF-positive sputum samples. The ROC curves show participants with symptomatic clinical prevalent TB versus symptomatic controls (Symptom+), and participants with asymptomatic subclinical prevalent TB versus asymptomatic controls (Symptom–). The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup> The box-and-whisker plots depict signature score (measured at enrolment) distribution by symptom status (each dot represents a participant) in asymptomatic and symptomatic participants with no TB, participants who progressed to incident TB, and participants with prevalent subclinical (asymptomatic) and clinical (symptomatic) TB. Prevalent and incident TB comprised all primary endpoint cases. Symptoms were recorded at the time of enrolment for participants without TB and those with prevalent TB. p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney *U* test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Boxes depict the IQR, the midline represents the median, and the whiskers indicate the  $IQR \pm (1.5 \times IQR)$ .



**Figure S5: Parsimonious signatures diagnostic performance for primary versus secondary endpoint TB in people without HIV**

Receiver operating characteristic (ROC) curves depicting primary (\*1) versus secondary (\*2) endpoint diagnostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Penn-Nicholson6 (RISK6), (e) Roe1 (BATF2), (f) Roe3, (g) Suliman4 (RISK4), (h) Sweeney3, and (i) Thompson5 (RESPONSE5) signatures in the CORTIS-01 study of people without HIV. The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup>

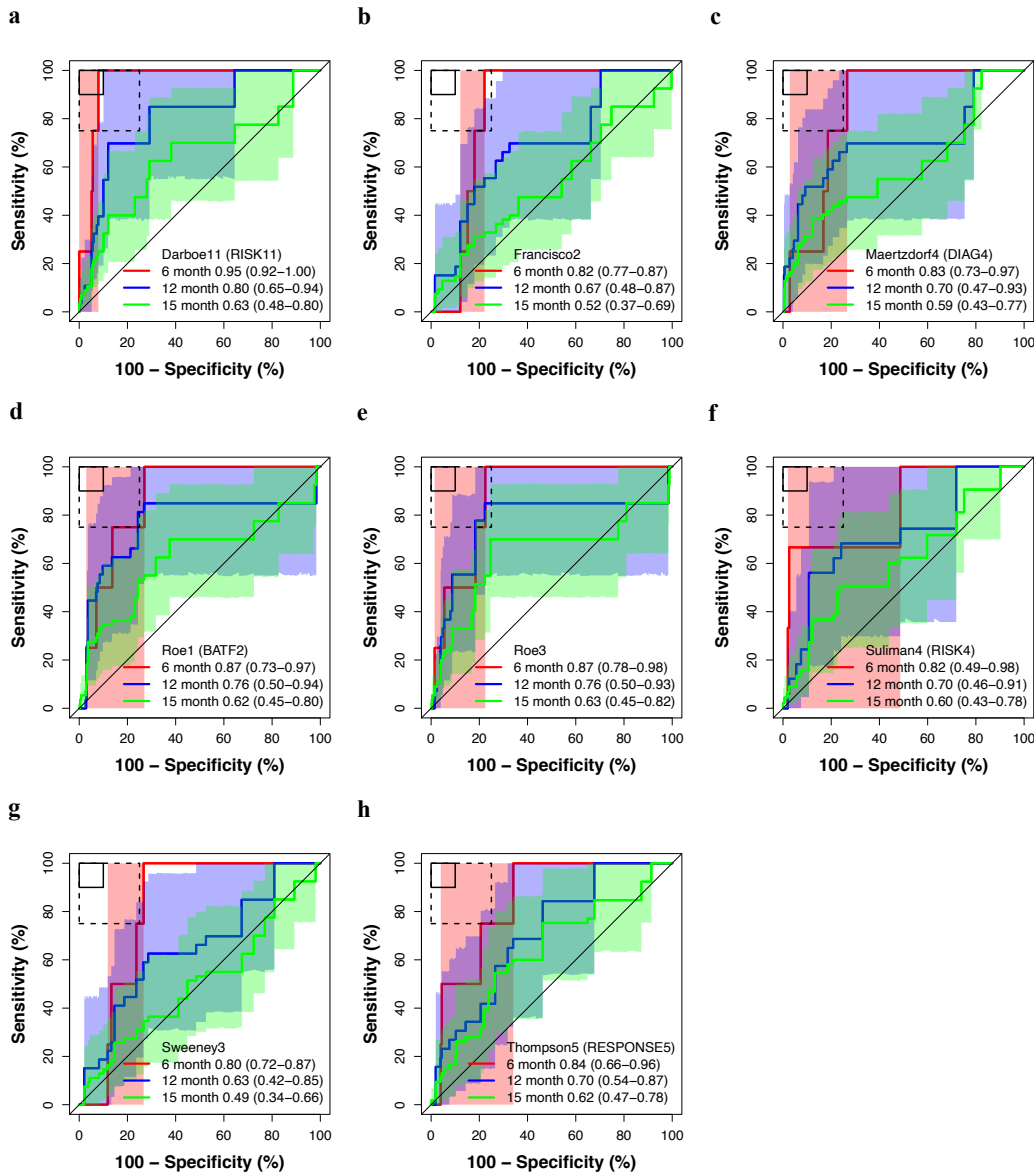
(j) Summary of signature diagnostic performance in order of primary endpoint AUC estimates. The diagnostic AUC estimates for the secondary endpoint are also shown. The midline indicates the AUC estimate, the error bars indicate the 95% CIs, and the red dotted line indicates the lower bound of the 95% CI for the best performing signature for the primary endpoint.



**Figure S6: Parsimonious signatures diagnostic performance for primary versus secondary endpoint TB in people living with HIV**

Receiver operating characteristic (ROC) curves depicting primary (\*1) versus secondary (\*2) endpoint diagnostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Penn-Nicholson6 (RISK6), (e) Roe1 (BATF2), (f) Roe3, (g) Suliman4 (RISK4), (h) Sweeney3, and (i) Thompson5 (RESPONSE5) signatures in the CORTIS-HR study of people living with HIV. The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup>

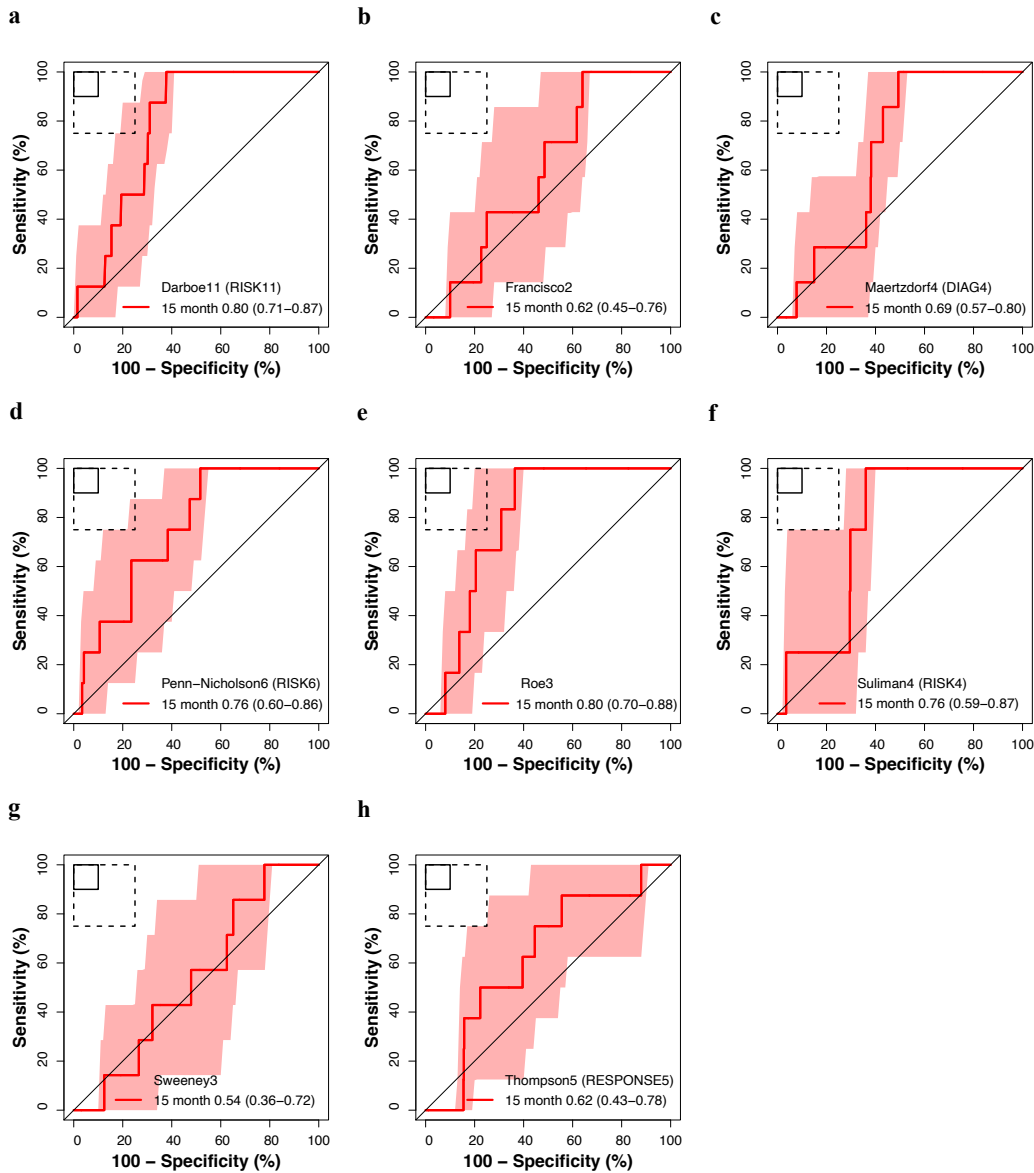
(j) Summary of signature diagnostic performance in order of primary endpoint AUC estimates. The diagnostic AUC estimates for the secondary endpoint are also shown. The midline indicates the AUC estimate, the error bars indicate the 95% CIs, and the red dotted line indicates the lower bound of the 95% CI for the best performing signature for the primary endpoint. The black dashed line indicates an AUC cut-off of 0.5.



**Figure S7: Parsimonious signature prognostic performance for incident TB in people without HIV**

Receiver operating characteristic curves depicting prognostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Roe1 (BATF2), (e) Roe3, (f) Suliman4 (RISK4), (g) Sweeney3, and (h) Thompson5 (RESPONSE5) signatures for incident TB diagnosed on two or more liquid culture-positive or Xpert MTB/RIF-positive sputum samples (primary endpoint) through 6, 12, and 15 months follow-up in the CORTIS-01 study of people without HIV.

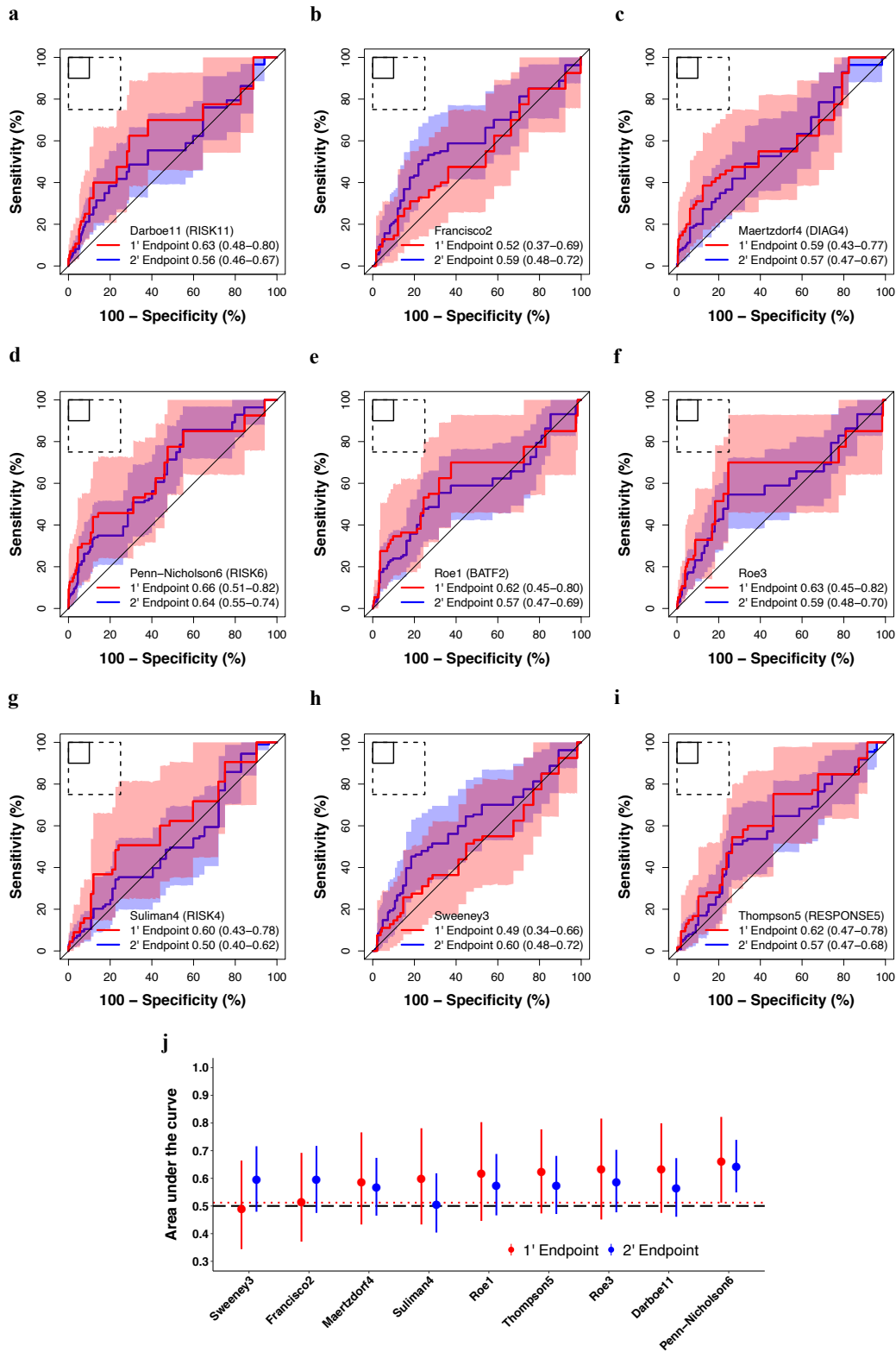
The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (90% sensitivity and 90% specificity) and the dashed box depicts the minimal criteria (75% sensitivity and 75% specificity) set out in the WHO Target Product Profile for an incipient TB test.<sup>4</sup>



**Figure S8: Parsimonious signature prognostic performance for prevalent TB in people living with HIV**

Receiver operating characteristic curves depicting prognostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Penn-Nicholson6 (RISK6), (e) Roe3, (f) Suliman4 (RISK4), (g) Sweeney3, and (h) Thompson5 (RESPONSE5) signatures for incident TB diagnosed on two or more liquid culture-positive or Xpert MTB/RIF-positive sputum samples (primary endpoint) through 15 months follow-up in the CORTIS-HR study of people living with HIV.

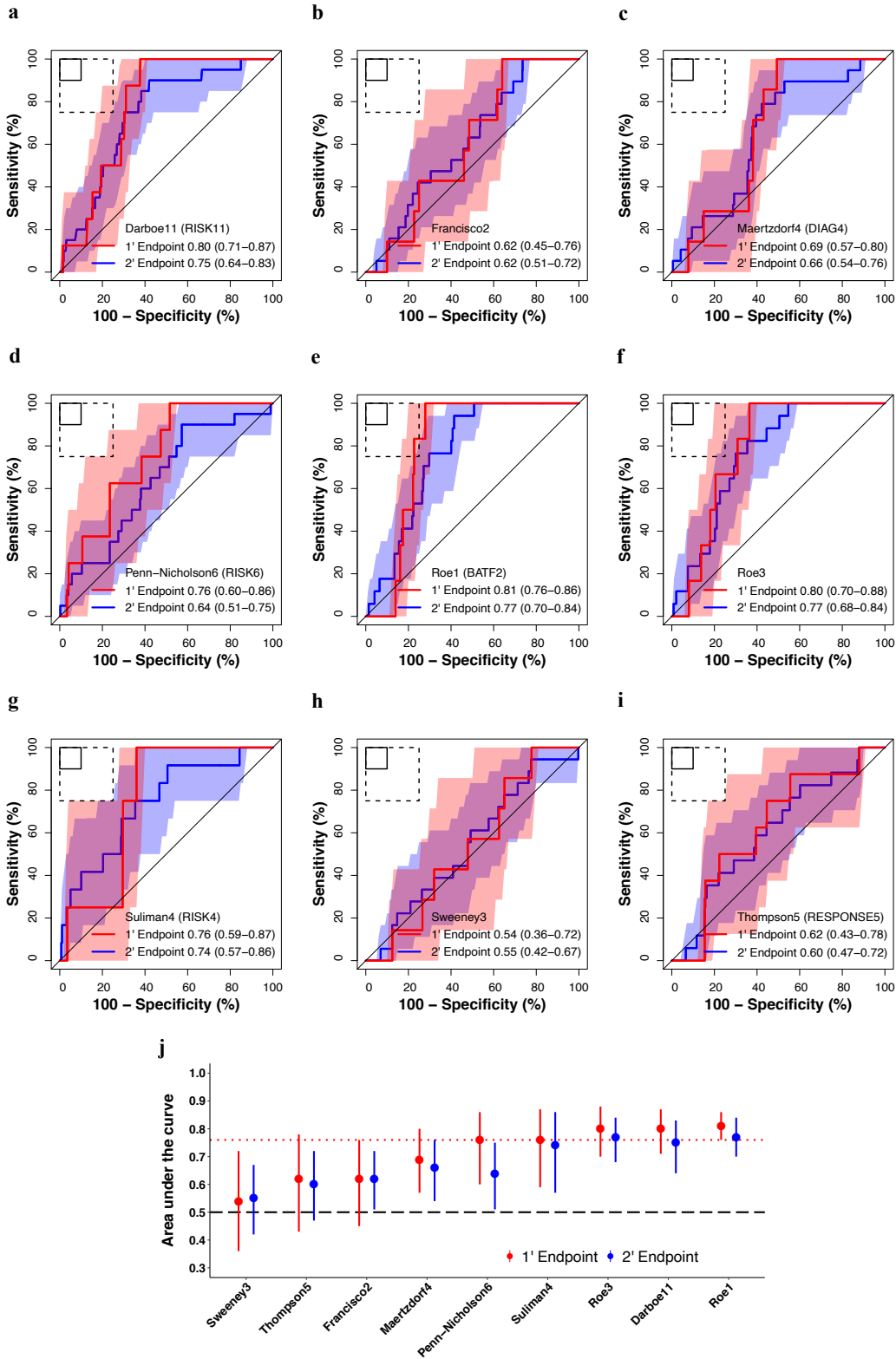
The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (90% sensitivity and 90% specificity) and the dashed box depicts the minimal criteria (75% sensitivity and 75% specificity) set out in the WHO Target Product Profile for an incipient TB test.<sup>4</sup>



**Figure S9: Parsimonious signatures prognostic performance for primary versus secondary endpoint TB through 15-months follow-up in people without HIV**

Receiver operating characteristic (ROC) curves depicting primary (‘1) versus secondary (‘2) endpoint prognostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Penn–Nicholson6 (RISK6), (e) Roe1 (BATF2), (f) Roe3, (g) Suliman4 (RISK4), (h) Sweeney3, and (i) Thompson5 (RESPONSE5) signatures through 15-months follow-up in the CORTIS-01 study of people without HIV. The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (90% sensitivity and 90% specificity) and the dashed box depicts the minimal criteria (75% sensitivity and 75% specificity) set out in the WHO Target Product Profile for an incipient TB test.<sup>4</sup>

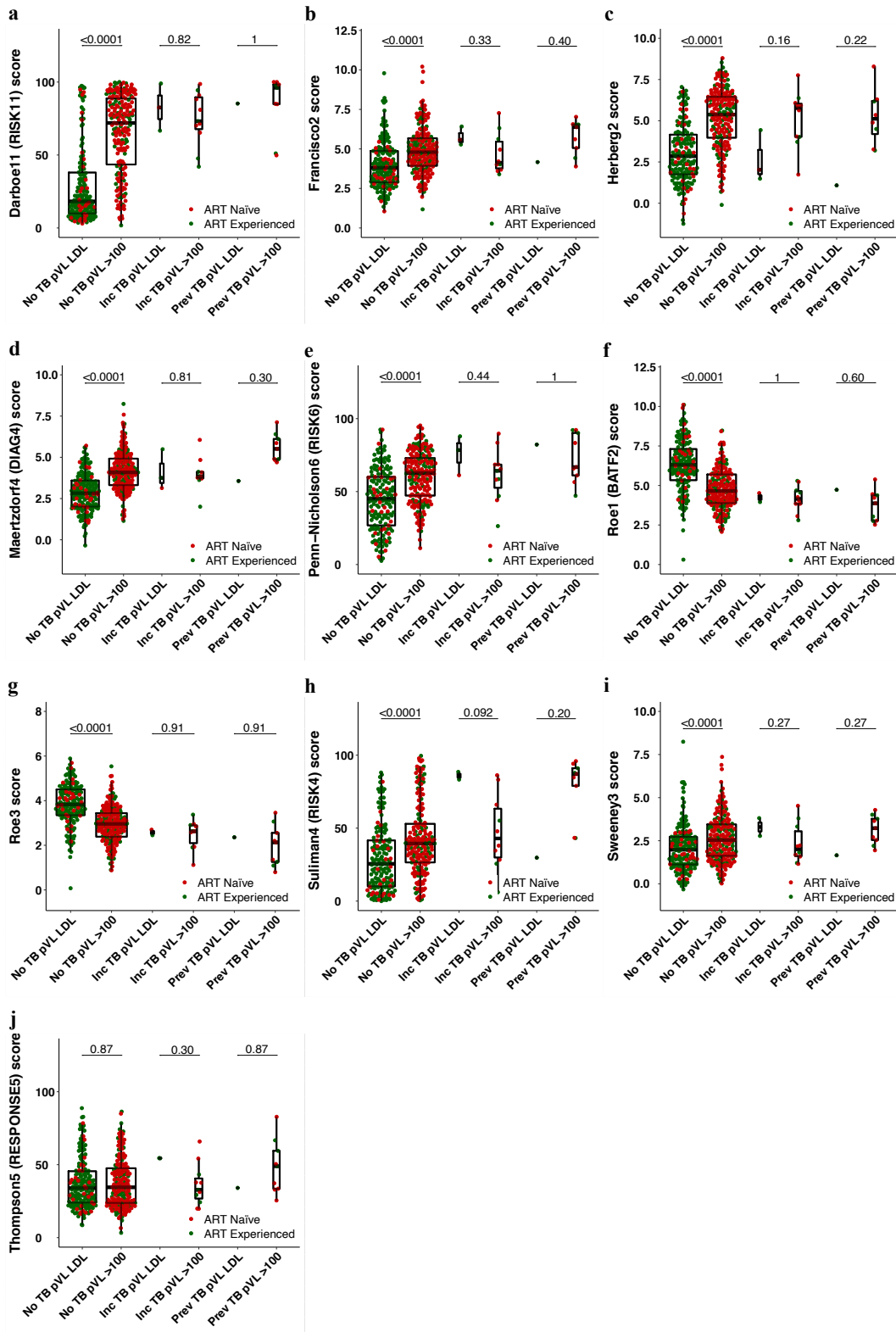
(j) Summary of signature prognostic performance in order of primary endpoint AUC estimates. The prognostic AUC estimates for the secondary endpoint are also shown. The midline indicates the AUC estimate, the error bars indicate the 95% CIs, the red dotted line indicates the lower bound of the 95% CI for the best performing signature for the primary endpoint, and the black dashed line indicates an AUC cut-off of 0.5.



**Figure S10: Parsimonious signatures prognostic performance for primary versus secondary endpoint TB through 15-months follow-up in people living with HIV**

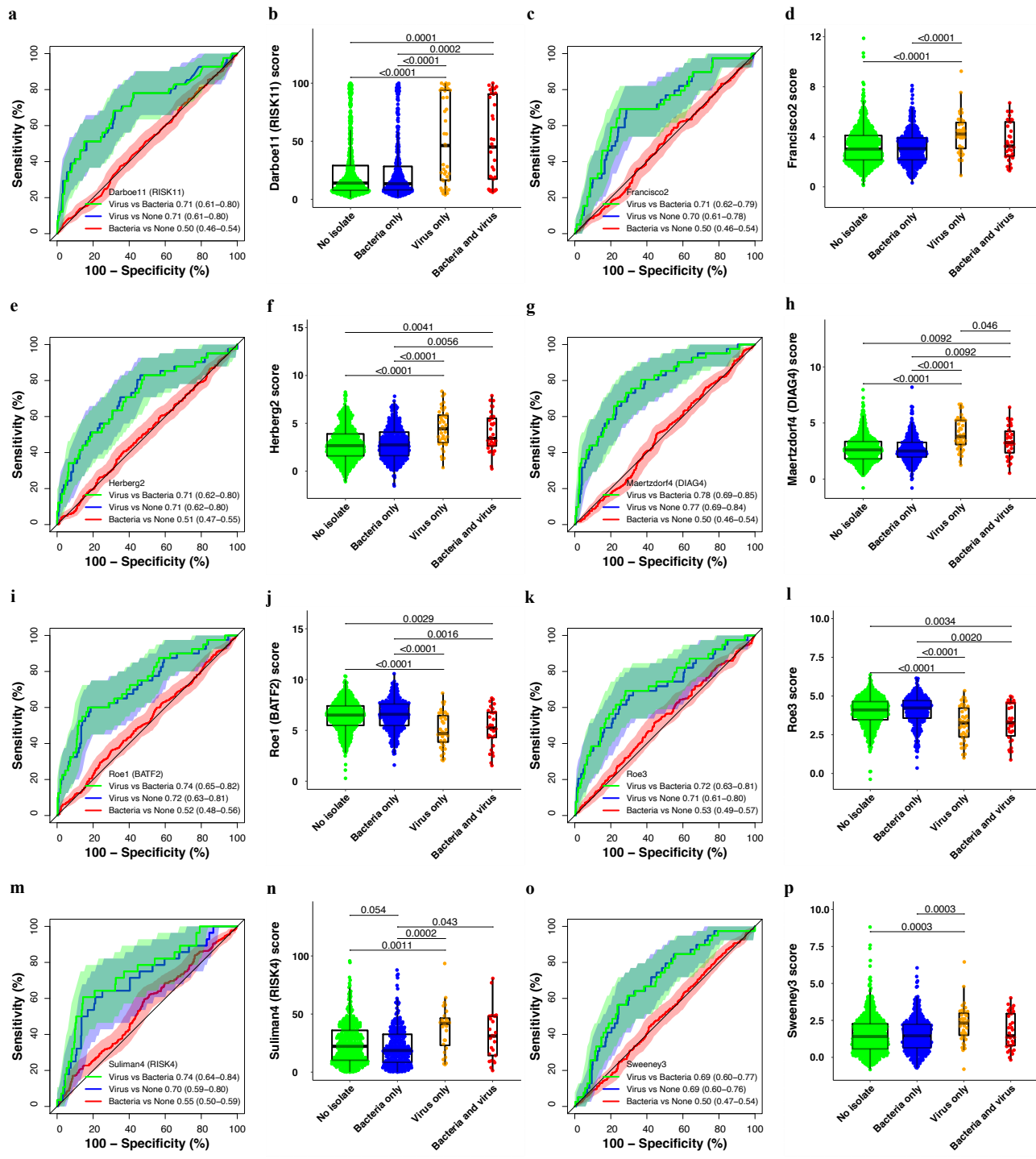
Receiver operating characteristic (ROC) curves depicting primary (1<sup>st</sup>) versus secondary (2<sup>nd</sup>) endpoint prognostic performance (area under the curve, AUC, with 95% CI) of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Maertzdorf4 (DIAG4), (d) Penn-Nicholson6 (RISK6), (e) Roe1 (BATF2), (f) Roe3, (g) Suliman4 (RISK4), (h) Sweeney3, and (i) Thompson5 (RESPONSE5) signatures through 15-months follow-up in the CORTIS-HR study of people living with HIV. The shaded areas represent the 95% CIs. The solid box depicts the optimal criteria (90% sensitivity and 90% specificity) and the dashed box depicts the minimal criteria (75% sensitivity and 75% specificity) set out in the WHO Target Product Profile for an incipient TB test.<sup>4</sup> (j) Summary of signature prognostic performance in order of primary endpoint AUC estimates. The prognostic AUC estimates for the secondary endpoint are also shown. The midline indicates the AUC estimate, the error bars indicate the 95% CIs, the red dotted line indicates the lower bound of the 95% CI for the best performing signature for the primary endpoint, and the black dashed line indicates an AUC cut-off of 0.5.





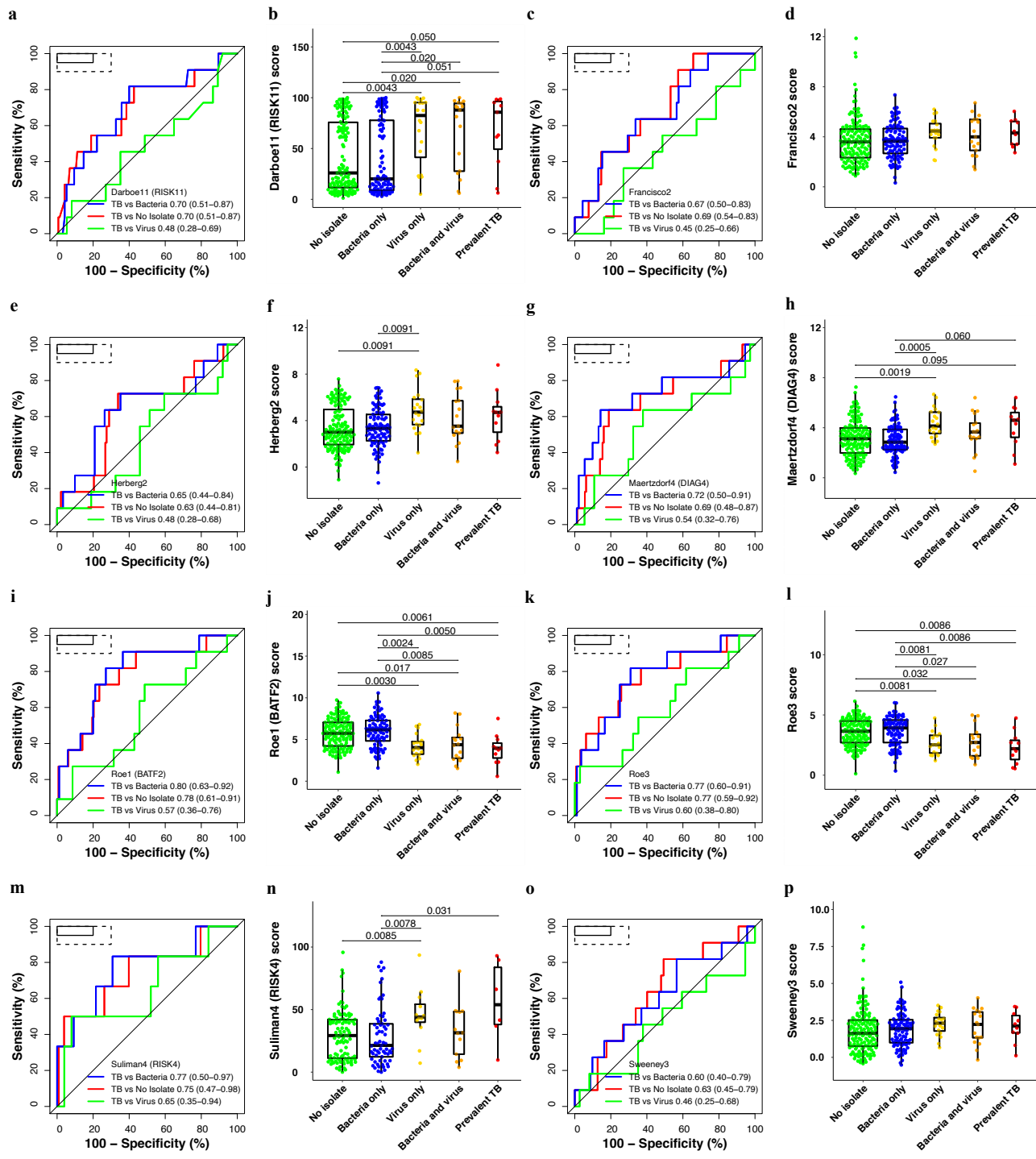
**Figure S11: Parsimonious signatures score distribution by HIV plasma viral load**

Box-and-whisker plots depicting distributions of the (a) Darboe11 (RISK11), (b) Francisco2, (c) Herberg2, (d) Maertzdorf4 (DIAG4), (e) Penn-Nicholson6 (RISK6), (f) Roet1 (BATF2), (g) Roe3, (h) Suliman4 (RISK4), (i) Sweeney3, and (j) Thompson5 (RESPONSE5) signature scores measured at baseline in the CORTIS-HR study stratified by HIV plasma viral load (copies per mL) and TB status. Prevalent (Prev) and incident (Inc) TB comprised all microbiologically confirmed secondary endpoint cases (i.e. TB confirmed on at least one sputum sample). HIV plasma viral load (pVL) is stratified into two groups: lower than the detectable limit (LDL) of 100 copies per mL and greater than 100 copies per mL (>100). Each dot represents a participant. p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney *U* test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Boxes depict the IQR, the midline represents the median, and the whiskers indicate the IQR  $\pm$  (1.5  $\times$  IQR).



**Figure S12: Parsimonious signature performance for differentiating participants with viral or bacterial upper respiratory tract pathogens, and those without any pathogens**

Paired receiver operating characteristic (ROC) curves and box-and-whisker plots for the (a-b) Darboe11 (RISK11), (c-d) Francisco2, (e-f) Herberg2, (g-h) Maertzdorf4 (DIAG4), (i-j) Roe1 (BATF2), (k-l) Roe3, (m-n) Suliman4 (RISK4), and (o-p) Sweeney3 signatures in all participants without prevalent TB in the respiratory pathogens sub-study (11 participants with secondary endpoint prevalent TB excluded). The box-and-whisker plots depict signature score distributions in participants with no upper respiratory pathogens ( $n=563$ ), bacterial upper respiratory pathogens only ( $n=352$ ), viral upper respiratory pathogens only ( $n=41$ ), both viral and bacterial upper respiratory pathogens ( $n=33$ ). p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney  $U$  test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Only p-values below 0.1 are shown. Each dot represents a participant. Boxes depict the IQR, the midline represents the median, and the whiskers indicate the  $IQR \pm (1.5 \times IQR)$ . The ROC curves depict performance (area under the curve, AUC, with 95% CI) of the parsimonious signatures in differentiating between participants with viral upper respiratory pathogens and participants with bacterial upper respiratory pathogens, between participants with viral pathogens and those with no pathogens, or between participants with bacterial pathogens and those with no pathogens. The shaded areas represent the 95% CIs.



**Figure S13: Parsimonious signatures performance for differentiating participants with TB from those viral or bacterial upper respiratory tract pathobionts, and those without any pathobionts**

Paired receiver operating characteristic (ROC) curves and box-and-whisker plots for the (a-b) Darboe11 (RISK11), (c-d) Francisco2, (e-f) Herberg2, (g-h) Maertzdorf4 (DIAG4), (i-j) Roe1 (BATF2), (k-l) Roe3, (m-n) Suliman4 (RISK4), and (o-p) Sweeney3 signatures in all participants randomised in the CORTIS-01 study and co-enrolled in the respiratory pathobionts sub-study. Participants who were not randomised in the CORTIS-01 study, and thus not investigated for TB, are not included.

The box-and-whisker plots depict signature score distribution in participants with no upper respiratory pathobionts (n=143), bacterial upper respiratory pathobionts only (n=95), viral upper respiratory pathobionts only (n=20), both viral and bacterial upper respiratory pathobionts (n=17), and *Mycobacterium tuberculosis* (n=11) detected on GeneXpert MTB/RIF or MGIT culture (microbiologically-confirmed secondary endpoint prevalent TB; i.e. TB confirmed on at least one sputum sample). p values for comparison of median signature scores between groups in box-and-whisker plots were calculated with the Mann-Whitney *U* test and corrected for multiple comparisons by use of the Benjamini-Hochberg Procedure.<sup>3</sup> Only p-values below 0.1 are shown. Each dot represents a participant. Boxes depict the IQR, the midline represents the median, and the whiskers indicate the IQR ± (1.5 × IQR).

The ROC curves depict performance (area under the curve, AUC, with 95% CI) of the parsimonious signatures in differentiating between participants with prevalent TB (n=11) and participants with viral upper respiratory pathobionts (n=37), participants with bacterial upper respiratory pathobionts only (n=95), and participants with no pathobionts (n=143). Participants with both viral and bacterial pathobionts (n=17) were included in the group with viral pathobionts only (n=20) as the presence of bacterial pathobionts did not appear to affect signature scores. The solid box depicts the optimal criteria (95% sensitivity and 80% specificity) and the dashed box depicts the minimal criteria (90% sensitivity and 70% specificity) set out in the WHO Target Product Profile for a triage test.<sup>2</sup>

## Supplementary References

1. Penn-Nicholson A, Mbandi SK, Thompson E, Mendelsohn SC, Suliman S, Chegou NN, et al. RISK6, a 6-gene transcriptomic signature of TB disease risk, diagnosis and treatment response. *Sci Rep.* 2020;10(1):8629. doi: 10.1038/s41598-020-65043-8.
2. WHO. High-priority target product profiles for new tuberculosis diagnostics: report of a consensus meeting. Geneva: World Health Organization, 2014. <https://apps.who.int/iris/handle/10665/135617> (accessed May 22, 2020).
3. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal statistical society: series B (Methodological).* 1995;57(1):289-300. doi: 10.1111/j.2517-6161.1995.tb02031.x.
4. WHO. Consensus Meeting Report: Development of a Target Product Profile (TPP) and a framework for evaluation for a test for predicting progression from tuberculosis infection to active disease. Geneva: World Health Organization, 2017. <http://apps.who.int/iris/handle/10665/259176> (accessed May 22, 2020).