Supporting Information for "Nanoformulation-by-design: An experimental and molecular dynamics study for polymer coated drug nanoparticles."

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34, COCH2), 3.40 (s,3, PEGCH3), 3.67 (m, 24, OCH2CH2O), 4.08 (t, 34, CH2CH2CH2O), 4.25 (t,2, CH2CH2OCO).. 18 SI Figure 13: GPC chromatograph of mPEG350PCL. ... 19 SI Figure 14: GPC chromatograph of mPEG550PCL. .. 20 SI Figure 15: GPC chromatograph of mPEG750PCL. .. 21 SI Figure 16: GPC chromatograph of mPEG2000PCL. .. 22 SI Figure 17: ATR IR spectra of the polymers. ATR-IR: u (cm-1) 3447, 2937, 2889, 2866, 1718, 1473, 1418, 1395, 1366, 1288, 1234, 1171, 1107, 1065, 1047, 957, 935, 733. 23 SI Figure 18: DSC of mPEG350PCL.. 23 SI Figure 19: DSC of mPEG550PCL... 24 SI Figure 20: DSC of mPEG750PCL... 24 SI Figure 21: DSC of mPEG2000PCL. .. 24 SI Figure 22: Contact angle measurements of the 4 diblock copolymers for the assessment of their wettability properties. Error bars correspond to the standard deviation calculated from four measurements. The θ° values decrease with the increase of the hydrophilic block reflecting the increased hydrophilicity of the polymers... 25 SI Figure 23: Analysis of ΔH measurements from DSC revealed a decrease in the percentage of crystallinity of the PCL block of the copolymers with an increase of the length of the hydrophilic block... 25 SI Figure 24: Reproducibility of the interfacial deposition method; Overlay of size measurements (DLS) of 4 batches of polymer NPs formed with PEG350PCL at a 0.1 mg polymer amount. ... 27 SI Figure 25: Zeta potential of the polymeric NPs. Key refers to PEG chain length of polymer.. 27 SI Figure 26: Stability of the PEG350PCL polymeric NPs (final polymer concentration in water 0.2 mg/ml): The original measurement is in red, the same sample after 1 month in green and the blue is after sonication of the latter for 5 minutes. The size distribution is the same in all 3 cases.. 28 SI Figure 27: TEM image of mPEG350- PCL 2000 polymeric NPs at 0.05 mg/ml initial concentration in acetone (polymer amount 1mg). Diameters of selected NP indicated. Scale bar 1000 nm... 28 SI Figure 28: DSC of indomethacin as received .. 29 SI Figure 29: Size distribution of Indomethacin in water (1mg/ml), just after sonication, by DLS. .. 29 SI Figure 30: Polarised optical microscopy images of indomethacin in aqueous phase prior to the coating experiments. (a) without and (b) with the polariser. Indomethacin is clearly crystalline. Magnification 10x... 30 SI Figure 31: mPEG350-PCL2000 coated indomethacin nanoparticle size distribution by DLS. (polymer amount 0.1mg)... 31

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Molecular Dynamics Section

Topologies and parameters for MD

Acetone ;----------------------------TITLE -- acetone ; ; This file was generated at 01:26 on 2014-09-24 by ; ; Automatic Topology Builder ; ; REVISION 2014-09-22 ;--- ; Authors : Alpeshkumar K. Malde, Le Zuo, Matthew Breeze, Martin Stroet, Alan E. Mark ; ; Institute : Molecular Dynamics group, School of Chemistry and Molecular Biosciences (SCMB), The University of Queensland, QLD 4072, Australia ; URL : http://compbio.biosci.uq.edu.au/atb ; Citation : Malde AK, Zuo L, Breeze M, Stroet M, Poger D, Nair PC, Oostenbrink C, Mark AE. An Automated force field Topology Builder (ATB) and repository: version 1.0. Journal of Chemical Theory and Computation, 2011, 7(12), 4026-4037. ; http://pubs.acs.org/doi/abs/10.1021/ct200196m ; ; Disclaimer : While every effort has been made to ensure the accuracy and validity of parameters provided below ; the assignment of parameters is being based on an automated procedure combining data provided by a given user as well as calculations performed using third party software. They are provided as a guide. The authors of the ATB cannot guarantee that the parameters are complete or that the parameters provided are appropriate for use in any specific application. Users are advised to treat these parameters with discretion and to perform additional validation tests for their specific application if required. Neither the authors of the ATB or The University of Queensland except any responsibly for how the parameters may be used. ; ; Release notes and warnings: ; (1) The topology is based on a set of atomic coordinates and other data provided by the user after after quantum mechanical optimization of the structure using different levels of theory depending on ; the nature of the molecule. ; (2) In some cases the automatic bond, bond angle and dihedral type assignment is ambiguous. ; In these cases alternative type codes are provided at the end of the line. ; (3) While bonded parameters are taken where possible from the nominated force field non-standard bond, angle and dihedral type code may be incorporated in cases where an exact match could not be found. These are marked as "non-standard" ; or "uncertain" in comments.

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; (4) In some cases it is not possible to assign an appropriate parameter 
automatically. "%" is used as a place holder
      for those fields that could not be determined automatically. The parameters
in these fields must be assigned manually
      before the file can be used.
;-----------------------------------------------------------------------------
; Input Structure : UAC
; Output : UNITED ATOM topology
      Use in conjunction with the corresponding united atom PDB file.
    ;----------------------------------------------------------------------------
; Final Topology Generation was performed using: 
; A B3LYP/6-31G* optimized geometry.
; Bonded and van der Waals parameters were taken from the GROMOS 53A6 parameter 
set.
; Initial charges were estimated using the ESP method of Merz-Kollman.
; Final charges and charge groups were generated by method described in the ATB 
paper.
; If required, additional bonded parameters were generated from a Hessian matrix 
calculated at the B3LYP/6-31G* level of theory.
;
; Topology file generated at 01:09 on 24 Sep 2014 for molecule
; acetone (IUPAC: propan-2-one, database identifier: UAC)
; by Automatic Topology Builder(revision 2014-09-22).
;
[ moleculetype ]
; Name nrexcl<br>UAC 3
UAC 3
[ atoms ]
; nr type resnr resid atom cgnr charge mass total_charge
   1 CH3 1 UAC C3 1 -0.083 15.0350<br>2 C 1 UAC C2 1 0.734 12.0110
    2 C 1 UAC C2 1 0.734 12.0110 
 3 O 1 UAC O1 1 -0.568 15.9994 ; 0.083
 4 CH3 1 UAC C1 2 -0.083 15.0350 ; -0.083
; total charge of the molecule: 0.000
[ bonds ]
; ai aj funct c0 c1<br>1 2 2 0.1520 5.43000
      1 2 2 0.1520 5.4300e+06
    2 3 2 0.1230 1.6600e+07
    2 4 2 0.1520 5.4300e+06
[ pairs ]
 ai aj funct ; all 1-4 pairs but the ones excluded in GROMOS itp
[ angles ]
; ai aj ak funct angle fc
    1 2 3 2 125.00 750.00
    1 2 4 2 120.00 560.00
    3 2 4 2 125.00 750.00
[ dihedrals ]
; GROMOS improper dihedrals
; ai aj ak al funct angle fc
    2 1 3 4 2 0.00 167.36
[ dihedrals ]
  ai aj ak al funct ph0 cp mult
[ exclusions ]
; ai aj funct ; GROMOS 1-4 exclusions
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Polymer residual topologies

[dihedrals]

[angles]

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Control Simulation: mPEG350-PCL2000 in acetone

SI Figure 1: Five mPEG350-PCL2000 polymer chains surrounded by acetone (not shown), a favourable solvent. Periodic boundary conditions (PBC) apply, where the molecules can move across the end of the box.

Control Simulation: mPEG350-PCL2000 in water

SI Figure 2: Five mPEG350-PCL2000 polymer chains surrounded by water (not shown). The polymers self aggregate within 4ns and then they form a larger polymeric NP within the first 32 ns of the simulation. PBC apply.

Control Simulation: Indomethacin NP in acetone

SI Figure 3: A 5nm diameter indomethacin NP surrounded by acetone (not shown), a favourable solvent. The NP dissolves. PBC apply.

Control Simulation: Indomethacin NP in water

SI Figure 4: A 5nm diameter indomethacin NP surrounded by water (not shown). The NP stays intact. PBC apply.

Control Simulation: Formation of polymeric NP in interfacial deposition

SI Figure 5: mPEG350 (light blue)-b-PCL2000 (dark blue) polymer chains in the phase 1 of the interfacial deposition simulation, for the formation of a polymer NP. Snapshots taken at every 20 ns, solvents not shown for clarity. The polymer chains start to aggregate,

SI Figure 6: mPEG350 (light blue)-b-PCL2000 (dark blue) polymer chains in the phase 2 of the interfacial deposition simulation, for the formation of a polymer NP. Snapshots taken at the end of every 10 ns MD run where acetone molecules (pink) are replaced by water molecules.

Control Simulation: Indomethacin NP in interfacial deposition

SI Figure 7: Control simulation of the indomethacin NP (grey) to evaluate the NP's stability in the biphasic system. The drug NP stays intact until 70 ns, however as the presence acetone (pink) in the NP's surrounding environment increases, the NP starts to swell up.

Main Simulation: Interfacial Deposition of mPEG350-PCL2000

SI Figure 8: Phase 1 of the interfacial deposition simulation of mPEG350 (light blue)-b-PCL2000 (dark blue) polymer chains in the presence of a 5 nm indomethacin NP (grey). Solvent molecules are not shown for clarity. PBC apply. Snapshots taken at various time points of the simulation. The polymer chains start to aggregate slowly as the acetone diffuses towards the water region. The indomethacin NP stays intact in the aqueous region until it reaches close proximity with the polymer chains and gets dissolved at 108 ns.

SI Figure 9: Phase 2 of the interfacial deposition for the formation of a polymer-coated drug NP. Snapshots taken at the end of every 10 ns MD run where acetone molecules (pink) are replaced by water molecules, mimicking evaporation of the acetone. A polymer-drug NP of 7nm in diameter is formed. Water is not shown for clarity, PBC conditions apply.

SI Figure 10: Number of contacts and the distance between the PCL blocks and the indomethacin molecules during phase 1 of the interfacial deposition.

SI Figure 11: Radius of gyration (a, c) and end-to-end distances (b, d) for both phases 1 and 2 of the polymer-drug NP formation simulation. c and d refer just to phase 2.

Experimental Section

Polymer Characterisation: NMR

SI Figure 12: NMR Spectra of the synthesised polymers. PEG350PCL: 1H NMR (figure 4.1) (400 MHz, CDCl3, ppm): 1.41 (m,34, CH2CH2CH2), 1.68 (m, 68, CH2CH2CH2), 2.33 (t, 34, COCH2), 3.40 (s,3, PEGCH3), 3.67 (m, 24, OCH2CH2O), 4.08 (t, 34, CH2CH2CH2O), 4.25 (t,2, CH2CH2OCO).

Polymer Characterisation: GPC

Sample:IDSA09

Date: 24/04/2015 17:32:03

Workbook: CHCl3_conv_150420.plw Inj File: 150424-0006.cgrm Path: C:\Cirrus Workbooks\2015\CHCl3_conv_150420\CHCl3_conv_150420.plw Res. File: 150424-0006.rst Batch: 150424 Method: PSeasy_CHCl3_150420 **Eluent: Chloroform**

SI Figure 13: GPC chromatograph of mPEG350PCL.

Sample:IDSB10

Date: 24/04/2015 15:44:17

Inj File: 150424-0003.cgrm Workbook: CHCl3_conv_150420.plw Path: C:\Cirrus Workbooks\2015\CHCl3_conv_150420\CHCl3_conv_150420.plw Res. File: 150424-0003-Repeat (01).rst Method: PSeasy_CHCl3_150420 Batch: 150424 Eluent: Chloroform

SI Figure 14: GPC chromatograph of mPEG550PCL.

Sample:IDSC11

Date: 24/04/2015 16:56:13

Workbook: CHCl3_conv_150420.plw Inj File: 150424-0005.cgrm Path: C:\Cirrus Workbooks\2015\CHCl3_conv_150420\CHCl3_conv_150420.plw Res. File: 150424-0005-Repeat (01).rst Method: PSeasy_CHCl3_150420 Batch: 150424 Eluent: Chloroform

SI Figure 15: GPC chromatograph of mPEG750PCL.

Sample:IDSF16

Date: 24/04/2015 15:08:27

Inj File: 150424-0002.cgrm Workbook: CHCl3_conv_150420.plw Path: C:\Cirrus Workbooks\2015\CHCl3_conv_150420\CHCl3_conv_150420.plw Res. File: 150424-0002-Repeat (01).rst Method: PSeasy_CHCl3_150420 Batch: 150424 Eluent: Chloroform

SI Figure 16: GPC chromatograph of mPEG2000PCL.

Polymer Characterisation: ATR-IR

SI Figure 17: ATR IR spectra of the polymers. ATR-IR: u (cm-1) 3447, 2937, 2889, 2866, 1718, 1473, 1418, 1395, 1366, 1288, 1234, 1171, 1107, 1065, 1047, 957, 935, 733.

Polymer Characterisation: DSC

SI Figure 19: DSC of mPEG550PCL.

SI Figure 20: DSC of mPEG750PCL.

SI Figure 21: DSC of mPEG2000PCL.

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Polymer Characterisation: Contact Angle

SI Figure 22: Contact angle measurements of the 4 diblock copolymers for the assessment of their wettability properties. Error bars correspond to the standard deviation calculated from four measurements. The θο values decrease with the increase of the hydrophilic block reflecting the increased hydrophilicity of the polymers.

Polymer Characterisation: DSC

SI Figure 23: Analysis of ΔH measurements from DSC revealed a decrease in the percentage of crystallinity of the PCL block of the copolymers with an increase of the length of the hydrophilic block.

The contribution of the mPEG chain in the ΔH of the copolymers was calculated by:

 $\varDelta H_{mPEG} = \varDelta H * \left(\frac{w}{w}\% (mPEG)\right) = \varDelta H * \frac{Mw_{mPEG}}{114.14*DP_{PCL}+Mw_{mPEG}}$ where DP_{PCL} is the number of CL units in the PCL blocks as calculated via H¹ NMR. Then $\Delta_{\sf HPCL}$ =ΔH-ΔH_{mPEG.}

Polymeric NP Characterisation: DLS

PEG350PCL Polymeric NPs different batches (polymer amount 0.1mg)

SI Figure 24: Reproducibility of the interfacial deposition method; Overlay of size measurements (DLS) of 4 batches of polymer NPs formed with PEG350PCL at a 0.1 mg polymer amount.

Polymeric NP Characterisation: ζ-potential

SI Figure 25: Zeta potential of the polymeric NPs. Key refers to PEG chain length of polymer.

SI Figure 26: Stability of the PEG350PCL polymeric NPs (final polymer concentration in water 0.2 mg/ml): The original measurement is in red, the same sample after 1 month in green and the blue is after sonication of the latter for 5 minutes. The size distribution is the same in all 3 cases.

Polymeric NP Characterisation TEM

SI Figure 27: TEM image of mPEG350- PCL 2000 polymeric NPs at 0.05 mg/ml initial concentration in acetone (polymer amount 1mg). Diameters of selected NP indicated. Scale bar 1000 nm.

Indomethacin bulk characterisation: DSC

SI Figure 28: DSC of indomethacin as received

Indomethacin NP Characterisation: DLS

SI Figure 29: Size distribution of Indomethacin in water (1mg/ml), just after sonication, by DLS.

Indomethacin NP Characterisation: POM

SI Figure 30: Polarised optical microscopy images of indomethacin in aqueous phase prior to the coating experiments. (a) without and (b) with the polariser. Indomethacin is clearly crystalline. Magnification 10x.

Polymer-coated indomethacin NP Characterisation: DLS

SI Figure 31: mPEG350-PCL2000 coated indomethacin nanoparticle size distribution by DLS. (polymer amount 0.1mg)

SI Figure 32: Overlay of two experiments with mPEG350-PCL2000 and mPEG350- PCL2000 coated indomethacin nanoparticles (0.1mg polymer amount) to demonstrate the difference between the populations: Red line corresponds to the polymeric NPs that are formed in the absence of indomethacin, while the green distribution relates to the polymer-coated indomethacin

SI Figure 33: Centrifugation of the polymer coated drug nanoparticles (bottom graph) gives a PDI of 0.116, compared to a PDI that ranged from 0.394-0.265 in the precentrifugation suspensions (top graph)

Polymer-coated indomethacin NP Characterisation: POM

SI Figure 34: Polarised optical microscopy images of the mPEG350PCL-coated indomethacin NPs. (a1) and (a2) prior to centrifugation, where uncoated drug particles form aggregates and (b1), (b2) after the purification where no crystals were observed (Magnification 10x).

Polymer-coated indomethacin NP Characterisation: TEM

SI Figure 35: TEM picture of mPEG-PCL coated indomethacin particles, (highest polymer starting concentration). Scale bar 200nm.

Polymer-coated indomethacin NP Characterisation: Reproducibility

SI Figure 36: Reproducibility of the coating method; Overlay of 5 batches of mPEG350PCL-coated indomethacin NPs (0.1 mg polymer amount, 1 mg/ml indomethacin concentration in water)

Polymer-coated indomethacin NP Characterisation: Stability

SI Figure 37: Stability of the produced polymer coated-drug NPs; Same batch of mPEG350PCL-coated indomethacin NPs (0.1 mg polymer amount, 1 mg/ml indomethacin concentration in water) measured after 10 days.

Polymer-coated indomethacin NP Characterisation: Effect of Drug amount

indomethacin concentration in water (mg/ml)

SI Figure 38: Analysis of the effect of decreasing indomethacin concentration in the starting drug-bearing aqueous phase. Y-axis:Size distribution (Intensity), Right Y-axis PDI of measurements.

Polymer-coated indomethacin NP Characterisation: Drug Loading

Table 1 - Calculation of drug loading and encapsulation efficiency via UV-Vis, after lyophilisation of the suspension

% Drug loading = polymer amount/ (polymer amount + drug amount in nanoparticles) *100 (Assuming all polymer is still present in drug coated nanoparticles)

In order to account for the drug loss during the loading of the aqueous bearing syringe and the pumping, Ep2 was amended into the following:

%Entrapment Efficiency ‡

= (amount of coated drug/amount of recovered control drug NPs)*100.