

## Supplemental data

**Supplemental Table 1** | Forward (F) and reverse (R) primers for molecular cloning, HRM analysis, sequencing and RT-PCR analysis.

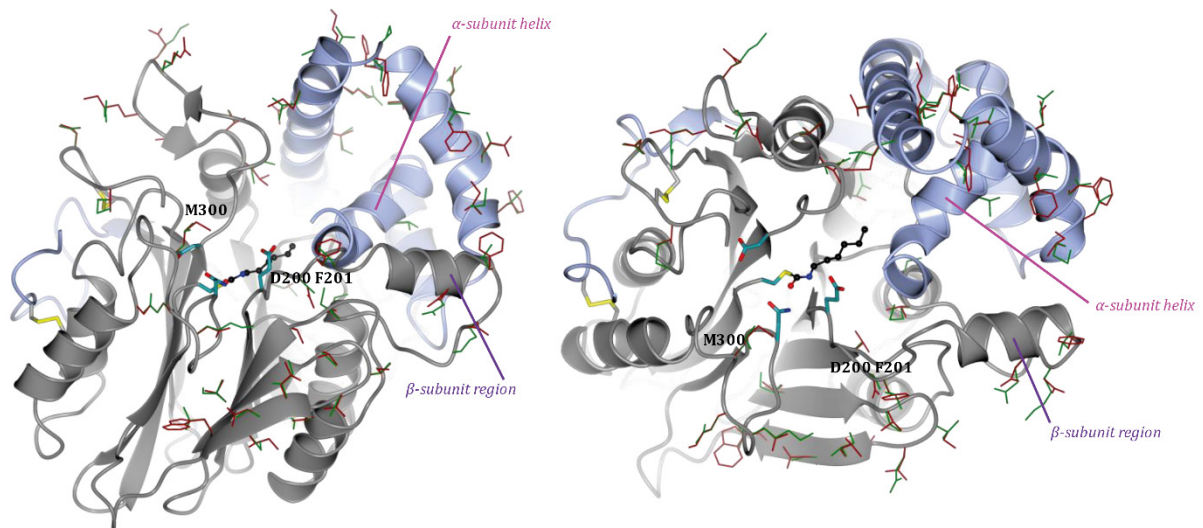
Target	Note	Forward primer sequence (5'→3')	Reverse primer sequence (5'→3')	
<i>asah1a</i>	Gateway cloning	GGGGACAAGTTTGTACAAAAAAGCAGGC TACCACCATGAAGCTTGTGTTCCGTTAC	GGGGACCACTTTGTACAAGAAAGCTGGGT cTTACCATGGTGTGCATGGATTTG	
<i>asah1b</i>	Gateway cloning	GGGGACAAGTTTGTACAAAAAAGCAGGC TACCACCATGAACAACAGATTAAACCTG	GGGGACCACTTTGTACAAGAAAGCTGGGT cTCACCAGGGCATGCAGGGATTTG	
<i>asah1a</i>	HRM	GTCTAGACTCGAATAAGTTCATG	TGGGAAACAGTTACCTCTGTG	
<i>asah1b</i>	HRM	TGCAAAGAGATGTGTTAGATTG	TCCTTCAGATGGCGAGCATG	
<i>asah1a</i>	Sequencing	TGGGATGTATCCACCTAAAGG	Same as HRM Rev	
<i>asah1b</i>	Sequencing	CAGCAAGCAAAGATGGACAG	TACGATTTTGGGAGATTTATCTC	
	NCBI code			
<i>asah1a</i>	NM_001006088	ATTAGGCCTGGTGAAGTGC	CTGCGAGTAAGAAAACCCGTC	125 bp
<i>asah1b</i>	NM_200577	TGGACTGTTTCATGGGATGGG	CCGGTCAACATCCCGACATA	150 bp
<i>gpnmb</i>	XM_009294247	GCAAGGGCGTAGAATTGAAA	TGGCAGGGACATGTCAGTAA	
<i>chia.6</i>	NM_199603	TCCACGGCTCATGGGAGAGTGTC	AGCGCCCTGATCTCGCCAGT	(73)
<i>catD</i>	NM_131710	TGGGTGGAAAGGTCTACTCG	CACTCAGGCAGATGTCGTGT	
<i>il1β</i>	NM_212844	TGGACTTCGCAGCACAAAATG	GTTCACTTCACGCTCTTGGATG	(74)
<i>tnfβ*</i>	NM_001024447	GCATGTGATGAAGCCAAACG	GATTGTCCTGAAGGGTCACC	(75)
<i>apoeb</i>	NM_131098	AAACTGACATGACCGACGCT	TAGGTTGCTACGGTGTTCGG	172 bp
<i>c1qa</i>	NM_001020527	CTCTGTTTCCCTTTTCCTTCTG	CTTTCTCTCCTTTTGGTCTGG	108 bp
<i>c3a.1</i>	NM_131242	CGCTGCACAAAGTACTTCCAC	GCCAGCTCCATGTCCTTGAC	197 bp
<i>c5aR1</i>	XM_005159274	CCGACAAGCTCGCATCCTAT	GCGAATGATGGTTATCGCCC	163 bp
<i>c5</i>	XM_001919191	CAAGGCCACGGTTCAATCAG	TCTTCATGCTTTTCGGCAGTCA	152 bp
<i>th1</i>	NM_131149	AGCTTTGTGGACGCTACTGA	GTGGGTTGTCCAGCACTTCT	112 bp
<i>th2</i>	NM_001001829	TACAAGCCATTCGACCCAGC	ATGCTGCAAGTGTAGGGGTC	173 bp
<i>sncβ</i>	NM_200969	GGAGTTTGGTCAGGAAGCCA	CCTCGGGCTCATAATCCTGG	107 bp
<i>sncyα</i>	NM_001017567	TGGAGGGGCTGGAGACTATG	AGCATCATGGGACATTCGGTT	123 bp
<i>sncyβ</i>	NM_001020652	ATGGTGAACCCGGGTGACTT	AGGCTTTGGAGCAGAAACGTA	129 bp

<i>mcpa</i>	XM_002665562	TGGTCATCTATCCTCCTCTCCA	CTTTCTCCCAGGCCCAATAGTTCT	150 bp
<i>efla</i>		CTGGAGGCCAGCTCAAACAT	ATCAAGAAGAGTAGTACCGCTAGCATTAC	(76)
<i>rpl13a</i>		TCTGGAGGACTGTAAGAGGTATGC	AGACGCACAATCTTGAGAGCAG	(76)

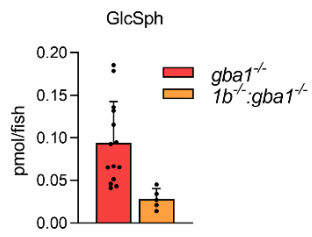
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\* Also known as *tnf- $\alpha$ 2*

Human ACase	MPGRSCVALV-LLAAAVSCAVAQHAPPWTEDCRKSTYPPSPGPTYRGAVPWYTIINLDDLPPY	59
Asah1a	---MKLVFRYNALFISIFIALYV-QGLEDCRSGMYPPKGPTRYRGNVTRYTVNLDLPPS	55
Asah1b	MNNRNLNLCFFI-LSYCMCLSAQYVFPFTEDCRSGMYPPNGPTFKGDVSWYTVDDLPPAS	59
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Human ACase	KRWHELMLDKAPVLRKVIIVNSLKNMINTFVPSGKIMQVVDEKLPGLLGNFPGPFEEEMKGI	119
Asah1a	ERWTQIIKDKNTELEIMVQTIKDMAKGFH-HGKLVNFVDKELPFIVDTLPNPFNEEIKGI	114
Asah1b	KRWTDVISDKKTEMASMIQAIRDLADAFVPSGKLIQLVDKDLPLMVDTLPLYPFNEEIRGI	119
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	<u>α-subunit helix</u>	
Human ACase	AAVTDIPLGELIISFNIFYELFTICTSIVAEDKKGHLIHGRNMDFGVFLGWNINNDTWVIT	179
Asah1a	AAVSGIPLGELIALFNIFYEVFTVCTSLVAEDNNGNIYHGRNLDFGLFMGWRQNKWTTLT	174
Asah1b	ASVSGVPLGEVVLFNIFYEVFTVCTSLVAEDVNGNLIHARNLDFGLFMGWDLKNRSWVIT	179
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	<u>β-subunit region</u>	
Human ACase	EQLKPLTVNLDFQRNKTQVFKASSFAGYVGLTGFKPGLESLTLNERFSINGGYLGILEW	239
Asah1a	EKLKPLVVNINFERKNQTVFKSTSFAGYVGLTGIRPGELELTMNERFDGDDGYYIGILDW	234
Asah1b	EKLKPLVVNIDFTRNGQTVFKSTNFAGYVGLTGIHQNSFTLTMNERFSLDGGYYIGILEW	239
	*:***.***:* * : *	
Human ACase	ILGKKDVMWIGFLTRTVLENSTSYEEAKNLLTKTKILAPAYFILGGNQSSEGCVITRDRK	299
Asah1a	IFGNRDGMWTGFLTRRVLENSTSYEDAKDQLSQTKLLAPVYFILGGNRTGGCVITRTRI	294
Asah1b	ILGKRDMWMSFLTRSVLENATSYESAKALLSDTKLLAPAYFILGGNQSSEACIITRSRT	299
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Human ACase	ESLDVYELDAKQGRWYVVQTNDRWKHPFLDDRRTPAKMCNRTSQENISFETMYDVLS	359
Asah1a	NTLDIWELEMLGRWYVLETNYDHWKPMFLDDRRTPAKCMNQTTQANISLASIYNVLS	354
Asah1b	QNISPLELVKNGRWYVLETNYDHWKEPLFLDDRRTPAKCMNQTTQTNISVKTVDVLS	359
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Human ACase	TKPVLNKLTVYTTLLIDVTKGQFETYLRDCPDPCIGW 395	
Asah1a	TKPVLNKLTTYTSLMAVSTGTLESYVRDCPNPCTPW 390	
Asah1b	TKPVLNKLTTYTTLMEVSKGTLESFIRDCPNPCMPW 395	
	*****.***: * : * : : : : : : : : * *	

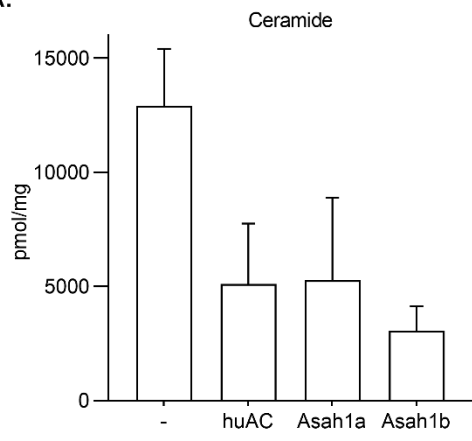


Ribbon diagram of the side (left) or top (right) of human ACCase (PDB 6MHM) with  $\alpha$ -subunit (light grey),  $\beta$ -subunit (dark grey) and disulfide bonds (yellow) indicated. Inhibitor Carmofur (black, ball and stick) and catalytic residues C143, D162, E225 or N320 (cyan sticks) are also visualized for clarity, as proposed by Dementiev et al.(32). Divergent residues of Asah1a (red) and Asah11b (green) surrounding the catalytic pocket are superimposed on the human structure. Only residues Asp 200, Phe 201 and Met 300 of Asah1a, located on the loops adjacent to the catalytic site, might be close enough to be of significance.

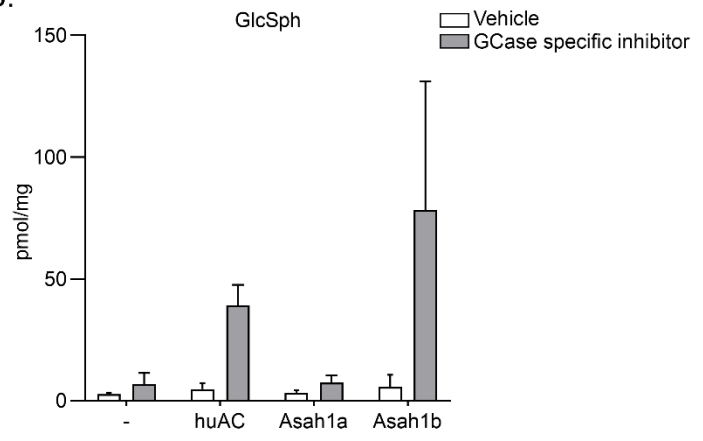


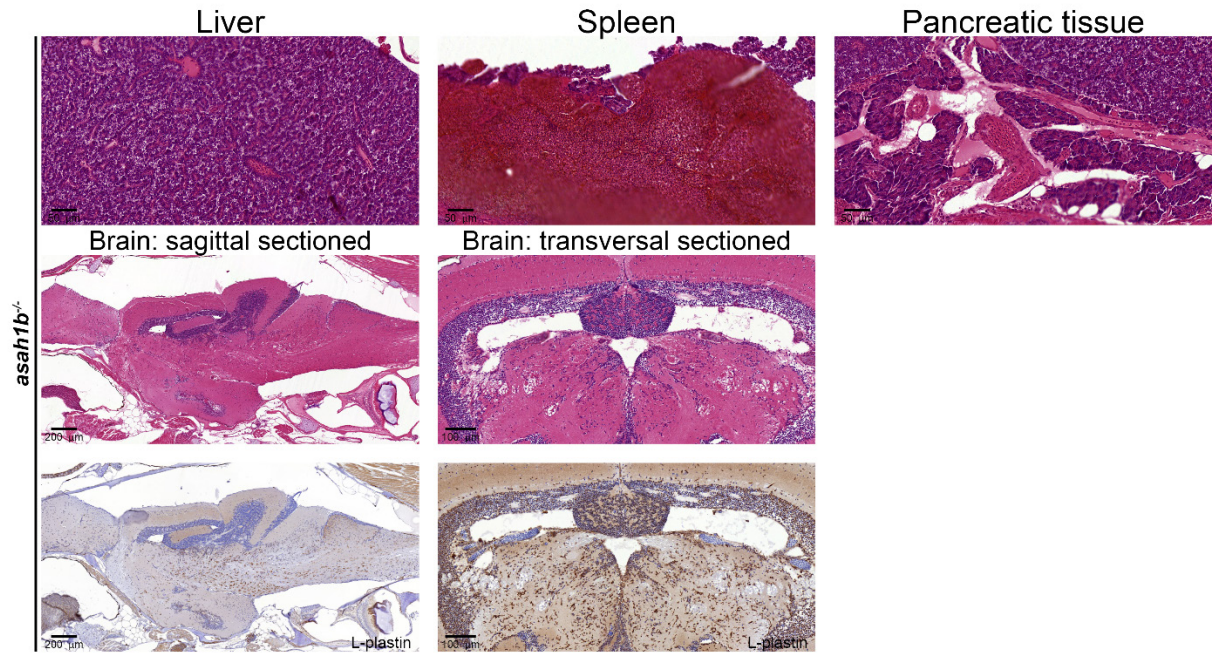
GlcSph levels of  $gba1^{-/-}$  (n = 14) and  $gba1^{-/-}:asah1b^{-/-}$  (n = 5) larvae KO at 5dpf in pmol/fish. Data of  $gba1^{-/-}$  is obtained from (33). Data is depicted as mean  $\pm$  SD.

A.



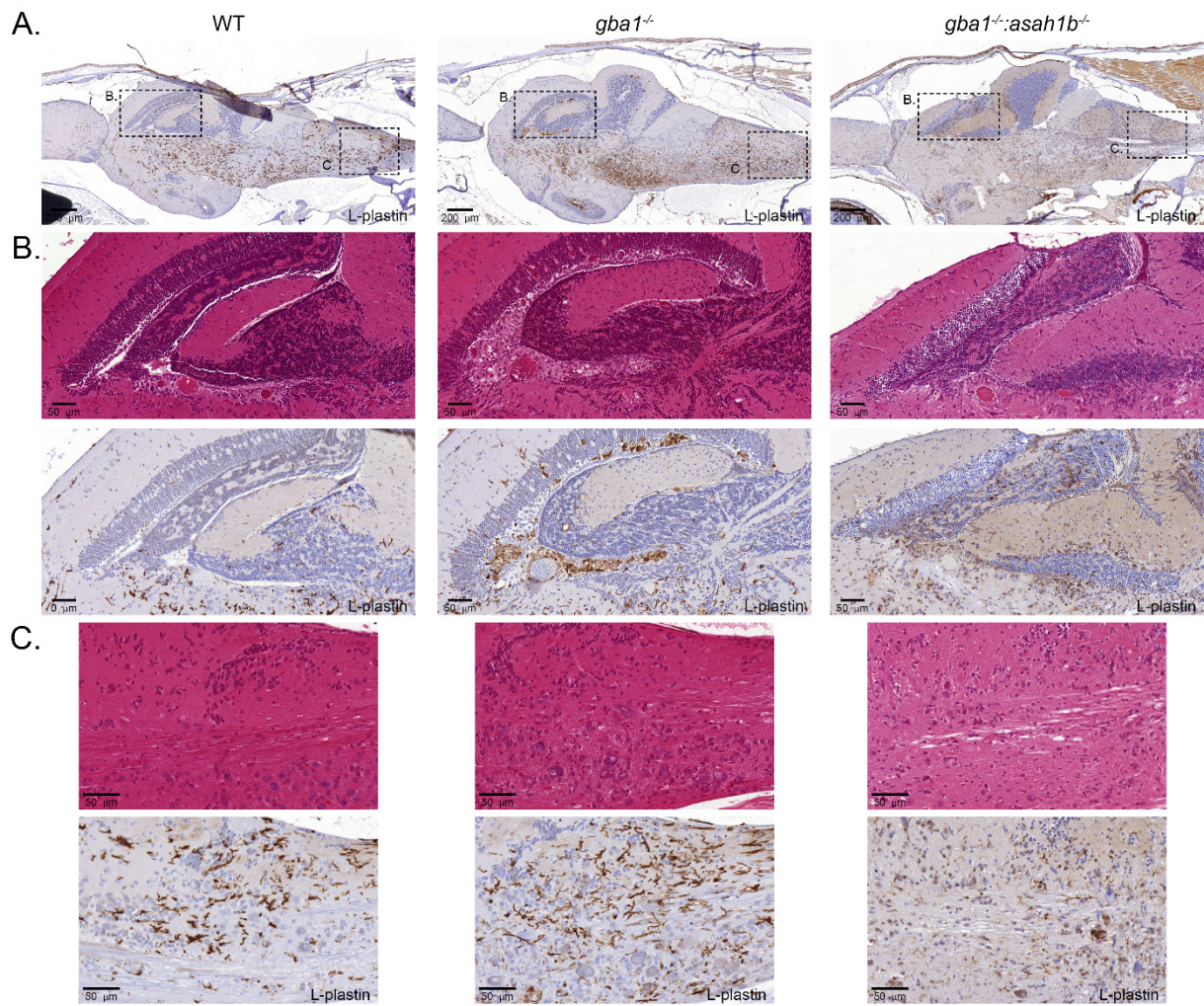
B.





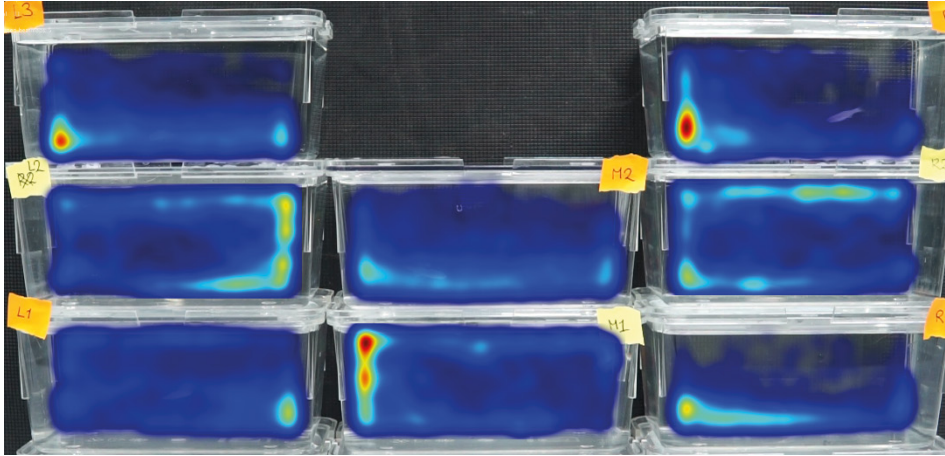
**Supplemental Figure S5:** Histopathology of *asah1b*<sup>-/-</sup> zebrafish including liver, spleen and pancreatic tissue (top panel) as well as sagittal section (left) and transversal section (right panel) of *asah1b*<sup>-/-</sup> zebrafish brain using H&E staining. Bottom panel: immunostaining of L-plastin in sagittal section (left) and transversal section (right) of *asah1b*<sup>-/-</sup> zebrafish brain. Scale bars: 50 µm for top panel, 200 µm and 100 µm for sagittal and transversal sectioned brain, respectively.





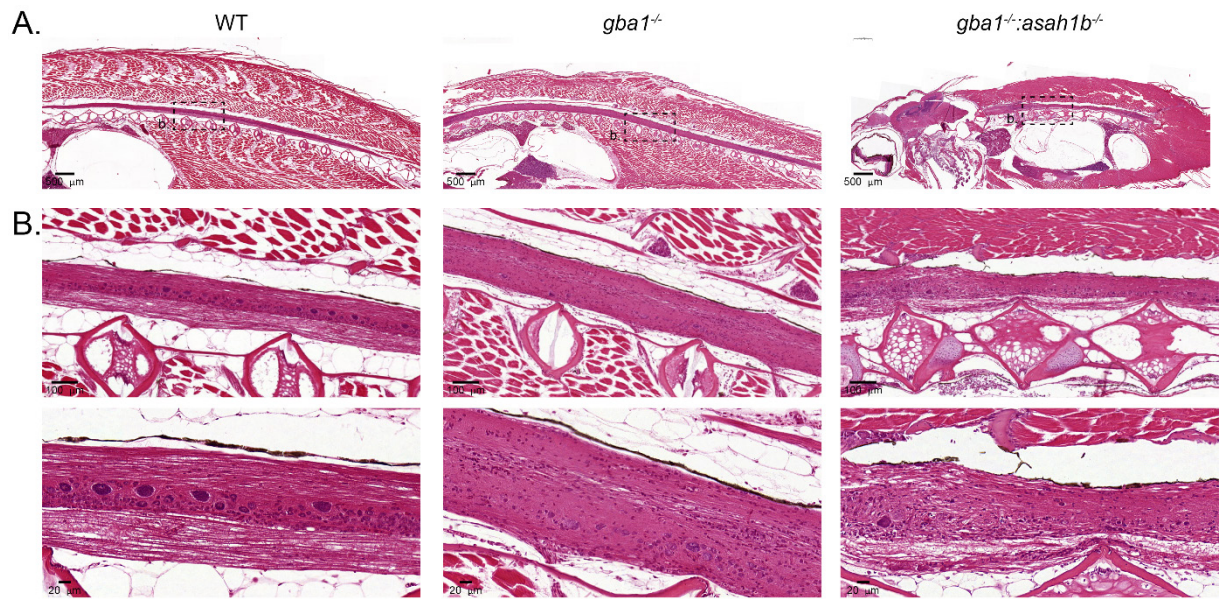
**Supplemental Figure S6:** Histopathology of WT, *gba1*<sup>-/-</sup> and *gba1*<sup>-/-</sup>:*asah1b*<sup>-/-</sup> of sagittal sections of zebrafish brains. **(A)** Immunostaining of L-plastin with low magnification. Scale bars: 200 μm. **(B)** Higher magnification focusing on periventricular grey zone with H&E staining (upper panel) and immunostaining of L-plastin (lower panel). To compare consecutive sections stained with H&E and L-plastin, H&E staining sections were the same as displayed in Figure 6B. Scale bars: 50 μm. **(C)** Higher magnification focusing on the medulla oblongata with H&E staining (upper panel) and immunostaining of L-plastin (lower panel). Scale bars: 50 μm.





**Supplemental Figure S7: Heatmaps of individually filmed *asah1b*<sup>-/-</sup> and *gba1*<sup>-/-</sup>:*asah1b*<sup>-/-</sup> zebrafish.**

Heatmaps of *asah1b*<sup>-/-</sup> (yellow stickers) and *gba1*<sup>-/-</sup>:*asah1b*<sup>-/-</sup> (orange stickers) at 15 wpf, with individual fish starting to show differences in swimming behaviour. Red indicates more time and blue less time spend at that location.



**Supplemental Figure S8:** (A) H&E staining of spinal cords of WT,  $gba1^{-/-}$  and  $gba1^{-/-}:asah1b^{-/-}$  including higher magnifications in (B). Scale bars: 500  $\mu\text{m}$  for (A) and 100  $\mu\text{m}$  and 20  $\mu\text{m}$  for the top and bottom panel in (B).

**Supplemental Video 1** | Short video of WT (white stickers),  $gba1^{-/-}$  (red),  $asah1b^{-/-}$  (yellow) and  $gba1^{-/-}:asah1b^{-/-}$  (orange stickers).

**Supplemental Video 2** | Short video of  $gba1^{-/-}$  (red),  $asah1b^{-/-}$  (yellow) and  $gba1^{-/-}:asah1b^{-/-}$  (orange stickers).

**Supplemental Video 3** | Short video of WT (white stickers),  $asah1b^{-/-}$  (yellow) and  $gba1^{-/-}:asah1b^{-/-}$  (orange stickers).