

## **Supplementary Information for**

# **RibU is an essential determinant of** *Listeria* **pathogenesis that mediates acquisition of FMN and FAD during intracellular growth**

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#### **Supplementary Information Figures**



Intracellular

**Fig. S1. Flavin metabolism in** *L. monocytogenes***.** *L. monocytogenes* encodes the energycoupling factor (ECF) transporter RibU which is composed of four subunits: RibU, the substrate binding subunit; EcfT, the transmembrane coupling subunit; and EcfA and EcfA', two ATPase subunits. RibU binds and transports riboflavin, as well as FMN and FAD (this study). The RibU subunit is dissociated in the membrane in its flavin unbound form and upon capturing a flavin molecule it interacts with the transport module (EcfT:EcfA:EcfA') to import the flavin across the membrane using energy from ATP hydrolysis (1). Upon riboflavin import, the bifunctional enzyme RibC catalyzes the phosphorylation of riboflavin to generate FMN and the conversion of FMN to FAD by attaching an adenosine monophosphate molecule (adenylylation) to FMN. The enzyme RibF can also adenylylate FMN to synthesize FAD (2, 3).



**Fig. S2. The ∆***ribU* **strain producing riboflavin grows in chemically defined media lacking flavins.** Broth growth curve of *L. monocytogenes* strains grown in chemically defined media lacking flavins. OD<sub>600</sub> was used to determine cell density. The data show the means and standard deviations of three independent experiments. In chemically defined media lacking flavins, wildtype *L. monocytogenes* grows until it depletes its flavin pool. In contrast, the ∆*ribU* + *ribDEAHT*  strain grows to higher densities. The picture shows the media supernatant of wild-type (left) and the ∆*ribU* + *ribDEAHT* strain (right) after 24 h of growth at 37 °C shaking. The change in color from colorless to bright yellow, the natural color of flavins, suggests the ∆*ribU* + *ribDEAHT* strain is producing riboflavin and allowing it to grow in media lacking flavins.



**Fig. S3. Growth dynamics of the ∆***ribU* **mutant in BMMs.** (A) Percentage of bacteria that colocalized with the autophagy receptor p62 in infected BMMs. In BMMs treated with cytochalasin D, bacteria that escape phagosomes are tagged with p62. Percent phagosomal escape is calculated by counting the number of  $p62<sup>+</sup>$  bacteria of total bacteria. The data show the means and standard errors of the mean of two independent experiments. (B-C) Intracellular growth curves of *L. monocytogenes* strains in BMMs. BMMs were infected at an MOI of 0.1 and CFUs were enumerated at the indicated times. (B) Intracellular growth curve of indicated *L. monocytogenes* strains in wild-type BMMs. The data show the means and standard errors of the mean of two independent experiments. (C) Intracellular growth curve of indicated flavin-starved *L. monocytogenes* strains in riboflavin-deprived wild-type BMMs (for 3 h) supplemented with 1 µM riboflavin just prior to infection. The data represent the means and standard errors of the mean of three independent experiments. (D) Generation time of intracellularly growing bacteria in riboflavin-sufficient and -deprived BMMs between 2 to 5 h. Negative values indicate that the number of recoverable bacteria was decreasing over time.

### **Table S1. Bacterial strains used in this study**



#### **SI References**

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