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#### **Supplemental Material**

Screening of Organophosphate Flame Retardants with Placentation-Disrupting Effects in Human Trophoblast Organoid Model and Characterization of Adverse Pregnancy Outcomes in Mice

Chenke Xu, Haojia Ma, Fumei Gao, Chenhao Zhang, Wenxin Hu, Yingting Jia, Jun Xu, and Jianying Hu

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**Figure S1. Induced and Characterized Trophoblast Organoids and Optimized Protocol.** (*A*) Human trophoblast organoid; (*B*) Immunofluorescence of villus markers KRT7 (green), GATA3 (red), and DAPI (blue) in trophoblast organoids following previous protocol (Sheridan, 2020); (*C*) Previous digestion (Sheridan, 2020) and optimized digestion (3 min in 37°C and pipetted gently); (*D*) Relative Ki67 intensity (means ± SDs) of organoids after 2-day culture in previous digestion (Sheridan, 2020) and optimized digestion (3 min in 37°C and pipetted gently); (*E*) Relative Ki67 intensity (means ± SDs) of organoids after 2-day culture in previous digestion (Sheridan, 2020) and optimized digestion (3 min in 37°C and pipetted gently); (*E*) Relative Ki67 intensity (means ± SDs) of organoids after 2-day culture in previous and optimized TOM (Table S5). Data in (*C*) and (*D*) are expressed relative to the levels of previous digestion and previous TOM, respectively, which were set to 1. *n* = 3. All organoids in (*C*) and (*D*) were from a single donor. Data were analyzed using an unpaired two-tailed Student's t-test. Indicated values are significantly different from control value. \**p* < 0.05. KRT7, keratin 7; GATA3, GATA binding protein 3; SD, standard deviation; TOM, trophoblast organoid medium. Scale bars, 60 µm. Numeric data in (*D*)-(*E*) were listed in Table S16.

**Figure S2. Additional measures of short- and long-term toxicity of EHDPP in organoids.** (*A*) Relative fluorescence intensity (means  $\pm$  SDs) of Sytox Green (green) in trophoblast organoids in 2-day EHDPP exposure; (*B*) Relative fluorescence intensity (means  $\pm$  SDs) of Sytox Green (green) in trophoblast organoids in 10-day EHDPP exposure; (*C*) Western blotting of PD in control and 2-day EHDPP exposure groups; (*D*) Relative protein levels of PD (means  $\pm$  SDs) in western blotting in control and 2-day EHDPP exposure groups; (*E*) CD71 (green) and DAPI (grey) in control and 2-day EHDPP (10,000 nM) exposure groups; (*F*) Relative intensity (means  $\pm$  SDs) of CD71 in (*E*); (*G*) HLA-G (red), F-actin (blue) and DAPI (grey) in 4-day culture. Data in (*A*)-(*G*) are expressed relative to the levels of DMSO-treated organoids, which were set to 1. *n* = 3. All organoids in (*A*)-(*G*) were from a single donor. Analyzed by the unpaired two-tailed Student's t-test. Indicated values are significantly different from control value. \**p* < 0.05. \*\**p* < 0.01. Scale bars, 20 µm. EHDPP, 2-ethylhexyl-diphenyl phosphate; PD, pyruvate dehydrogenase complex; CD71, transferrin receptor; HLA-G, human leucocyte antigen protein-G; DAPI, 4',6-diamidino-2-phenylindole; DMSO, dimethyl sulfoxide. Numeric data in (*A*), (*B*), (*D*) and (*F*) were listed in Table S16.

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Table S18. Numeric data for Fig S4B, S4C, S4D, S4E, S4G, S4H.

Sequence of HA-GFP

References

Additional File- Excel Document

#### **Supplementary Text**



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Classification	Full name (Abbreviation)	CAS	Company
	Triethyl phosphate (TEP)	78-40-0	TCI Corp., P0270
	Triisopropyl phosphate (TiPP)	513-02-0	Sigma, 554669
	Tributyl phosphate (TBP)	126-73-8	Sigma, 8186040100
	Tripropyl phosphate (TPrP)	513-08-6	Macklin, T819409
	Triisobutyl phosphate (TiBP)	126-71-6	Macklin, T834115
	Tris(2-ethylhexyl) phosphate (TEHP)	78-42-2	TCI Corp., P1022
	Dimethyl hydrogen phosphate (DMP)	813-78-5	Macklin, D885090
	Diethyl hydrogen phosphate (DEP)	598-02-7	Macklin, D839196
Alkyl-OPFRs	Dibutyi phosphate (DBP) "(He et al. 2021) Pis(2 athylhawyl) hydrogon phosphate (PEUD) *(Hyo at al. 2020)	10/-00-4	Macklin, D8359/6 Macklin, D802752
	Tribevul phosphate (T6CP) *(Wang, 2020)	298-07-7	Macklin, B002732 Macklin, T008781
	Timexyl phosphate (10e1) (wang, 2020)	2328-37-4	Wackini, 1908/81
	Isopropylphenyl Phenyl Phosphate (ip-PPP)	68782-95-6	Macklin, I860749
	2-Ethylhexyl diphenyl phosphate (EHDPP)	1241-94-7	TCI Corp., P1021
	Isodecyl diphenyl phosphate (IDDPP)	29761-21-5	AccuStandard, PFRS-008S
	Triphenyl phosphate (TPhP)	115-86-6	TCI Corp., P0272
	Tris(3,5-xylenyl) phosphate (TXP)	25155-23-1	Synthesized
	Tris(4-tert-butylphenyl) phosphate (T4tBPPP)	78-33-1	Synthesized
	Tricresyl phosphate (TCrP)	1330-78-5	Macklin, T819410
	Diisodecyl phenyl phosphate (DIDPP)	51363-64-5	Synthesized
	Tri-O-cresyl phosphate (o-TCrP)	78-30-8	Macklin, T820120
	Tricresyl phosphate (p-TCrP)	78-32-0	Macklin, T862850
	Tri-M-cresyl phosphate (m-TCrP)	563-04-2	Macklin, T867695
	Tris(4-isopropylphenyl) phosphate (T4IPPP)	2502-15-0	Wellington Laboratories
Arvl_OPFRs	Bis(p-tert-butylphenyl) phenyl phosphate (B4tBPPP)	115-87-7	Wellington Laboratories
/ ilyi-of i ks	Resorcinol bis(diphenyl phosphate) (RDP)	57583-54-7	Macklin, R860747
	Cresyl diphenyl phosphate (CDP)	26444-49-5	Macklin, C8341114
	Diphenyl phosphate (DPP) *(Funk et al. 2019)	838-85-7	Macklin, D831185
	Tris(nonylphenyl) phosphate (TNPP) *(Wang et al. 2020)	26569-53-9	Synthesized
	Isopropylphenyl diphenyl phosphate (2IPPDPP) *(Wang et al. 2020)	64532-94-1	Wellington Laboratories
	Bis(2-isopropylphenyl) phenyl phosphate (B2IPPPP) *(Wang et al. 2020)	69500-29-4	Wellington Laboratories
	4-tert-butylphenyl diphenyl phosphate (4tBPDPP) *(Wang et al. 2020)	981-40-8	Wellington Laboratories
	Bis(2-ethylhexyl) phenyl phosphate (BEHPP) *(Wang et al. 2020)	16368-97-1	Macklin, B905605
	Bisphenol A bis(diphenyl phosphate) (BPADP) *(Wang et al. 2020)	5945-33-5	Aladdin, B304092
	Tris(2,4-ditert-butylphenyl) phosphate (T4DtBPP) *(Wang et al. 2020)	95906-11-9	Wellington Laboratories
	Bis(2-methylphenyl) hydrogen phosphate (DoCP) *(Gao, 2020)	35787-74-7	TRC, D494575

### Table S1. OPFRs list for screening

Most of OPFRs were picked from Toxicity Forecaster (Toxcast) list (<u>https://comptox.epa.gov/dashboard/chemical-lists/FLAMERETARD</u>). \* Chemicals that are not on the Toxicity Forecaster (Toxcast)

### Continued

Classification	Full name (Abbreviation)	CAS	Company
	Tris(1,3-dichloro-2-propyl)phosphate (TDCIPP)	13674-87-8	TCI Corp., P0269
	Tris(2-chloroethyl) phosphate (TCEP)	115-96-8	Sigma-Aldrich, 119660
	tert-Butylphenyl diphenyl phosphate (BPDPP)	56803-37-3	Johnlong, CFRT111308
	Bis(2,3-dibromopropyl) hydrogen phosphate (BDBPP)	5412-25-9	Bioruler, RH132419
	Tris(1-chloro-2-propyl) phosphate (TCIPP)	13674-84-5	Rowan, R010190
Halogenated-	2,2-Bis-(bromomethyl)-3-bromo-1-propanol phosphate (TTBNPP)	19186-97-1	Macklin, T859201
OPFRs	Tris(2,3-dibromopropyl) phosphate (TDBPP)	126-72-7	AccuStandard, PFS-008N
	Oxydi-2,1-Ethanediyl-Phosphoric Acid Tetrakis(2-Chloro-1-Methylethyl) Ester (RDT905)* (Wang et al. 2021)	52186-00-2	Omnistab, FR-RDT 9
	Phosphoric acid, 2,2-bis(chloromethyl)-1,3-propanediyl tetrakis(2-chloroethyl) ester (V6)* (Wang et al. 2021)	38051-10-4	AccuStandard, PFS-020S
	3,9-Bis(2,4-di-tert-butylphenoxy)-2,4,8,10-tetraoxa-3,9- diphosphaspiro[5.5]undecane (AO626=O2)*(Wang et al. 2021)	26741-53-7	Macklin, A832583
	Bis(2-chloroethyl) phosphate (BCEP)*(He et al. 2021)	3040-56-0	Johnlong, CCHM701085

Most of OPFRs were picked from Toxicity Forecaster (Toxcast) list

(<u>https://comptox.epa.gov/dashboard/chemical-lists/FLAMERETARD</u>). \* Chemicals that are not on the Toxicity Forecaster (Toxcast)

Abbreviation	Geographical information	Reference
DBP	the USA	(He et al. 2021)
BEHP	China	(Huo et al. 2020)
T6CP	China	(Gao et al. 2020)
DPP	China; the USA; Canada	(Gao et al. 2020; Carignan et al. 2017; Funk et al.
		2019)
TNPP	Canada; China	(Liu et al. 2019; Wang et al. 2020)
2IPPDPP	China	(Wang et al. 2020)
B2IPPPP	China	(Wang et al. 2020)
4tBPDPP	China	(Wang et al. 2020)
BEHPP	China	(Wang et al. 2020)
BPADP	China	(Wang et al. 2020)
T4DtBPP	China	(Wang et al. 2020)
DoCP	China	(Gao et al. 2020)
RDT905	China	(Wang et al. 2021)
V6	China; the USA	(Wang et al. 2021, Stapleton et al. 2011)
AO626=O2	China	(Wang et al. 2021)
BCEP	the USA	(He et al. 2021)

Table S2. Geographical information of exposure in 16 OPFRs

Note: For definition of abbreviations, see Table S1.

## Table S3. Chemicals used in this paper

Chemicals	Company	Final concentration
OSI-906 (Linsitinib)	MCE, HY-10191	50 nM
Testosterone	NOVUS, NBP2-45187	40 µM

### Table S4. Information of donors

Donor Number	Age	Gestational age	Karyotype
1#	24	7 weeks	44+XX
2#	28	8 weeks	44+XY
3#	30	7 weeks	44+XX
4#	39	7 weeks	44+XY
5#	28	8 weeks	44+XY

Product	Company and Product	Optimized (Previous)
	Number	concentration
Advanced DMEM/F12	Life	1X
	Technologies, 12634010	
N2 supplement	Life Technologies,	1X
	17502048	
B27 supplement	Life Technologies,	1X
	12587010	
L-glutamine	Life Technologies, 25030	2 mM
N-acetyl-L-cysteine	Sigma, A9165	1.25 mM
ALK-4, -5, -7 inhibitor, A83-0.1	Tocris, 2939	500 nM
CHIR99021	Tocris, 4423	5 μΜ (1.5 μΜ)
Recombinant human EGF	Peprotech, AF-100-15	50 ng/mL
Recombinant human R-spondin	BioTechne, 4645-RS	80 ng/mL
1		
Recombinant human FGF2	Peprotech, 100-18C	100 ng/mL
Recombinant human HGF	Peprotech, 100-39	50 ng/mL
Y27632	Tocris, 1254	10 μM (2 μM)
PGE2	Sigma, P0409	10 μM (2.5 μM)
Penicillin Streptomycin	Gibco, 15140122	100X

 Table S5. Trophoblast organoid medium (TOM)

# Table S6. Primer sequences in RT-qPCR

Primer	Forward	Reverse
Human IGF1	GCTCTTCAGTTCGTGTGTGGA	GCCTCCTTAGATCACAGCTCC
Human IGF2	GTGGCATCGTTGAGGAGTG	CACGTCCCTCTCGGACTTG
Human IGF1R	TCGACATCCGCAACGACTATC	CCAGGGCGTAGTTGTAGAAGAG
Human $\beta$ -actin	CATGTACGTTGCTATCCAGGC	CTCCTTAATGTCACGCACGAT

Antibody	Company	Product number	Dilution
anti-Ki67	CST	12075S	1:100 (IF)
anti-HLA-G	Abcam	ab52454	1:100 (IF)
anti-p63	Abcam	ab124762	1:100 (IF)
anti-CD71	Abcam	ab38171	1:100 (IF)
anti-Cdx2	Abcam	ab576541	1:50 (IF)
anti-rabbit lgG H&L	Abcam	ab150075	1:100 (IF)
anti-mouse lgG H&L	CST	8890S	1:100 (IF)
anti-trophoblast specific protein $\alpha$	Abcam	ab104401	1:30 (IF)
anti-HAND1	Bioss	bs-9459R	1:30 (IF)
anti-transcription factor AP-2y	Santa Cruz	sc-8977	1:30 (IF)
DAPI	SIGMA	D9542	1:5000 (IF)
Alexa fluor 488 Phallo	Molecular Probes	A12379	1:200 (IF)
anti-pyruvate dehydrogenase	CST	32508	1:1000 (WB)
anti-cytochrome c	CST	4280S	1:1000 (WB)
anti-β-Actin	CST	3700S	1:2000 (WB)
anti-mouse IgG HRP-linked	CST	7076S	1:5000 (WB)
anti-rabbit IgG HRP-linked	CST	14708S	1:1000 (WB)
anti-phospho-Akt (Ser <sub>473</sub> )	CST	4060S	1:1000 (WB)
anti-Akt (pan)	CST	7076S	1:5000 (WB)
anti-phospho-IGF1Rβ (Tyr1135)	CST	3918S	1:1000 (WB)
anti-IGF1Rβ	CST	9750S	1:1000 (WB)
anti-KRT7	CST	4465S	1:50 (IF)
anti-GATA3	Abcam	ab100428	1:50 (IF)

Table S7. Antibodies and DAPI

\* IF, immunofluorescence; WB, western blotting.

Chemicals	Average	Average
	Intensity of Ki67	Intensity of Sytox Green
DMSO	$7,832.3 \pm 896.1$	$2,336.7 \pm 29.1$
TEP	$7,129.2 \pm 1,478.2$	$2,381.1 \pm 127.4$
TiPP	$8,214.9 \pm 1,151.1$	$2,280.8 \pm 72.5$
TBP	$7,\!168.4\pm1,\!726.7$	$2,383.9 \pm 20.6$
TPrP	$7,560.9 \pm 392.4$	$2,422.3 \pm 131.7$
TiBP	$7,168.4 \pm 889.5$	$2,348.1 \pm 38.6$
TEHP	$7,390.8 \pm 13.1$	$2,312.1 \pm 35.0$
DMP	$8,057.9 \pm 1268.9$	$2,339.5 \pm 65.7$
DEP	$7,\!822.5\pm1046.5$	$2,265.2 \pm 25.1$
DBP	$7,220.8 \pm 758.7$	$2,316.5 \pm 31.9$
BEHP	$7,142.3 \pm 444.8$	$2,384.7 \pm 27.3$
T6CP	$7,338.5 \pm 1517.4$	$2,342.0 \pm 56.2$
ip-PPP	$6,\!370.5 \pm 130.8$	$2,431.7 \pm 13.0$
EHDPP	$5{,}598.7 \pm 601.7$	$2,\!622.7\pm159.5$
IDDPP	$5,808.0 \pm 1,216.5$	$2{,}568.9\pm2.0$
TPhP	$7,966.4 \pm 366.3$	$2,506.6 \pm 56.3$
ТХР	$6{,}906.8\pm797.9$	$2,526.0 \pm 35.4$
T4tBPPP	$7,534.7 \pm 745.6$	$2,590.1 \pm 240.0$
TCrP	$7,403.9 \pm 222.4$	$2,557.9 \pm 45.0$
DIDPP	$6,\!148.1\pm954.9$	$2,\!407.0\pm92.2$
o-TCrP	$5,232.4 \pm 811.0$	$2,391.9 \pm 102.7$
p-TCrP	$8,633.5 \pm 366.3$	$2,\!409.9\pm89.7$
m-TCrP	$6{,}645.2 \pm 994.2$	$2,\!388.4\pm93.6$
T4IPPP	$6,\!434.3\pm 497.1$	$2,469.0 \pm 131.1$
B4tBPPP	$5,546.4 \pm 484.0$	$2,534.9 \pm 58.0$
RDP	$9222.2\pm457.8$	$2,303.1 \pm 18.6$
CDP	$6,370.5 \pm 143.9$	$2,422.5 \pm 76.8$
DPP	$7,521.6 \pm 706.4$	$2,403.1 \pm 137.2$
TNPP	$11,092.8 \pm 1,420.7$	$2,302.5 \pm 41.2$

 Table S8. Screening of OPFRs

Continued	l
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Chemicals	Average Intensity of Ki67 (AU)	Average Intensity of Sytox Green (AU)
2IPPDPP	$7,\!168.4 \pm 1,\!530.5$	$2,299.6 \pm 64.5$
B2IPPPP	$6,684.4 \pm 26.2$	$2,612.4 \pm 26.0$
4tBPDPP	$7,430.1 \pm 2,106.1$	$2,115.2 \pm 230.8$
BEHPP	$7,\!050.7\pm797.9$	$2,334.5 \pm 115.3$
BPADP	$7,\!142.3\pm1,\!151.1$	$2,365.4 \pm 92.7$
T4DtBPP	$6{,}815.2\pm706.4$	$2,524.9 \pm 63.0$
DoCP	$7,273.1 \pm 876.4$	$2,362.6 \pm 39.6$
TDCIPP	$7,757.1 \pm 1,203.5$	$2,401.5 \pm 31.4$
TCEP	$8,\!829.7\pm706.4$	$2,\!289.7\pm41.4$
BPDPP	$8,254.2 \pm 1,412.8$	$2,361.0 \pm 115.7$
BDBPP	$7,076.9 \pm 1,177.3$	$2,384.2 \pm 73.9$
TCIPP	$6,933.0 \pm 811.0$	$2,354.0 \pm 60.4$
TTBNPP	$6,370.5 \pm 196.2$	$2,461.6 \pm 84.5$
TDBPP	$7,377.7 \pm 2,145.3$	2,511.9 ±86.0
RDT905	$7,678.6 \pm 627.9$	$2,487.8 \pm 73.5$
V6	$6,854.5 \pm 340.1$	$2,566.0 \pm 41.3$
AO626=O2	$8,097.2 \pm 1,151.1$	$2,543.0 \pm 113.4$
BCEP	$7,704.8 \pm 1,007.2$	$2,424.8 \pm 271.4$

Note: numerical data for Figure 2C, D (means  $\pm$  SDs); AU, arbitrary units.

**Table S9.** Numeric data for Fig 3B, 3D, 3E, 3G, 3I, 3K, 3L, 3M, 3O, 3P (means ± SDs)

Experiment	Dose	Numeric data
	DMSO	$187.3 \pm 15.0$
Average intensity of	100 nM EHDPP	$166.7 \pm 18.7$
Ki67 (2 d; AU)	1,000 nM EHDPP	$148.0\pm18.7$
	10.000 nM EHDPP	$89.9 \pm 24.3$
	DMSO	$3.018.1 \pm 740.3$
<b>Basic Metabolism</b>	100 nM EHDPP	$2.042.5 \pm 560.4$
(pmol/min)	1.000 nM EHDPP	$1.344.5 \pm 380.9$
	10,000 nM EHDPP	$1,269.1 \pm 252.0$
	DMSO	$3,012.2\pm477.8$
Respiratory Capacity	100 nM EHDPP	$2,383.6 \pm 790.2$
(pmol/min)	1,000 nM EHDPP	$1,643.9 \pm 655.9$
	10,000 nM EHDPP	$1,528.9 \pm 383.4$
	DMSO	$103.3 \pm 16.5$
Average intensity of	100 nM EHDPP	$90.9 \pm 13.4$
TP63 (2 d; AU)	1,000 nM EHDPP	$67.1 \pm 12.4$
	10,000 nM EHDPP	$59.9 \pm 18.6$
	DMSO	$172.9\pm8.6$
Average intensity of	100 nM EHDPP	$164.3 \pm 1.7$
Ki67 (10 d; (AU)	1,000 nM EHDPP	$138.3\pm19.0$
	10,000 nM EHDPP	$115.9\pm20.8$
	DMSO	$97.4\pm6.8$
Average intensity of	100 nM EHDPP	$84.7\pm26.7$
TP63 (10 d; AU)	1,000 nM EHDPP	$69.1\pm15.9$
	10,000 nM EHDPP	$63.8 \pm 11.4$
	DMSO	$100.8\pm15.1$
Average intensity of	100 nM EHDPP	$67.5\pm4.0$
CD71 (10 d; AU)	1,000 nM EHDPP	$55.4 \pm 16.1$
	10,000 nM EHDPP	$51.4 \pm 11.1$
	DMSO	$105.7 \pm 12.7$
Average intensity of	100 nM EHDPP	$94.1 \pm 22.2$
HLA-G(10 d; AU)	1,000 nM EHDPP	$75.0 \pm 16.9$
	10,000 nM EHDPP	$68.7 \pm 12.7$
	DMSO	$10,188.3 \pm 611.3$
Concentration of hCG	100 nM EHDPP	$10,780.0 \pm 305.6$
(mIU/mL)	1,000 nM EHDPP	$8,\!456.3\pm203.8$
	10,000 nM EHDPP	$7946.9 \pm 101.9$
Concentration of E2	DMSO	$24,585.7 \pm 245.9$
Concentration of E2 $(n \alpha/m T)$	100 nM EHDPP	$22,864.7 \pm 2,704.4$
(pg/mL)	1,000 nM EHDPP	$19{,}422.7 \pm 1966.9$
	10,000 nM EHDPP	$17701.7 \pm 2950.3$

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AU, arbitrary units; TP63, tumor protein 63; CD71, transferrin receptor; HLA-G, human leucocyte antigen protein-G; hCG, human chorionic gonadotropin; E2, estradiol

Dose	Time	Numeric data	Dose	Time	Numeric data
	(min)	$(means \pm SDs)$		(min)	$(means \pm SDs)$
	1.40	$615.06 \pm 132.10$		1.40	$420.45 \pm 119.32$
	9.97	$588.07 \pm 149.15$		9.97	$401.99 \pm 105.11$
	18.55	$582.39 \pm 153.41$		18.55	$393.47\pm98.01$
	27.21	$394.89 \pm 123.58$		27.21	$295.45\pm80.97$
	35.78	$306.82\pm98.01$		35.78	$248.58\pm85.23$
	44.31	$257.10 \pm 86.65$		44.31	$207.39\pm66.76$
DMSO	52.99	$640.63 \pm 83.81$	EHDPP 100	52.99	$549.72 \pm 150.57$
	61.55	$602.27\pm98.01$	nM	61.55	$480.11 \pm 180.40$
	70.12	$627.84 \pm 132.10$		70.12	$453.13 \pm 149.15$
	78.80	$197.44 \pm 89.49$		78.80	$176.14 \pm 39.77$
	87.39	$130.68\pm36.93$		87.39	$132.10\pm36.93$
	95.93	$123.58\pm22.73$		95.93	$126.42 \pm 39.77$
	1.40	$278.41 \pm 82.39$		1.40	$262.78\pm38.35$
	9.97	$264.20 \pm 78.13$		9.97	$264.20\pm45.45$
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	18.55	$257.10 \pm 79.55$		18.55	$258.52\pm49.72$
	27.21	$213.07 \pm 39.77$			
	35.78	$139.20\pm58.24$		35.78	$178.98\pm22.73$
	44.31	$122.16\pm42.61$		44.31	$153.41\pm24.15$
EHDPP	52.99	$356.53 \pm 133.64$	EHDPP	52.99	$365.06 \pm 88.07$
1,000 nM	61.55	$338.07 \pm 153.41$	10,000 nM	61.55	$318.18\pm69.06$
	70.12	$321.02 \pm 154.83$		70.12	$296.88\pm62.50$
	78.80	$142.05\pm8.52$		78.80	$125.00 \pm 39.77$
	87.39	$93.75\pm7.10$		87.39	$90.91\pm8.52$
	95.93	$79.55\pm2.84$		95.93	$78.13\pm5.68$

 Table S10. Numeric data for Fig 3C (oxygen consumption rate; pmol/min)

Table S11. Numeric data for Fig 3Q (means  $\pm$  SDs)

Experiment	Strain of organoids	Group	Numeric data (AU)
	1#	DMSO	$459.5\pm41.2$
	$1\pi$	EHDPP	$354.4\pm16.0$
	2#	DMSO	$383.5\pm24.7$
	2#	EHDPP	$287.9\pm12.3$
Average intensity of	24	DMSO	$388.5\pm42.8$
Ki67	5#	5# EHDPP	$282.4\pm14.5$
	14	DMSO	$402.2\pm12.6$
	<b>4</b> #	EHDPP	$332.7\pm24.7$
	5#	DMSO	$349.7\pm25.9$
	5#	EHDPP	$281.8\pm\!\!14.0$

AU, arbitrary units

Experiment	Dose	Numeric data		
		1#	2#	3#
	DMSO	153.1	149.7	150.5
Gray value of p-Akt (Ser473;	100 nM EHDPP	142.7	141.9	147.8
Intden/Area, AU)	1,000 nM EHDPP	136.9	144.5	142.2
	10,000 nM EHDPP	142.7	140.2	147.2
	Background	128.6	128.9	136.4
		1#	2#	3#
	DMSO	201.2	201.8	209.5
Gray value of Akt (Intden/Area; AU)	100 nM EHDPP	196.0	202.9	207.2
	1,000 nM EHDPP	190.0	192.0	210.0
	10,000 nM EHDPP	194.7	206.9	219.0
	Background	123.3	125.1	122.5
		1#	2#	3#
Conservation of a ICE1D (Intelliged A	DMSO	194.1	205.0	211.1
Gray value of p-IGFTR (Intden/Area;	100 nM EHDPP	177.8	178.1	180.0
AO)	1,000 nM EHDPP	168.5	169.5	163.6
	10,000 nM EHDPP	174.3	172.1	167.2
	Background	141.5	144.8	131.5
		1#	2#	3#
Creative of ICE1D (Intdam/Arrest	DMSO	211.6	202.8	202.4
Gray value of IGFTR (Iniden/Area;	100 nM EHDPP	205.0	209.3	194.2
AO)	1,000 nM EHDPP	206.8	200.8	183.8
	10,000 nM EHDPP	200.3	224.6	217.6
	Background	144.5	155.9	158.7
	HA-GFP transfection (1,000 nM $0.418 \pm 0.0$ EHDPP)		)34	
EHDPP (ng/mL; means $\pm$ SDs)	HA-IGF1R transfection (1,000 nM EHDPP)	F1R transfection (1,000 nM $1.162 \pm 0.384$ EHDPP)		
	HA-GFP transfection (10,000 nM EHDPP)	$3.177\pm0.062$		
	HA-IGF1R transfection (10,000 nM EHDPP)	$10.166\pm5.433$		

Table	<b>S12</b>	Numeric	data	for	Fig	$\Delta C$	4D	4F
Lavic	014.	INUITOITO	uata	101	rig	<del>т</del> С,	+D,	ΗL

AU, arbitrary units

Chemical	Dose	Numeric data (means ± SDs)
	0.1 nM	$116.4 \pm 8.5\%$
	1 nM	$120.3\pm4.2\%$
	10 nM	$95.6\pm5.0\%$
EHDPP	100 nM	$70.2\pm7.7\%$
	1,000 nM	$50.1 \pm 11.1\%$
	10,000 nM	$10.5\pm3.5\%$
	0.001 nM	$108.5\pm5.2\%$
	0.01 nM	$102.4 \pm 14.2\%$
	0.1 nM	$76.1\pm5.3\%$
081-906	1 nM	$62.3\pm4.2\%$
	10 nM	$50.1\pm6.3\%$
	100 nM	$15.7\pm0.4\%$

Table S13. Numeric data for Fig 4F (% inhibition)

Table S14. Numeric data for Fig 5D, 5E, 5J, 5L, 5M

Experiments	Dose	Numeric data (means ± SDs)
Average intensity of	Control	$343.8\pm58.4$
Cdx2 (E7.5; AU)	10 mg/kg EHDPP	$240.7\pm75.64$
Average intensity of	Control	$572.6\pm40.3$
Ki67 (E7.5; AU)	10 mg/kg EHDPP	$503.9\pm40.1$
	Control	$1.495\pm0.105$
	0.4 mg/kg EHDPP	$1.490\pm0.145$
Body weight (g)	2 mg/kg EHDPP	$1.429\pm0.142$
	10 mg/kg EHDPP	$1.401\pm0.146$
	Control	$156 (15.6 \pm 1.58)$
Implanted embryonic	0.4 mg/kg EHDPP	$152 (15.2 \pm 1.99)$
number (Total (Mean	2 mg/kg EHDPP	$142 (14.2 \pm 2.10)$
± SD))	10 mg/kg EHDPP	$137 (13.7 \pm 1.77)$
	Control	$152 (15.2 \pm 1.87)$
Surviving embryonic	0.4 mg/kg EHDPP	$143 (14.3 \pm 1.70)$
number (Total (Mean	2 mg/kg EHDPP	$128 (12.8 \pm 2.62)$
± SD))	10 mg/kg EHDPP	$123 (12.3 \pm 2.54)$

AU, arbitrary units; Cdx2, caudal-type homeobox 2

Dage	Time (min)	Blood glucose
Dose	Time (min)	(mmol/L)
	0	$2.46\pm0.70$
	15	$11.54 \pm 2.77$
Control	30	$14.83\pm2.57$
Control	60	$12.04\pm3.02$
	90	$6.82\pm3.18$
	0	$2.77 \pm 1.03$
	15	$12.29 \pm 1.15$
0.4  mg/ltg EUDDD	30	$13.60 \pm 2.29$
0.4 mg/kg EnDPP	60	$12.09 \pm 3.24$
	90	$8.97 \pm 3.72$
	0	$1.93\pm0.95$
	15	$9.22 \pm 3.55$
2 ma/lta ELIDDD	30	$11.93 \pm 4.19$
2 mg/kg EHDPP	60	$11.98 \pm 5.06$
	90	$8.24\pm3.94$
	0	$2.07\pm0.53$
	15	$13.04\pm1.12$
10 mg/kg EHDPP	30	$14.19\pm2.21$
00-	60	$15.25\pm1.93$
	90	$11.73\pm2.91$

**Table S15.** Numeric data for Fig 5K (means  $\pm$  SDs)

Experiments	Group	Numeric data		ata		
Average intensity of	Previous digestion	47	$78.3 \pm 29$	0.2		
Ki67 (AU)	Optimized digestion	$406.6\pm28.7$				
Average intensity of	Previous TOM	34	$17.7 \pm 13$	.9		
Ki67 (AU)	Optimized TOM	40	$406.8\pm13.4$			
Average intensity of	DMSO	$1579.1 \pm 41.2$				
Sytox Green (2 d· AU)	100 nM EHDPP	163	$37.7 \pm 17$	5.3		
5 y tox 6100n (2 u, 110)	1,000 nM EHDPP	17	$01.5 \pm 44$	4.0		
	10,000 nM EHDPP	16	$42.9 \pm 42$	3.7		
Average intensity of	DMSO	15	$82.4 \pm 33$	8.4		
Sytox Green (2 d· AU)	100 nM EHDPP	16	$17.3 \pm 52$	2.7		
5 y tox 6100n (2 u, 110)	1,000 nM EHDPP	16	$1621.6 \pm 21.1$			
	10,000 nM EHDPP	$1621.0\pm21.9$		1.9		
		1#	2#	3#		
~ 1 175	DMSO	189.5	152.9	148.4		
Gray value of PD	100 nM EHDPP	170.2	152.6	146.3		
(Intden/Area; AU)	1,000 nM EHDPP	161.6	146.8	145.8		
	10,000 nM EHDPP	160.0	148.0	145.6		
	Background	137.4	134.6	137.1		
		1#	2#	3#		
	DMSO	225.1	227.0	214.3		
Gray value of $\beta$ -actin	100 nM EHDPP	224.6	226.8	206.3		
(Intden/Area; AU)	1,000 nM EHDPP	219.7	228.1	205.7		
	10,000 nM EHDPP	222.6	224.5	212.4		
	Background	158.7	153.7	151.6		
Average intensity of	Control (DMSO)	1	$38.6 \pm 5$	.5		
CD71 (2 d; AU)	EHDPP (10, 000 nM)	) $141.4 \pm 20.8$		).8		

Table S16. Numeric data for Fig S1D, S1E, S2A, S2B, S2F (means  $\pm$  SDs), and S2D

TOM, trophoblast organoid medium; AU, arbitrary units; PD, pyruvate dehydrogenase complex; CD71, transferrin receptor

Experiments	Dose	Numeric data
	DMSO	$1.00\pm0.20$
Polative evenession of ICE1 (DNA level)	100 nM EHDPP	$0.98\pm0.12$
Relative expression of IGF1 (KINA level;	1,000 nM EHDPP	$0.86\pm0.19$
KO)	10,000 nM EHDPP	$0.91\pm0.22$
	DMSO	$1.00\pm0.33$
Polative expression of IGE2 (DNA level)	100 nM EHDPP	$1.84\pm0.40$
RUD	1,000 nM EHDPP	$2.11\pm0.24$
KO)	$\begin{array}{c c} & Dose & Nun \\ & DMSO & 1.0 \\ & 100 nM EHDPP & 0.9 \\ & 1,000 nM EHDPP & 0.9 \\ & 1,000 nM EHDPP & 0.9 \\ & 10,000 nM EHDPP & 0.9 \\ & 100 nM EHDPP & 0.9 \\ & 100 nM EHDPP & 0.9 \\ & 100 nM EHDPP & 1.8 \\ & 1,000 nM EHDPP & 1.4 \\ & 1,000 nM EHDPP & 1.4 \\ & 1,000 nM EHDPP & 1.4 \\ & 10,000 nM & 10,000 nM & 10 \\ & 10,000 nM & 10,0$	$1.47\pm0.21$
Average Intensity of Ki67 (2 d. AII)	Control (DMSO)	$204.6\pm12.3$
Average intensity of Ki07 (2 d, AO)	EHDPP (10, 000 nM)	$69.6\pm32.7$
Average Intensity of TP63 (2 d: AU)	Control (DMSO)	$113.7\pm12.5$
Average intensity of 11 05 (2 d, AO)	EHDPP (10, 000 nM)	$44.3\pm15.9$
Average Intensity of CD71 (2 d: AII)	Control (DMSO)	$126.1\pm5.0$
Average intensity of CD/1 (2 d, AO)	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$122.3\pm15.1$
Average Intensity of Ki67 (10 d. AII)	Control (DMSO)	$273.5\pm10.9$
Average intensity of Ki07 (10 d, AO)	Dose           DMSO           el;         100 nM EHDPP           1,000 nM EHDPP           100 nM EHDPP           1000 nM EHDPP           10,000 nM EHDPP           0         Control (DMSO)           EHDPP (10,000 nM)           0         Control (DMSO)           EHDPP (10,000 nM)           0         EHDPP (10,000 nM)           0         Control (DMSO)           EHDPP (10,000 nM)           0         EHDPP (10,000 nM)	$60.1\pm30.1$
Average Intensity of TP63 (10 d. AU)	Control (DMSO)	$142.6\pm7.13$
Average intensity of 1105 (10 d, AO)	EHDPP (10, 000 nM)	$81.3\pm21.4$
Average Intensity of CD71 (10 d: AU)	Control (DMSO)	$132.3\pm11.9$
Average intensity of CD/1 (10 d, AO)	EHDPP (10, 000 nM)	$62.2\pm17.2$
Average Intensity of HI A-G (10 d: AII)	Control (DMSO)	$62.2\pm17.2$
Average intensity of fille- O (10 d, AO)	EHDPP (10, 000 nM)	$41.6\pm10.7$

Table S17. Numeric data for Fig S3A, S3B, S3D, S3E, S3G, S3H, S3I, S3J, S3K (mean  $\pm$  SDs)

IGF1, insulin-like growth factor 1; IGF2, insulin-like growth factor 2; TP63, tumor protein 63; AU, arbitrary units; RU, relative unit; CD71, transferrin receptor; HLA-G, human leucocyte antigen protein-G

Experiments	Dose	]	Numeric data	a
Average Intensity of TPBPA	Control		$39.9\pm6.0$	
$(mean \pm SDs; AU)$	10 mg/kg EHDPP		$35.2\pm2.8$	
Average Intensity of TFAP2C	Control		$48.2\pm8.2$	
$(mean \pm SDs; AU)$	10 mg/kg EHDPP		$42.4\pm3.4$	
Average Intensity of HAND1	Control		$31.0\pm4.3$	
$(mean \pm SDs; AU)$	10 mg/kg EHDPP		$21.7\pm6.8$	
		1#	2#	3#
	Control	159.2	139.9	168.8
Placenta p-IGF1R (Gray value;	0.4 mg/kg EHDPP	148.4	130.5	160.8
Intden/Area; AU)	2 mg/kg EHDPP	138.2	130.1	160.5
	10 mg/kg EHDPP	145.8	132.9	159.6
	Background	108.6	120.4	147.9
		1#	2#	3#
	Control	184.6	137.7	183.8
Placenta IGF1R (Gray value;	0.4 mg/kg EHDPP	179.8	138.7	205.9
Intden/Area; AU)	2 mg/kg EHDPP	207.9	173.6	179.3
	10 mg/kg EHDPP	184.4	158.2	193.5
	Background	97.6	104.2	161.9
		1#	2#	3#
	Control	198.9	184.6	212.6
Placenta PD (Gray value;	0.4 mg/kg EHDPP	188.6	173.9	177.7
Intden/Area; AU)	2 mg/kg EHDPP	175.0	154.4	192.5
	10 mg/kg EHDPP	169.1	153.0	189.0
	Background	138.0	126.8	129.1
		1#	2#	3#
	Control	200.0	224.5	228.9
Placenta $\beta$ -actin (Gray value,	0.4 mg/kg EHDPP	188.6	223.0	225.9
with PD; Intden/Area; AU)	2 mg/kg EHDPP	192.1	218.7	221.3
	10 mg/kg EHDPP	191.5	224.6	223.1
	Background	127.5	113.3	160.9
		1#	2#	3#
	Control	204.7	145.6	178.1
Placenta cytochrome C (Gray	0.4 mg/kg EHDPP	193.9	139.9	176.3
value; Intden/Area; AU)	2 mg/kg EHDPP	171.8	144.2	175.2
	10 mg/kg EHDPP	169.0	135.9	174.9
	Background	141.8	125.8	163.8
		1#	2#	3#
	Control	197.6	225.6	226.9
Placenta p-actin (Gray value,	0.4 mg/kg EHDPP	196.8	223.0	225.1
Intden/Arca: AU	2 mg/kg EHDPP	195.3	226.7	223.0
much/Arca, AU)	10 mg/kg EHDPP	191.0	228.0	222.2
	Background	136.0	152.8	164.1

### Table S18. Numeric data for Fig S4B, S4C, S4D, S4E, S4G, S4H

TPBPA, trophoblast-specific protein  $\alpha$ ; AU, arbitrary units; TFAP2C, transcription factor AP-2 $\gamma$ ; HAND1, crest derivatives-expressed protein 1; p-IGF1R, phosphorylated-IGF1R; IGF1R, insulin-like growth factor 1 receptor; PD, pyruvate dehydrogenase complex

#### **Sequence of HA-GFP**

gacattgatt attgactagt tattaatagt aatcaattac ggggtcatta gttcatagcc catatatgga gttccgcgtt acataactta eggtaaatgg eeegeetgge tgacegeeca acgaceeeg eeeattgacg teaataatga egtatgttee eatagtaacg ccaataggga ctttccattg acgtcaatgg gtggagtatt tacggtaaac tgcccacttg gcagtacatc aagtgtatca tatgccaagt acgcccccta ttgacgtcaa tgacggtaaa tggcccgcct ggcattatgc ccagtacatg accttatggg acttteetae ttggeagtae atetaegtat tagteatege tattaeeatg gtgatgeggt tttggeagta eateaatggg cgtggatagc ggtttgactc acggggattt ccaagtctcc accccattga cgtcaatggg agtttgtttt ggcaccaaaa tcaacgggac tttccaaaat gtcgtaacaa ctccgcccca ttgacgcaaa tgggcggtag gcgtgtacgg tgggaggtct atataagcag agetetetgg etaactagag aacceaetge ttaetggett ategaaatta ataegaetea etatagggag acceaagetg getageatgt acceataega egtaceagat taegetagea agggagaaga actetttaet ggtgttgtee caattetggt tgagetggat ggtgatgtga atggecacaa attetetgtg tetggtgaag gtgaaggaga tgeaacttat ggaaagetga etetgaagtt eatttgtaea acaggaaage tgeeagtgee ttggeeaaet etggtgaeea eeetgaetta tggtgttcaa tgtttcagca ggtaccctga ccacatgaag cagcatgact tctttaaatc tgcaatgcca gaaggttatg ttcaggagag gacaatette tttaaggatg atggaaatta taagacaagg gcagaagtga agtttgaagg tgatacaetg gttaacagaa ttgagctgaa aggcattgat tttaaggaag atggaaacat tctgggtcac aagctggagt acaactataa tteteacaat gtttacatta tggcagataa gcagaagaat ggaattaagg ttaattteaa gattagacac aacattgagg atggatetgt ceaaetggea gaceattace ageagaacae ecetattggt gatggeeeag tteteeteee agataateae tateteegea eteaatetge tetgteeaaa gaecetaatg agaaaagaga ceacatggte eteetggagt ttgtgacage agcaggaatt actctgggaa tggatgagct gtacaaggga tccaaggaca acaccgtgcc cctgaagctg atcgccctgc tggccaacgg cgagttccac tctggcgagc agctgggaga gaccctggga atgagcagag ccgccatcaa caagcacatc cagacactga gagactgggg agtggacgtg ttcaccgtgc ctggcaaggg ctacagcctg cctgagccta tccagctgct gaacgecaag cagateetgg gacagetgga tggeggaage gtggeegtge tgeetgtgat egacteeace aateagtaee tgctggacag aatcggagag ctgaagtccg gcgacgcctg catcgccgag taccagcagg ctggcagagg aggcagagga eggaagtggt teageceatt eggagecaae etgtacetgt ceatgttetg gagaetggag eagggaeetg etgetgecat cggactgagt ctggtgatcg gaatcgtgat ggccgaggtg ctgagaaagc tgggagccga caaggtgaga gtgaagtggc ctaatgacct gtacctccag gaccgcaagc tggctggcat cctggtggag ctgacaggca agacaggcga tgccgctcag atcgtgatcg gagccggaat caacatggcc atgagaagag tggaggagag cgtggtgaac cagggctgga tcaccctgca ggaggctggc atcaacctgg accggaacac cctggccgcc atgctgatca gagagctgag agccgctctg gagctgttcg agcaggaggg actggcteet taeetgagea gatgggagaa getggacaae tteateaaca gaeetgtgaa getgateate ggcgacaagg aaatettegg cateteeaga ggaategaca ageagggage tetgetgetg gageaggaeg gaateateaa gccctggatg ggcggagaaa tctccctgag aagcgcagag aaggcttacc cttacgatgt accggattac gcataggcgg cctaagttta aaccgctgat cagcctcgac tgtgccttct agttgccagc catctgttgt ttgcccctcc cccgtgcctt cettgaceet ggaaggtgee acteecactg teettteeta ataaaatgag gaaattgeat egeattgtet gagtaggtgt cattctattc tggggggtgg ggtggggcag gacagcaagg gggaggattg ggaagacaat agcaggcatg ctggggatgc ggtgggctct atggcttctg aggcggaaag aaccagctgg ggctctaggg ggtatcccca cgcgccctgt agcggcgcat taagegegge gggtgtggtg gttaegegea gegtgaeege taeaettgee agegeeetag egeeegetee tttegettte tteeetteet ttetegeeae gttegeegge ttteeeegte aagetetaaa tegggggete eetttagggt teegatttag tgetttaegg cacetegace ecaaaaaaet tgattagggt gatggtteae gtagtgggee ategeeetga tagaeggttt ttcgcccttt gacgttggag tccacgttct ttaatagtgg actcttgttc caaactggaa caacactcaa ccctatctcg gtctattctt ttgatttata agggattttg ccgatttcgg cctattggtt aaaaaatgag ctgatttaac aaaaatttaa cgcgaattaa ttctgtggaa tgtgtgtcag ttagggtgtg gaaagteece aggeteecea geaggeagaa gtatgeaaag eatgeatete aattagteag caaccaggtg tggaaagtcc ccaggctccc cagcaggcag aagtatgcaa agcatgcatc tcaattagtc agcaaccata gteeegeeee taacteegee eateeegeee etaacteege eeagtteege eeatteteeg eeeeatgget gaetaatttt

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#### References

- Carignan CC, Mínguez-Alarcón L, Butt CM, Williams PL, Meeker JD, Stapleton HM, et al. 2017. Urinary concentrations of organophosphate flame retardant metabolites and pregnancy outcomes among women undergoing *in vitro* fertilization. Environ Health Perspect 125(8):087018, PMID: 288858831, <u>https://doi.org/10.1289/EHP1021</u>.
- Funk SP, Duffin L, He Y, McMullen C, Sun C, Utting N, et al. 2019. Assessment of impacts of diphenyl phosphate on groundwater and near-surface environments: Sorption and toxicity.
   J Contam Hydrol 221:50-57, PMID:30642690, https://doi.org/10.1016/j.jconhyd.2019.01.002.
- Gao D, Yang J, Bekele TG, Zhao S, Zhao H, Li J, et al. 2020. Organophosphate esters in human serum in Bohai Bay, North China. Environ Sci Pollut Res 27(3):2721-2729, PMID: 31836969, <u>https://doi.org/10.1007/s11356-019-07204-5</u>.
- He M, Jin K, Qiu S, Liao X, Zheng X, Chen Z, et al. 2021. The associations between organophosphate esters and urinary incontinence in the general us population. Environ Sci Pollut Res, online ahead of print, PMID: 34523086, <u>https://doi.org/10.1007/s11356-021-14153-5</u>
- Hou M, Shi Y, Jin Q, Cai Y. 2020. Organophosphate esters and their metabolites in paired human whole blood, serum, and urine as biomarkers of exposure. Environ Intl 39:105698, PMID: 32278199, <u>https://doi.org/10.1016/j.envint.2020.105698</u>.
- Liu R, Mabury SA. 2019. Organophosphite antioxidants in indoor dust represent an indirect source of organophosphate esters. Environ Sci Technol 53(4):1805-1811, PMID: 30657667, <u>http://doi.org/10.1021/acs.est.8b05545</u>.
- Sheridan MA, Fernando RC, Gardner L, Holinshead MS, Burton GJ, Moffett A, et al. 2020. Establishment and differentiation of long-term trophoblast organoid cultures from the human placenta. Nat Protoc 15(10):3441-3463, PMID: 32908314, https://doi.org/10.1038/s41596-020-0381-x.
- Stapleton HM, Klosterhaus S, Keller A, Ferguson PL, van Bergen S, Cooper E, et al. 2011. Identification of flame retardants in polyurethane foam collected from baby products. Environ Sci Technol 45(12):5323-31, PMID: 21591615, <u>http://doi.org/10.1021/es2007462</u>.
- Wang L, Jia Y, Kang Q, Song W, Hu J. 2020. Nontarget discovery of 11 aryl organophosphate triesters in house dust using high-resolution mass spectrometry. Environ Sci Technol 54(18):11376-11385, PMID: 32830962, <u>https://doi.org/10.1021/acs.est.0c01970</u>.
- Wang L, Kang Q, Jia Y, Li X, Hu J. 2021. Identification of three novel chloroalkyl organophosphate triesters in house dust using halogenation-guided nontarget screening combined with suspect. Environ Sci Technol 55(4):2482-2490 PMID: 33502167, <u>https://doi.org/ 10.1021/acs.est.0c07278</u>.