Supplementary Online Content

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This supplementary material has been provided by the authors to give readers additional information about their work.

eMethods

Data Selection

In order to find genome-wide association (GWA) summary statistics, we searched the published literature (Pubmed, SCOPUS), data repositories (dbGaP, GWAS ATLAS [https://atlas.ctglab.nl], GWAS catalogue [https://www.ebi.ac.uk/gwas], opentargets.org), and the Psychiatric Genomics Consortium (PGC) website (https://www.med.unc.edu/pgc/). We sought data from studies with the largest available European ancestry, due to the limited availability of well-powered GWA data from other ancestry groups.¹ Most of the selected summary data were produced via meta-analyses of large case-control samples by multi-institutional collaborative consortia; detailed descriptions of selection criteria, quality control, and analyses are described within the individual studies (see references in Table 1 and S. Table 1). We included 14 psychiatric phenotypes, including attention deficit hyperactivity disorder (ADHD; $N = 53,293)^2$, anorexia nervosa (AN; N =72,517)³, bipolar disorder (N = 51,710)⁴, the PGC cross-disorder phenotype (PGC-CD; N = 107,785)⁵, cannabis use disorder (CUD; N = 357,806)⁶, major depressive disorder (MDD; N = 500,199)⁷, obsessivecompulsive disorder (OCD; N = 9,725)⁸, opioid use disorder (OUD; N = 82,707)⁹, problematic alcohol use $(PAU; N = 435,563)^{10}$, post-traumatic stress disorder $(PTSD; N = 174,659)^{11}$, schizophrenia $(SZ; N = 100)^{11}$ $105,318)^{12}$, and Tourette's syndrome (TS; N = 14,307).¹³ We also included three continuous phenotypes: generalized anxiety disorder (N = 175, 163)¹⁴ was assessed based on the GAD-2 Score, ¹⁵ a 2-item self-report assessment of the frequency of worrying and related physical sensations;¹⁵ the personality trait of neuroticism (N = 168, 105);¹⁶ and another dispositional trait reflecting risk tolerance (RT; N = 315, 894).¹⁷ We selected 13 immune-related phenotypes, including allergic rhinitis (AR; N = 289,307),¹⁸ asthma (N = 385,822),¹⁸ atopic dermatitis (AD; N = 40,835),¹⁹ celiac disease (N = 15,283),²⁰ Crohn's disease (N = 40,266),²¹ hypothyroidism (HYPO; N = 244.890; primarily autoimmune-mediated in developed countries).¹⁸ primary biliary cholangitis (PBC; N = 13,239),²² primary sclerosing cholangitis (PSC; N = 14,890),²³ rheumatoid arthritis (RA; N =58,284),²⁴ systemic lupus erythematosus (SLE; N = 23,210),²⁵ type 1 diabetes (T1D; N = 26,890),²⁶ ulcerative

colitis (UC; N = 45,975),²¹ and vitiligo (N = 44,266).²⁷ Data for multiple sclerosis could not be shared at the time of our search.

We searched the literature using the following SCOPUS for articles identifying risk factors for psychiatric, allergic, autoimmune, and inflammatory disorders. We identified numerous risk factors that influence liability to both psychiatric and immune-related disorders, including age,^{28,29} sex,^{30,31} alcohol and tobacco consumption,^{32–37} cognitive processing,³⁸ diet,^{39–42} exercise,^{43,44} early life stress or trauma,^{45–48}, educational attainment,⁴⁹ infection and microbial dysbiosis,^{50–53} perinatal factors,^{54–59} obesity,^{60,61} sleep,^{62,63} socioeconomic status and neighborhood deprivation, ^{64–67} social connectedness, ⁶⁸ and toxic exposures. ^{69,70} Additionally, our review identified considerable evidence that chronic stress leads to dysregulation of autonomic and neuroendocrine signaling mechanisms, which can have direct effects on liability to psychiatric and immune-related disorders, but are additionally proposed to exert effects through changes in function of certain immune cell populations, chronic low-grade inflammation, gastrointestinal dysbiosis, and changes in gastrointestinal and vascular permitablity.^{71–75} As such, we sought to include phenotypes that capture aspects of the human stress response. We included resting heart rate,¹⁸ heart rate reactivity and recovery during exercise,⁷⁶ and heart rate variability,⁷⁶ all of which are influenced by cardiac autonomic signaling. We sought well-powered GWA data relevant to these phenotypes and ultimately selected 15 phenotypes, including body mass index (BMI; N = 806,834)⁷⁷, cigarettes per day (CPD; N = 263,954)⁷⁸, cognitive processing (CP; N = $(257,828)^{79}$, alcoholic drinks per week (DPW; $N = 537,349)^{78}$, educational attainment (EA; $N = 766,345)^{79}$, the frequency of moderate-intensity exercise (EXER; N = 367,908)¹⁸, annual income (N = 332,594)¹⁸, sleep duration (N = 446,118)⁸⁰, social deprivation (*via* the Townsend Index;⁸¹ SOCD; N = 420,035),⁸² and the frequency of social interactions (SOCI: N = 383.941).¹⁸ We also selected phenotypes that capture aspects of the human autonomic response to stress, including resting heart rate (HR; N = 361,411),¹⁸ HR increase during and recovery after exercise (HRI and HRR; N = 58,818),⁸³ heart rate variability (HRV; N = 28,122).⁸³ We sought to include cortisol-related phenotypes, 84,85 but this was precluded by low heritability (h^2) Z-statistics and relatively small sample size. In order to restrict the scope of the study and preserve statistical power, we made the somewhat arbitrary decision to exclude a number of well-powered GWA studies capturing a variety of dietary consumption phenotypes,^{86,87} and we also did not examine studies of gut microbiome composition and related metabolites.^{88,89} The final list of phenotypes is provided in S. Table 1.

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Genetic Correlations via LD-Score Regression

We removed SNPs with MAF < 1% based on the 1000 Genomes Phase reference panel for European populations and we excluded the major histocompatibility region. We used *LD-score regression* (LDSC)⁹⁰ with default settings and the HapMap3 reference panel to estimate SNP h^2 .

Mendelian Randomization

We briefly review the assumptions and limitations of MR: 1) The instrument SNPs are associated and have a causal effect on the exposure phenotype. For exposures with sparse genetic architecture, weak associations between instrument SNP and exposure are likely to bias toward null findings. For polygenic phenotypes, the inclusion of a larger set of lower confidence instrument SNPs could increase the likelihood of selecting instrument SNPs that have a pleiotropic mechanism relevant to both the exposure and outcome phenotype. One approach to cope with potential bias introduced by weak instruments and highly polygenic phenotypes is to perform MR with robust associated profile scores.⁹¹ 2) The exposure phenotype should share no common cause with the outcome phenotype (*i.e.*, there are no confounding factors). When this possibility is not excluded, multivariable MR can be used to reassess exposure-outcome relationships while adjusting for genetic associations of a third variable. 3) The instrument SNPs only exert effects on the outcome phenotype through the exposure phenotype, as opposed to shared biological mechanisms or mediation through some other phenotype. Outlier removal, heterogeneity testing, sensitivity analyses, and comparison across multiple MR methods can help evaluate this assumption. 4) Two-sample MR rests on the assumption of independent samples drawn from the same population and sample overlap can bias toward rejecting the null hypothesis, especially when there are weak instruments with overestimated effect sizes.

For the present study, the *TwoSampleMR* (TSMR)⁹² package was used to harmonize data and threshold and clump instrument SNPs ($p_1 < 5x10^{-8}$, $p_2 < 0.001$, window = $1x10^5$). For phenotypes denoted with \dagger in Table 1, this identified < 15 LD-independent SNPs, so we instead used a suggestive threshold of $p < 1x10^{-5}$. Bidirectional MR analyses were performed using inverse variance-weighted (IVW) method with significance set at FDR q < 0.05 and we verified findings with Egger-based models (reported in S. Tables). We performed single-SNP, heterogeneity, and leave-one-out (LOO) sensitivity analyses. We verified effects with the *mr.raps* package,⁹¹ which uses adjusted profile scores that are more robust to outliers in the genetic instrument and © 2022 America Medical Association. All rights reserved. estimates effects under assumptions of pervasive balanced pleiotropy, which can account for weak instrument bias. We preferentially report results after removal of potential outlier SNPs (|standardized residual| > 2.5 *s.d.* of the mean). We assessed the effects of adjustment for each of the third variables separately using the *MVMR* package with default settings.⁹³ To check for possible sample overlap among the MR effects, we examined LDSC covariance intercepts. For those with significant positive values, we repeated bidirectional IVW analyses using the *MRlap* package (default settings) to adjust for additional bias introduced by sample overlap.⁹⁴

Characterization of Loci

For six phenotype pairs with robust MR effects (*i.e.*, FDR q < 0.05 and consistent across sensitivity and MVMR-adjusted analyses), a two-sided meta-analysis was performed using the ASSETT package.⁹⁵ Because we encountered an unresolvable error when combining continuous and dichotomous phenotypes, the effect of RT on AR was not assessed. SNPs showing significant effects for only one of the phenotypes were removed from the analysis. Subsets of SNPs with concordant and discordant effects were separately advanced for enrichment analysis using FUMA's SNP2GENE function. The concordant subsets included information about the direction of the effects, while the latter subsets contained only the overall model p-values. FUMA analysis was performed with clumping setting of $r^2 < 0.1$, p-value thresholds corresponding to the threshold used to define the MR instrument (*i.e.*, 5x10⁻⁸ and in some cases 1x10⁻⁵) and the second threshold of 0.05, with the window size of 250kB, using the default settings for both positional and eQTL-based gene mapping. For celltype analysis, we included available human single-cell RNA-seq reference datasets.^{96–105} For cell types surviving multiple test correction (family-wise FDR q < 0.05), we examined the pair-wise conditional analyses within- (i.e. Step 2) and across-reference datasets (i.e. Step 3) to prioritize among potentially correlated cell types. High proportional significance (PS; *i.e.*, \geq 0.8) indicates complete independence, while lower values suggest partial or complete dependence on other correlated cell types. In S. Figure 5B, the numeral 2 denotes a cell type with > 0.8 in Step 2, while 2' denotes the cell type with the highest PS despite being < 0.8. A similar relationship applies between numerals 3 and 3' with respect to Step 3 conditioning PS.

eDiscussion

The main findings of our study supported modest positive effects of psychiatric on immune liability, where the relevant loci were primarily enriched with brain tissues and cell types, but also contained signals for © 2022 America Medical Association. All rights reserved. peripheral leukocytes and lymphoid tissue. These loci were also enriched in signal for various behavioral traits, immune cell counts, and biological terms including cell adhesion, calcium-binding, and response to viral infection. Below, we provide a literature search of individual genes implicating these biological terms. When we considered the six robust positive MR effects (*i.e.*, FDR *q* < 0.05 and consistent across sensitivity and MVMR-adjusted analyses), 54 psychiatric instrument loci were independent of third variable effects. These were significantly enriched (FDR *q* < 0.05) for several annotations, including cellular adhesion (comprised of genes *ATXN1*, *ADD3*, and the protocadherin- α family within a locus in 5q31.3; S. Table 5).

For *ATXN1*, the relevant protein participates in complex interactions to regulate transcription and splicing, though its exact roles are not well understood. Our review identified considerable mechanistic evidence linking polyglutamine expansion mutations in this gene (and others) with various neurodegenerative disorders, including spinocerebellar ataxias and amyotrophic lateralsclerosis,^{106–108} but its less clear how it may relate to the penetrance of autosomal dominant Alzheimer's phenotypes.¹⁰⁹ This gene was identified in bioinformatic analyses based on enrichmed protein-protein interactions with 19 candidate genes for SZ.¹¹⁰ Additionally, *ATXN1* appears to have relevance to immune-related diseases, as loss of function is associated with more severe disease in a mouse model of multiple sclerosis, and this was mediated by effects on a potentially pathogenic B-cell subpopulation.^{111,112} It plays a role in extracellular matrix remolding during lung development and is down-regulated in a model of asthma.^{113,114} Additionally, it was identified in prior pharmacogenetic GWAS of IBD treatment response.¹¹⁵ GWAS studies support associations of ATXN1 with cognitive function, intelligence, and educational attainment.^{116,117}

ADD3 encodes γ-adducins, which are ubiquitously expressed, heteromeric proteins that participate in membrane-associated spectrin-actin cytoskeletal networks and interact with calmodulin.¹¹⁸ Homozygous loss-of-function mutations within a consanguineous family were associated with inherited cerebral palsy.¹¹² Loss-of-function mutations have also been associated with renal podocyte dysfunction and related chronic kidney disease.¹¹⁹ GWAS data suggest this gene may be relatively specific to bipolar disorder¹²⁰ and it was identified in an earlier study comparing bipolar I and II.¹²¹ There is literature linking γ-adducins to electrophysiological plasticity induced by environmental stimuli (*e.g.*, effects of cocaine exposure on striatal neurons) and learning behavior (e.g., Morris water maze).^{122,123} Another line of research associated gamma-adducin expression with blood pressure homeostasis and with cerebrovascular and blood-brain-barrier dysfunction.^{119,124} A GWAS © 2022 America Medical Association. All rights reserved.

study in East Asian ancestry identified *ADD3* in association with neonatal biliary atresia.¹²⁵ *ADD3* was identified among differentially expressed genes that discriminated intestinal biopsies from Celiac-affected individuals and subsets of first degree relatives that lacked serological evidence of disease activity.¹²⁶ Furthermore, ADD3 was down-regulated in a mouse model of spontaneous colitis caused by macrophage-restricted knockout of the *IL10RA* gene.¹²⁷ ADD3 was also identified among CD4+ T cell genes whose expression discriminated high-and low-atopy subtypes of asthma and predicted differences in eosinophil counts and IgE levels.¹²⁸

Protocadherins are cell adhesion molecules that undergo alternative splicing and combinatorial heteromerization to allow specificity of cell recognition, though their exact roles in the developing and adult CNS are not well understood yet.^{129–131} Multiple subtypes of protocadherins have been identified, and members of the clustered α -protocadherin family have been associated with psychiatric and neurodevelopmental phenotypes.¹³⁰ Deletion of this family was associated with reduced post-injury axon outgrowth, myelination, and expression of *BDNF* in rodent model.¹³² Deletion of this family also impacted cortical neuron migration¹³³ and serotonergic fiber development and maturation.^{134,135} One study found α -protocadherins were upregulated during stimulation of Th₂ lymphocytes.¹³⁶ We also found relevant associations with other types of protocadherins. *PCDH1* plays a well-characterized role in asthma pathophysiology and is important in the response to glucocorticioids.^{137,138} Roles in GI epithelial structure and function have been observed for members of the other protocadherin families.¹³⁹

We also saw enrichment for genes involved in calcium-binding (including *LTBP2*, *PLCB2*, and *MATN4*). *LTBP2* encodes a latent transforming growth factor (TGF) beta binding protein, which is found in association with fibrillin in the extracellular matrix and thought to help limit the availability of TGF-β.¹⁴⁰ TGF-β plays a role in regulating inflammation and promoting airway and extracellular matrix remodeling in asthma.¹⁴⁵ Both TGF-β and *LTBP2* genes play roles in inflammation-induced fibrosis in other organs, including the intestines.^{146,147} Mutations of *LTBP2* are associated with congenital ophthalmic disorders,¹⁴¹¹⁴² but there is less evidence linking it with psychiatric phenotypes. Research in pulmonary development identifies roles for several members of the LTBP gene family.¹⁴³ Secreted protein product of *LTBP2* was identified as a biomarker of idiopathic pulmonary fibrosis.¹⁴⁴ Interestingly, *LTBP2* was identified among genes differentially expressed after antibiotic treatment designed to alter the gut microbiome.¹⁴⁸ Research in oncology identifies a role for *LTBP2* in tumor/metastasis suppression for several malignancies.^{149,150} *PLCB2* encodes phospholipase C beta-2, an enzyme that © 2022 America Medical Association. All rights reserved.

participates in G-protein-coupled signaling pathways relevant to taste receptors and regulation of platelet response.^{151,152} This gene was identified in secondary analyses of SZ GWAS data.¹⁵³ It was also found among down-regulated genes in the amygdala of rats subjected to chronic stress.¹⁵⁴ This gene also participates in pathways that influence differentiation, activation, and chemotaxis of multiple immune cells.^{155,156} One study identified a role in negative regulation of inflammatory signaling in a model of viral infection.¹⁶⁷ It was also among over-expressed genes in childhood asthma.¹⁵⁸ *MATN4* encodes matrilin-4, a member of the von Willebrand factor A domain-containing protein family, which are involved in formation of filamentous extracellular matrix-associated in various tissues. Mutations in this gene have been found in neurogenetic syndromes within consanguineous families.¹⁵⁹ The gene was also identified among those differentially methylated in children in relation to history of early life adversity and adults in relation to air pollution.^{160,161} Matrilin-4 has been shown experimentally to regulate stress-induced hematopoietic stem cell proliferation.¹⁶²

We also observed enrichment among genes downregulated in a fibroblast model of human cytomegalovirus infection, including *AKT3*, *ADD3*, *CHMP2A*, *TMCO6*, *REV3L*, *TRAF3IP2*, and *TNKS*. *AKT3* encodes a serine/threonine kinase that regulates signaling in response to growth factors like platelet-derived growth factor and insulin-like growth factor 1. In humans, mutations in *AKT3* are associated with profound neurodevelopmental phenotypes¹⁶³ and experimental models also suggest roles in behavioral traits.^{164–167} In T-regulatory cells, *AKT3* inhibits interferon-γ, and loss of this function may promote loss of this cell population's suppressive function.¹⁶⁸ Loss of function is also associated with more severe phenotypes in models of multiple sclerosis.^{169,170} *AKT3* was investigated as a potential susceptibility gene in an association analysis between Danish and Genetics of Asthma International Network families, but associations were non-significant.¹⁷¹

CHMP2A encodes a protein that functions within the endosomal sorting systems and within exosomes. Loss of function mutations are thought to contribute to proteostatic stress that causes neurodegenerative phenotypes.¹⁷² *TMCO*6 is a transmembrane protein whose function is not well characterized, but loss-offunction mutations are associated with a subtype of mitochondrial complex I deficiency. Within bipolar-affected and unaffected individuals, this gene was near a locus associated with soluble CD14 levels (a non-specific marker of monocyte activation) within cerebrospinal fluid samples.¹⁷³

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REV3L encodes a catalytic subunit of DNA polymerase zeta, which is found in the mitochondria and plays a role in the response to DNA damage. Numerous studies have examined its role in malignancies,^{174,175} including cervical cancer,¹⁷⁴ chemoresistant squamous cell carcinoma of the head and neck,^{176,177} and non-small cell lung cancer.¹⁷⁸ Defects in this gene are associated with genotoxic stress and interferon activation,¹⁷⁹ and are disruptive to mammalian embryonic development.^{180–182} SNPs in intergenic regions including *REV3L* and *TRAF3IP2* were associated with rheumatoid arthritis in a large-scale study of black South Africans.¹⁸³

TRAF3IP2 encodes an adaptor protein that plays a central role in innate immunity response to pathogens, inflammation, and stress via regulating responses of the Rel/NF-kappaB transcription factor family members in their response to cytokines.¹⁸⁴ TRAF3IP2 facilitates pro-inflammatory IL-17 signaling,^{185–187} which has been implicated in endothelial dysfunction and cardiovascular disease,^{188–191} as well as obesity-related vascular insulin resistance.¹⁹² GWA studies have identified a locus in *TRAF3IP2* with common variants affecting susceptibility to psoriasis and psoriatic arthritis,¹⁹⁴¹⁹⁵ as well as an association with cutaneous manifestations of inflammatory bowel disease,¹⁹⁶ and susceptibility to mucocutaneous adverse reactions among patients treated with Nevirapine.¹⁹⁷ TNKS encodes a poly-ADP-ribosyltransferase involved in several processes, including activation of Wnt/beta-catenin signaling pathway,^{198,199} as well as in regulating telomere length²⁰⁰ and vesicle trafficking.²⁰¹ It's been studied for its role in malignancy,^{202,203} herpesvirus infection,^{204,205} and obesity.²⁰⁶

Among the 54 loci involved in six robust psychiatric-immune MR effects (described in main text and S. Table 4), we observed nominal enrichment (uncorrected p < 0.05) for multiple immunologic signatures (*e.g.,* genes down-regulated by IFN-gamma in microglia, $p = 1.3 \times 10^{-4}$, targets of several transcription factors (*e.g.,* ELK1 $p = 1.5 \times 10^{-4}$; E2A p = 0.01; STAT1 p = 0.01), and transcripts upregulated by suppression of JAK2 (p = 0.01; S. Table 5). Overall, these findings support the idea that genes with pleiotropic effects potentially relevant to both psychiatric and immune-related disorders were identified in association with some of the loci contributing to significant MR effects.

eReferences

 G S, SM W, SA T. The Missing Diversity in Human Genetic Studies. *Cell*. 2019;177(1):26-31. doi:10.1016/J.CELL.2019.02.048

Demontis D, Walters RK, Martin J, et al. Discovery of the first genome-wide significant risk loci for
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attention deficit/hyperactivity disorder. Nat Genet. 2019;51(1):63-75. doi:10.1038/s41588-018-0269-7

- Watson HJ, Yilmaz Z, Thornton LM, et al. Genome-wide association study identifies eight risk loci and implicates metabo-psychiatric origins for anorexia nervosa. *Nat Genet*. 2019;51(8):1207-1214. doi:10.1038/s41588-019-0439-2
- 4. Stahl EA, Breen G, Forstner AJ, et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nat Genet*. 2019;51(5):793-803. doi:10.1038/s41588-019-0397-8
- Lee PH, Anttila V, Won H, et al. Genomic Relationships, Novel Loci, and Pleiotropic Mechanisms across Eight Psychiatric Disorders. *Cell*. 2019;179(7):1469-1482.e11. doi:10.1016/j.cell.2019.11.020
- Johnson EC, Demontis D, Thorgeirsson TE, et al. A large-scale genome-wide association study metaanalysis of cannabis use disorder. *Lancet Psychiatry*. 2020;7(12):1032-1045. doi:10.1016/s2215-0366(20)30339-4
- Howard DM, Adams MJ, Clarke TK, et al. Genome-wide meta-analysis of depression identifies 102 independent variants and highlights the importance of the prefrontal brain regions. *Nat Neurosci*. 2019;22(3):343-352. doi:10.1038/s41593-018-0326-7
- Arnold PD, Askland KD, Barlassina C, et al. Revealing the complex genetic architecture of obsessive– compulsive disorder using meta-analysis. *Mol Psychiatry*. Published online August 1, 2017. doi:10.1038/mp.2017.154
- Zhou H, Rentsch CT, Cheng Z, et al. Association of OPRM1 Functional Coding Variant With Opioid Use Disorder: A Genome-Wide Association Study. *JAMA Psychiatry*. 2020;77(10):1072-1080. doi:10.1001/jamapsychiatry.2020.1206
- Zhou H, Sealock JM, Sanchez-Roige S, et al. Genome-wide meta-analysis of problematic alcohol use in 435,563 individuals yields insights into biology and relationships with other traits. *Nat Neurosci*. 2020;23(7):809-818. doi:10.1038/s41593-020-0643-5
- Nievergelt CM, Maihofer AX, Klengel T, et al. International meta-analysis of PTSD genome-wide association studies identifies sex- and ancestry-specific genetic risk loci. *Nat Commun*. 2019;10(1):4558. doi:10.1038/s41467-019-12576-w
- 12. Pardiñas AF, Holmans P, Pocklington AJ, et al. Common schizophrenia alleles are enriched in mutationintolerant genes and in regions under strong background selection. *Nat Genet*. 2018;50(3):381-389.
- © 2022 America Medical Association. All rights reserved.

doi:10.1038/s41588-018-0059-2

- Yu D, Sul JH, Tsetsos F, et al. Interrogating the Genetic Determinants of Tourette's Syndrome and Other Tic Disorders Through Genome-Wide Association Studies. *Am J Psychiatry*. 2019;176(3):217-227. doi:10.1176/appi.ajp.2018.18070857
- Levey DF, Gelernter J, Polimanti R, et al. Reproducible Genetic Risk Loci for Anxiety: Results From ~200,000 Participants in the Million Veteran Program. *Am J Psychiatry*. 2020;177(3):223-232. doi:10.1176/appi.ajp.2019.19030256
- Kroenke K, Spitzer RL, Williams JBW, Löwe B. The Patient Health Questionnaire Somatic, Anxiety, and Depressive Symptom Scales: a systematic review. *Gen Hosp Psychiatry*. 2010;32(4):345-359. doi:10.1016/J.GENHOSPPSYCH.2010.03.006
- 16. Turley P, Walters RK, Maghzian O, et al. Multi-trait analysis of genome-wide association summary statistics using MTAG. *Nat Genet*. 2018;50(2):229-237. doi:10.1038/s41588-017-0009-4
- Karlsson Linnér R, Biroli P, Kong E, et al. Genome-wide association analyses of risk tolerance and risky behaviors in over 1 million individuals identify hundreds of loci and shared genetic influences. *Nat Genet*. 2019;51(2):245-257. doi:10.1038/s41588-018-0309-3
- Watanabe K, Stringer S, Frei O, et al. A global overview of pleiotropy and genetic architecture in complex traits. *Nat Genet*. 2019;51(9):1339-1348. doi:10.1038/s41588-019-0481-0
- Paternoster L, Standl M, Waage J, et al. Multi-ancestry genome-wide association study of 21,000 cases and 95,000 controls identifies new risk loci for atopic dermatitis. *Nat Genet*. 2015;47(12):1449-1456. doi:10.1038/ng.3424
- 20. Dubois PCA, Trynka G, Franke L, et al. Multiple common variants for celiac disease influencing immune gene expression. *Nat Genet*. 2010;42(4):295-302. doi:10.1038/ng.543
- de Lange KM, Moutsianas L, Lee JC, et al. Genome-wide association study implicates immune activation of multiple integrin genes in inflammatory bowel disease. *Nat Genet*. 2017;49(2):256-261. doi:10.1038/ng.3760
- Cordell HJ, Han Y, Mells GF, et al. International genome-wide meta-analysis identifies new primary biliary cirrhosis risk loci and targetable pathogenic pathways. *Nat Commun*. 2015;6:8019. doi:10.1038/ncomms9019
- © 2022 America Medical Association. All rights reserved.

- Ji SG, Juran BD, Mucha S, et al. Genome-wide association study of primary sclerosing cholangitis identifies new risk loci and quantifies the genetic relationship with inflammatory bowel disease. *Nat Genet*. 2017;49(2):269-273. doi:10.1038/ng.3745
- 24. Okada Y, Wu D, Trynka G, et al. Genetics of rheumatoid arthritis contributes to biology and drug discovery. *Nature*. 2014;506(7488):376-381. doi:10.1038/nature12873
- 25. Bentham J, Morris DL, Cunninghame Graham DS, et al. Genetic association analyses implicate aberrant regulation of innate and adaptive immunity genes in the pathogenesis of systemic lupus erythematosus. *Nat Genet*. 2015;47(12):1457-1464. doi:10.1038/ng.3434
- 26. Bradfield JP, Qu HQ, Wang K, et al. A genome-wide meta-analysis of six type 1 diabetes cohorts identifies multiple associated loci. *PLoS Genet*. 2011;7(9). doi:10.1371/journal.pgen.1002293
- Jin Y, Andersen G, Yorgov D, et al. Genome-wide association studies of autoimmune vitiligo identify 23 new risk loci and highlight key pathways and regulatory variants. *Nat Genet*. 2016;48(11):1418-1424. doi:10.1038/ng.3680
- Solmi M, Radua J, Olivola M, et al. Age at onset of mental disorders worldwide: large-scale metaanalysis of 192 epidemiological studies. *Mol Psychiatry 2021*. 2021;17:1-15. doi:10.1038/s41380-021-01161-7
- 29. Amador-Patarroyo MJ, Rodriguez-Rodriguez A, Montoya-Ortiz G. How Does Age at Onset Influence the Outcome of Autoimmune Diseases? *Autoimmune Dis*. 2012;2012(1). doi:10.1155/2012/251730
- Gleicher N, Barad DH. Gender as risk factor for autoimmune diseases. *J Autoimmun*. 2007;28(1):1-6.
 doi:10.1016/J.JAUT.2006.12.004
- 31. Riecher-Rössler A. Sex and gender differences in mental disorders. *The Lancet Psychiatry*. 2017;4(1):89. doi:10.1016/S2215-0366(16)30348-0
- Mahid SS, Minor KS, Soto RE, Hornung CA, Galandiuk S. Smoking and Inflammatory Bowel Disease: A Meta-analysis. *Mayo Clin Proc.* 2006;81(11):1462-1471. doi:10.4065/81.11.1462
- 33. Saulyte J, Regueira C, Montes-Martínez A, Khudyakov P, Takkouche B. Active or passive exposure to tobacco smoking and allergic rhinitis, allergic dermatitis, and food allergy in adults and children: a systematic review and meta-analysis. *PLoS Med.* 2014;11(3). doi:10.1371/JOURNAL.PMED.1001611
- 34. Prochaska JJ, Das S, Young-Wolff KC. Smoking, Mental Illness, and Public Health. Annu Rev Public

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Health. 2017;38:165. doi:10.1146/ANNUREV-PUBLHEALTH-031816-044618

- Yuan S, Yao H, Larsson SC. Associations of cigarette smoking with psychiatric disorders: evidence from a two-sample Mendelian randomization study. *Sci Reports 2020 101*. 2020;10(1):1-9. doi:10.1038/s41598-020-70458-4
- Jiang X, Zhu Z, Manouchehrinia A, Olsson T, Alfredsson L, Kockum I. Alcohol Consumption and Risk of Common Autoimmune Inflammatory Diseases—Evidence From a Large-Scale Genetic Analysis Totaling 1 Million Individuals. *Front Genet*. 2021;12. doi:10.3389/FGENE.2021.687745
- Palzes VA, Parthasarathy S, Chi FW, et al. Associations Between Psychiatric Disorders and Alcohol Consumption Levels in an Adult Primary Care Population. *Alcohol Clin Exp Res*. 2020;44(12):2536. doi:10.1111/ACER.14477
- McTeague LM, Goodkind MS, Etkin A. Transdiagnostic Impairment of Cognitive Control in Mental Illness. *J Psychiatr Res*. 2016;83:37. doi:10.1016/J.JPSYCHIRES.2016.08.001
- Manzel A, Muller DN, Hafler DA, Erdman SE, Linker RA, Kleinewietfeld M. Role of "Western Diet" in Inflammatory Autoimmune Diseases. *Curr Allergy Asthma Rep*. 2014;14(1):404. doi:10.1007/S11882-013-0404-6
- 40. Li Y, Lv MR, Wei YJ, et al. Dietary patterns and depression risk: A meta-analysis. *Psychiatry Res*. 2017;253:373-382. doi:10.1016/J.PSYCHRES.2017.04.020
- Firth J, Marx W, Dash S, et al. The Effects of Dietary Improvement on Symptoms of Depression and Anxiety: A Meta-Analysis of Randomized Controlled Trials. *Psychosom Med*. 2019;81(3):265. doi:10.1097/PSY.000000000000673
- Probst Y, Mowbray E, Svensen E, Thompson K. A Systematic Review of the Impact of Dietary Sodium on Autoimmunity and Inflammation Related to Multiple Sclerosis. *Adv Nutr.* 2019;10(5):902-910. doi:10.1093/ADVANCES/NMZ032
- 43. Sharif K, Watad A, Bragazzi NL, Lichtbroun M, Amital H, Shoenfeld Y. Physical activity and autoimmune diseases: Get moving and manage the disease. *Autoimmun Rev.* 2018;17(1):53-72.
 doi:10.1016/J.AUTREV.2017.11.010
- 44. Firth J, Solmi M, Wootton RE, et al. A meta-review of "lifestyle psychiatry": the role of exercise, smoking, diet and sleep in the prevention and treatment of mental disorders. *World Psychiatry*. 2020;19(3):360-
- © 2022 America Medical Association. All rights reserved.

380. doi:10.1002/WPS.20773

- LeMoult J, Humphreys KL, Tracy A, Hoffmeister JA, Ip E, Gotlib IH. Meta-analysis: Exposure to Early Life Stress and Risk for Depression in Childhood and Adolescence. *J Am Acad Child Adolesc Psychiatry*. 2020;59(7):842-855. doi:10.1016/J.JAAC.2019.10.011
- 46. Nelson CA, Scott RD, Bhutta ZA, Harris NB, Danese A, Samara M. Adversity in childhood is linked to mental and physical health throughout life. *BMJ*. 2020;371. doi:10.1136/BMJ.M3048
- 47. Copeland WE, Shanahan L, Hinesley J, et al. Association of Childhood Trauma Exposure With Adult Psychiatric Disorders and Functional Outcomes. *JAMA Netw Open*. 2018;1(7):e184493-e184493. doi:10.1001/JAMANETWORKOPEN.2018.4493
- Dube SR, Fairweather D, Pearson WS, Felitti VJ, Anda RF, Croft JB. Cumulative childhood stress and autoimmune diseases in adults. *Psychosom Med*. 2009;71(2):243-250. doi:10.1097/PSY.0b013e3181907888
- 49. Erickson J, El-Gabalawy R, Palitsky D, et al. EDUCATIONAL ATTAINMENT AS A PROTECTIVE FACTOR FOR PSYCHIATRIC DISORDERS: FINDINGS FROM A NATIONALLY REPRESENTATIVE LONGITUDINAL STUDY. *Depress Anxiety*. 2016;33(11):1013-1022. doi:10.1002/DA.22515
- Nikolova VL, Hall MRB, Hall LJ, Cleare AJ, Stone JM, Young AH. Perturbations in Gut Microbiota Composition in Psychiatric Disorders: A Review and Meta-analysis. *JAMA psychiatry*. 2021;78(12):1343-1354. doi:10.1001/JAMAPSYCHIATRY.2021.2573
- 51. Wucherpfennig KW. Mechanisms for the induction of autoimmunity by infectious agents. *J Clin Invest*.
 2001;108(8):1097. doi:10.1172/JCI14235
- Volkova A, Ruggles K V. Predictive Metagenomic Analysis of Autoimmune Disease Identifies Robust Autoimmunity and Disease Specific Microbial Signatures. *Front Microbiol*. 2021;12. doi:10.3389/FMICB.2021.621310/FULL
- 53. Khandaker GM, Zimbron J, Dalman C, Lewis G, Jones PB. Childhood infection and adult schizophrenia: A meta-analysis of population-based studies. *Schizophr Res*. 2012;139(1-3):161-168. doi:10.1016/J.SCHRES.2012.05.023
- 54. Froehlich-Santino W, Londono Tobon A, Cleveland S, et al. Prenatal and perinatal risk factors in a twin study of autism spectrum disorders. *J Psychiatr Res*. 2014;54(1):100-108.
- © 2022 America Medical Association. All rights reserved.

doi:10.1016/j.jpsychires.2014.03.019

- 55. Davies C, Segre G, Estradé A, et al. Prenatal and perinatal risk and protective factors for psychosis: a systematic review and meta-analysis. *The lancet Psychiatry*. 2020;7(5):399-410. doi:10.1016/S2215-0366(20)30057-2
- Gardener H, Munger KL, Chitnis T, Michels KB, Spiegelman D, Ascherio A. Prenatal and perinatal factors and risk of multiple sclerosis. *Epidemiology*. 2009;20(4):611-618. doi:10.1097/EDE.0B013E31819ED4B9
- 57. Räisänen L, Viljakainen H, Sarkkola C, Kolho KL. Perinatal risk factors for pediatric onset type 1 diabetes, autoimmune thyroiditis, juvenile idiopathic arthritis, and inflammatory bowel diseases. *Eur J Pediatr*. 2021;180(7):2115-2123. doi:10.1007/S00431-021-03987-3
- Metsälä J, Kilkkinen A, Kaila M, et al. Perinatal Factors and the Risk of Asthma in Childhood—A Population-based Register Study in Finland. *Am J Epidemiol*. 2008;168(2):170-178. doi:10.1093/AJE/KWN105
- 59. Kolevzon A, Gross R, Reichenberg A. Prenatal and perinatal risk factors for autism: a review and integration of findings. *Arch Pediatr Adolesc Med*. 2007;161(4):326-333. doi:10.1001/archpedi.161.4.326
- 60. Versini M, Jeandel PY, Rosenthal E, Shoenfeld Y. Obesity in autoimmune diseases: not a passive bystander. *Autoimmun Rev.* 2014;13(9):981-1000. doi:10.1016/J.AUTREV.2014.07.001
- 61. Simon GE, Von Korff M, Saunders K, et al. Association Between Obesity and Psychiatric Disorders in the US Adult Population. *Arch Gen Psychiatry*. 2006;63(7):824-830. doi:10.1001/ARCHPSYC.63.7.824
- 62. Li L, Wu C, Gan Y, Qu X, Lu Z. Insomnia and the risk of depression: A meta-analysis of prospective cohort studies. *BMC Psychiatry*. 2016;16(1):1-16. doi:10.1186/S12888-016-1075-3/FIGURES/4
- Hsiao YH, Chen YT, Tseng CM, et al. Sleep Disorders and Increased Risk of Autoimmune Diseases in Individuals without Sleep Apnea. *Sleep*. 2015;38(4):581. doi:10.5665/SLEEP.4574
- Calixto OJ, Anaya JM. Socioeconomic status. The relationship with health and autoimmune diseases.
 Autoimmun Rev. 2014;13(6):641-654. doi:10.1016/J.AUTREV.2013.12.002
- Li X, Sundquist J, Hamano T, Sundquist K. Neighborhood Deprivation and Risks of Autoimmune Disorders: A National Cohort Study in Sweden. *Int J Environ Res Public Health*. 2019;16(20). doi:10.3390/IJERPH16203798
- © 2022 America Medical Association. All rights reserved.

- Fone D, White J, Farewell D, et al. Effect of neighbourhood deprivation and social cohesion on mental health inequality: a multilevel population-based longitudinal study. *Psychol Med*. 2014;44(11):2449-2460. doi:10.1017/S0033291713003255
- 67. McElroy E, McIntyre JC, Bentall RP, et al. Mental Health, Deprivation, and the Neighborhood Social Environment: A Network Analysis: *https://doi.org/101177/2167702619830640*. 2019;7(4):719-734. doi:10.1177/2167702619830640
- Saeri AK, Cruwys T, Barlow FK, Stronge S, Sibley CG. Social connectedness improves public mental health: Investigating bidirectional relationships in the New Zealand attitudes and values survey. *Aust N Z J Psychiatry*. 2018;52(4):365-374. doi:10.1177/0004867417723990
- Nilsen FM, Frank J, Tulve NS. A Systematic Review and Meta-Analysis Investigating the Relationship between Exposures to Chemical and Non-Chemical Stressors during Prenatal Development and Childhood Externalizing Behaviors. *Int J Environ Res Public Health*. 2020;17(7). doi:10.3390/IJERPH17072361
- Khan A, Plana-Ripoll O, Antonsen S, et al. Environmental pollution is associated with increased risk of psychiatric disorders in the US and Denmark. *PLOS Biol*. 2019;17(8):e3000353. doi:10.1371/JOURNAL.PBIO.3000353
- 71. Liu YZ, Wang YX, Jiang CL. Inflammation: The Common Pathway of Stress-Related Diseases. *Front Hum Neurosci*. 2017;11. doi:10.3389/FNHUM.2017.00316
- McEwen BS. Neurobiological and Systemic Effects of Chronic Stress. *Chronic Stress*. 2017;1. doi:10.1177/2470547017692328
- 73. Dhabhar FS. Effects of stress on immune function: the good, the bad, and the beautiful. *Immunol Res*.
 2014;58(2-3):193-210. doi:10.1007/S12026-014-8517-0
- 74. Gong Y, Niu W, Tang Y, et al. Aggravated mucosal and immune damage in a mouse model of ulcerative colitis with stress. *Exp Ther Med*. 2019;17(3). doi:10.3892/ETM.2019.7162
- Amano H, Negishi I, Akiyama H, Ishikawa O. Psychological Stress can Trigger Atopic Dermatitis in NC/Nga Mice: An Inhibitory Effect of Corticotropin-Releasing Factor. *Neuropsychopharmacol 2008 333*. 2007;33(3):566-573. doi:10.1038/sj.npp.1301435
- 76. Verweij N, van de Vegte YJ, van der Harst P. Genetic study links components of the autonomous
- © 2022 America Medical Association. All rights reserved.

nervous system to heart-rate profile during exercise. *Nat Commun*. 2018;9(1):898. doi:10.1038/s41467-018-03395-6

- 77. Pulit SL, Stoneman C, Morris AP, et al. Meta-analysis of genome-wide association studies for body fat distribution in 694 649 individuals of European ancestry. *Hum Mol Genet*. 2019;28(1):166-174. doi:10.1093/hmg/ddy327
- 78. Liu M, Jiang Y, Wedow R, et al. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. *Nat Genet*. 2019;51(2):237-244. doi:10.1038/s41588-018-0307-5
- Lee JJ, Wedow R, Okbay A, et al. Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. *Nat Genet*. 2018;50(8):1112-1121. doi:10.1038/s41588-018-0147-3
- Dashti HS, Jones SE, Wood AR, et al. Genome-wide association study identifies genetic loci for selfreported habitual sleep duration supported by accelerometer-derived estimates. *Nat Commun*. 2019;10(1):1100. doi:10.1038/s41467-019-08917-4
- 81. Townsend P. Deprivation. *J Soc Policy*. Published online 1987:125-146.
- 82. Pan-UKB_Team. Pan-UKB. Published online 2020. https://pan.ukbb.broadinstitute.org
- 83. Nolte IM, Munoz ML, Tragante V, et al. Genetic loci associated with heart rate variability and their effects on cardiac disease risk. *Nat Commun.* 2017;8:15805. doi:10.1038/ncomms15805
- 84. Bolton JL, Hayward C, Direk N, et al. Genome wide association identifies common variants at the SERPINA6/SERPINA1 locus influencing plasma cortisol and corticosteroid binding globulin. *PLoS Genet*. 2014;10(7):e1004474. doi:10.1371/journal.pgen.1004474
- Neumann A, Direk N, Crawford AA, et al. The low single nucleotide polymorphism heritability of plasma and saliva cortisol levels. *Psychoneuroendocrinology*. 2017;85:88-95. doi:10.1016/j.psyneuen.2017.08.011
- Meddens SFW, de Vlaming R, Bowers P, et al. Genomic analysis of diet composition finds novel loci and associations with health and lifestyle. *Mol Psychiatry 2020 266*. 2020;26(6):2056-2069. doi:10.1038/s41380-020-0697-5
- 87. Kho M, Smith JA, Verweij N, et al. Genome-Wide Association Meta-Analysis of Individuals of European © 2022 America Medical Association. All rights reserved.

Ancestry Identifies Suggestive Loci for Sodium Intake, Potassium Intake, and Their Ratio Measured from 24-Hour or Half-Day Urine Samples. *J Nutr*. 2020;150(10):2635-2645. doi:10.1093/JN/NXAA241

- 88. Awany D, Allali I, Dalvie S, et al. Host and microbiome genome-wide association studies: Current state and challenges. *Front Genet*. 2019;10(JAN):637. doi:10.3389/FGENE.2018.00637/BIBTEX
- Kurilshikov A, Medina-Gomez C, Bacigalupe R, et al. Large-scale association analyses identify host factors influencing human gut microbiome composition. *Nat Genet 2021 532*. 2021;53(2):156-165. doi:10.1038/s41588-020-00763-1
- 90. Bulik-Sullivan BK, Loh P-R, Finucane HK, et al. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*. 2015;47(3):291-295. doi:10.1038/ng.3211
- 91. Zhao Q, Wang J, Hemani G, Bowden J, Small DS. Statistical inference in two-sample summary-data Mendelian randomization using robust adjusted profile score. *Ann Stat.* 2020;48(3):1742-1769. doi:10.1214/19-AOS1866
- 92. G H, K T, G DS. Orienting the causal relationship between imprecisely measured traits using GWAS summary data. *PLoS Genet.* 2017;13(11). doi:10.1371/JOURNAL.PGEN.1007081
- 93. Sanderson E, Spiller W, Bowden J. Testing and correcting for weak and pleiotropic instruments in twosample multivariable Mendelian randomization. *Stat Med*. Published online 2021. doi:10.1002/SIM.9133
- 94. Mounier N, Kutalik Z. Bias correction for inverse variance weighting Mendelian randomization. *bioRxiv*.
 Published online December 18, 2021:2021.03.26.437168. doi:10.1101/2021.03.26.437168
- 95. S B, P R, KB J, et al. A subset-based approach improves power and interpretation for the combined analysis of genetic association studies of heterogeneous traits. *Am J Hum Genet*. 2012;90(5):821-835. doi:10.1016/J.AJHG.2012.03.015
- 96. Breton G, Zheng S, Valieris R, Tojal da Silva I, Satija R, Nussenzweig MC. Human dendritic cells (DCs) are derived from distinct circulating precursors that are precommitted to become CD1c+ or CD141+ DCs. *J Exp Med*. 2016;213(13):2861-2870. doi:10.1084/JEM.20161135
- M B, A V, SL W, et al. A Single-Cell Transcriptomic Map of the Human and Mouse Pancreas Reveals Inter- and Intra-cell Population Structure. *Cell Syst.* 2016;3(4):346-360.e4. doi:10.1016/J.CELS.2016.08.011
- 98. S D, SA S, Y Z, et al. A survey of human brain transcriptome diversity at the single cell level. *Proc Natl*© 2022 America Medical Association. All rights reserved.

Acad Sci U S A. 2015;112(23):7285-7290. doi:10.1073/PNAS.1507125112

- 99. S Z, S Z, X F, et al. A single-cell RNA-seq survey of the developmental landscape of the human prefrontal cortex. *Nature*. 2018;555(7697):524-528. doi:10.1038/NATURE25980
- G LM, D G, S C, et al. Molecular Diversity of Midbrain Development in Mouse, Human, and Stem Cells. *Cell*. 2016;167(2):566-580.e19. doi:10.1016/J.CELL.2016.09.027
- 101. H H, P L, R H, et al. STRT-seq-2i: dual-index 5' single cell and nucleus RNA-seq on an addressable microwell array. *Sci Rep*. 2017;7(1). doi:10.1038/S41598-017-16546-4
- 102. D W, S L, J W, et al. Comprehensive functional genomic resource and integrative model for the human brain. *Science*. 2018;362(6420). doi:10.1126/SCIENCE.AAT8464
- 103. GX Z, JM T, P B, et al. Massively parallel digital transcriptional profiling of single cells. *Nat Commun*.
 2017;8. doi:10.1038/NCOMMS14049
- 104. N H, I A-D, A B, et al. Massively parallel single-nucleus RNA-seq with DroNc-seq. *Nat Methods*.2017;14(10):955-958. doi:10.1038/NMETH.4407
- 105. M E, HE A, M M, et al. Single-Cell Analysis of Human Pancreas Reveals Transcriptional Signatures of Aging and Somatic Mutation Patterns. *Cell*. 2017;171(2):321-330.e14. doi:10.1016/J.CELL.2017.09.004
- Skipper M. A two-pronged model of polyglutamine disease. *Nat Rev Genet 2008 95*. 2008;9(5):322-322.
 doi:10.1038/nrg2366
- 107. Tazelaar GHP, Boeynaems S, De Decker M, et al. ATXN1 repeat expansions confer risk for amyotrophic lateral sclerosis and contribute to TDP-43 mislocalization. *Brain Commun*. 2020;2(2):14. doi:10.1093/BRAINCOMMS/FCAA064
- Crespo-Barreto J, Fryer JD, Shaw CA, Orr HT, Zoghbi HY. Partial Loss of Ataxin-1 Function Contributes to Transcriptional Dysregulation in Spinocerebellar Ataxia Type 1 Pathogenesis. *PLOS Genet*. 2010;6(7):e1001021. doi:10.1371/JOURNAL.PGEN.1001021
- 109. Gardiner SL, Harder AVE, Campman YJM, et al. Repeat length variations in ATXN1 and AR modify disease expression in Alzheimer's disease. *Neurobiol Aging*. 2019;73:230.e9-230.e17. doi:10.1016/J.NEUROBIOLAGING.2018.09.007
- 110. Liu J, Su B. Integrated analysis supports ATXN1 as a schizophrenia risk gene. *Schizophr Res*.2018;195:298-305. doi:10.1016/J.SCHRES.2017.10.010
- © 2022 America Medical Association. All rights reserved.

- 111. Didonna A, Puig EC, Ma Q, et al. Ataxin-1 regulates B cell function and the severity of autoimmune experimental encephalomyelitis. *Proc Natl Acad Sci U S A*. 2020;117(38):23742-23750.
 doi:10.1073/PNAS.2003798117/-/DCSUPPLEMENTAL
- 112. Ma Q, Didonna A. The novel multiple sclerosis susceptibility gene ATXN1 regulates B cell receptor signaling in B-1a cells. *Mol Brain*. 2021;14(1):1-5. doi:10.1186/S13041-020-00715-0/FIGURES/1
- Plank MW, Maltby S, Tay HL, et al. MicroRNA Expression Is Altered in an Ovalbumin-Induced Asthma Model and Targeting miR-155 with Antagomirs Reveals Cellular Specificity. *PLoS One*. 2015;10(12):e0144810. doi:10.1371/JOURNAL.PONE.0144810
- 114. Lee Y, Fryer JD, Kang H, et al. ATXN1 protein family and CIC regulate extracellular matrix remodeling and lung alveolarization. *Dev Cell*. 2011;21(4):746-757. doi:10.1016/J.DEVCEL.2011.08.017
- 115. Dubinsky MC, Mei L, Friedman M, et al. Genome Wide Association (GWA) Predictors Of Anti-TNFα Therapeutic Responsiveness In Pediatric Inflammatory Bowel Disease (IBD). *Inflamm Bowel Dis*. 2010;16(8):1357. doi:10.1002/IBD.21174
- Polushina T, Banerjee N, Giddaluru S, et al. Identification of pleiotropy at the gene level between psychiatric disorders and related traits. *Transl Psychiatry 2021 111*. 2021;11(1):1-9. doi:10.1038/s41398-021-01530-4
- 117. Rizzi TS, Arias-Vasquez A, Rommelse N, et al. The ATXN1 and TRIM31 genes are related to intelligence in an ADHD background: evidence from a large collaborative study totaling 4,963 subjects. *Am J Med Genet B Neuropsychiatr Genet*. 2011;156(2):145-157. doi:10.1002/AJMG.B.31149
- 118. Kiang KMY, Leung GKK. A Review on Adducin from Functional to Pathological Mechanisms: Future Direction in Cancer. *Biomed Res Int.* 2018;2018. doi:10.1155/2018/3465929
- 119. Thomas KN, Wang S, Zhang H, et al. Abstract 35: Gamma Adducin Dysfunction Leads To Cerebrovascular Distention, Blood Brain Barrier Leakage, And Cognitive Deficits In The Fawn-hooded Hypertensive Rats. *Hypertension*. 2021;78(Suppl_1). doi:10.1161/HYP.78.SUPPL_1.35
- 120. Byrne EM, Zhu Z, Qi T, et al. Conditional GWAS analysis to identify disorder-specific SNPs for psychiatric disorders. *Mol Psychiatry*. 2021;26(6):2070-2081. doi:10.1038/S41380-020-0705-9
- 121. Charney AW, Ruderfer DM, Stahl EA, et al. Evidence for genetic heterogeneity between clinical subtypes of bipolar disorder. *Transl Psychiatry*. 2017;7(1):e993. doi:10.1038/TP.2016.242
- © 2022 America Medical Association. All rights reserved.

- 122. Rabenstein RL, Addy NA, Caldarone BJ, et al. Impaired Synaptic Plasticity and Learning in Mice
 Lacking β-Adducin, an Actin-Regulating Protein. *J Neurosci*. 2005;25(8):2138-2145.
 doi:10.1523/JNEUROSCI.3530-04.2005
- 123. Engmann O, Giralt A, Gervasi N, et al. DARPP-32 interaction with adducin may mediate rapid environmental effects on striatal neurons. *Nat Commun 2015* 61. 2015;6(1):1-14. doi:10.1038/ncomms10099
- Yang H, Francis SC, Sellers K, et al. Hypertension-linked decrease in the expression of brain γ-adducin.
 Circ Res. 2002;91(7):633-639. doi:10.1161/01.RES.0000036749.73316.73
- 125. Cheng G, Tang CSM, Wong EHM, et al. Common genetic variants regulating ADD3 gene expression alter biliary atresia risk. *J Hepatol*. 2013;59(6):1285-1291. doi:10.1016/J.JHEP.2013.07.021
- 126. Acharya P, Kutum R, Pandey R, et al. First Degree Relatives of Patients with Celiac Disease Harbour an Intestinal Transcriptomic Signature that Might Protect them from Enterocyte Damage. *Clin Transl Gastroenterol.* 2018;9(10). doi:10.1038/S41424-018-0059-7
- 127. Zigmond E, Bernshtein B, Friedlander G, et al. Macrophage-restricted interleukin-10 receptor deficiency, but not IL-10 deficiency, causes severe spontaneous colitis. *Immunity*. 2014;40(5):720-733. doi:10.1016/J.IMMUNI.2014.03.012
- 128. Howrylak JA, Moll M, Weiss ST, Raby BA, Wu W, Xing EP. Gene expression profiling of asthma phenotypes demonstrates molecular signatures of atopy and asthma control. *J Allergy Clin Immunol*. 2016;137(5):1390-1397.e6. doi:10.1016/J.JACI.2015.09.058
- 129. Weiner JA, Jontes J. Protocadherins, not prototypical: A complex tale of their interactions, expression, and functions. *Front Mol Neurosci*. 2013;0(MARCH 2013):4. doi:10.3389/FNMOL.2013.00004/BIBTEX
- Jia Z, Wu Q. Clustered Protocadherins Emerge as Novel Susceptibility Loci for Mental Disorders. *Front Neurosci.* 2020;14. doi:10.3389/FNINS.2020.587819
- 131. Pancho A, Aerts T, Mitsogiannis MD, Seuntjens E. Protocadherins at the Crossroad of Signaling Pathways. *Front Mol Neurosci*. 2020;13:117. doi:10.3389/FNMOL.2020.00117/BIBTEX
- Lu, Zhou Y, Qiao P, Zheng J, Wu Q, Shen Q. The protocadherin alpha cluster is required for axon extension and myelination in the developing central nervous system. *Neural Regen Res*. 2018;13(3):427. doi:10.4103/1673-5374.228724
- © 2022 America Medical Association. All rights reserved.

- 133. Fan L, Lu Y, Shen X, Shao H, Suo L, Wu Q. Alpha protocadherins and Pyk2 kinase regulate cortical neuron migration and cytoskeletal dynamics via rac1 GTPase and WAVE complex in mice. *Elife*. 2018;7. doi:10.7554/ELIFE.35242
- 134. Katori S, Noguchi-Katori Y, Okayama A, et al. Protocadherin-αC2 is required for diffuse projections of serotonergic axons. *Sci Reports 2017 71*. 2017;7(1):1-14. doi:10.1038/s41598-017-16120-y
- 135. Katori S, Hamada S, Noguchi Y, et al. Protocadherin-α Family Is Required for Serotonergic Projections to Appropriately Innervate Target Brain Areas. *J Neurosci*. 2009;29(29):9137-9147.
 doi:10.1523/JNEUROSCI.5478-08.2009
- 136. Xue L, Fergusson J, Salimi M, et al. Prostaglandin D2 and leukotriene E4 synergize to stimulate diverse TH2 functions and TH2 cell/neutrophil crosstalk. *J Allergy Clin Immunol*. 2015;135(5):1358-1366.e11. doi:10.1016/J.JACI.2014.09.006
- 137. Tellez GF, Willemse BWM, Brouwer U, et al. Protocadherin-1 Localization and Cell-Adhesion Function in Airway Epithelial Cells in Asthma. *PLoS One*. 2016;11(10):e0163967. doi:10.1371/JOURNAL.PONE.0163967
- 138. Kozu Y, Gon Y, Maruoka S, et al. Protocadherin-1 is a glucocorticoid-responsive critical regulator of airway epithelial barrier function. *BMC Pulm Med 2015 151*. 2015;15(1):1-12. doi:10.1186/S12890-015-0078-Z
- 139. SW C, DA S, NE G-L, et al. Intestinal brush border assembly driven by protocadherin-based intermicrovillar adhesion. *Cell*. 2014;157(2):433-446. doi:10.1016/J.CELL.2014.01.067
- 140. Robertson IB, Horiguchi M, Zilberberg L, Dabovic B, Hadjiolova K, Rifkin DB. Latent TGF-β-binding proteins. *Matrix Biol*. 2015;47:44. doi:10.1016/J.MATBIO.2015.05.005
- 141. Rauf B, Irum B, Khan SY, et al. Novel mutations in LTBP2 identified in familial cases of primary congenital glaucoma. *Mol Vis*. 2020;26:14. Accessed February 4, 2022. /labs/pmc/articles/PMC7043638/
- 142. Désir J, Sznajer Y, Depasse F, et al. LTBP2 null mutations in an autosomal recessive ocular syndrome with megalocornea, spherophakia, and secondary glaucoma. *Eur J Hum Genet 2010 187*. 2010;18(7):761-767. doi:10.1038/ejhg.2010.11
- 143. Mižíková I, Morty RE. The extracellular matrix in bronchopulmonary dysplasia: Target and source. *Front* © 2022 America Medical Association. All rights reserved.

Med. 2015;2(DEC):91. doi:10.3389/FMED.2015.00091/BIBTEX

- 144. Enomoto Y, Matsushima S, Shibata K, et al. LTBP2 is secreted from lung myofibroblasts and is a potential biomarker for idiopathic pulmonary fibrosis. *Clin Sci (Lond)*. 2018;132(14):1565-1580. doi:10.1042/CS20180435
- 145. Huo R, Tian X, Chang Q, et al. Targeted inhibition of β-catenin alleviates airway inflammation and remodeling in asthma via modulating the profibrotic and anti-inflammatory actions of transforming growth factor-β1. *Ther Adv Respir Dis*. 2021;15. doi:10.1177/1753466620981858
- Speca S, Giusti I, Rieder F, Latella G. Cellular and molecular mechanisms of intestinal fibrosis. World J Gastroenterol. 2012;18(28):3635. doi:10.3748/WJG.V18.I28.3635
- 147. Troncone E, Marafini I, Stolfi C, Monteleone G. Transforming growth factor-β1/Smad7 in intestinal immunity, inflammation, and cancer. *Front Immunol*. 2018;9(JUN):1407.
 doi:10.3389/FIMMU.2018.01407/BIBTEX
- Schokker D, Jansman AJM, Veninga G, et al. Perturbation of microbiota in one-day old broiler chickens with antibiotic for 24 hours negatively affects intestinal immune development. *BMC Genomics*. 2017;18(1):1-14. doi:10.1186/S12864-017-3625-6/FIGURES/5
- Chen J, Gao G, Wang H, Ye X, Zhou J, Lin J. Expression and clinical significance of latent-transforming growth factor beta-binding protein 2 in primary hepatocellular carcinoma. *Medicine (Baltimore)*.
 2019;98(39). doi:10.1097/MD.00000000017216
- 150. Wang J, Jiang C, Li N, et al. The circEPSTI1/mir-942-5p/LTBP2 axis regulates the progression of OSCC in the background of OSF via EMT and the PI3K/Akt/mTOR pathway. *Cell Death Dis 2020 118*. 2020;11(8):1-18. doi:10.1038/s41419-020-02851-w
- 151. Ahmad R, Dalziel JE. G Protein-Coupled Receptors in Taste Physiology and Pharmacology. *Front Pharmacol.* 2020;11:1771. doi:10.3389/FPHAR.2020.587664/BIBTEX
- 152. Mao GF, Jin J, Kunapuli SP, Rao AK. Nuclear factor-κB regulates expression of platelet phospholipase
 C-β2 (PLCB2). *Thromb Haemost*. 2016;116(5):931-940. doi:10.1160/TH15-09-0749
- Lin JR, Cai Y, Zhang Q, Zhang W, Nogales-Cadenas R, Zhang ZD. Integrated post-GWAS analysis sheds new light on the disease mechanisms of schizophrenia. *Genetics*. 2016;204(4):1587-1600. doi:10.1534/GENETICS.116.187195/-/DC1/FIGURES18.PDF
- © 2022 America Medical Association. All rights reserved.

- 154. Li H, Li X, Smerin SE, et al. Mitochondrial gene expression profiles and metabolic pathways in the amygdala associated with exaggerated fear in an animal model of PTSD. *Front Neurol*. 2014;5 AUG. doi:10.3389/FNEUR.2014.00164/ABSTRACT
- 155. Kawakami T, Xiao W. Phospholipase C-β in immune cells. *Adv Biol Regul.* 2013;53(3):249-257.
 doi:10.1016/J.JBIOR.2013.08.001
- 156. Bach TL, Chen Q-M, Kerr WT, et al. Phospholipase cbeta is critical for T cell chemotaxis. *J Immunol*. 2007;179(4):2223-2227. doi:10.4049/JIMMUNOL.179.4.2223
- 157. Wang L, Zhou Y, Chen Z, et al. PLCβ2 negatively regulates the inflammatory response to virus infection by inhibiting phosphoinositide-mediated activation of TAK1. *Nat Commun 2019 101*. 2019;10(1):1-13. doi:10.1038/s41467-019-08524-3
- 158. Zhang NZ, Chen XJ, Mu YH, Wang H. Identification of differentially expressed genes in childhood asthma. *Medicine (Baltimore)*. 2018;97(21). doi:10.1097/MD.0000000000010861
- 159. Alazami AM, Patel N, Shamseldin HE, et al. Accelerating Novel Candidate Gene Discovery in Neurogenetic Disorders via Whole-Exome Sequencing of Prescreened Multiplex Consanguineous Families. *Cell Rep.* 2015;10(2):148-161. doi:10.1016/J.CELREP.2014.12.015
- Dunn EC, Soare TW, Zhu Y, et al. Sensitive periods for the effect of childhood adversity on DNA methylation: Results from a prospective, longitudinal study. *Biol Psychiatry*. 2019;85(10):838. doi:10.1016/J.BIOPSYCH.2018.12.023
- Gondalia R, Baldassari A, Holliday KM, et al. Methylome-wide association study provides evidence of particulate matter air pollution-associated DNA methylation. *Environ Int*. 2019;132:104723. doi:10.1016/J.ENVINT.2019.03.071
- 162. Uckelmann H, Blaszkiewicz S, Nicolae C, et al. Extracellular matrix protein Matrilin-4 regulates stressinduced HSC proliferation via CXCR4. *J Exp Med*. 2016;213(10):1961-1971. doi:10.1084/JEM.20151713
- Alcantara D, Timms AE, Gripp K, et al. Mutations of AKT3 are associated with a wide spectrum of developmental disorders including extreme megalencephaly. *Brain*. 2017;140(10):2610-2622. doi:10.1093/BRAIN/AWX203
- 164. Matsuda S, Ikeda Y, Murakami M, Nakagawa Y, Tsuji A, Kitagishi Y. Roles of PI3K/AKT/GSK3 Pathway © 2022 America Medical Association. All rights reserved.

Involved in Psychiatric Illnesses. *Dis 2019, Vol 7, Page 22*. 2019;7(1):22. doi:10.3390/DISEASES7010022

- 165. Howell KR, Floyd K, Law AJ. PKBγ/AKT3 loss-of-function causes learning and memory deficits and deregulation of AKT/mTORC2 signaling: Relevance for schizophrenia. *PLoS One*. 2017;12(5). doi:10.1371/JOURNAL.PONE.0175993
- 166. Wong H, Levenga J, Laplante L, et al. Isoform-specific roles for AKT in affective behavior, spatial memory, and extinction related to psychiatric disorders. *Elife*. 2020;9:1-27. doi:10.7554/ELIFE.56630
- Bergeron Y, Bureau G, Laurier-Laurin MÉ, Asselin E, Massicotte G, Cyr M. Genetic Deletion of Akt3
 Induces an Endophenotype Reminiscent of Psychiatric Manifestations in Mice. *Front Mol Neurosci*.
 2017;10. doi:10.3389/FNMOL.2017.00102
- 168. Kitz A, Marcken M de, Gautron A-S, Mitrovic M, Hafler DA, Dominguez-Villar M. AKT isoforms modulate Th1-like Treg generation and function in human autoimmune disease. *EMBO Rep*. 2016;17(8):1169-1183. doi:10.15252/EMBR.201541905
- 169. DuBois JC, Ray AK, Gruber RC, et al. Akt3-Mediated Protection Against Inflammatory Demyelinating Disease. *Front Immunol.* 2019;10:1738. doi:10.3389/FIMMU.2019.01738/FULL
- 170. Tsiperson V, Gruber RC, Goldberg MF, et al. Suppression of Inflammatory Responses during Myelin Oligodendrocyte Glycoprotein–Induced Experimental Autoimmune Encephalomyelitis Is Regulated by AKT3 Signaling. *J Immunol.* 2013;190(4):1528-1539. doi:10.4049/JIMMUNOL.1201387
- 171. White JH, Chiano M, Wigglesworth M, et al. Identification of a novel asthma susceptibility gene on chromosome 1qter and its functional evaluation. *Hum Mol Genet*. 2008;17(13). doi:10.1093/hmg/ddn087
- 172. Ugbode C, West RJH. Lessons learned from CHMP2B, implications for frontotemporal dementia and amyotrophic lateral sclerosis. *Neurobiol Dis*. 2021;147:105144. doi:10.1016/J.NBD.2020.105144
- 173. Zhang R, Song J, Isgren A, et al. Genome-wide study of immune biomarkers in cerebrospinal fluid and serum from patients with bipolar disorder and controls. *Transl Psychiatry*. 2020;10(1). doi:10.1038/S41398-020-0737-6
- 174. Yang L, Shi T, Liu F, et al. REV3L, a promising target in regulating the chemosensitivity of cervical cancer cells. *PLoS One*. 2015;10(3). doi:10.1371/journal.pone.0120334
- 175. Wittschieben JP, Patil V, Glushets V, Robinson LJ, Kusewitt DF, Wood RD. Loss of DNA polymerase ζ

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enhances spontaneous tumorigenesis. Cancer Res. 2010;70(7). doi:10.1158/0008-5472.CAN-09-4267

- 176. Huang KK, Jang KW, Kim S, et al. Exome sequencing reveals recurrent REV3L mutations in cisplatinresistant squamous cell carcinoma of head and neck. *Sci Rep*. 2016;6. doi:10.1038/srep19552
- 177. Zhu X, Zou S, Zhou J, et al. REV3L, the catalytic subunit of DNA polymerase ζ, is involved in the progression and chemoresistance of esophageal squamous cell carcinoma. *Oncol Rep.* 2016;35(3). doi:10.3892/or.2016.4549
- 178. Jamwal RS, Mahajan N, Bhat GR, et al. REV3L single nucleotide variants lead to increased susceptibility towards non-small cell lung cancer in the population of Jammu and Kashmir. *Cancer Epidemiol.* 2021;75. doi:10.1016/j.canep.2021.102047
- Martin SK, Tomida J, Wood RD. Disruption of DNA polymerase ζ engages an innate immune response.
 Cell Rep. 2021;34(8). doi:10.1016/j.celrep.2021.108775
- 180. Lange SS, Wittschieben JP, Wood RD. DNA polymerase zeta is required for proliferation of normal mammalian cells. *Nucleic Acids Res*. 2012;40(10). doi:10.1093/nar/gks054
- Tomas-Roca L, Tsaalbi-Shtylik A, Jansen JG, et al. De novo mutations in PLXND1 and REV3L cause
 Möbius syndrome. *Nat Commun*. 2015;6. doi:10.1038/ncomms8199
- 182. O-Wang J, Kajiwara K, Kawamura K, et al. An essential role for REV3 in mammalian cell survival:
 Absence of REV3 induces p53-independent embryonic death. *Biochem Biophys Res Commun.* 2002;293(3). doi:10.1016/S0006-291X(02)00341-8
- 183. Govind N, Choudhury A, Hodkinson B, et al. Immunochip identifies novel, and replicates known, genetic risk loci for rheumatoid arthritis in black South Africans. *Mol Med*. 2014;20(1). doi:10.2119/molmed.2014.00097
- 184. Li X, Commane M, Nie H, et al. Act1, an NF-κB-activating protein. *Proc Natl Acad Sci U S A*.
 2000;97(19). doi:10.1073/pnas.160265197
- 185. Qian Y, Liu C, Hartupee J, et al. The adaptor Act1 is required for interleukin 17 Dependent signaling associated with autoimmune and inflammatory disease. *Nat Immunol*. 2007;8(3). doi:10.1038/ni1439
- 186. Hunter CA. Act1-ivating IL-17 inflammation. Nat Immunol. 2007;8(3). doi:10.1038/ni0307-232
- 187. Mummidi S, Das NA, Carpenter AJ, et al. RECK suppresses interleukin-17/TRAF3IP2-mediated MMP13 activation and human aortic smooth muscle cell migration and proliferation. *J Cell Physiol*.
- © 2022 America Medical Association. All rights reserved.

2019;234(12). doi:10.1002/jcp.28792

- 188. Valente AJ, Irimpen AM, Siebenlist U, Chandrasekar B. OxLDL induces endothelial dysfunction and death via TRAF3IP2: Inhibition by HDL3 and AMPK activators. *Free Radic Biol Med*. 2014;70. doi:10.1016/j.freeradbiomed.2014.02.014
- 189. Sakamuri SSVP, Higashi Y, Sukhanov S, et al. TRAF3IP2 mediates atherosclerotic plaque development and vulnerability in ApoE-/- mice. *Atherosclerosis*. 2016;252. doi:10.1016/j.atherosclerosis.2016.05.029
- 190. Padilla J, Carpenter AJ, Das NA, et al. TRAF3IP2 mediates high glucose-induced endothelin-1 production as well as endothelin-1-induced inflammation in endothelial cells. *Am J Physiol - Hear Circ Physiol*. 2018;314(1). doi:10.1152/ajpheart.00478.2017
- 191. Venkatesan B, Valente AJ, Das NA, et al. CIKS (Act1 or TRAF3IP2) mediates high glucose-induced endothelial dysfunction. *Cell Signal*. 2013;25(1). doi:10.1016/j.cellsig.2012.10.009
- 192. Grunewald ZI, Ramirez-Perez FI, Woodford ML, et al. TRAF3IP2 (TRAF3 interacting protein 2) mediates obesity-associated vascular insulin resistance and dysfunction in male mice. *Hypertension*. Published online 2020. doi:10.1161/HYPERTENSIONAHA.120.15262
- 193. Shafer S, Yao Y, Comrie W, et al. Two patients with chronic mucocutaneous candidiasis caused by TRAF3IP2 deficiency. *J Allergy Clin Immunol*. 2021;148(1). doi:10.1016/j.jaci.2020.12.629
- 194. Hüffmeier U, Uebe S, Ekici AB, et al. Common variants at TRAF3IP2 are associated with susceptibility to psoriatic arthritis and psoriasis. *Nat Genet*. 2010;42(11):996-999. doi:10.1038/ng.688
- 195. Ellinghaus E, Ellinghaus D, Stuart PE, et al. Genome-wide association study identifies a psoriasis susceptibility locus at TRAF3IP2. *Nat Genet*. 2010;42(11):991-995. doi:10.1038/ng.689
- 196. Ciccacci C, Biancone L, Fusco D Di, et al. TRAF3IP2 gene is associated with cutaneous extraintestinal manifestations in Inflammatory Bowel Disease. *J Crohn's Colitis*. 2013;7(1). doi:10.1016/j.crohns.2012.02.020
- 197. Ciccacci C, Rufini S, Mancinelli S, et al. A pharmacogenetics study in Mozambican patients treated with Nevirapine: Full resequencing of TRAF3IP2 gene shows a novel association with SJS/TEN susceptibility. *Int J Mol Sci.* 2015;16(3). doi:10.3390/ijms16035830
- 198. DaRosa PA, Wang Z, Jiang X, et al. Allosteric activation of the RNF146 ubiquitin ligase by a poly(ADPribosyl)ation signal. *Nature*. 2014;517(7533). doi:10.1038/nature13826
- © 2022 America Medical Association. All rights reserved.

- 199. Huang SMA, Mishina YM, Liu S, et al. Tankyrase inhibition stabilizes axin and antagonizes Wnt signalling. *Nature*. 2009;461(7264). doi:10.1038/nature08356
- 200. Smith S, Giriat I, Schmitt A, Lange T De. Tankyrase, a poly(ADP-ribose) polymerase at human telomeres. *Science (80-)*. 1998;282(5393). doi:10.1126/science.282.5393.1484
- 201. Chi NW, Lodish HF. Tankyrase is a Golgi-associated mitogen-activated protein kinase substrate that interacts with IRAP in GLUT4 vesicles. *J Biol Chem*. 2000;275(49). doi:10.1074/jbc.M007635200
- 202. Haikarainen T, Krauss S, Lehtio L. Tankyrases: Structure, Function and Therapeutic Implications in Cancer. *Curr Pharm Des*. 2014;20(41). doi:10.2174/1381612820666140630101525
- 203. Fuchs U, Rehkamp GF, Slany R, Follo M, Borkhardt A. The formin-binding protein 17, FBP17, binds via a TNKS binding motif to tankyrase, a protein involved in telomere maintenance. *FEBS Lett*. 2003;554(1-2). doi:10.1016/S0014-5793(03)01063-9
- 204. Li Z, Yamauchi Y, Kamakura M, et al. Herpes Simplex Virus Requires Poly(ADP-Ribose) Polymerase Activity for Efficient Replication and Induces Extracellular Signal-Related Kinase-Dependent Phosphorylation and ICP0-Dependent Nuclear Localization of Tankyrase 1. *J Virol.* 2012;86(1). doi:10.1128/jvi.05897-11
- 205. Deng Z, Atanasiu C, Zhao K, et al. Inhibition of Epstein-Barr Virus OriP Function by Tankyrase, a Telomere-Associated Poly-ADP Ribose Polymerase That Binds and Modifies EBNA1. *J Virol*. 2005;79(8). doi:10.1128/jvi.79.8.4640-4650.2005
- 206. Scherag A, Dina C, Hinney A, et al. Two new loci for body-weight regulation identified in a joint analysis of genome-wide association studies for early-onset extreme obesity in French and German study groups. *PLoS Genet*. 2010;6(4). doi:10.1371/journal.pgen.1000916

Supplementary Figures

eFigure 1. Flow Chart of the Study Design and Analysis Plan



S. Figure 1 Legend. Study design and analysis plan with miniaturized depictions of the corresponding Figures and S. Figures to orient and guide readers. The top-left panel depicts literature and databases searched for relevant GWAS summary statistics. The mid-left panel depicts LD-score regression (LDSC) analysis to estimate SNP heritability Z-score, resulting in a final set of phenotypes. The bottom-left panel depicts pairwise evaluation of genetic correlations among psychiatric, immune-related, and third variable phenotypes using LDSC. A set of 44 correlated psychiatric-immune pairs (FDR q < 0.05) were investigated for bidirectional Mendelian randomization (MR) effects with sensitivity analyses. Multivariable MR (MVMR) was used to adjust for effects of additional phenotypes (depicted in the bottom-center panel). Significant psychiatric instrument loci with positive effects on immune outcomes were interrogated for enriched genes, tissues, and gene-sets using FUMA's *GENE2FUNCTION* analysis (middle-center panel). For phenotype pairs demonstrating robust MR effects, two-sided GWAS meta-analysis identified subsets of SNPs with concordant and discordant effects (bottom right panel), and these SNP subsets were characterized for enriched genes tissues, cell-types, gene-sets, and genes was performed with FUMA SNP2GENE function (middle-center panel).

eFigure 2. Genetic Correlations Among Psychiatric Disorders and Immune-Related Traits and Other Risk Factors



Genetic correlation coefficients (text) and z-score (color) between psychiatric (X-axis), immune-related (Y-axis), and third variable phenotypes (both axes). Increasing intensity of red color indicates positive correlations and increasing intensity of blue indicates negative correlations. Correlations reaching uncorrected p < 0.05 are denoted with *, while those reaching false discovery rate q < 0.05 (reflecting correction for a total 587 unique tests) are denoted with **. Abbreviations include allergic rhinitis (AR), anorexia nervosa (AN), atopic dermatitis (AD), attention-deficit hyperactivity disorder (ADHD), body mass index (BMI), cannabis use disorder (CUD), cigarettes per day (CPD), cognitive processing (CP), alcoholic drinks per week (DPW), educational attainment (EA), frequency of moderate intensity exercise (EXER), heart rate (HR), heart rate increase during exercise (HRI), heart rate recovery after exercise (HRR), heart rate variability (HRV), hypothyroidism (HYPO), obsessive compulsive disorder (OCD), post-traumatic stress disorder (PTSD), primary biliary cirrhosis (PBC),

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primary sclerosing cholangitis (PSC), problematic alcohol use (PAU), rheumatoid arthritis (RA), risk-tolerance (RT), sleep duration (SLEEP), social deprivation *via* the Townsend Index (SOCD), social interaction frequency (SOCI), schizophrenia (SZ), systemic lupus erythematosus (SLE), Tourette's syndrome (TS), type 1 diabetes (T1D), ulcerative colitis (UC). Full results are also provided in S. Table 2.

eFigure 3. MVMR Effects After Adjustment for Other Risk Factors



Multivariable Mendelian randomization results, depicting effect sizes with 95% confidence intervals with and without adjustment for the additional phenotypes. Full results of these analyses are provided in S. Table 3. Abbreviations include body mass index (BMI), cognitive processing (CP), heart rate variability (HRV), hypothyroidism (HYPO), major depressive disorder (MDD), schizophrenia (SZ), ulcerative colitis (UC).





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Meta-Analysis Crohn's and Cross-Disorder: Discordant

4B.









4F.

S. Figures 4 Legend. Miami plots depicting the results of ASSET two-sided meta-analysis of psychiatric-immune phenotype pairs (limited to casecontrol data only), such that the top of the plot reflects SNPs with significantly concordant effect for both phenotypes, and the bottom plot reflects SNPs with significantly discordant effect for both phenotypes. Y-axes reflect the magnitude of the $-\log_{10}(p$ -value) and all plots have the same y-axis limits to facilitate comparison. The red line depicts the threshold for genome-wide significance (5x10⁻⁸). Details for each of the LD-independent genome-wide loci are provided in S. Table 6. eFigure 5. A & B. Tissue and Cell Type Enrichment Analyses of Loci With Concordant and Discordant Effects Within the Psychiatric-Immune Associations Identified



A CONDON 26 CD COM¹ COM¹ Per Coconsister Rectain Contraction St. como MOOT SHIPPED Poconestine ori St. Score St. Composition



Cell Type Enrichment





Allen_LGN_Level1_GABAergic Allen_LGN_Level2_LGN_Inh_LAMP5 Allen_MTG_Level1_Glutamatergic Allen MTG Level2 Exc L3.5 RORB COL22A1 Allen_MTG_Level2_Exc_L6_FEZF2_SCUBE1 Allen_MTG_Level2_Inh_L1.2_LAMP5_DBP Allen_MTG_Level2_Inh_L1.2_PAX6_CDH12 DroNc_Hippocampus_GABA2 GSE101601_Linnarsson_Temporal_Cortex_GABA_1 GSE101601 Linnarsson Temporal Cortex Glut 5 GSE104276_PFC_GW26_GABAergic_Neurons GSE67835 Cortex Fetal Quiescent GSE67835_Cortex_Fetal_Replicating GSE67835_Cortex_woFetal_Neurons GSE84133 Pancreas Endothelial PsychENCODE_Adult_Ex3e PsychENCODE_Adult_Ex4 PsychENCODE Adult Ex9 PsychENCODE_Dev_Fetal_Astrocytes PsychENCODE_Dev_Fetal_Endothelial PsychENCODE_Dev_Fetal_ExN PsychENCODE_Dev_Fetal_IntN PsychENCODE_Dev_Fetal_IPC PsychENCODE Dev Fetal NEP PsychENCODE_Dev_Fetal_Oligo PsychENCODE_Dev_Fetal_OPC PsychENCODE_Dev_Fetal_Pericytes PsychENCODE_Dev_Fetal_Quiescent PsychENCODE_Dev_Fetal_Replicating PsychENCODE_Dev_Fetal_Trans PsychENCODE_Dev_Neurons



Figure 5 Legend. Tissue and cell type enrichment results performed on concordant (Conc.) and discordant (Disc.) SNP subsets generated with respect to psychiatric-immune phenotype pairs. Color intensity reflects $-\log_{10}(p)$ of the enrichment *p*-value. *P*-values were corrected by applying the false discover rate (FDR) multiple testing correction per database and significant findings (FDR q < 0.05) are denoted with *. Differences in enrichment effects between respective concordant and discordant effects based on two-tailed Z-test are indicated using "Z" to denote the more significant enrichment. We display all terms with significant enrichment among at least 1 concordant SNP subsets, will full results in S. Table 7. Panel A depicts tissue-level enrichment analyses conducted using eQTL data from the GTEXv8 database. Panel B depicts cell type enrichment analyses using all human datasets provided within the FUMA interface. Numerical symbols within the heatmap denote the results of conditioned significance analyses; cell types for which Step 2 (*i.e.*, within reference data set conditioning) average proportional significance [PS] > 0.8 are denoted with 2, whereas 2' denotes the cell type with the highest average Step 2 PS when none reached the threshold of 0.8. Similarly, 3 and 3' indicate annotations with an average PS > 0.8 and the highest average PS when none reached this threshold. Full names of gene-sets can be searched within the Molecular Signature Database for detailed descriptions. Abbreviations include: ACC (anterior cingulate cortex), Brodmann's area (BA), the PGC cross-disorder phenotype (PGC-CD), developmental (Dev.), excitatory (Ex), destational week (GW), hemisphere (Hemi), inhibitory (Inh), lateral geniculate nucleus (LGN), major depressive disorder (MDD), middle temporal gyrus (MTG), neuroepithelial cells (NEP), oligodendrocyte precursor cells (OPC), prefrontal cortex (PFC), schizophrenia (SZ), and ulcerative colitis (UC).