Supplemental Figures, Tables, Methods, and References for Komachi and Burgess, 2022



Figure S1. Effect of hemizygosity on localization of mar(K101E), mar(F120Y), and mar(89-130) GFP fusions. Top panels show fluorescent images of the homozygous strains, bottom panels show hemizygous (*mar/nup2* Δ) strains. The MAR allele depicted is indicated at the top of each column. The scale bar in the wild-type MAR-GFP panel represents 5 µm.



Figure S2. Biological replicates of the MAR interacting with full-length Nup60(1-539) or Nup60(188-388) in the Y2H assay. (A) A pGAL1-HIS3 strain was transformed with *MAR-GAL4DBD* and *NUP60(1-539)-GAL4AD* plasmids. Cultures grown from four independent transformants were spotted onto -Trp -Leu (left) and -Trp -Leu -His + 5 mM 3-aminotriazole plates (right, 3-AT). (B) Same as (A), except with *NUP60(188-388)-GAL4AD* instead of *NUP60(1-539)-GAL4AD*.



Figure S3. Quantitation of fluorescence from meiotic chromosome spreads from strains expressing wild-type or mutant versions of the MAR fused to GFP. Total corrected fluorescence was measured as described in the Methods. Each panel shows results for two combined sets of mutant and wild-type spreads stained on the same day. P < 0.0001 for all mutant/WT comparisons using a two-tailed t-test, except for mar(71-130), in which P = 0.95.





 YPD
 100 mM HU

 NUP60
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Figure S4. Complementation of the growth defect and HU sensitivity by *NUP60* fragments. (A) Strains carrying the indicated alleles of *NUP60* were streaked onto YPD plates and grown for 48 hours at 30°C. The *nup60* Δ and *nup60*(Δ 189-388) strains exhibit poor growth, forming smaller colonies with a nibbled morphology. (B) Strains were grown to mid-log phase in liquid YPD and diluted to an OD₆₀₀ of 1. Serial 10-fold dilutions were spotted onto YPD (left) or YPD plus 100 mM hydroxyurea (right) and grown at

В

30°C for 48 hours and 96 hours, respectively. For both (A) and (B), the strains are isogenic to the wildtype (*NUP60*) strain: *NUP60* (SBY1903), *nup60*Δ (SBY5217), *nup60(1-388)* (SBY6423), *nup60(189-539)* (SBY6426), *nup60*(Δ189-388) (SBY6429), *nup60(189-388)* (SBY6420).

	Relevant genotype	Growth	Sporulation efficiency
NUP60-GFP NUP60 ∆(189-388)- NUP60	NUP60-GFP	normal	72 ± 7
GFP	GFP	poor	15 ± 5
	(189-388)-GFP	normal	20 ± 6
	(1-388)-GFP	normal	68 ± 7
(189-388)- (1-388)-	(189-539)-GFP	normal	15 ± 3
GFP GFP GFP GFP	(∆189-388)-GFP	poor	16 ± 4

Figure S5. Complementation of growth and sporulation by truncated Nup60-GFP fusions. (A) Growth was assessed by streaking strains onto YPD plates and growing at 30°C for 48 hours. Poorly growing strains formed smaller colonies with a nibbled morphology. All strains are isogenic to the wild-type (*NUP60*) strain: *NUP60* (SBY1899), *NUP60-GFP* (SBY6304), *GFP* (SBY6307), *nup60(189-388)-GFP* (SBY6310), *nup60(1-388)* (SBY6313), *nup60(189-539)* (SBY6316), *nup60(△189-388)-GFP* (SBY6382).
(B) The growth column is a summary of the results in (A). The strains used to assay sporulation efficiency are in an *ndj1* background and are all isogenic to the *NUP60-GFP* strain: *NUP60-GFP* (SBY6320), *GFP* (SBY6323), *nup60(189-388)-GFP* (SBY6326), *nup60(1-388)* (SBY6329), *nup60(189-388)-GFP* (SBY6332), *nup60(189-388)-GFP* (SBY6335).





Figure S6. AlphaFold predictions of the structures of Nup60 and Nup2. (A) Nup60 predicted structure from the AlphaFold v2.0 database updated on 1 July 2021 (Jumper *et al.* 2021). Regions are color-coded according to AlphaFold confidence scores based on the local distance difference test (LDDT) (Mariani *et al.* 2013): Very high (pLDDT > 90); Confident (90 > pLDDT > 70); Low (70 > pLDDT > 50); Very low (pLDDT < 50). The first and last amino acids are numbered. (B) Nup60 predicted structure, with the region sufficient for sporulation function (aa 1-388) colored gray and the region containing the inhibitory domain (aa 389-539) colored green. A predicted helix near the extreme C-terminus is colored magenta; the FxF repeats (aa 399-401, 427-429, 469-471, 509-511), purple;

SUMOylation sites (aa 440, 442, 505), yellow; a known acetylation site (aa 467), white. (C) AlphaFold v2.0 prediction of the structure of Nup2, with regions color-coded according to confidence scores described in (A). The MAR (indicated by the red arrow) and Ran GTPase binding domain are the only two regions where structure can be predicted with high confidence. (D) Blow-up of the helices that make up the MAR, rotated approximately 90° around the x-axis with respect to (C). The positions of the mutations are marked by different colors: mutations that do not disrupt localization to the nuclear envelope (K101 and F120) are colored magenta; mutations that disrupt localization but not binding to full-length Nup60 in Y2H, cyan; mutations that disrupt localization and all Y2H interaction, green. (E) Helices of the MAR in the same orientation as in (C), with side chains shown. The top panel highlights mutations that disrupt Y2H to full-length Nup60. Images in B, D, and E were created using AlphaFold v2.0 coordinates and UCSF ChimeraX version 1.2.5 (Pettersen *et al.* 2021).

				* *	* * *	* * *		*	;		* *	*	*	*	*			
S.	<i>cerevisiae</i> Nup2	89	SNSR	KAI	LN L	2F <mark>K</mark> A	K <mark>V</mark>	DDL <mark>V</mark>	LGK	PLA	DL	R <mark>PI</mark>	FI	'R <mark>Y</mark>	ELY	IKN	I	133
Η.	sapiens Nup50	144	AYHKQ	LAAI	LNC:	sv <mark>r</mark> d	DW <mark>I</mark>	VKH <mark>v</mark>	NTN	PLC	DL	Τ <mark>ΡΙ</mark>	FF	(D <mark>y</mark>	EKY	LAN	IEQQ	189
М.	<i>musculus</i> Nup50	144	AYHKQ	LAGI	LNC:	sv <mark>r</mark> d	DW <mark>I</mark>	VKH <mark>v</mark>	NTN	PLC	DL	Τ <mark>ΡΙ</mark>	FF	(D <mark>y</mark>	ERY	LAT	IEKQ	189
Χ.	<i>laevis</i> Nup50	144	EYNKQ	LTSI	LNC:	sv <mark>r</mark> d	DW <mark>I</mark>	VKH <mark>v</mark>	NAN	PLC	DL	Τ <mark>ΡΙ</mark>	F	KD <mark>Y</mark>	ERK	LSA	I <mark>eqk</mark>	189

Figure S7. Fragment of *Xenopus laevis* Nup50 sufficient for binding to Nup153 and MEL28/ELYS (Holzer *et al.* 2021) aligned with mammalian Nup50s and the MAR from *S. cerevisiae* Nup2.

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Relevant genotype	Sporulation efficiency
3HA-NUP60	82 ± 8
3HA-NUP60 ndj1	22 ± 4
3HA-NUP60 nup2	75 ± 5
3HA-NUP60 ndj1 nup2	14 ± 2

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Unlike *nup60 nup2* strains, the *NUP60-3HA nup2* strains grow well, suggesting that the Nup60-HA fusion is being partially degraded, leaving enough Nup60 to support cell growth. It is unclear why Nup60-3HA is only unstable in the absence of Nup2.



Figure S9. Binding of meiotic chromosomes by Nup60-mCherry and Nup60-3HA. (A) Representative meiotic chromosome spreads from a strain expressing Nup60-mCherry and MAR-GFP (SBY6481). Left to right are the merged image and images from the DAPI, TRITC, and FITC channels. (B) Representative meiotic chromosome spreads from a strain expressing Nup60-3HA and MAR-mCherry (SBY6410). Left to right are the merged image and images from the DAPI, FITC, and TRITC channels. The GFP fusions were detected using a polyclonal antibody to GFP and an Alexa Fluor 488-conjugated secondary antibody. Nup60-mCherry was detected using a polyclonal antibody to mCherry and an Alexa Fluor 594-conjugated secondary. Nup60-3HA was detected using a polyclonal antibody to HA and Alexa Fluor 594-conjugated secondary. The scale bar in the upper left panel represents 5 μm.

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Table S1. Yeast strains used in this study

Strain	Genotype	Background	Figure Panel	References
SBY6259	MATa ho::hisG leu2::hisG ura3::hisG GAL3(?) his4::LEU2-(NBam) nup2(∆51- 175) ndj1::TRP1	SK1	Figure 1B (Screen)	
SBY6260	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pstl) HIS4::LEU2- (NBam) nup2::KanMx ndj1::KanMx	SK1	Figure 1B (Screen)	
SBY1899	MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3	SK1	Wild-type parent of SBY1903	(Chu <i>et al.</i> 2017)
SBY1900	MATa ho∷hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam)	SK1	Wild-type parent of SBY1903	(Chu <i>et al.</i> 2017)
SBY1903	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3	SK1	Wild-type diploid strain	(Chu <i>et al.</i> 2017)
SBY6062	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2(51- 175)-GFP::CaURA3 MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam) ndj1::KanMx nup2(51- 175)-GFP::CaURA3	SK1	Table 1 (sporulation, <i>ndj1</i> background) wt MAR control	
SBY6062*	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2(51- 175)*-GFP::CaURA3 MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2(51-175)*-GFP::CaURA3	SK1	Table 1 (sporulation, <i>ndj1</i> background) Point mutant versions of SBY6062	
SBY6059	MATa ho::hisG leu2::hisG ura3(Δ Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2(Δ 1- 720)-GFP::CaURA3 MATalpha ho::hisG leu2::hisG ura3(Δ Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2(Δ 1-720)-GFP::CaURA3	SK1	Table 1 (sporulation, <i>ndj1</i> background) GFP (no MAR) control	
SBY6065	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2(71- 130)-GFP::CaURA3	SK1	Table 1 (sporulation, <i>ndj1</i> background)	

	MATalpha ho::hisG leu2::hisG			
	ura3(Δ Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
00)/0000	nup2(71-130)-GFP::CaURA3	01/4	Table 4	
SBY6266	MATa no::nisG leuz::nisG	SK1		
	ura3(\alphaSma-Pst) HIS4::LEU2-		(sporulation, <i>naj1</i>	
	(NBam) ndj1::KanMx nup2(89-		background)	
	130)-GFP::CaURA3	-		
	MATalpha ho::hisG leu2::hisG			
	ura3(Δ Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
	nup2(89-130)-GFP::CaURA3	0.44		
SBY6414	MATa ho::hisG leu2::hisG	SK1	Table 1	
	ura3(∆Sma-Pst) HIS4::LEU2-		(sporulation, <i>ndj1</i>	
	(NBam) ndj1::KanMx nup2(89-		background)	
	175)-GFP::CaURA3	-		
	MATalpha ho::hisG leu2::hisG			
	ura3(∆Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
	nup2(89-175)-GFP::CaURA3			
SBY3981	MATalpha ho::hisG leu2::hisG	SK1	Table 1	
X	ura3(∆Sma-Pst) his4-X::LEU2-		(hemizygote	
SBY6114	(NBam)-URA3 nup2:KanMx		sporulation in <i>ndj1</i>	
	ndj1::KanMx	-	background)	
	MATa ho::hisG leu2::hisG			
	ura3(∆Sma-Pst) HIS4::LEU2-			
	(NBam) nup2(51-175) ^{k101E} -			
	GFP::CaURA3 ndj1::KanMx			
SBY3981	MATalpha ho::hisG leu2::hisG	SK1	Table 1	
X	ura3(∆Sma-Pst) his4-X::LEU2-		(hemizygote	
SBY6081	(NBam)-URA3 nup2:KanMx		sporulation in <i>ndj1</i>	
	ndj1::KanMx	_	background)	
	MATa ho::hisG leu2::hisG			
	ura3(∆Sma-Pst) HIS4::LEU2-			
	(NBam) nup2(51-175) ^{F120Y} -			
	GFP::CaURA3 ndj1::KanMx			
SBY3981	MATalpha ho::hisG leu2::hisG	SK1	Table 1	
X	ura3(∆Sma-Pst) his4-X::LEU2-		(hemizygote	
SBY6264	(NBam)-URA3 nup2:KanMx		sporulation in <i>ndj1</i>	
	ndj1::KanMx	-	background)	
	MATa ho::hisG leu2::hisG			
	ura3(∆Sma-Pst) HIS4::LEU2-			
	(NBam) nup2(89-130)-			
	GFP::CaURA3 ndj1::KanMx			
SBY3981	MATalpha ho::hisG leu2::hisG	SK1	Table 1	
X	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-		(hemizygote	
SBY6413	(NBam)-URA3 nup2:KanMx		sporulation in <i>ndj1</i>	
	ndj1::KanMx	4	background)	
	MATa ho::hisG leu2::hisG			
	ura3(⊿Sma-Pst) HIS4::LEU2-			
	(NBam) nup2(89-175)-			
	GFP::CaURA3 ndj1::KanMx			

SBY6068	MATa ho::hisG leu2::hisG	SK1	Table 1, Figure 1D
	$ura3(\Delta Sma-Pst)$ HIS4::LEU2-		(sporulation,
	GFP::CaURA3		localization
	MATalpha ho::hisG leu2::hisG	-	
	ura3(Δ Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup2(51-175)- GEP::CaURA3		
SBY6068*	MATa ho::hisG leu2::hisG	SK1	Table 1, Figure 1D
	ura3(∆Sma-Pst) HIS4::LEU2-		(sporulation,
	(NBam) nup2(51-175)*-		localization)
	GFP::CaURA3 MATalpha horrhisG leu2rrhisG		versions of
	ura3(\DeltaSma-Pst) his4-X::LEU2-		SBY6068
	(NBam)-URA3 nup2(51-175)*-		
00074	GFP::CaURA3	01/4	
SBY6074	MATA NO::NISG IEU2::NISG ura3(ASma-Pst) HIS4I FLI2-	SKI	I able 1, Figure 1D (sporulation
	$(NBam) nup2(\Delta 1-720)$ -		localization)
	GFP::CaURA3		
	MATalpha ho::hisG leu2::hisG		
	$ura3(\Delta Sma-Pst)$ his4-X::LEU2-		
	GFP::CaURA3		
SBY6071	MATa ho::hisG leu2::hisG	SK1	Table 1, Figure 1D
	ura3(∆Sma-Pst) HIS4::LEU2-		(sporulation,
	(NBam) nup2(71-130)-		localization)
	MATalpha ho::hisG leu2::hisG	-	
	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup2(71-130)-		
SBV6262	GFP::CaURA3	SK1	Toble 1 Figure 1D
3010203	ura3(/Sma-Pst) HIS4::LEU2-	SKI	(sporulation.
	(NBam) nup2(89-130)-		localization)
	GFP::CaURA3	-	
	MATalpha ho::hisG leu2::hisG		
	(NBam)-URA3 nup2(89-130)-		
	GFP::CaURA3		
SBY3943	MATalpha ho::hisG leu2::hisG	SK1	Table1, Figure S1
X SBV6111	ura3(\DeltaSma-Pst) his4-X::LEU2-		(hemizygote
0010111	(INBall)-ORAS hupzRahimx		NDJ1 background,
	MATa ho::hisG leu2::hisG		localization)
	ura3(<i>Δ</i> Sma-Pst) HIS4::LEU2-		
	GFP::CaURA3		
SBY3983	MATalpha ho::hisG leu2::hisG	SK1	Table1, Figure S1
X	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-		(hemizygote
SB16084	(NBam)-URA3 nup2::KanMx		sporulation in

	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{F120Y} - GFP::CaURA3		NDJ1 background, localization)	
SBY3983 x SBY6261	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2::KanMx	SK1	Table1, Figure S1 (hemizygote sporulation in <i>NDJ1</i> background,	
	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup2(89-130)- GFP::CaURA3		localization)	
SBY3983 x SBY6535	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2::KanMx	SK1	Table1, Figure S1 (hemizygote sporulation in <i>NDJ1</i> background,	
	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(89-175)- GFP::CaURA3		localization)	
AH109	MATa, trp1–901, leu2–3, 112, ura3–52, his3–200, gal4Δ, gal80Δ, LYS2:: GAL1UAS- GAL1TATA-HIS3, GAL2UAS- GAL2TATA-ADE2, URA3:: MEL1UAS-MEL1TATA-lacZ.		Figures 2, S2 (Y2H)	Clontech
SBY6054	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175)- GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51-175)- GFP::CaURA3 ndt80::Hph	SK1	Figures 3A, B (chromosome spreads) wt control	
SBY6119	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{K101E} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{K101E} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A (chromosome spreads) K101E version of SBY6054	
SBY6107	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{F120Y} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{F120Y} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A (chromosome spreads) F120Y version of SBY6054	

SBY6496	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{№97S} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{№97S} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A (chromosome spreads) N97S version of SBY6054	
SBY6499	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{V108D} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{V108D} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (V108D version of SBY6054)	
SBY6502	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{R117W} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{R117W} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (R117W version of SBY6054)	
SBY6490	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{L93P} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{L93P} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (L93P version of SBY6054)	
SBY6493	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175) ^{A102P} - GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51- 175) ^{A102P} -GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (A102P version of SBY6054)	
SBY6101	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(∆1-720)- GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2-	SK1	Figures 3A, B chromosome spreads (GFP control)	

	(NBam)-URA3 nup2(∆1-720)- GFP::CaURA3 ndt80::Hph			
SBY6269	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2(71-130)- GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(71-130)- GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (mar(71-130) version of SBY6054)	
SBY6272	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup2(89-130)- GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(89-130)- GFP::CaURA3 ndt80::Hph	SK1	Figure 3A chromosome spreads (mar(89-130) version of SBY6054)	
SBY6278	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup2(51-175)- GFP::CaURA3 ndt80::Hph nup60::KanMx MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2(51-175)- GFP::CaURA3 ndt80::Hph nup60::KanMx	SK1	Figure 3B chromosome spreads (<i>nup60∆</i> version of SBY6054)	
SBY5217	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60::Hph MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60::Hph	SK1	Figures 4, S4 (growth, sporulation, HU sensitivity)	(Chu <i>et al.</i> 2017)
SBY6420	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(189-388) MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189-388)	SK1	Figures 4, S4, 5 (growth, sporulation, HU sensitivity, meiotic time course)	
SBY6423	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(1-388) MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(1-388)	SK1	Figures 4, S4 (growth, sporulation, HU sensitivity)	
SBY6426	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(189-539)	SK1	Figures 4, S4 (growth,	

	MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189-539)		sporulation, HU sensitivity)	
SBY6429	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(⊿189-388) MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(⊿189- 388)	SK1	Figures 4, S4 (growth, sporulation, HU sensitivity)	
SBY1904	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx	SK1	Figure 4 (sporulation)	(Chu <i>et al.</i> 2017)
SBY5222	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60::Hph	SK1	Figure 4 (sporulation)	
SBY6293	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(189-388) MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(189-388)	SK1	Figures 4, 5 (sporulation, meiotic time courses)	
SBY6296	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(1- 388) MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(1-388)	SK1	Figure 4 (sporulation)	
SBY6299	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(189-539) MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(189-539)	SK1	Figure 4 (sporulation)	
SBY6302	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(⊿189-388)	SK1	Figure 4 (sporulation)	

	MATalpha ho∷hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(⊿189-388)			
SBY6304	MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- GFP::KanMx	SK1	Figure S5A (GFP fusion growth)	
SBY6307	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(∆1-539)- GFP::KanMx	SK1	Figure S5A (GFP fusion growth)	
SBY6310	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189- 388)-GFP::KanMx	SK1	Figure S5A (GFP fusion growth)	
SBY6313	MATa ho::hisG leu2::hisG MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(1-388)- GFP::KanMx	SK1	Figure S5A (GFP fusion growth)	
SBY6316	MATalpha ho∷hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189- 539)-GFP::KanMx	SK1	Figure S5A (GFP fusion growth)	

SBY6382	MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(∆189- 388)-GFP::KanMx	SK1	Figure S5A (GFP fusion growth)
SBY6320	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx NUP60- GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx NUP60-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)
SBY6323	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(∆1-539)-GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(∆1-539)-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)
SBY6326	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(189-388)-GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(189-388)-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)
SBY6329	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(1- 388)-GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(1-388)-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)
SBY6332	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(189-539)-GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(189-539)-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)
SBY6335	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(∆189-388)-GFP::KanMx	SK1	Figure S5B (GFP fusion sporulation)

	MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(∆189-388)-GFP::KanMx			
SBY3945	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2::KanMx	SK1	Figure 5 (Meiotic time courses)	(Chu <i>et al.</i> 2017)
SBY6432	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2::KanMx nup60(189-388) MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2::KanMx nup60(189-388)	SK1	Figure 5, Table 2 (Meiotic time courses, <i>nup60</i> epistasis)	
SBY3983	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup2::KanMx ndj1::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup2::KanMx ndj1::KanMx	SK1	Figure 5, Table 2 (Meiotic time courses, <i>nup60</i> epistasis)	(Chu <i>et al.</i> 2017)
SBY6296	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup60(189-388) MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup60(189-388)	SK1	Figure 5, Table 2 (Meiotic time courses, <i>nup60</i> epistasis)	
SBY6254	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) NUP60-GFP::KanMx nup2(51-175)-mCherry::Nat ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph	SK1	Figures 6 and 7 (NUP60-GFP localization and spreads)	
SBY6338	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(⊿1-539)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph	SK1	Figures 6 and 7 (NUP60-GFP localization and spreads)	

	MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(∆1-539)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph			
SBY6344	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) nup60(1-388)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(1-388)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph	SK1	Figures 6 and 7 (NUP60-GFP localization and spreads)	
SBY6341	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup60(189-388)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189- 388)-GFP::KanMx nup2(51- 175)-mCherry::Nat ndt80::Hph	SK1	Figures 6 and 7 (NUP60-GFP localization and spreads)	
SBY6347	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup60(189-539)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(189- 539)-GFP::KanMx nup2(51- 175)-mCherry::Nat ndt80::Hph	SK1	Figures 6 and 7 (NUP60-GFP localization and spreads)	
SBY6350	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) nup60(∆189-388)- GFP::KanMx nup2(51-175)- mCherry::Nat ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 nup60(∆189- 388)-GFP::KanMx nup2(51- 175)-mCherry::Nat ndt80::Hph	SK1	Figure 6 (NUP60-GFP localization)	
SBY4959	MATa ho::hisG leu2::hisG ura3(DSma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2::Nat MATalpha ho::hisG leu2::hisG ura3(DSma-Pst) his4-X::LEU2-	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	

	(NBam)-URA3 ndj1::KanMx nup2::Nat			
SBY6365	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2::Nat NUP60- GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2::Nat NUP60- GFP::KanMx	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	
SBY6371	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2::Nat nup60(1-388)- GFP::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2::Nat nup60(1-388)- GFP::KanMx	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	
SBY6356	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2::Nat nup60(1-388)MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2::Nat nup60(1-388)	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	
SBY6436	MATa ho::hisG leu2::hisGura3(∆Sma-Pst) HIS4::LEU2-(NBam) ndj1::KanMxnup2::Nat NUP60-mCherry::KanMxMATalpha ho::hisG leu2::hisGura3(∆Sma-Pst) his4-X::LEU2-(NBam)-URA3 ndj1::KanMxnup2::Nat NUP60-mCherry::KanMx	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	
SBY6439	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) ndj1::KanMx nup2::Nat NUP60- 13myc::KanMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 ndj1::KanMx nup2::Nat NUP60- 13myc::KanMx	SK1	Table 3 (Suppression of <i>nup2 ndj1</i>)	

SBY3982 x	MATa ho::hisG leu2::hisG	SK1	Table 3	
SBY6364	ura3(⊿Sma-Pst) HIS4::LEU2-		(Suppression of	
	(NBam) ndj1::KanMx		nup2 ndj1)	
	nup2::Nat			
	MATalpha ho::hisG leu2::hisG			
	ura3(<i>ASma-Pst</i>) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
	nup2::Nat NUP60-			
	GFP::KanMxB	01//	T 1 1 0	
SBY3982 x	MATa ho::hisG leu2::hisG	SK1	l able 3	
5610435	Ura3(ASma-Pst) HIS4::LEU2-		(Suppression of	
	(INBarri) naj L.:Nanivix		nupz nuj r)	
	MATalpha horrhisG leu2rrhisG	-		
	$ura3(\Lambda Sma_Pst)$ his Λ -X··I FI/2-			
	(NBam)-I IRA3 ndi1KanMy			
	nup2::Nat NUP60-			
	mCherry::KanMx			
SBY3982 x	MATa ho::hisG leu2::hisG	SK1	Table 3	
SBY6370	ura3(⊿Sma-Pst) HIS4::LEU2-		(Suppression of	
	(NBam) ndj1::KanMx		nup2 ndj1)	
	nup2::Nat	-		
	MATalpha ho::hisG leu2::hisG			
	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
	nup2:://at/nup60(1-388)-			
SBV3082 v	MATa horrhisC lou2rthisC	SK1	Table 3	
SBY6355	μ	SKI	(Suppression of	
0010000	(NBam) ndi1KanMx		nun2 ndi1)	
	nup2::Nat			
	MATalpha ho::hisG leu2::hisG	-		
	ura3(△Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 ndj1::KanMx			
	nup2::Nat nup60(1-388)			
SBY6484	MATa ho::hisG leu2::hisG	SK1	Figure 8	
	ura3(<i>∆</i> Sma-Pst) HIS4::LEU2-		(Nup60-GFP	
	(NBam) NUP60-GFP::KanMx		binding with and	
	ndt80::Hph	-	without Nup2)	
	MA l alpha ho::hisG leu2::hisG			
	$ura3(\Delta Sma-Pst)$ nis4-X::LEU2-			
	CEP:KanMy ndtROUnb			
SBY6487	MATa horhisG leu2rhisG	SK1	Figure 8	
	$\mu ra3(\Lambda Sma-Pst)$ HIS4. I FU2-		(Nup60-GFP	
	(NBam) NUP60-GFP::KanMx		binding with and	
	nup2::NatMx ndt80::Hph		without Nup2)	
	MATalpha ho::hisG leu2::hisG			
	ura3(^A Sma-Pst) his4-X::LEU2-			
	(NBam)-URA3 NUP60-			
	GFP::KanMx nup2::NatMx			
	ndt80::Hph			

	1		
SBY6505	MATa ho::hisG leu2::hisG	SK1	Figure 8
	ura3(∆Sma-Pst) HIS4::LEU2-		(Nup60-GFP
	(NBam) nup60(189-388)-		binding with and
	GFP::KanMx ndt80::Hph		without Nup2)
	MATalpha ho::hisG leu2::hisG	-	
	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup60(189-		
	388)-GFP::KanMx ndt80::Hph		
SBY6442	MATa horhisG leu2rhisG	SK1	Table 2
0010112	$\mu ra3(\Lambda Sma-Pst)$ HISA. I FI 12-	U.C.	(nun60 epistasis)
	(NBam) csm4··Hoh		(napoe opieracie)
	MATalpha ho::bisG leu2::bisG		
	$\mu ra2(4 \text{Sm}_2 \text{-} \text{Pet})$ his $4 \text{-} \text{Y} \cdot 1 \text{-} \text{E} 12$		
	(NPam) IIPA2 com 4: Uph		
	MATe heybiaC lou2ubiaC	<u> </u>	
SD10440		SNI	
	ura3(ASma-Pst) HIS4::LEU2-		(nupou epistasis)
	(NBam) nup60(189-388)		
		-	
	MATalpha no::nisG leu2::nisG		
	ura3(<i>A</i> Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup60(189-388)		
	csm4::Hph		
SBY6445	MATa ho::hisG leu2::hisG	SK1	Table 2
	ura3(⊿Sma-Pst) HIS4::LEU2-		(<i>nup60</i> epistasis)
	(NBam) nup60::Hph		
	csm4::Hph	_	
	MATalpha ho::hisG leu2::hisG		
	ura3(⊿Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup60::Hph		
	csm4::Hph		
SBY6451	MATa ho::hisG leu2::hisG	SK1	Table 2
	ura3(⊿Sma-Pst) HIS4::LEU2-		(<i>nup60</i> epistasis)
	(NBam) nup2::NatMx		
	csm4::Hph		
	MATalpha ho::hisG leu2::hisG		
	ura3(^A Sma-Pst) his4-X::LEU2-		
	(NBam)-URA3 nup2::NatMx		
	csm4::Hph		
SBY6353	MATa ho::hisG leu2::hisG	SK1	Table 2
	ura3(⊿Sma-Pst) HIS4::LEU2-		(nup60 epistasis)
	(NBam) nup60(189-388)		
	nup2::KanMx ndi1::KanMx		
	MATalpha ho::hisG leu2::hisG	1	
	ura3(ASma-Pst) his4-X··I FII2-		
	(NBam)-URA3 nun60(189-388)		
	nup2::KanMx ndi1::KanMx		
SBY6457	MATa horrhisG leu2rrhisG	SK1	Figure S8A
	$\mu ra3(\Lambda Sma-Pst)$ HISA···I FI 12-		(HA-Nup60
	(NBam) 3HΔ-NI IP60		sporulation
	MaTalnha howhis Cloud whis C	-	
	ura2/ASma_Det his A V. I EU2		
	(NIPam) LIPA2 24A MUDEO		
	INDAIN/OKAS SHA-NUPOU		

SBY6460	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) 3HA-NUP60 ndj1::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 3HA-NUP60 ndj1::Hph	SK1	Figure S8A (HA-Nup60 sporulation)
SBY6463	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) 3HA-NUP60 nup2::NatMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 3HA-NUP60 nup2::NatMx	SK1	Figure S8A (HA-Nup60 sporulation)
SBY6466	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) 3HA-NUP60 ndj1::Hph nup2::NatMx MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 3HA-NUP60 ndj1::Hph nup2::NatMx	SK1	Figure S8A (HA-Nup60 sporulation)
SBY6469	MATa ho::hisG leu2::hisG ura3(ΔSma-Pst) HIS4::LEU2- (NBam) NUP60- 3HA(Δ1F)::KanMx MATalpha ho::hisG leu2::hisG ura3(ΔSma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- 3HA(Δ1F)::KanMx	SK1	Figure S8B (Nup60-HA western)
SBY6472	MATa ho::hisG leu2::hisG ura3(ΔSma-Pst) HIS4::LEU2- (NBam) NUP60- 3HA(Δ1F)::KanMx ndj1::Hph MATalpha ho::hisG leu2::hisG ura3(ΔSma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- 3HA(Δ1F)::KanMx ndj1::Hph	SK1	Figure S8B (Nup60-HA western)
SBY6475	MATa ho::hisG leu2::hisG ura3(ΔSma-Pst) HIS4::LEU2- (NBam) NUP60- 3HA(Δ1F)::KanMx nup2::NatMx MATalpha ho::hisG leu2::hisG ura3(ΔSma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- 3HA(Δ1F)::KanMx nup2::NatMx	SK1	Figure S8B (Nup60-HA western)
SBY6478	MATa ho::hisG leu2::hisG ura3(⊿Sma-Pst) HIS4::LEU2- (NBam) NUP60-	SK1	Figure S8B (Nup60-HA western)

	3HA(∆1F)::KanMx nup2::NatMx ndj1::Hph			
	MATalpha ho::hisG leu2::hisG ura3(⊿Sma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- 3HA(∆1F)::KanMx nup2::NatMx ndj1::Hph			
SBY6481	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) NUP60- mCherry::KanMx nup2(51- 175)-GFP::CaURA3 ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- mCherry::KanMx nup2(51- 175)-GFP::CaURA3 ndt80::Hph	SK1	Figure S9A (Nup60-mCherry spreads)	
SBY6410	MATa ho::hisG leu2::hisG ura3(∆Sma-Pst) HIS4::LEU2- (NBam) NUP60-3HA::KanMx nup2(51-175)-mCherry::NatMx ndt80::Hph MATalpha ho::hisG leu2::hisG ura3(∆Sma-Pst) his4-X::LEU2- (NBam)-URA3 NUP60- 3HA::KanMx nup2(51-175)- mCherry::NatMx ndt80::Hph	SK1	Figure S9B (Nup60-3HA spreads)	

Table S2. Plasmids used in this study

Plasmid name	Description	Source/ Reference
pCR-BluntII- TOPO	cloning vector	Thermo Fisher/Invitrogen
pSB740	MAR-GFP::URA3 subcloned into pCR-Blunt II-TOPO	
pGBKT7	Gal4 DBD vector	Clontech
pACT2-2	Gal4 AD vector	(Arora <i>et al.</i> 2004)
pSB655	SK1 Nup2(51-175) coding sequence subcloned into pGBKT7	
pSB745	Nup2(51-175) ^{F100S} coding sequence subcloned into pGBKT7	
pSB746	Nup2(51-175) ^{K101E} coding sequence subcloned into pGBKT7	
pSB747	Nup2(51-175) ^{R117W} coding sequence subcloned into pGBKT7	
pSB748	Nup2(51-175) ^{L98P} coding sequence subcloned into pGBKT7	
pSB749	Nup2(51-175) ^{A102P} coding sequence subcloned into pGBKT7	
pSB750	Nup2(51-175) ^{L119P} coding sequence subcloned into pGBKT7	
pSB751	Nup2(51-175) ^{Y123C} coding sequence subcloned into pGBKT7	
pSB752	Nup2(51-175) ^{D115V} coding sequence subcloned into pGBKT7	
pSB753	Nup2(51-175) ^{L93P} coding sequence subcloned into pGBKT7	
pSB754	Nup2(51-175) ^{L96S} coding sequence subcloned into pGBKT7	
pSB755	Nup2(51-175) ^{N97S} coding sequence subcloned into pGBKT7	
pSB756	Nup2(51-175) ^{V108D} coding sequence subcloned into pGBKT7	
pSB757	Nup2(51-175) ^{F120Y} coding sequence subcloned into pGBKT7	
pSB758	Nup2(51-175) ^{Y126N} coding sequence subcloned into pGBKT7	
pSB759	Nup2(51-175) ^{L116S} coding sequence subcloned into pGBKT7	
pSB743	Nup2(71-130) coding sequence subcloned into pGBKT7	
pSB744	Nup2(89-130) coding sequence subcloned into pGBKT7	
pSB741	SK1 Nup60 coding sequence subcloned into pACT2-2	
pSB742	SK1 Nup60(188-388) coding sequence subcloned into pACT2-2	

Primer name	Sequence	Description
oSB1274	GGGTTCGACTCCCCGTATC	forward primer for amplifying the full MAR-GFP::URA3 fragment
		MAR mutagenesis: forward primer for amplifying the 5' fragment containing the MAR
oSB1774	TGCGCGATGTTTTAACGAAG	reverse primer for amplifying the full MAR-GFP::URA3 fragment
		MAR mutagenesis: reverse primer for amplifying the 3' fragment containing GFP-URA3
oSB1918	TAAACCAGCACCGTCACCC	MAR mutagenesis: reverse primer for amplifying the 5' fragment containing MAR
oSB1919	GGGTGACGGTGCTGGTTTA	MAR mutagenesis: forward primer for amplifying the 3' fragment containing GFP-URA3
oSB1616	ACTGGAATTCATGAAACCTTTTGGTTCTGCAA	Y2H: forward primer for insertion of Nup2(51-175) coding sequence into pGBKT7
oSB1617	ACTGGGATCCTTACTTGGGTCCCTCCACCTTAACC	Y2H: reverse primer for insertion of Nup2(51-175) coding sequence into pGBKT7
oSB1920	ACTGCAACTAGTATGCATCGTAAATCATTGAGGAGGGCTA G	Y2H: forward primer for insertion of Nup60 coding sequence into pACT2-2
oSB1921	TAAGCACCATGGCAAAGGTATATAGGGACTTGAAAGCCTC AAC	Y2H: reverse primer for insertion of Nup60 coding sequence into pACT2-2
oSB1922	ACTGCAACTAGTATGCCGACCTTCAACCCAAAATATGATAC TTCAAATG	Y2H: forward primer for insertion of Nup60(188-388)

 Table S3. Primers for PCR mutagenesis and Y2H plasmid construction

		coding sequence into pACT2-2
oSB1923	TAAGCACCATGGCACCATCCTTCTTTCAGGTGAGGTTTCA G	Y2H: reverse primer for insertion of Nup60(188-388) coding sequence into pACT2-2
oSB1924	ACTGGAATTCATGAACCGGGCGGACGGCACTG	Y2H: forward primer for insertion of Nup2(71-130) coding sequence into pGBKT7
oSB1925	ACTGGAATTCATGAGCAATTCCAGACTAAAAGCATTGAACC	Y2H: forward primer for insertion of Nup2(89-130) coding sequence into pGBKT7
oSB1926	ACTGGGATCCTTAGATATTCTTTATGTATAATTCGTACCTG GTG	Y2H: reverse primer for insertion of Nup2(71-130) and Nup2(89-130) coding sequences into pGBKT7

Supplemental methods

Western blot

Yeast extracts were prepared by alkaline lysis (von der Haar 2007) and resolved on a 4-15% Mini-PROTEAN TGX gel (BioRad, 4561085). Gel electrophoresis and wet transfer to PVDF were carried out according to standard procedures (Bolt and Mahoney 1997). Western blot detection was performed using ECL reagents and the manufacturer's instructions for PBS-based buffers (Amersham, RPN2108). Membranes were imaged with an ImageQuant LAS4000.

Antibodies

Primary antibodies used were mouse monoclonal antibody to HA (Santa Cruz Biotechnology SC-7392; RRID:AB_627809), rabbit polyclonal antibody to β-tubulin (Abcam AB15568; RRID:AB_2210952), chicken polyclonal antibody to GFP (Novus NB100-1614; RRID:AB_10001164), and rabbit polyclonal antibody to mCherry (Novus NBP2-25157; RRID:AB_2753204). The secondary antibodies used were goat anti-chicken 488 (Thermofisher A11039; RRID:AB_2534096), goat anti-rabbit 594 (ThermofisherA11012; RRID:AB_2534079), donkey anti-mouse 488 (Invitrogen A32766; RRID:AB_2762823), and the anti-rabbit and anti-mouse HRP-conjugated secondary antibodies provided in the ECL kit. All antibodies were used at a 1:1000 dilution.

- Arora C., K. Kee, S. Maleki, and S. Keeney, 2004 Antiviral protein Ski8 is a direct partner of Spo11 in meiotic DNA break formation, independent of its cytoplasmic role in RNA metabolism. Mol. Cell 13: 549–559.
- Bolt M. W., and P. A. Mahoney, 1997 High-efficiency blotting of proteins of diverse sizes following sodium dodecyl sulfate-polyacrylamide gel electrophoresis. Anal. Biochem. 247: 185–192.
- Chu D. B., T. Gromova, T. A. C. Newman, and S. M. Burgess, 2017 The Nucleoporin Nup2 Contains a Meiotic-Autonomous Region that Promotes the Dynamic Chromosome Events of Meiosis.
 Genetics 206: 1319–1337.
- Haar T. von der, 2007 Optimized protein extraction for quantitative proteomics of yeasts. PLoS One 2: e1078.
- Holzer G., P. De Magistris, C. Gramminger, R. Sachdev, A. Magalska, *et al.*, 2021 The nucleoporin Nup50 activates the Ran guanine nucleotide exchange factor RCC1 to promote NPC assembly at the end of mitosis. EMBO J. 40: e108788.
- Jumper J., R. Evans, A. Pritzel, T. Green, M. Figurnov, *et al.*, 2021 Highly accurate protein structure prediction with AlphaFold. Nature 596: 583–589.
- Mariani V., M. Biasini, A. Barbato, and T. Schwede, 2013 IDDT: a local superposition-free score for comparing protein structures and models using distance difference tests. Bioinformatics 29: 2722–2728.
- Pettersen E. F., T. D. Goddard, C. C. Huang, E. C. Meng, G. S. Couch, *et al.*, 2021 UCSF ChimeraX: Structure visualization for researchers, educators, and developers. Protein Sci. 30: 70–82.

Saiz-Baggetto S., E. Méndez, I. Quilis, J. C. Igual, and M. C. Bañó, 2017 Chimeric proteins tagged with specific 3xHA cassettes may present instability and functional problems. PLoS One 12: e0183067.