

Supplementary Materials for

Interactive effects of climate and land use on pollinator diversity differ among taxa and scales

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Supplementary Text

Effectivity of Malaise traps for catching flower-visiting insects

Apart from estimating the proportion of pollinator taxa caught with Malaise traps (see Results), we compared the richness obtained with our Malaise traps with other studies from Bavaria that used methods more targeted to specific insect groups.

For instance, one study caught a total of 87 bee species in grasslands in and in the vicinity of the National Park Berchtesgaden with sweep netting, finding between 6 and 24 species per site (**81**). In our only grassland plot in the vicinity of Berchtesgaden, we found 38 wild bee species: *Andrena carantonica*, *A. fucata*, *A. minutula*, *A. ovatula*, *B. campestris*, *B. hortorum*, *B. pascuorum*, *B. sylvestris*, *Hylaeus communis*, *H. gredleri*, *H. punctatus*, *H. sinuatus*, *Halictus rubicundus*, *H. tumulorum*, *Lasioglossum calceatum*, *L. fulvicorne*, *L. laticeps*, *L. leucopus*, *L. leucozonium*, *L. morio*, *L. nitidulum*, *L. pauxillum*, *L. villosulum*, *L. zonulum*, *Sphecodes crassus*, *S. gibbus*, *S. hyalinatus*, *S. longulus*, *S. monilicornis*, *S. pellucidus*, *S. reticulatus*, *S. scabriocollis*, *Anthidiellum strigatum*, *Anthidium oblongatum*, *Hoplitis leucomelana*, *Megachile centuncularis*, *M. ligniseca*, *M. nigriventris*. This high diversity could be attributed to the long exposure of Malaise traps (active all-day long for six weeks), which are more likely to catch less abundant species (**82**).

We also compared our data with a study that used several different methods to analyze 12 taxonomic groups in different agri-environmental schemes and calcareous grasslands in the north-west of Bavaria (**61**). This study found in their 27 plots: 143 bee species (transect walks + pan traps), 91 flower visiting beetles (pan traps), 56 butterfly species and 120 moth species (transect walks + light traps), and 66 syrphids (Malaise traps and DNA metabarcoding). While our study covers a larger area, this previous study captured insects in calcareous grasslands, which are considered biodiversity hotspots in Europe. Since they used the same method to survey the syrphid taxa, we can use the difference in syrphid catches as a baseline to compare both studies. While they got 70% of the number of syrphid taxa we caught, the combination of pan traps and sweep netting used by (**61**) was effective for bees (74% of our catches), while they got 24% of the number of butterflies and moths and 35% of the flower-visiting beetles. This means that, although the combination of pan traps and sweep netting is likely the best to survey wild bees, Malaise traps are also an effective and much more representative method across pollinator taxa.

	Pollinators	Bees	Hymenoptera	Syrphids	Diptera	Butterflies	Moths	Beetles
Species	3218	194	869	95	1060	96	646	258
Genera	1452	31	435	47	459	58	314	108
Families	136	6	34	1	47	10	18	20

Table S1. Total number of species, genera and families found from each pollinator taxon (n samples = 534, n sites = 179). ‘Butterflies’ includes all butterflies plus the moth families in which all species are known to be diurnal (Choreutidae, Micropterigidae, Scythrididae, Sesiidae and Zygaenidae).

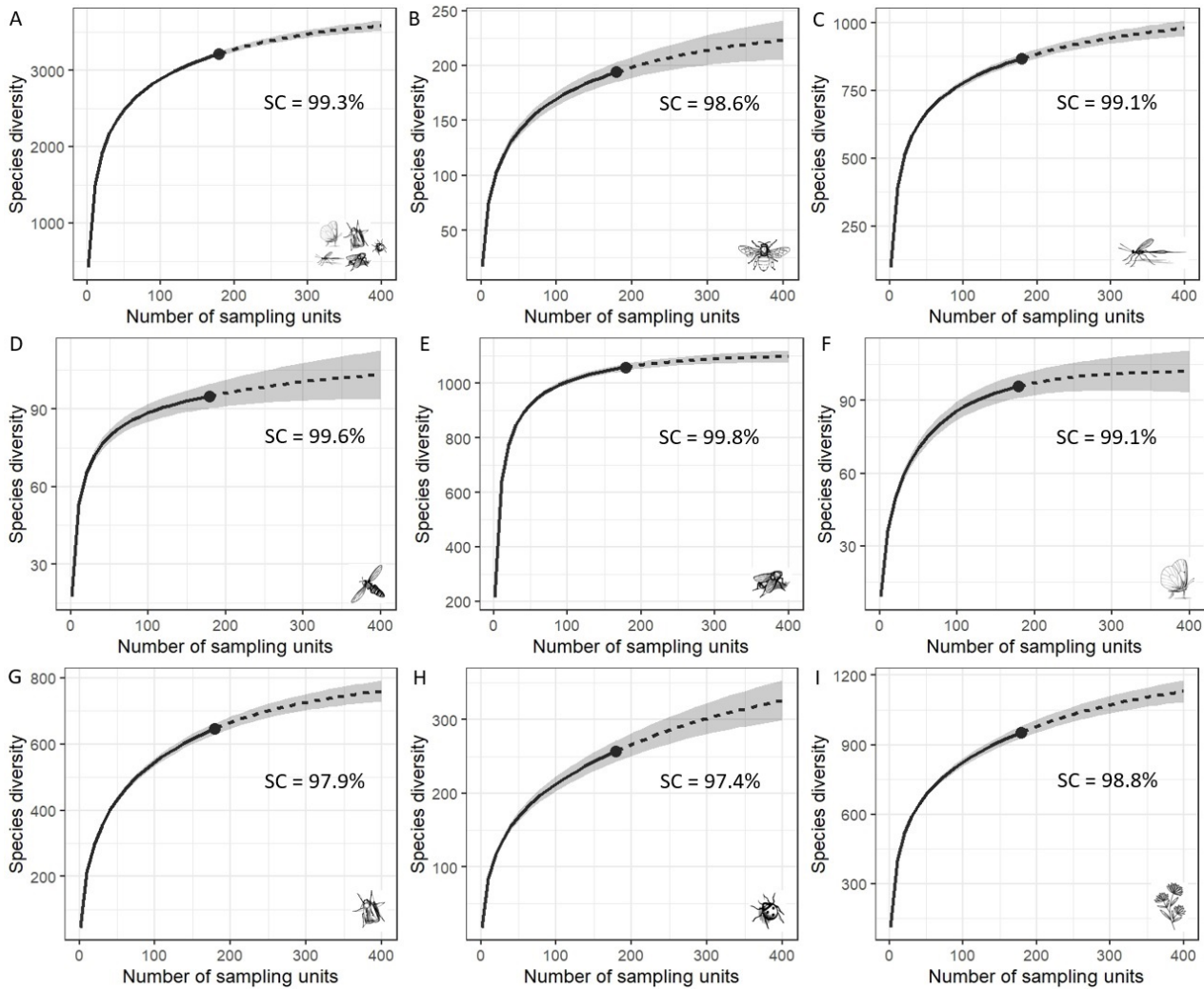


Fig. S1. Rarefaction interpolation curves for the different pollinator communities and flowering plants. We compared diversity estimates for our study plots with rarefaction and extrapolation sampling curves of Hill numbers (i.e. the effective number of species; $q = 0$, species richness) via the iNEXT function (iNEXT package (83)). Continuous lines indicate interpolated data and dashed lines extrapolated data. The sample completeness percentage (SC) is indicated in the panels. The different panels show: (A) whole pollinator community, (B) bees, (C) non-bee Hymenoptera, (D) syrphids, (E) non-syrphid Diptera, (F) butterflies, (G) moths, (H) beetles, (I) flowering plants.

Response: Community composition - whole pollinator community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.573	0.287	1.627	0.051	0.002 **
MAT	1	0.687	0.687	3.904	0.061	0.001 ***
RLU:MAT	2	0.456	0.228	1.296	0.041	0.015 *
Residuals	54	9.507	0.176		0.847	
Total	59	11.224			1.000	

Response: Community composition - bee community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.631	0.315	1.670	0.051	0.043 *
MAT	1	0.747	0.747	3.956	0.060	0.001 ***
RLU:MAT	2	0.819	0.409	2.168	0.066	0.003 **
Residuals	54	10.197	0.189		0.823	
Total	59	12.394			1.000	

Response: Community composition - Hymenoptera community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.606	0.303	1.555	0.049	0.003 **
MAT	1	0.588	0.588	3.016	0.048	0.001 ***
RLU:MAT	2	0.591	0.296	1.516	0.048	0.006 **
Residuals	54	10.529	0.195		0.855	
Total	59	12.314			1.000	

Response: Community composition - syrphid community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.428	0.214	1.645	0.049	0.075 .
MAT	1	0.615	0.615	4.731	0.070	0.001 ***
RLU:MAT	2	0.673	0.337	2.588	0.077	0.008 **
Residuals	54	7.024	0.130		0.804	
Total	59	8.740			1.000	

Response: Community composition - Diptera community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.467	0.233	1.766	0.054	0.004 **
MAT	1	0.615	0.615	4.651	0.071	0.001 ***
RLU:MAT	2	0.396	0.198	1.497	0.046	0.011 *
Residuals	54	7.138	0.132		0.829	
Total	59	8.615			1.000	

Response: Community composition - butterfly community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.491	0.245	1.576	0.048	0.122
MAT	1	0.410	0.410	2.630	0.040	0.022 *
RLU:MAT	2	0.952	0.476	3.057	0.093	0.002 **
Residuals	54	8.411	0.156		0.819	
Total	59	10.264			1.000	

Response: Community composition - moth community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.688	0.344	1.581	0.050	0.016 *
MAT	1	0.746	0.746	3.428	0.054	0.001 ***
RLU:MAT	2	0.699	0.349	1.606	0.050	0.007 **
Residuals	54	11.748	0.218		0.846	
Total	59	13.880			1.000	

Response: Community composition - beetle community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.622	0.311	1.590	0.048	0.040 *
MAT	1	0.877	0.877	4.488	0.068	0.001 ***
RLU:MAT	2	0.841	0.420	2.150	0.065	0.005 **
Residuals	54	10.557	0.196		0.819	
Total	59	12.897			1.000	

Response: Community composition - flowering plant community

	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr(>F)
RLU	2	0.703	0.351	2.115	0.064	0.001 ***
MAT	1	0.700	0.700	4.213	0.064	0.001 ***
RLU:MAT	2	0.601	0.301	1.809	0.055	0.003 **
Residuals	54	8.970	0.166		0.817	
Total	59	10.973			1.000	

Table S2. The community composition of most pollinator taxa and flowering-plant species changed significantly across land-use intensity and temperature gradients, and with the interaction between both gradients. Shown are results of permutational multivariate analyses of variance (PERMANOVA), performed with the adonis function (vegan package (72)), using Jaccard distances and the standard number of permutations (n = 999).

Response: Distances (whole pollinator community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.00	0.00	0.43	0.652
Residuals	57	0.04	0.00		

Response: Distances (bee community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.03	0.02	1.35	0.268
Residuals	57	0.64	0.01		

Response: Distances (non-bee Hymenoptera community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.00	0.00	0.14	0.866
Residuals	57	0.11	0.00		

Response: Distances (syrphid community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.03	0.01	1.22	0.301
Residuals	57	0.61	0.01		

Response: Distances (non-syrphid Diptera community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.00	0.00	0.03	0.970
Residuals	57	0.06	0.00		

Response: Distances (butterfly community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.02	0.01	0.75	0.476
Residuals	57	0.97	0.02		

Response: Distances (moth community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.00	0.00	0.06	0.944
Residuals	57	0.19	0.00		

Response: Distances (beetle community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.02	0.01	0.89	0.416
Residuals	57	0.61	0.01		

Response: Distances (flowering plant community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	2	0.00	0.00	0.82	0.446
Residuals	57	0.11	0.00		

Table S3. Different regional land-use types have a similar compositional variance of the different pollinator taxa. Shown are ANOVA tables displaying the non-significant results of the betadisper function models for the three regional land uses (semi-natural, agricultural, urban).

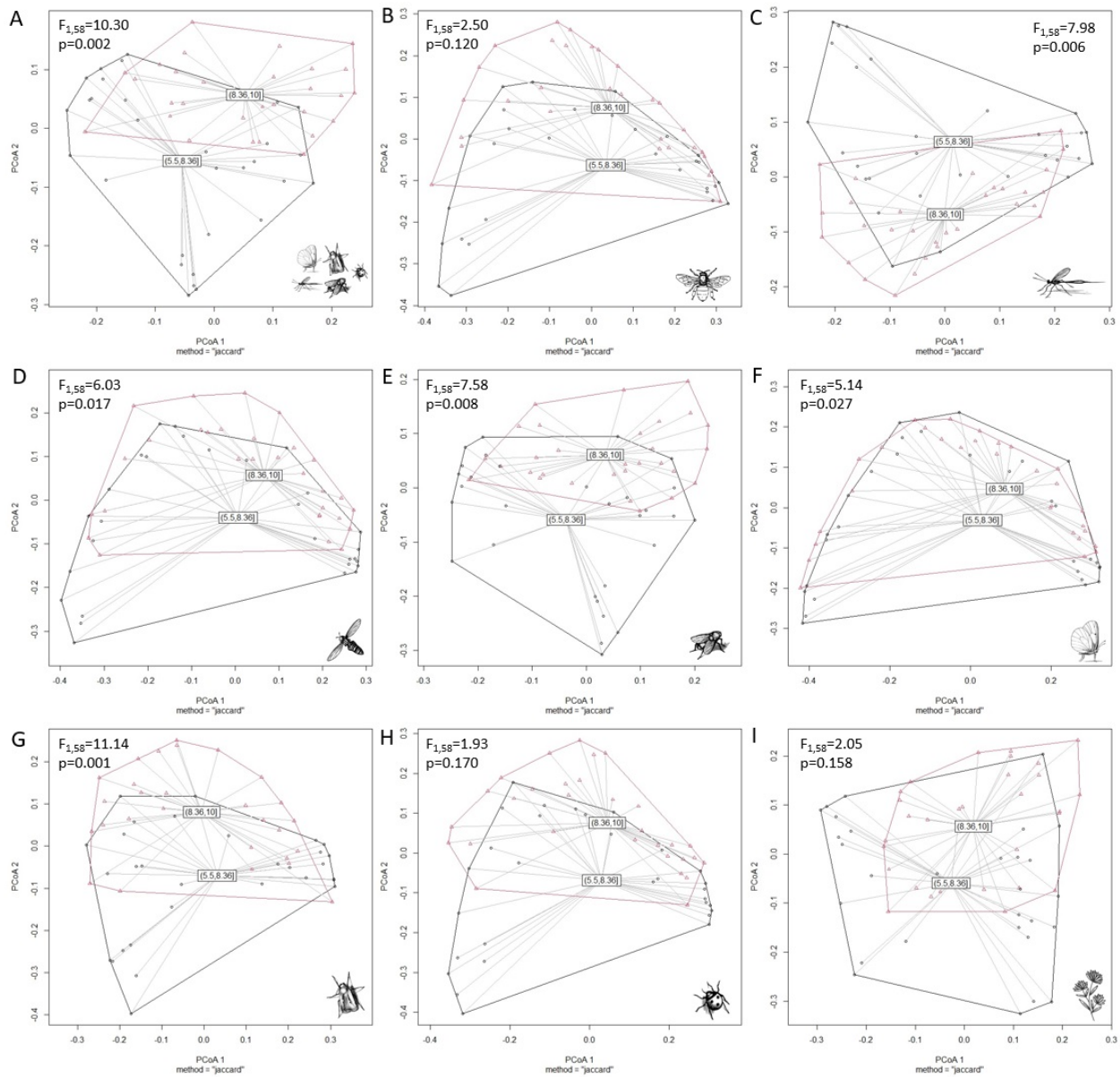


Fig. S2. Difference in compositional variance (inter-regional dissimilarity) between cooler and warmer regions for the different pollinator taxa. Ordinations (NMDS, Jaccard's method) show the homogeneity of dispersion of the different pollinator taxa in cooler (multi-annual mean temperature between 5.6 and 8.4 °C) versus warmer (multi-annual mean temperature between 8.4 and 9.8 °C) regions. The position of regions (dots) in the NMDS space represents the similarity in pollinator community composition in relation to other regions: the closer the dots, the higher the proportion of species shared. Rectangles represent centroids of the two temperature regimes; polygons delimit the NMDS space occupied by regions with the same temperature category. The different panels show: **(A)** whole pollinator community, **(B)** bees, **(C)** non-bee Hymenoptera, **(D)** syrphids, **(E)** non-syrphid Diptera, **(F)** butterflies, **(G)** moths, **(H)** beetles, **(I)** flowering plants. Results from betadisper function models (F and p values from ANOVA) indicating whether the compositional variance of the different pollinator taxa is significantly different between temperature regimes are indicated in the upper corner of each panel.

Response: Distances (whole pollinator community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.01	0.00	1.79	0.131
Residuals	54	0.03	0.00		

Response: Distances (bee community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.07	0.01	1.26	0.296
Residuals	54	0.61	0.01		

Response: Distances (non-bee Hymenoptera community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.02	0.00	1.97	0.098 .
Residuals	54	0.12	0.00		

Response: Distances (syrphid community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.10	0.02	1.65	0.164
Residuals	54	0.64	0.01		

Response: Distances (non-syrphid Diptera community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.01	0.00	1.17	0.337
Residuals	54	0.05	0.00		

Response: Distances (butterfly community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.13	0.03	1.38	0.247
Residuals	54	1.04	0.02		

Response: Distances (moth community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.03	0.01	1.70	0.150
Residuals	54	0.18	0.00		

Response: Distances (beetle community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.05	0.01	0.95	0.455
Residuals	54	0.57	0.01		

Response: Distances (flowering plant community)

	Df	Sum Sq	Mean Sq	F value	Pr (>F)
Groups	5	0.01	0.00	0.77	0.573
Residuals	54	0.10	0.00		

Table S4. The interaction between land-use intensity and temperature does not affect the compositional variance of the different pollinator taxa across regions. Shown are ANOVA tables displaying the non-significant results of the betadisper function models for the interaction between regional land-use types and temperature.

Response: Pollinator Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	6.687	0.013	531.84	0.132	699.0	3.3
For _{Region}	0.030	0.013	2.37			

Response: Bee Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	3.476	0.042	83.73	0.611	451.1	25.5
MAT	0.201	0.044	4.52			
MAP	-0.014	0.048	-0.30			
MAT:MAP	-0.134	0.036	-3.71			

Response: Hymenoptera Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	5.310	0.020	266.41	0.131	588.5	3.2
For _{Region}	0.047	0.020	2.36			

Response: Syrphid Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	3.429	0.023	147.26	0.299	374.7	11.7
MAP	0.083	0.022	3.78			

Response: Diptera Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	5.939	0.012	494.99	0.322	603.8	12.0
MAP	0.048	0.012	3.98			

Response: Butterfly Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	2.815	0.043	66.11	0.395	370.3	7.8
MAT	-0.009	0.044	-0.20			
MAP	-0.097	0.050	-1.93			
For _{Region}	-0.006	0.038	-0.14			
MAT:MAP	-0.180	0.052	-3.43			
MAT:For _{Region}	0.215	0.051	4.24			

Response: Moth Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	4.574	0.023	203.24	0.340	514.1	13.0
MAP	-0.096	0.023	-4.14			

Response: Beetle Gamma diversity

	Estimate	Std. Error	Z value	Pseudo-R ²	AIC	ΔAIC_{null}
(Intercept)	3.644	0.024	152.22	0.307	412.0	9.2
MAT	0.066	0.025	2.66			
For _{Region}	0.079	0.024	3.25			

Table S5. Summary tables of the best gamma diversity models (n = 60). Gamma diversity models were analyzed with generalized linear models (GLMs) with a Poisson or a negative binomial error distribution when Poisson models showed overdispersion. ΔAIC_{null} indicate the difference in AIC between the best model and the null model.

Response: Pollinator Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.707	0.005	156.66	0.418/0.364	-239.3	22.5
MAT	-0.009	0.005	-1.79			
MAP	0.003	0.005	0.50			
For _{Region}	0.009	0.004	2.18			
Flower β	0.017	0.004	4.12			
MAT:MAP	0.019	0.004	4.49			

Response: Bee Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.800	0.009	88.800	0.297/0.259	-153.7	15.1
MAT	-0.017	0.009	-1.83			
Urb _{Region}	0.009	0.013	0.70			
MAT:Urb _{Region}	-0.038	0.011	-3.39			

Response: Hymenoptera Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.775	0.005	152.51	0.243/0.216	-213.0	12.7
MAT	-0.018	0.005	-3.57			
Flower β	0.012	0.005	2.29			

Response: Syrphid Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.628	0.009	70.33	0.205/0.178	-145.3	9.8
MAT	0.025	0.009	2.81			
Flower β	0.023	0.009	2.52			

Response: Diptera Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.639	0.005	123.86	0.376/0.319	-223.1	18.3
MAT	-0.007	0.005	-1.34			
MAP	0.009	0.006	1.53			
For _{Region}	0.011	0.005	2.21			
Flower β	0.020	0.005	4.14			
MAT:MAP	0.016	0.005	3.40			

Response: Butterfly Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.7546	0.013	58.81	0.289/0.251	-100.9	14.5
MAT	-0.038	0.013	-2.85			
Ag _{Region}	-0.036	0.013	-2.72			
Flower β	0.026	0.013	2.00			

Response: Moth Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.788	0.005	145.25	0.467/0.417	-217.1	27.7
MAT	-0.014	0.006	-2.42			
MAP	0.014	0.007	2.00			
Ag _{Region}	0.012	0.006	-2.16			
Flower β	0.018	0.005	3.69			
MAT:MAP	0.013	0.005	2.63			

Response: Beetle Beta diversity

	Estimate	Std. Error	t value	R ² /adj. R ²	AIC	ΔAIC _{null}
(Intercept)	0.792	0.008	103.02	0.299/0.248	-163.5	13.3
MAT	-0.006	0.008	-0.76			
For _{Region}	0.012	0.008	1.46			
Flower β	0.028	0.008	3.62			
MAT:For _{Region}	0.020	0.007	2.77			

Table S6. Summary tables of the best intra-regional beta diversity models (n = 60). Beta diversity models were analyzed with simple linear models (LMs). ΔAIC_{null} indicate the difference in AIC between the best model and the null model.

Response: Pollinator Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	6.204	0.026	237.49	0.335/0.403	2022.1	62.1
HabGra _{Local}	-0.220	0.036	-6.07			
HabAgr _{Local}	-0.329	0.037	-8.93			
HabUrb _{Local}	-0.216	0.047	-4.55			
Flower α	0.063	0.017	3.67			

Response: Bee Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	3.369	0.081	41.46	0.370/0.487	1345.8	59.8
T _{Local}	0.162	0.051	3.21			
HabGra _{Local}	-0.413	0.109	-3.77			
HabAgr _{Local}	-0.636	0.114	-5.57			
HabUrb _{Local}	-0.316	0.162	-1.95			
Urb _{Landsc}	0.154	0.055	2.81			
Flower α	0.173	0.056	3.12			

Response: Hymenoptera Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	4.817	0.040	120.09	0.301/0.399	1653.0	54.8
HabGra _{Local}	-0.362	0.055	-6.61			
HabAgr _{Local}	-0.450	0.056	-8.05			
HabUrb _{Local}	-0.329	0.073	-4.53			
Flower α	0.083	0.027	3.12			

Response: Syrphid Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	2.994	0.040	73.96	0.175/0.254	1072.4	22.8
T _{Local}	-0.059	0.025	-2.33			
HabGra _{Local}	-0.193	0.057	-3.40			
HabAgr _{Local}	-0.235	0.061	-3.87			
HabUrb _{Local}	-0.208	0.075	-2.75			
Flower α	0.105	0.028	3.78			

Response: Diptera Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	5.474	0.023	235.58	0.292/0.329	1755.8	51.2
HabGra _{Local}	-0.141	0.033	-4.28			
HabAgr _{Local}	-0.231	0.038	-6.15			
HabUrb _{Local}	-0.094	0.036	-2.58			
Agr _{Landsc}	-0.034	0.015	-2.34			

Response: Butterfly Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	2.126	0.050	42.84	0.073/0.517	1021.0	14.5
EdgeD _{Landsc}	0.069	0.031	2.27			
Flower α	0.102	0.031	3.26			

Response: Moth Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	4.052	0.045	89.95	0.248/0.396	1419.0	38.8
T _{Local}	0.065	0.028	2.31			
HabGra _{Local}	-0.147	0.065	-2.29			
HabAgr _{Local}	-0.395	0.064	-6.21			
HabUrb _{Local}	-0.315	0.079	-4.01			
Gra _{Landsc}	-0.351	0.161	-2.19			
Flower α	0.062	0.030	2.11			

Response: Beetle Alpha diversity

	Estimate	Std. Error	Z value	Marg./Cond. R ²	AIC	Δ AIC _{null}
(Intercept)	3.078	0.046	67.08	0.187/0.327	1114.7	30.2
HabGra _{Local}	-0.294	0.063	-4.64			

HabAgr _{Local}	-0.386	0.066	-5.86
HabUrb _{Local}	-0.316	0.084	-3.76
Flower α	0.088	0.031	2.81

Table S7. Summary tables of the best alpha diversity models (n = 175). Alpha diversity models were analyzed with generalized linear mixed models (GLMMs, glmmTMB function (73)), with a Poisson or a negative binomial error distribution and the term ‘region’ included as random effect. ΔAIC_{null} indicate the difference in AIC between the best model and the null model.

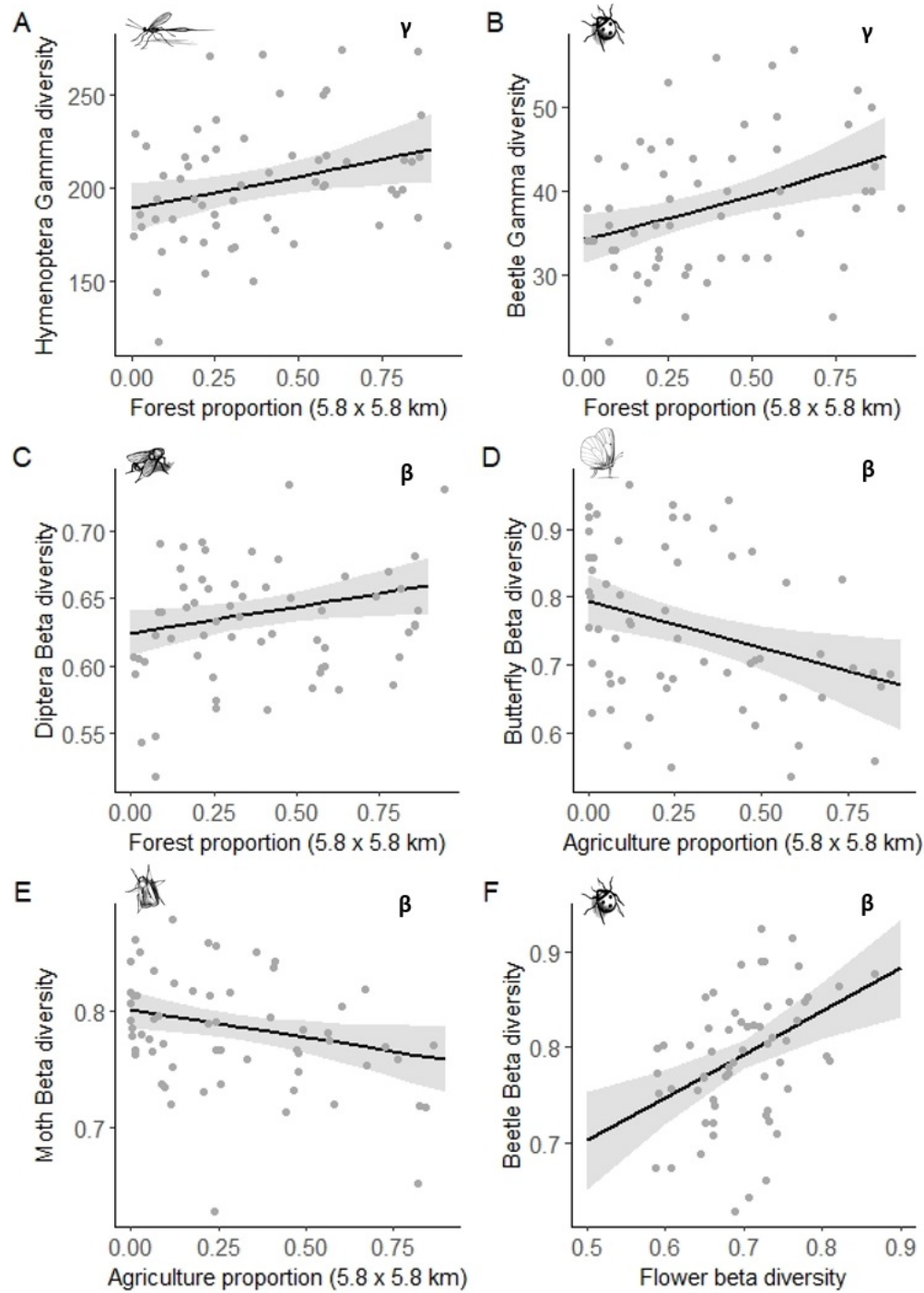


Fig. S3. Response of the gamma and beta diversity of different pollinator taxa to land-use variables. Graphs show predictions of the relationships selected in the best models between gamma diversity ($n = 60$) and (A, B) the proportion of forest in the region, and between beta diversity ($n = 60$) and (C) the proportion of forest and (D, E) agriculture in the region, and (F) flowering-plant beta diversity.

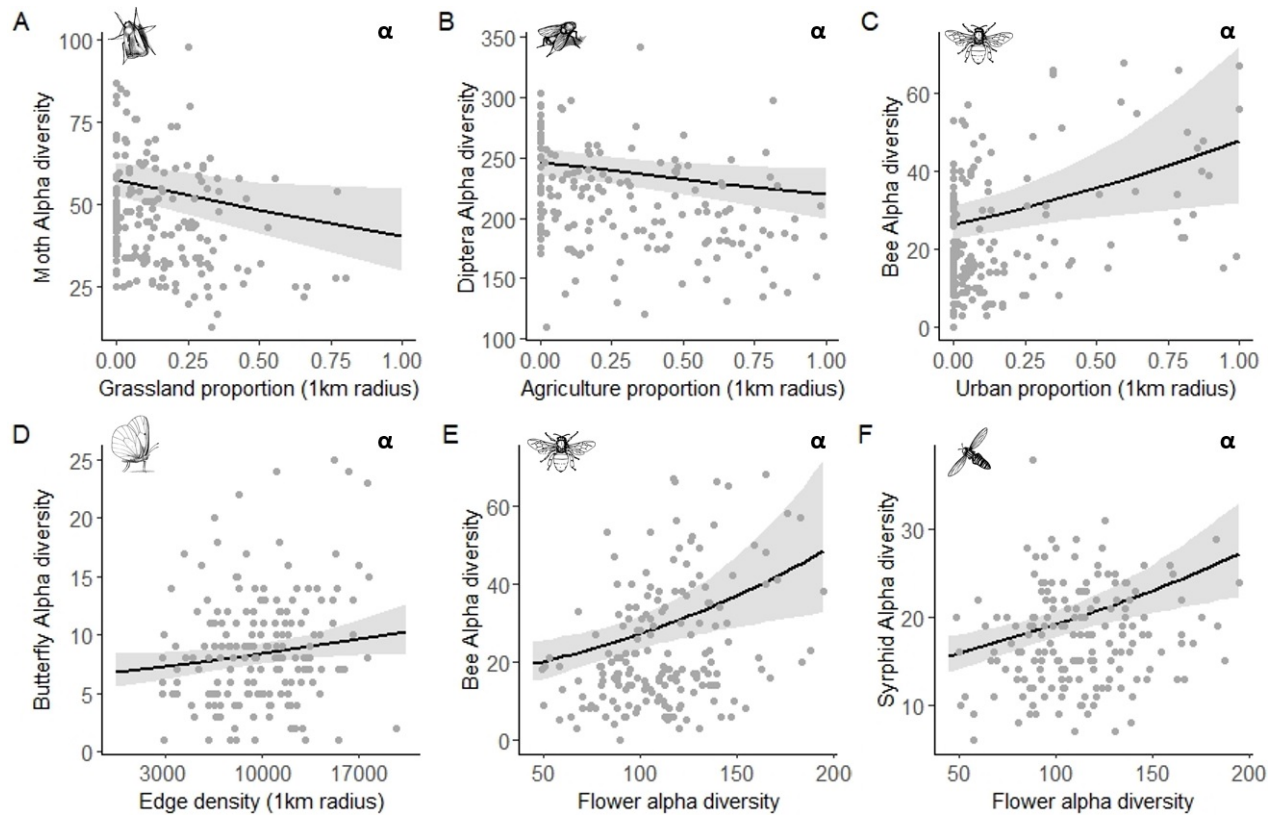


Fig. S4. Response of the alpha diversity of different pollinator taxa to land-use variables. Graphs show predictions of the relationships selected in the best models between alpha diversity ($n = 175$) and **(A)** the proportion of grassland, **(B)** agriculture and **(C)** urban in the landscape, **(D)** edge density, and **(E, F)** flowering-plant alpha diversity.

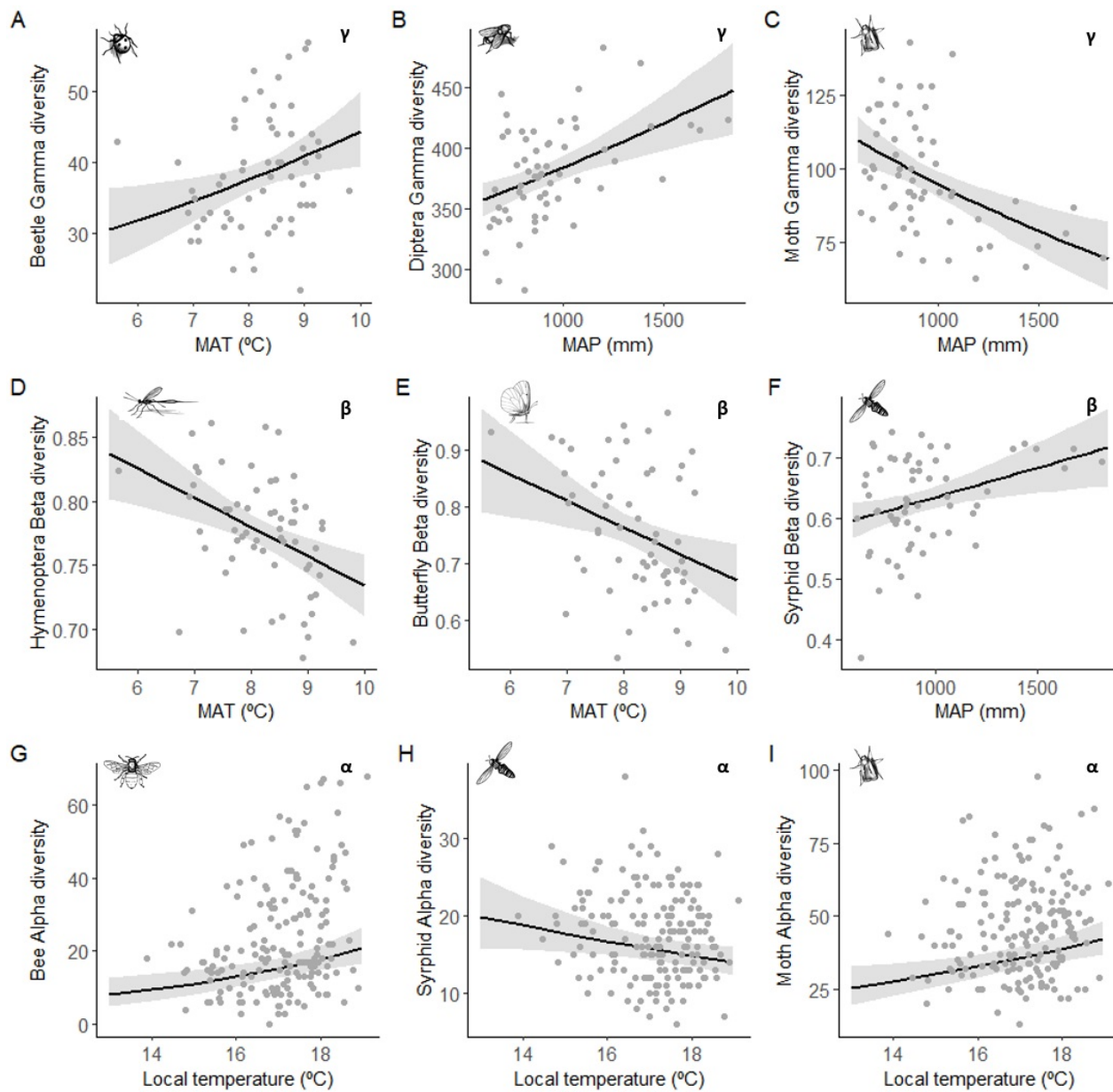


Fig. S5. Response of the gamma, beta and alpha diversity of different pollinator taxa to climate variables. Graphs show predictions of the relationships selected in the best models between gamma diversity ($n = 60$) and **(A)** multi-annual mean temperature and **(B, C)** precipitation, between beta diversity ($n = 60$) and **(D, E)** multi-annual mean temperature and **(F)** precipitation, and between alpha diversity ($n = 175$) and **(G-I)** local temperature.

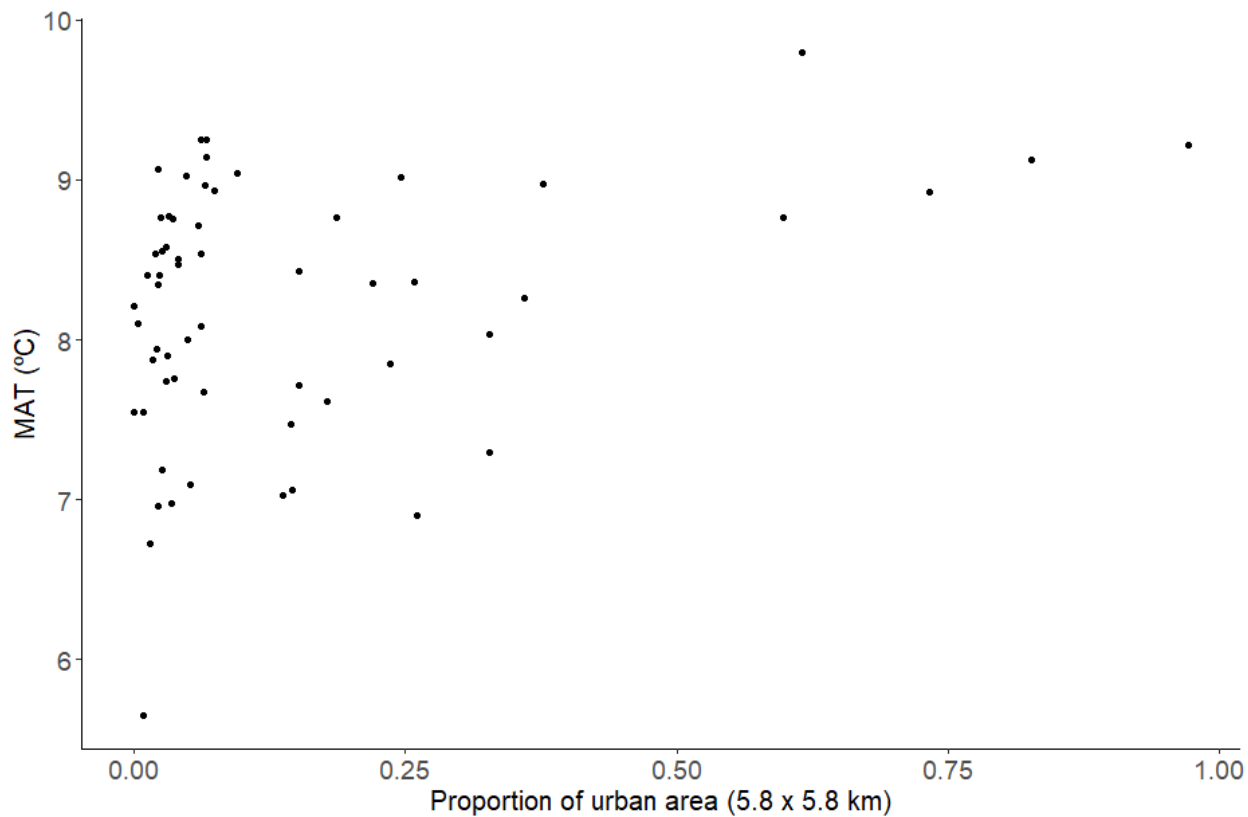


Fig. S6. Relationship between multi-annual mean temperature (MAT) and the proportion of urban area in the region (n = 60). Since we are lacking regions with a high proportion of urban and low temperatures, we must be careful with the interpretation of the interaction between multi-annual mean temperature and the proportion of urban area. If only the proportion of urban area is included in the model, highly urbanized regions show more homogeneous communities (LM, $F_{1,58} = 9.30$, $p = 0.003$).

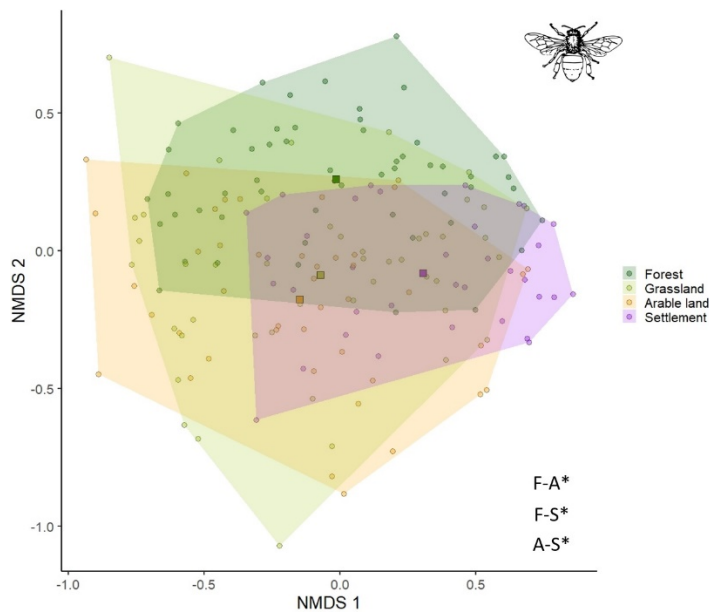


Fig. S7. Overlap in bee species composition among the four habitat types. The ordination diagram is based on non-metric multidimensional scaling (NMDS) of Jaccards dissimilarity matrices. The position of plots (dots, $n = 179$) in the NMDS space represents the similarity in bee community composition in relation to other plots: the closer the dots, the higher the proportion of species shared. Squares represent centroids of the four habitat types and polygons delimit the NMDS space occupied by plots with the same local habitat type. Significant differences in species composition between habitat types based on permutational multivariate analysis of variance (adonis function in the vegan package (72)) are shown in the downer right corner. Forest: $n = 54$, grassland: $n = 46$, arable land: $n = 41$, settlement: $n = 34$. Significance levels: *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$. The diagram shows a big overlap of grassland communities with the communities of other habitat types, especially forest habitats (see Table S8). This is not surprising, since all our plots were located on grassy strips. The high bee richness found in forest habitats can be explained by the fact that forest clearings may offer resources that are typical for both forest and grassland habitats: bee communities in forest habitats have most species in common with grasslands, and fewer with arable land and settlements; and more importantly, they have a higher number of non-overlapping plots compared to the other habitat types, which highlights the singularity of some bee communities of forest habitats.

Response: Bee species composition in forest and grassland habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_FG	1	0.329	0.329	1.633	0.016	0.119
Residuals	99	19.923	0.201		0.984	
Total	100	20.251			1.000	
Response: Bee species composition in forest and arable habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_FA	1	0.556	0.556	2.818	0.029	0.015 *
Residuals	96	18.938	0.197		0.971	
Total	97	19.494			1.000	
Response: Bee species composition in grassland and arable habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_GA	1	0.357	0.357	1.753	0.020	0.082
Residuals	87	17.699	0.203		0.980	
Total	88	18.056			1.000	
Response: Bee species composition in settlements and forest habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_SF	1	0.498	0.498	2.466	0.027	0.028 *
Residuals	88	17.768	0.202		0.973	
Total	89	18.266			1.000	
Response: Bee species composition in settlements and grassland habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_SG	1	0.413	0.412	1.946	0.024	0.054
Residuals	79	16.721	0.212		0.976	
Total	80	17.133			1.000	
Response: Bee species composition in settlements and arable habitats						
	Df	SumsOfSqs	MeanSqs	F.Model	R2	Pr (>F)
Habitat_SA	1	0.565	0.565	2.730	0.035	0.014 *
Residuals	76	15.729	0.207		0.9653	
Total	77	16.294			1.000	

Table S8. Results of permutational multivariate analysis of variance (PERMANOVA) to analyse the difference in bee species composition among habitat types. PERMANOVAs where performed with the adonis function (vegan package (72)), using Jaccard distances and the standard number of permutations (n = 999). Prior to analysis, we checked the homogeneity of variance among habitat types with the betadisper function (F = 0.22, P = 0.881).

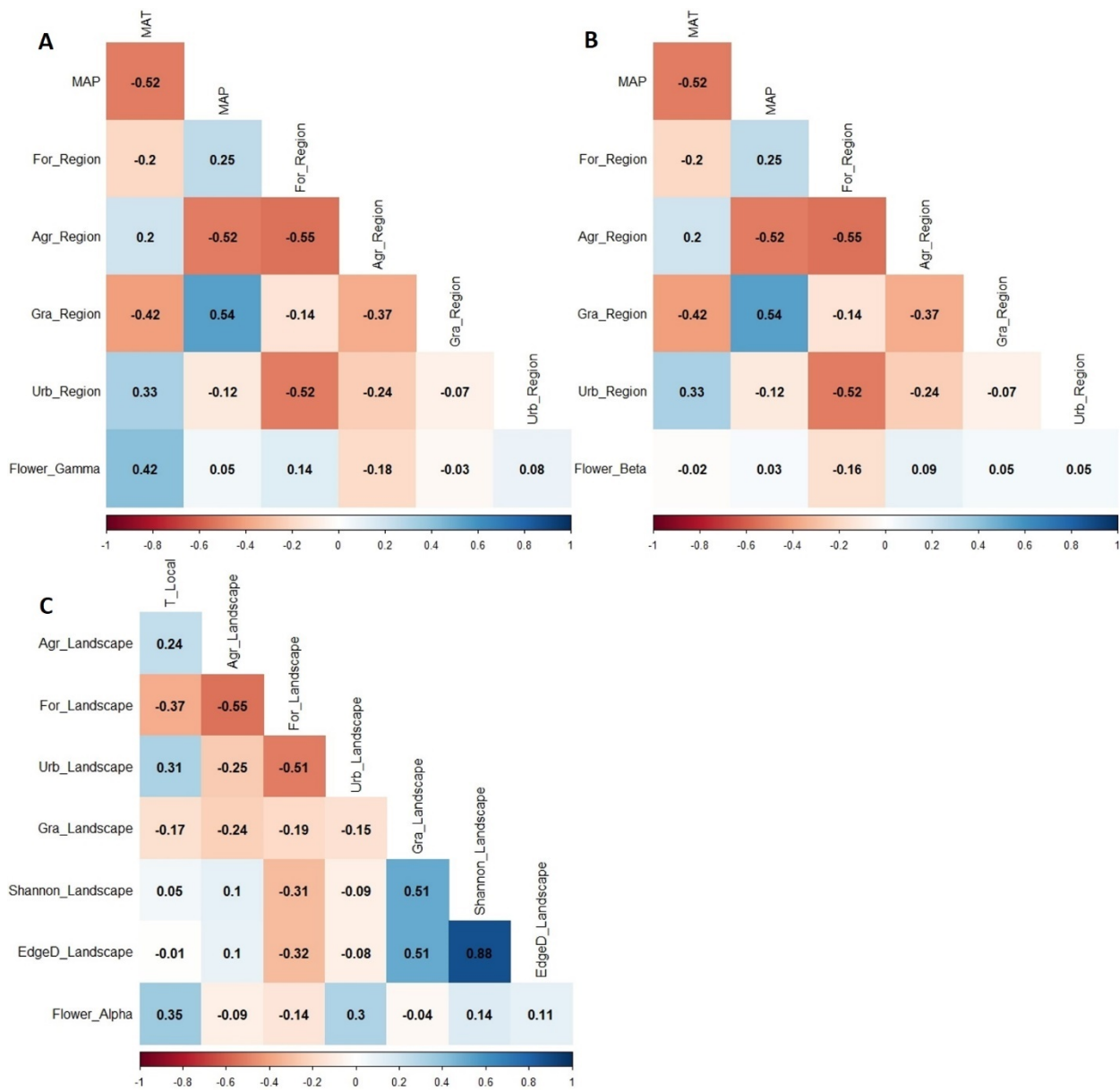


Fig. S8. Correlation plots of the variables used in (A) gamma, (B) beta, and (C) alpha diversity models. Positive correlations are displayed in blue and negative correlations in red. Color intensity is proportional to the correlation coefficients. Figures are produced with the corrplot package (84).

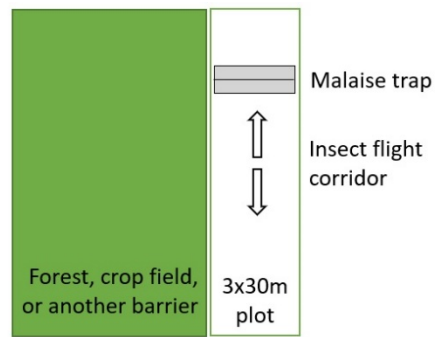


Fig. S9. Malaise trap placement within study plots. To ensure the maximum catchability of the traps in the different habitat types, we oriented traps toward insect corridors, i.e. perpendicular to barriers such as forest edges, hedgerows, crop fields, fences, or walls. When no barrier was present, traps were placed randomly, but in protected places rather than in open and windy areas (85). When establishing the traps, we made sure that the lowest part of the net touched the ground, to prevent low-flying insects escaping the trap, and we maintained the surrounding vegetation regularly to ensure that the entrance was not blocked by grass.

REFERENCES AND NOTES

1. S. Seibold, M. M. Gossner, N. K. Simons, N. Blüthgen, J. Müller, D. Ambarlı, C. Ammer, J. Bauhus, M. Fischer, J. C. Habel, K. E. Linsenmair, T. Nauss, C. Penone, D. Prati, P. Schall, E.-D. Schulze, J. Vogt, S. Wöllauer, W. W. Weisser, Arthropod decline in grasslands and forests is associated with landscape-level drivers. *Nature* **574**, 671–674 (2019).
2. R. van Klink, D. E. Bowler, K. B. Gongalsky, A. B. Swengel, A. Gentile, J. M. Chase, Meta-analysis reveals declines in terrestrial but increases in freshwater insect abundances. *Science* **368**, 417–420 (2020).
3. M. Dainese, E. A. Martin, M. A. Aizen, M. Albrecht, I. Bartomeus, R. Bommarco, L. G. Carvalheiro, R. Chaplin-Kramer, V. Gagic, L. A. Garibaldi, J. Ghazoul, H. Grab, M. Jonsson, D. S. Karp, C. M. Kennedy, D. Kleijn, C. Kremen, D. A. Landis, D. K. Letourneau, L. Marini, K. Poveda, R. Rader, H. G. Smith, T. Tscharntke, G. K. S. Andersson, I. Badenhausser, S. Baensch, A. D. M. Bezerra, B. Caballero-Lopez, P. Cavigliasso, A. Classen, S. Cusser, J. H. Dudenhöffer, J. Ekroos, T. Fijen, P. Franck, B. M. Freitas, M. P. D. Garratt, C. Gratton, J. Hipólito, A. Holzschuh, L. Hunt, A. L. Iverson, S. Jha, T. Keasar, T. N. Kim, M. Kishinevsky, B. K. Klatt, A.-M. Klein, K. M. Krewenka, S. Krishnan, A. E. Larsen, C. Lavigne, H. Liere, B. Maas, R. E. Mallinger, E. M. Pachon, A. Martínez-Salinas, T. D. Meehan, M. G. E. Mitchell, G. A. R. Molina, M. Nesper, L. Nilsson, M. E. O'Rourke, M. K. Peters, M. Ple, D. De L. Ramos, J. A. Rosenheim, M. Rundlöf, A. Rusch, A. Sáez, J. Scheper, M. Schleuning, J. M. Schmack, A. R. Sciligo, C. Seymour, D. A. Stanley, R. Stewart, J. C. Stout, L. Sutter, M. B. Takada, H. Taki, G. Tamburini, M. Tschumi, B. F. Viana, C. Westphal, B. K. Willcox, S. D. Wratten, A. Yoshioka, C. Zaragoza-Trello, W. Zhang, Y. Zou, I. Steffan-Dewenter, A global synthesis reveals biodiversity-mediated benefits for crop production. *Sci. Adv.* **5**, eaax0121 (2019).
4. J. Ollerton, R. Winfree, S. Tarrant, How many flowering plants are pollinated by animals? *Oikos* **120**, 321–326 (2011).
5. A.-M. Klein, B. E. Vaissière, J. H. Cane, I. Steffan-Dewenter, S. A. Cunningham, C. Kremen, T. Tscharntke, Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B* **274**, 303–313 (2007).

6. S. G. Potts, V. Imperatriz-Fonseca, H. T. Ngo, M. A. Aizen, J. C. Biesmeijer, T. D. Breeze, L. V. Dicks, L. A. Garibaldi, R. Hill, J. Settele, A. J. Vanbergen, Safeguarding pollinators and their values to human well-being. *Nature* **540**, 220–229 (2016).
7. F. Jauker, B. Bondarenko, H. C. Becker, I. Steffan-Dewenter, Pollination efficiency of wild bees and hoverflies provided to oilseed rape. *Agric. For. Entomol.* **14**, 81–87 (2012).
8. R. Rader, I. Bartomeus, L. A. Garibaldi, M. P. D. Garratt, B. G. Howlett, R. Winfree, S. A. Cunningham, M. M. Mayfield, A. D. Arthur, G. K. S. Andersson, R. Bommarco, C. Brittain, L. G. Carvalheiro, N. P. Chacoff, M. H. Entling, B. Foully, B. M. Freitas, B. Gemmill-Herren, J. Ghazoul, S. R. Griffin, C. L. Gross, L. Herbertsson, F. Herzog, J. Hipólito, S. Jaggar, F. Jauker, A.-M. Klein, D. Kleijn, S. Krishnan, C. Q. Lemos, S. A. M. Lindström, Y. Mandelik, V. M. Monteiro, W. Nelson, L. Nilsson, D. E. Pattermore, N. De O. Pereira, G. Pisanty, S. G. Potts, M. Reemer, M. Rundlöf, C. S. Sheffield, J. Scheper, C. Schüepp, H. G. Smith, D. A. Stanley, J. C. Stout, H. Szentgyörgyi, H. Taki, C. H. Vergara, B. F. Viana, M. Woyciechowski, Non-bee insects are important contributors to global crop pollination. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 146–151 (2016).
9. F. Sánchez-Bayo, K. A. G. Wyckhuys, Worldwide decline of the entomofauna: A review of its drivers. *Biol. Conserv.* **232**, 8–27 (2019).
10. D. Senapathi, M. A. Goddard, W. E. Kunin, K. C. R. Baldock, Landscape impacts on pollinator communities in temperate systems: Evidence and knowledge gaps. *Funct. Ecol.* **31**, 26–37 (2017).
11. S. Bae, L. Heidrich, S. R. Levick, M. M. Gossner, S. Seibold, W. W. Weisser, P. Magdon, A. Serebryanyk, C. Bässler, D. Schäfer, E. Schulze, I. Doerfler, J. Müller, K. Jung, M. Heurich, M. Fischer, N. Roth, P. Schall, S. Boch, S. Wöllauer, S. C. Renner, J. Müller, Dispersal ability, trophic position and body size mediate species turnover processes: Insights from a multi-taxa and multi-scale approach. *Divers. Distrib.* **27**, 439–453 (2021).
12. M. M. Gossner, T. M. Lewinsohn, T. Kahl, F. Grassein, S. Boch, D. Prati, K. Birkhofer, S. C. Renner, J. Sikorski, T. Wubet, H. Arndt, V. Baumgartner, S. Blaser, N. Blüthgen, C. Börschig, F. Buscot, T. Diekötter, L. R. Jorge, K. Jung, A. C. Keyel, A.-M. Klein, S. Klemmer, J. Krauss, M.

- Lange, J. Müller, J. Overmann, E. Pašalić, C. Penone, D. J. Perović, O. Purschke, P. Schall, S. A. Socher, I. Sonnemann, M. Tschapka, T. Tschardt, M. Türke, P. C. Venter, C. N. Weiner, M. Werner, V. Wolters, S. Wurst, C. Westphal, M. Fischer, W. W. Weisser, E. Allan, Land-use intensification causes multitrophic homogenization of grassland communities. *Nature* **540**, 266–269 (2016).
13. P. Theodorou, R. Radzevičiūtė, G. Lentendu, B. Kahnt, M. Husemann, C. Bleidorn, J. Settele, O. Schweiger, I. Grosse, T. Wubet, T. E. Murray, R. J. Paxton, Urban areas as hotspots for bees and pollination but not a panacea for all insects. *Nat. Commun.* **11**, 576 (2020).
14. K. C. R. Baldock, M. A. Goddard, D. M. Hicks, W. E. Kunin, N. Mitschunas, L. M. Osgathorpe, S. G. Potts, K. M. Robertson, A. V. Scott, G. N. Stone, I. P. Vaughan, J. Memmott, Where is the UK's pollinator biodiversity? The importance of urban areas for flower-visiting insects. *Proc. R. Soc. B* **282**, 20142849 (2015).
15. S. Kühnel, N. Blüthgen, High diversity stabilizes the thermal resilience of pollinator communities in intensively managed grasslands. *Nat. Commun.* **6**, 7989 (2015).
16. T. H. Oliver, M. D. Morecroft, Interactions between climate change and land use change on biodiversity: Attribution problems, risks, and opportunities. *WIREs Clim. Chang.* **5**, 317–335 (2014).
17. J. J. Williams, T. Newbold, Local climatic changes affect biodiversity responses to land use: A review. *Divers. Distrib.* **26**, 76–92 (2020).
18. C. A. Halsch, A. M. Shapiro, J. A. Fordyce, C. C. Nice, J. H. Thorne, D. P. Waetjen, M. L. Forister, Insects and recent climate change. *Proc. Natl. Acad. Sci. U.S.A.* **118**, e2002543117 (2021).
19. L. Hannah, M. Steele, E. Fung, P. Imbach, L. Flint, A. Flint, Climate change influences on pollinator, forest, and farm interactions across a climate gradient. *Clim. Change* **141**, 63–75 (2017).
20. L. Marshall, J. C. Biesmeijer, P. Rasmont, N. J. Vereecken, L. Dvorak, U. Fitzpatrick, F. Francis, J. Neumayer, F. Ødegaard, J. P. T. Paukkunen, T. Pawlikowski, M. Reemer, S. P. M. Roberts, J.

Straka, S. Vray, N. Dendoncker, The interplay of climate and land use change affects the distribution of EU bumblebees. *Glob. Chang. Biol.* **24**, 101–116 (2018).

21. T. A. Revilla, F. Encinas-Viso, M. Loreau, Robustness of mutualistic networks under phenological change and habitat destruction. *Oikos* **124**, 22–32 (2015).
22. J. K. Hill, Y. C. Collingham, C. D. Thomas, D. S. Blakeley, R. Fox, D. Moss, B. Huntley, Impacts of landscape structure on butterfly range expansion. *Ecol. Lett.* **4**, 313–321 (2001).
23. T. H. Oliver, H. H. Marshall, M. D. Morecroft, T. Brereton, C. Prudhomme, C. Huntingford, Interacting effects of climate change and habitat fragmentation on drought-sensitive butterflies. *Nat. Clim. Chang.* **5**, 941–945 (2015).
24. T. H. Oliver, T. Brereton, D. B. Roy, Population resilience to an extreme drought is influenced by habitat area and fragmentation in the local landscape. *Ecography* **36**, 579–586 (2013).
25. T. H. Oliver, S. Gillings, J. W. Pearce-Higgins, T. Brereton, H. Q. P. Crick, S. J. Duffield, M. D. Morecroft, D. B. Roy, Large extents of intensive land use limit community reorganization during climate warming. *Glob. Chang. Biol.* **23**, 2272–2283 (2017).
26. M. K. Peters, A. Hemp, T. Appelhans, J. N. Becker, C. Behler, A. Classen, F. Detsch, A. Ensslin, S. W. Ferger, S. B. Frederiksen, F. Gebert, F. Gerschlauser, A. Gütlein, M. Helbig-Bonitz, C. Hemp, W. J. Kindeketa, A. Kühnel, A. V. Mayr, E. Mwangomo, C. Ngereza, H. K. Njovu, I. Otte, H. Pabst, M. Renner, J. Röder, G. Rutten, D. Schellenberger Costa, N. Sierra-Cornejo, M. G. R. Vollstädt, H. I. Dulle, C. D. Eardley, K. M. Howell, A. Keller, R. S. Peters, A. Ssymank, V. Kakengi, J. Zhang, C. Bogner, K. Böhning-Gaese, R. Brandl, D. Hertel, B. Huwe, R. Kiese, M. Kleyer, Y. Kuzyakov, T. Nauss, M. Schleuning, M. Tschapka, M. Fischer, I. Steffan-Dewenter, Climate–land-use interactions shape tropical mountain biodiversity and ecosystem functions. *Nature* **568**, 88–92 (2019).
27. J. Aguirre-Gutiérrez, W. D. Kissling, J. C. Biesmeijer, M. F. WallisDeVries, M. Reemer, L. G. Carvalheiro, Historical changes in the importance of climate and land use as determinants of Dutch pollinator distributions. *J. Biogeogr.* **44**, 696–707 (2017).

28. F. Duchenne, E. Thébault, D. Michez, M. Gérard, C. Devaux, P. Rasmont, N. J. Vereecken, C. Fontaine, Long-term effects of global change on occupancy and flight period of wild bees in Belgium. *Glob. Change Biol.* **26**, 6753–6766 (2020).
29. M. Kammerer, S. C. Goslee, M. R. Douglas, J. F. Tooker, C. M. Grozinger, Wild bees as winners and losers: Relative impacts of landscape composition, quality, and climate. *Glob. Chang. Biol.* **27**, 1250–1265 (2021).
30. J. T. Kerr, A. Pindar, P. Galpern, L. Packer, S. G. Potts, S. M. Roberts, P. Rasmont, O. Schweiger, S. R. Colla, L. L. Richardson, D. L. Wagner, L. F. Gall, D. S. Sikes, A. Pantoja, Climate change impacts on bumblebees converge across continents. *Science* **349**, 177–180 (2015).
31. A. D. Papanikolaou, I. Kühn, M. Frenzel, O. Schweiger, Semi-natural habitats mitigate the effects of temperature rise on wild bees. *J. Appl. Ecol.* **54**, 527–536 (2017).
32. B. J. Graae, P. De Frenne, A. Kolb, J. Brunet, O. Chabrerie, K. Verheyen, N. Pepin, T. Heinken, M. Zobel, A. Shevtsova, I. Nijs, A. Milbau, On the use of weather data in ecological studies along altitudinal and latitudinal gradients. *Oikos* **121**, 3–19 (2012).
33. L. O. Frishkoff, D. S. Karp, J. R. Flanders, J. Zook, E. A. Hadly, G. C. Daily, L. K. M’Gonigle, Climate change and habitat conversion favour the same species. *Ecol. Lett.* **19**, 1081–1090 (2016).
34. A. P. Williams, E. R. Cook, J. E. Smerdon, B. I. Cook, J. T. Abatzoglou, K. Bolles, S. H. Baek, A. M. Badger, B. Livneh, Large contribution from anthropogenic warming to an emerging North American megadrought. *Science* **368**, 314–318 (2020).
35. C. R. Pyke, Habitat loss confounds climate change impacts. *Front. Ecol. Environ.* **2**, 178–182 (2004).
36. Zoologischen Staatssammlung München, Checklisten der Tierarten Bayerns, *DNA-Barcoding an der Zoologischen Staatssammlung München* (Zoologischen Staatssammlung München, 2022); www.barcoding-zsm.de/bayernfauna.

37. K. von der Dunk, Syrphidae of Bavaria—An annotated checklist. *Beiträge zur Bayerischen Entomofaunistik* **7**, 97–114 (2005).
38. M. Kuussaari, J. Heliölä, M. Luoto, J. Pöyry, Determinants of local species richness of diurnal Lepidoptera in boreal agricultural landscapes. *Agric. Ecosyst. Environ.* **122**, 366–376 (2007).
39. G. Jia, E. Shevliakova, P. Artaxo, N. D. Noblet-Ducoudré, R. Houghton, J. House, K. Kitajima, C. Lennard, A. Popp, A. Sirin, R. Sukumar, L. Verchot, Land–climate interactions, in *Climate Change and Land: an IPCC special report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems*, P. R. Shukla, J. Skea, E. Calvo Buendia, V. Masson-Delmotte, H.-O. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, E. Haughey, S. Luz, S. Neogi, M. Pathak, J. Petzold, J. Portugal Pereira, P. Vyas, E. Huntley, K. Kissick, M. Belkacemi, J. Malley, Eds. (IPCC, 2019), chap. 2; https://www.ipcc.ch/site/assets/uploads/2019/11/05_Chapter-2.pdf.
40. R. Nakadai, K. Hashimoto, T. Iwasaki, Y. Sato, Geographical co-occurrence of butterfly species: The importance of niche filtering by host plant species. *Oecologia* **186**, 995–1005 (2018).
41. P. De Frenne, F. Zellweger, F. Rodríguez-Sánchez, B. R. Scheffers, K. Hylander, M. Luoto, M. Vellend, K. Verheyen, J. Lenoir, Global buffering of temperatures under forest canopies. *Nat. Ecol. Evol.* **3**, 744–749 (2019).
42. C. Stefanescu, I. Torre, J. Jubany, F. Páramo, Recent trends in butterfly populations from north-east Spain and Andorra in the light of habitat and climate change. *J. Insect Conserv.* **15**, 83–93 (2011).
43. A. J. Suggitt, R. J. Wilson, N. J. B. Isaac, C. M. Beale, A. G. Auffret, T. August, J. J. Bennie, H. Q. P. Crick, S. Duffield, R. Fox, J. J. Hopkins, N. A. Macgregor, M. D. Morecroft, K. J. Walker, I. M. D. Maclean, Extinction risk from climate change is reduced by microclimatic buffering. *Nat. Clim. Chang.* **8**, 713–717 (2018).
44. F. Guo, J. Lenoir, T. C. Bonebrake, Land-use change interacts with climate to determine elevational species redistribution. *Nat. Commun.* **9**, 1315 (2018).

45. Y. Fourcade, M. F. WallisDeVries, M. Kuussaari, C. A. M. Swaay, J. Heliölä, E. Öckinger, Habitat amount and distribution modify community dynamics under climate change. *Ecol. Lett.* **24**, 950–957 (2021).
46. A. L. Hamblin, E. Youngsteadt, S. D. Frank, Wild bee abundance declines with urban warming, regardless of floral density. *Urban Ecosyst.* **21**, 419–428 (2018).
47. N. Deguines, R. Julliard, M. Flores, C. Fontaine, Functional homogenization of flower visitor communities with urbanization. *Ecol. Evol.* **6**, 1967–1976 (2016).
48. F. C. García, E. Bestion, R. Warfield, G. Yvon-Durocher, Changes in temperature alter the relationship between biodiversity and ecosystem functioning. *Proc. Natl. Acad. Sci. U.S.A.* **115**, 10989–10994 (2018).
49. A. Cormont, A. H. Malinowska, O. Kostenko, V. Radchuk, L. Hemerik, M. F. WallisDeVries, J. Verboom, Effect of local weather on butterfly flight behaviour, movement, and colonization: Significance for dispersal under climate change. *Biodivers. Conserv.* **20**, 483–503 (2011).
50. M. Leduc, A. Mailhot, A. Frigon, J.-L. Martel, R. Ludwig, G. B. Brietzke, M. Giguère, F. Brissette, R. Turcotte, M. Braun, J. Scinocca, The ClimEx Project: A 50-member ensemble of climate change projections at 12-km resolution over Europe and Northeastern North America with the Canadian Regional Climate Model (CRCM5). *J. Appl. Meteorol. Climatol.* **58**, 663–693 (2019).
51. Deutscher Wetterdienst, Multi-Year Temperature and Precipitation Data (Deutscher Wetterdienst, 2020); <https://opendata.dwd.de>.
52. M. S. Warren, D. Maes, C. A. M. van Swaay, P. Goffart, H. Van Dyck, N. A. D. Bourn, I. Wynhoff, D. Hoare, S. Ellis, The decline of butterflies in Europe: Problems, significance, and possible solutions. *Proc. Natl. Acad. Sci. U.S.A.* **118**, e2002551117 (2021).
53. European Commission, “Sustainable use of pesticides” (Overview Report, European Commission, European Union, 2017); https://ec.europa.eu/food/audits-analysis/overview_reports/act_getPDF.cfm?PDF_ID=1070.

54. S. Bailey, F. Requier, B. Nusillard, S. P. M. Roberts, S. G. Potts, C. Bouget, Distance from forest edge affects bee pollinators in oilseed rape fields. *Ecol. Evol.* **4**, 370–380 (2014).
55. D. Kleijn, I. Raemakers, A retrospective analysis of pollen host plant use by stable and declining bumble bee species. *Ecology* **89**, 1811–1823 (2008).
56. J. Mangels, K. Fiedler, F. D. Schneider, N. Blüthgen, Diversity and trait composition of moths respond to land-use intensification in grasslands: Generalists replace specialists. *Biodivers. Conserv.* **26**, 3385–3405 (2017).
57. N. E. Sjödin, J. Bengtsson, B. Ekbom, The influence of grazing intensity and landscape composition on the diversity and abundance of flower-visiting insects. *J. Appl. Ecol.* **45**, 763–772 (2008).
58. Federal Agency for Nature Conservation (BfN), Nature conservation areas in Germany (2019): www.bfn.de/en/service/facts-and-figures/nature-conservation/nature-conservation-areas/nature-conservation-areas-in-germany.html.
59. G. C. Daily, P. R. Ehrlich, Nocturnality and species survival. *Proc. Natl. Acad. Sci. U.S.A.* **93**, 11709–11712 (1996).
60. M. J. Skvarla, J. L. Larson, J. R. Fisher, A. P. G. Dowling, A review of terrestrial and canopy Malaise traps. *Ann. Entomol. Soc. Am.* **114**, 27–47 (2021).
61. F. A. Boetzi, J. Krauss, J. Heinze, H. Hoffmann, J. Juffa, S. König, E. Krimmer, M. Prante, E. A. Martin, A. Holzschuh, I. Steffan-Dewenter, A multitaxa assessment of the effectiveness of agri-environmental schemes for biodiversity management. *Proc. Natl. Acad. Sci. U.S.A.* **118**, e2016038118 (2021).
62. A. Hausmann, H. C. J. Godfray, P. Huemer, M. Mutanen, R. Rougerie, E. J. van Nieukerken, S. Ratnasingham, P. D. N. Hebert, Genetic patterns in European geometrid moths revealed by the barcode index number (BIN) system. *PLOS ONE* **8**, e84518 (2013).
63. Bundesministerium für Ernährung und Landwirtschaft, Forest condition report (Ergebnisse der Waldzustandserhebung) (Bundesministerium für Ernährung und Landwirtschaft, 2020):

www.bmel.de/SharedDocs/Downloads/DE/Broschueren/ergebnisse-waldzustandserhebung-2020.pdf?__blob=publicationFile&v=4.

64. S. Redlich, J. Zhang, C. Benjamin, M. S. Dhillon, J. Englmeier, J. Ewald, U. Fricke, C. Ganuza, M. Haensel, T. Hovestadt, J. Kollmann, T. Koellner, C. Kübert-Flock, H. Kunstmann, A. Menzel, C. Moning, W. Peters, R. Riebl, T. Rummler, S. Rojas-Botero, C. Tobisch, J. Uhler, L. Uphus, J. Müller, I. Steffan-Dewenter, Disentangling effects of climate and land use on biodiversity and ecosystem services—A multi-scale experimental design. *Methods Ecol. Evol.* **13**, 514–527 (2022).
65. A. Hausmann, A. H. Segerer, T. Greifenstein, J. Knubben, J. Morinière, V. Bozicevic, D. Doczkal, A. Günter, W. Ulrich, J. C. Habel, Toward a standardized quantitative and qualitative insect monitoring scheme. *Ecol. Evol.* **10**, 4009–4020 (2020).
66. M. Leray, N. Knowlton, DNA barcoding and metabarcoding of standardized samples reveal patterns of marine benthic diversity. *Proc. Natl. Acad. Sci. U.S.A.* **112**, 2076–2081 (2015).
67. J. Uhler, S. Redlich, J. Zhang, T. Hothorn, C. Tobisch, J. Ewald, S. Thorn, S. Seibold, O. Mitesser, J. Morinière, V. Bozicevic, C. S. Benjamin, J. Englmeier, U. Fricke, C. Ganuza, M. Haensel, R. Riebl, S. Rojas-Botero, T. Rummler, L. Uphus, S. Schmidt, I. Steffan-Dewenter, J. Müller, Relationship of insect biomass and richness with land use along a climate gradient. *Nat. Commun.* **12**, 5946 (2021).
68. P. A. Van Zandt, D. D. Johnson, C. Hartley, K. A. LeCroy, H. W. Shew, B. T. Davis, M. S. Lehnert, Which moths might be pollinators? Approaches in the search for the flower-visiting needles in the Lepidopteran haystack. *Ecol. Entomol.* **45**, 13–25 (2020).
69. M. J. Anderson, T. O. Crist, J. M. Chase, M. Vellend, B. D. Inouye, A. L. Freestone, N. J. Sanders, H. V. Cornell, L. S. Comita, K. F. Davies, S. P. Harrison, N. J. B. Kraft, J. C. Stegen, N. G. Swenson, Navigating the multiple meanings of β diversity: A roadmap for the practicing ecologist. *Ecol. Lett.* **14**, 19–28 (2011).
70. J. M. Chase, M. A. Leibold, Spatial scale dictates the productivity–biodiversity relationship. *Nature* **416**, 427–430 (2002).

71. R Core Team, *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2020); www.R-project.org/.
72. J. Oksanen, F. G. Blanchet, M. Friendly, R. Kindt, P. Legendre, D. McGlenn, P. R. Minchin, R. B. O'Hara, G. L. Simpson, P. Solymos, M. H. H. Stevens, E. Szoecs, H. Wagner, *vegan: Community Ecology Package*. R package version 2.5-6 (2019); <https://CRAN.R-project.org/package=vegan>.
73. M. E. Brooks, K. Kristensen, K. J. van Benthem, A. Magnusson, C. W. Berg, A. Nielsen, H. J. Skaug, M. Mächler, B. M. Bolker, *glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling*. *R J.* **9**, 378–400 (2017).
74. R. M. O'Brien, A caution regarding rules of thumb for variance inflation factors. *Qual. Quant.* **41**, 673–690 (2007).
75. D. Lüdtke, M. S. Ben-Shachar, P. Waggoner, P. Waggoner, D. Makowski, *performance: An R package for assessment, comparison and testing of statistical models*. *J. Open Source Softw.* **6**, 3139 (2021).
76. D. Senapathi, L. G. Carvalheiro, J. C. Biesmeijer, C.-A. Dodson, R. L. Evans, M. McKerchar, R. D. Morton, E. D. Moss, S. P. M. Roberts, W. E. Kunin, S. G. Potts, The impact of over 80 years of land cover changes on bee and wasp pollinator communities in England. *Proc. R. Soc. B* **282**, 20150294 (2015).
77. C. E. Grueber, S. Nakagawa, R. J. Laws, I. G. Jamieson, Multimodel inference in ecology and evolution: Challenges and solutions. *J. Evol. Biol.* **24**, 699–711 (2011).
78. K. Barton, *MuMIn: Multi-Model Inference*. R package version 1.43.17 (2020); <https://CRAN.R-project.org/package=MuumIn>.
79. F. Hartig, *Residual diagnostics for hierarchical (multi-level/mixed) regression models*. R package version 0.3.3.0 (2020); <https://CRAN.R-project.org/package=DHARMA>.
80. T. Hothorn, F. Bretz, P. Westfall, Simultaneous inference in general parametric models. *Biom. J.* **50**, 346–363 (2008).

81. B. Hoiss, J. Krauss, I. Steffan-Dewenter, Interactive effects of elevation, species richness and extreme climatic events on plant-pollinator networks. *Glob. Chang. Biol.* **21**, 4086–4097 (2015).
82. A. Smith-Pardo, V. H. Gonzalez, Bee diversity (Hymenoptera: Apoidea) in a tropical rainforest succession. *Acta Biol. Colomb.* **12**, 43–56 (2007).
83. T. C. Hsieh, K. H. Ma, A. Chao, iNEXT: An R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods Ecol. Evol.* **7**, 1451–1456 (2016).
84. T. Wei, V. Simko, R package “corrplot”: Visualization of a Correlation Matrix (2021); <https://github.com/taiyun/corrplot>.
85. K. van Achterberg, Can Townes type Malaise traps be improved? Some recent developments. *Entomol. Ber.* **69**, 129–135 (2009).