

1
2 **SUPPLEMENTARY INFORMATION**
3
4

5 **Fibronectin binding protein B binds to loricrin and promotes corneocyte**
6 **adhesion by *Staphylococcus aureus***
7

8 Thaina M. da Costa^{1§}, Albertus Viljoen^{2§}, Aisling M. Towell¹, Yves F. Dufrêne^{2,3*}, Joan
9 A. Geoghegan^{1,4*}
10

11 ¹Department of Microbiology, Moyne Institute of Preventive Medicine, School of
12 Genetics and Microbiology, Trinity College Dublin, Dublin 2, Ireland.

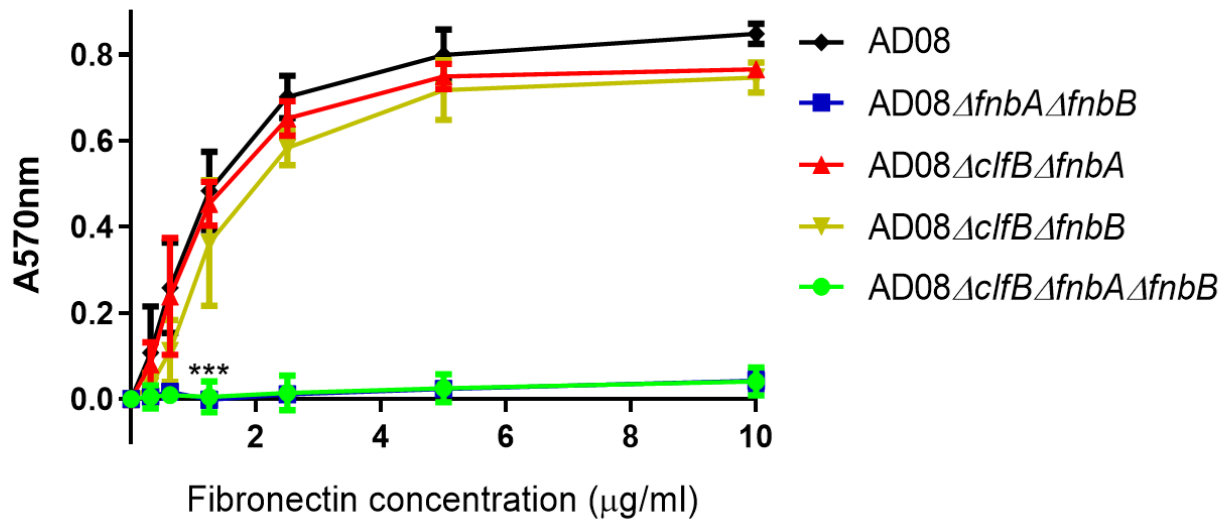
13 ²Louvain Institute of Biomolecular Science and Technology, UC Louvain, Croix du Sud,
14 4-5, bte L7.07.07, B-1348 Louvain-la-Neuve, Belgium.

15 ³Walloon Excellence in Life sciences and Biotechnology (WELBIO), Wavre, Belgium.

16 ⁴Institute of Microbiology and Infection, University of Birmingham, Edgbaston,
17 Birmingham, B15 2TT, United Kingdom.
18

19 *§These authors contributed equally*

20 *Corresponding authors: Joan A. Geoghegan (j.geoghegan@bham.ac.uk), Yves F.
21 Dufrêne (yves.dufrene@uclouvain.be)
22
23
24



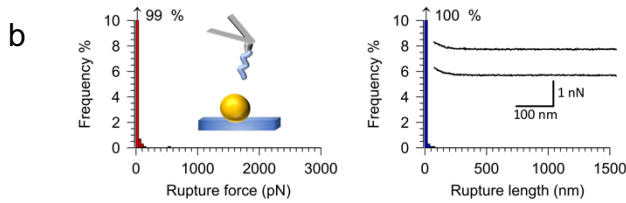
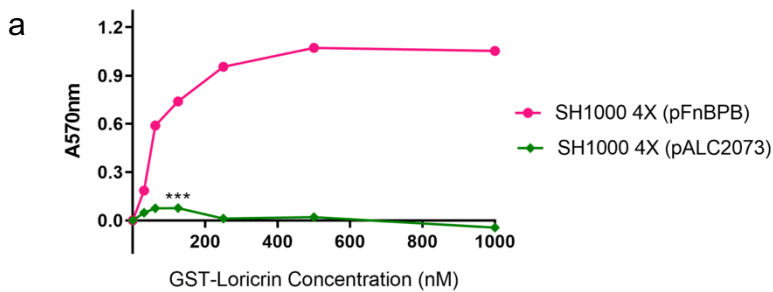
25

26 **Supplementary Figure 1. Adherence of AD08 mutants to fibronectin.**

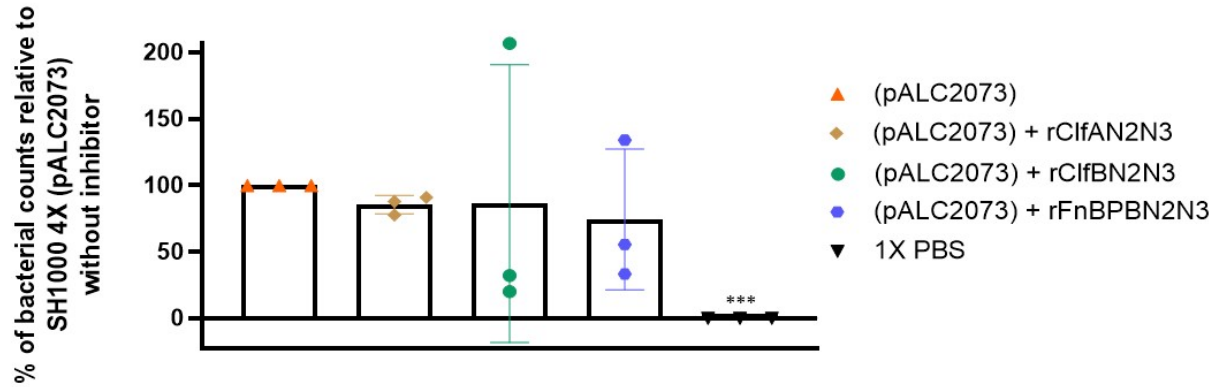
27 AD08 or mutants deficient in ClfB and/or FnBPA and/or FnBPB were grown to
 28 exponential phase ($OD_{600} = 0.35$), adjusted to an OD_{600} of 1.0 and incubated in microtiter
 29 plates coated with fibronectin. Following incubation, the wells were washed, adherent
 30 cells were stained with crystal violet, and the absorbance was read at 570 nm. The datum
 31 points on the graph represent the mean values of three independent biological
 32 experiments and error bars show the standard deviation. Statistical analysis was
 33 performed using a two-way ANOVA with a Dunnett's multiple comparison test to compare
 34 variances between AD08 and the mutants at the 1.25 μg/mL fibronectin concentration.
 35 No symbol indicates $P > 0.05$. ***, $P = 0.0001$.

36

37

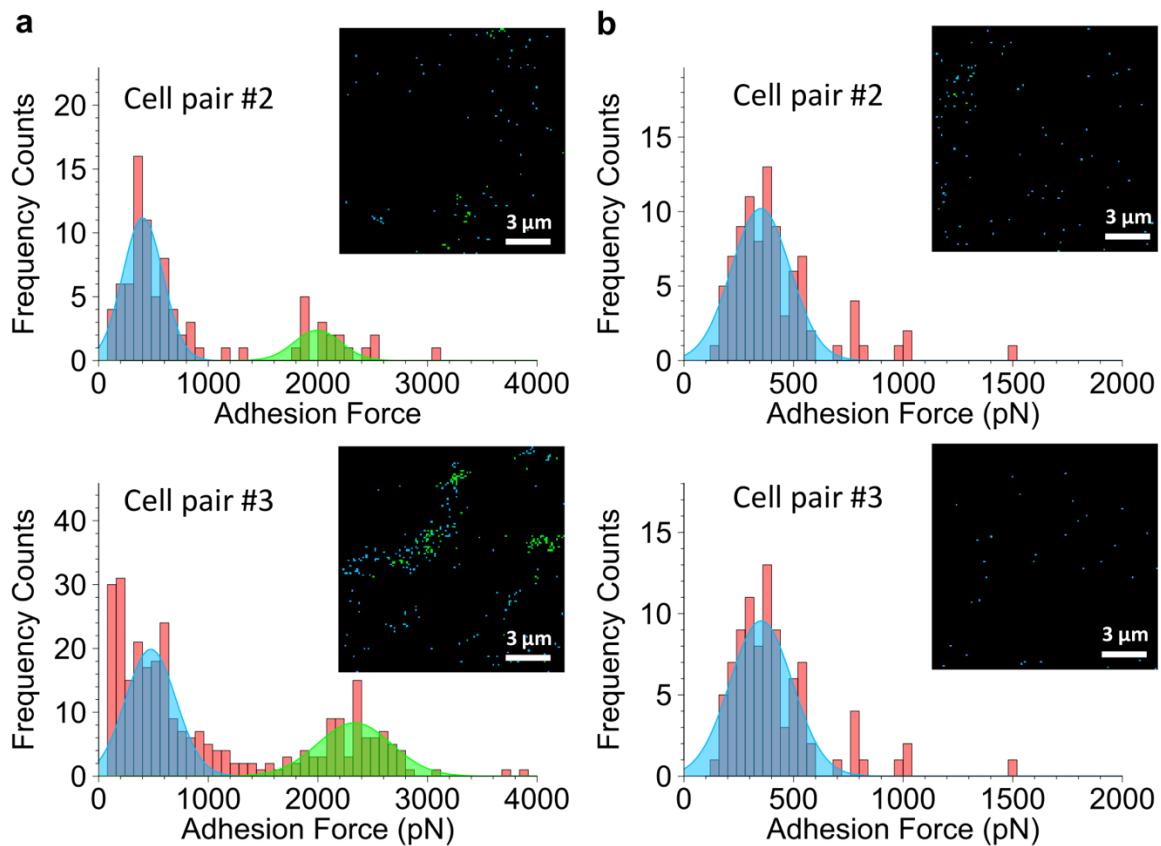


38
 39 **Supplementary Figure 2. *S. aureus* SH1000 deficient in ClfA, ClfB, FnBPA and**
 40 **FnBPB does not bind to loricrin. a) SH1000 and the mutant deficient in ClfA, ClfB,**
 41 **FnBPA and FnBPB (SH1000 4X) were grown to exponential phase ($OD_{600} = 0.35$),**
 42 **adjusted to an OD_{600} of 1.0 and incubated in microtiter plates coated with GST-loricrin.**
 43 **Following incubation, the wells were washed, adherent cells were stained with crystal**
 44 **violet, and the absorbance was read at 570 nm. The graph shows results from a single**
 45 **experiment that is representative of three independent experiments. The ligand**
 46 **concentration given on the x-axis is the concentration of the solution used to coat the**
 47 **wells. Statistical analysis was performed using a two-way ANOVA with a Sidak's multiple**
 48 **comparison test to compare variances between the wild type and the mutant at the 125**
 49 **nM loricrin concentration. ***, $P = 0.00078$. b) AFM tips modified with loricrin were used**
 50 **to probe single exponentially grown ($OD_{600} = 0.35$) staphylococcal cells. Data for one**
 51 **representative *S. aureus* SH1000 4X cell showing the absence of interaction with loricrin.**
 52 **Left. Histogram plot of rupture forces. Right. Histogram plot of rupture lengths with inset**
 53 **showing three representative curves.**



54
55 **Supplementary Figure 3. Recombinant proteins do not affect *S. aureus* SH1000 4X**
56 **(pALC2073) adherence to healthy human skin corneocytes.** Bacteria were grown in
57 TSB to exponential phase ($OD_{600} = 0.35$), adjusted to an $OD_{600} = 1.0$ and incubated with
58 tape strips containing corneocytes previously treated with rClfAN2N3 or rClfBN2N3 or
59 rFnBPBN2N3. Each datum point is from a different biological replicate, and the bars
60 represent the means of three independent experiments, with error bars showing the
61 standard deviation. A no bacteria control was used where PBS alone was incubated with
62 the corneocytes. Tape strips were washed, and adherent bacteria were stained with
63 crystal violet. The number of bacteria cells adhering to 10 corneocytes was counted and
64 the percentage of adherence is shown as comparison with total adherence of SH1000 4X
65 (pFnBPB) to corneocytes that were not pre-treated with recombinant protein. Statistical
66 analysis was performed using a one-way ANOVA with a Dunnett's multiple comparison
67 test to compare variances between *S. aureus* SH1000 4X (pALC2073) adhering to
68 corneocytes without recombinant protein to the *S. aureus* SH1000 4X (pALC2073)
69 adhering to corneocytes treated with recombinant protein, and the no bacteria control.
70 ***, $P = 0.000091$.

71



72

73 **Supplementary Figure 4. FnBPB expressed on living *S. aureus* cells forms**
 74 **ultrastrong single molecular complexes with ligands exposed on corneocyte**

75 **surfaces.** (a) Data for two more *S. aureus* SH1000 4X (pFnBPB) cells used to probe
 76 corneocytes in quantitative imaging mode. Histograms of binding forces with inset of the

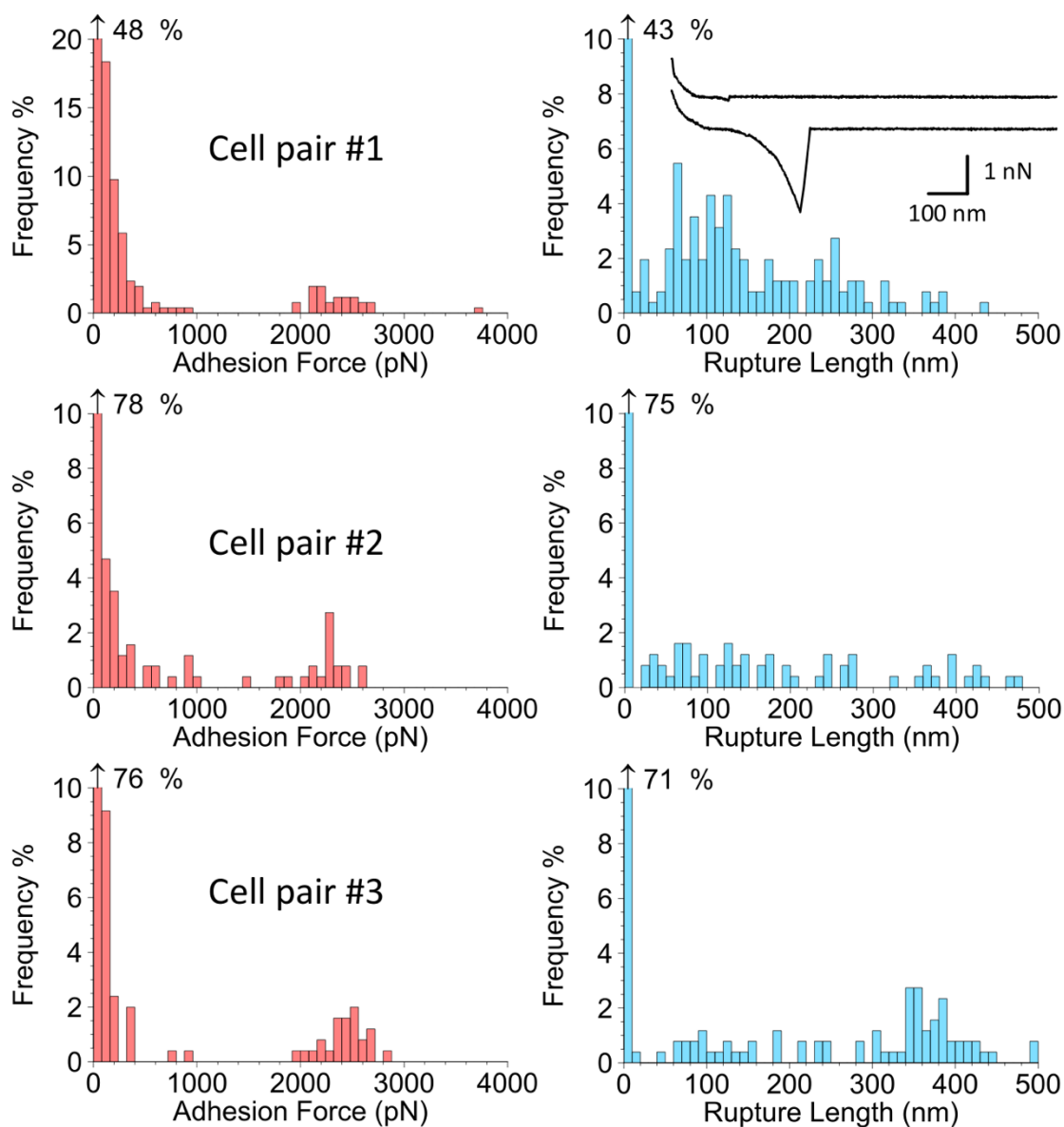
77 corresponding adhesion map. Black pixels show where zero adhesion was measured,
 78 blue pixels show where adhesive interactions between 0 and 1,000 pN were detected.

79 Green pixels are where adhesive interactions greater than 1,000 pN were detected. (b)

80 Data obtained when two more control cells that do not express FnBPB [*S. aureus* SH1000
 81 4X (pALC2073)] were used to probe corneocytes.

82

83



84

85 **Supplementary Figure 5.** Low retraction-speed ($1 \mu\text{m}\cdot\text{s}^{-1}$) force volume mode
 86 recordings using *S. aureus* SH1000 4X (pFnBPB) cell probes to probe corneocytes. A
 87 force volume map was recorded on a $2 \times 2 \mu\text{m}$ area (16×16 pixels) of a corneocyte
 88 where ultrastrong binding ($> 2,000$ pN) was previously detected in QI mode (for cell pair
 89 1, see Figure 7b; and for cell pairs 2 and 3, see Supplementary Figure 4a). Shown are
 90 adhesion force (left) and rupture length (right) histograms. The inset shows two
 91 representative retraction profiles.

92 **Supplementary Table 1. Bacterial strains.**

Strain	Description	Reference
Topp3	<i>E. coli</i> strain deficient in proteases for expression of recombinant proteins	Stratagene
XL-1 Blue	<i>E. coli</i> strain for deficient in proteases expression of recombinant proteins	Stratagene
IM01B	<i>E. coli</i> strain for plasmid propagation. SA08BQPN25-hsdS	1
AD08	<i>S. aureus</i> atopic dermatitis clinical isolate	2
AD08 Δ <i>clfB</i> Δ <i>fnbA</i>	AD08 deficient in <i>clfB</i> and <i>fnbA</i>	3
AD08 Δ <i>clfB</i> Δ <i>fnbB</i>	AD08 deficient in <i>clfB</i> and <i>fnbB</i>	3
AD08 Δ <i>fnbA</i> Δ <i>fnbB</i>	AD08 deficient in <i>fnbA</i> and <i>fnbB</i>	3
AD08 Δ <i>clfB</i> Δ <i>fnbA</i> Δ <i>fnbB</i>	AD08 deficient in <i>clfB</i> and <i>fnbA</i> and <i>fnbB</i>	3
SH1000	<i>S. aureus</i> strain 8325-4 derivative with a restored <i>rsbU</i> gene	4
SH1000 4X	SH1000 deficient in <i>clfA</i> and <i>clfB</i> and <i>fnbA</i> and <i>fnbB</i>	5

93

94

95 **Supplementary Table 2. Plasmids**

Plasmid	Description	Reference
pGEX-4T2	<i>E. coli</i> vector for the expression of glutathione S-transferase. Amp ^r .	GE Lifesciences
pGEX-4T2::HLor	pGEX-4T2 carrying the full-length cDNA encoding human loricrin. Amp ^r .	6
pQE30::rClfB ₂₀₁₋₅₄₂	pQE30 carrying the His-tagged N2 N3 recombinant ClfB. Amp ^r .	6
pQE30::rFnBPB ₁₆₂₋₄₈₀	pQE30 carrying the His-tagged N2 N3 recombinant FnBPB. Amp ^r .	7
pQE30::rClfA ₂₂₁₋₅₅₉	pQE30 carrying the His-tagged N2 N3 recombinant FnBPB. Amp ^r .	8
pALC2073	Tetracycline inducible expression plasmid. Cm ^r .	9
pFnBPB	pALC2073 carrying the <i>fnbB</i> gene. Cm ^r .	3

96

97

98 **SUPPLEMENTARY REFERENCES**

- 99 1. Monk, I. R., Tree, J. J., Howden, B. P., Stinear, T. P. & Foster, T. J. Complete
100 bypass of restriction systems for major *Staphylococcus aureus* lineages. *mBio* **6**,
101 e00308-15 (2015).
- 102 2. Fleury, O. M. *et al.* Clumping factor B promotes adherence of *Staphylococcus*
103 *aureus* to corneocytes in atopic dermatitis. *Infect. Immun.* **85**, e00994-16 (2017).
- 104 3. Towell, A. M. *et al.* *Staphylococcus aureus* binds to the N-terminal region of
105 corneodesmosin to adhere to the stratum corneum in atopic dermatitis. *Proc. Natl.*
106 *Acad. Sci. U. S. A.* **118**, e2014444118 (2021).
- 107 4. Horsburgh, M. J. *et al.* σ^B modulates virulence determinant expression and stress
108 resistance: characterization of a functional *rsbU* strain derived from
109 *Staphylococcus aureus* 8325-4. *J. Bacteriol.* **184**, 5457–5467 (2002).
- 110 5. O'Neill, E. *et al.* A novel *Staphylococcus aureus* biofilm phenotype mediated by
111 the fibronectin-binding proteins, FnBPA and FnBPB. *J. Bacteriol.* **190**, 3835–3850
112 (2008).
- 113 6. Mulcahy, M. E. *et al.* Nasal colonisation by *Staphylococcus aureus* depends upon
114 clumping factor B binding to the squamous epithelial cell envelope protein loricrin.
115 *PLoS Pathog.* **8**, e1003092 (2012).
- 116 7. Burke, F. M., McCormack, N., Rindi, S., Speziale, P. & Foster, T. J. Fibronectin-
117 binding protein B variation in *Staphylococcus aureus*. *BMC Microbiol.* **10**, (2010).
- 118 8. O'Connell, D. P. *et al.* The fibrinogen-binding MSCRAMM (clumping factor) of
119 *Staphylococcus aureus* has a Ca²⁺-dependent inhibitory site. *J. Biolog. Chem.*
120 **273**, 6821–6829 (1998).
- 121 9. Bateman, B. T., Donegan, N. P., Jarry, T. M., Palma, M. & Cheung, A. L.
122 Evaluation of a Tetracycline-Inducible Promoter in *Staphylococcus aureus* In Vitro
123 and In Vivo and Its Application in Demonstrating the Role of *sigB* in Microcolony
124 Formation. *Infect. Immun.* **69**, 7851–7857 (2001).

125

126