

1 **Supplementary Information for “Stable, diverse, fecal-derived *in vitro* microbial**
2 **communities that model the intestinal microbiota response to antibiotics”**

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4 **Authors:** Andrés Aranda-Díaz¹, Katharine Michelle Ng¹, Tani Thomsen¹, Imperio Real-
5 Ramírez¹, Dylan Dahan², Susannah Dittmar¹, Carlos Gutierrez Gonzalez³, Taylor
6 Chavez¹, Kimberly S. Vasquez², Taylor H. Nguyen¹, Feiqiao Brian Yu⁴, Steven K.
7 Higginbottom², Norma F. Neff⁴, Joshua E. Elias⁴, Justin L. Sonnenburg^{2,4}, Kerwyn Casey
8 Huang^{2,4*}

9

10 ¹Department of Bioengineering, Stanford University, Stanford, CA 94305

11 ²Department of Microbiology and Immunology, Stanford University School of
12 Medicine, Stanford, CA 94305

13 ³Department of Chemical and Systems Biology, Stanford University School of Medicine,
14 Stanford, CA 94305

15 ⁴Chan Zuckerberg Biohub, San Francisco, CA 94158

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17 *Correspondence: kchuang@stanford.edu

18 **Supplementary Tables**

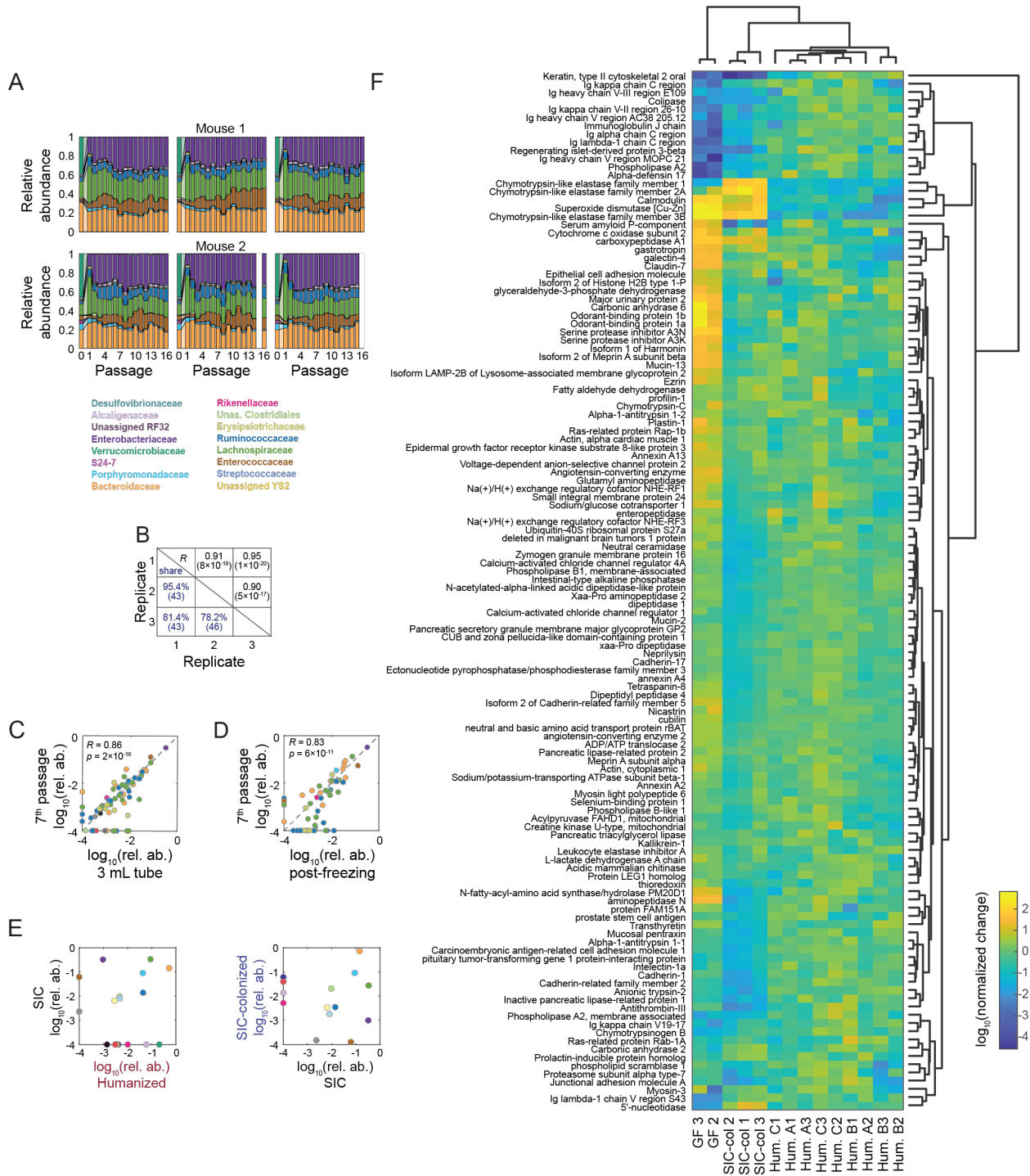
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Strain	Closest relative in NCBI 16S ribosomal RNA sequence database	Ciprofloxacin MIC ($\mu\text{g/mL}$)
TT1	<i>Enterococcus hirae</i> ATCC 9790	2
TT2	<i>Escherichia fergusonii</i> ATCC 35469	<0.5
TT3	[<i>Clostridium</i>] <i>symbiosum</i> ATCC 14940	16
TT4	<i>Bacteroides thetaiotaomicron</i> JCM 5827	16
TT5	[<i>Clostridium</i>] <i>clostridioforme</i> ATCC 25537	32
TT6	<i>Blautia producta</i> JCM 1471	32
TT7	[<i>Clostridium</i>] <i>scindens</i> strain DSM 5676	32
TT8	<i>Enterococcus faecium</i> strain DSM 20477	4
TT9	[<i>Clostridium</i>] <i>hylemonae</i> TN-272	32
TT10	<i>Enterococcus faecalis</i> NBRC 100480	4
TT11	[<i>Clostridium</i>] <i>hathewayi</i> 1313	32
TT12	<i>Bacteroides fragilis</i> NCTC 9343	4
TT13	<i>Flavonifractor plautii</i> 265	8
TT14	<i>Bacteroides uniformis</i> JCM 5828	16
TT15	<i>Parabacteroides distasonis</i> ATCC 8503	4

20 **Table S1: Strains isolated in this study, related to Figure 5.**

Medium	Formulation	Sterilization	Storage
BHI	Commercially available (BD 211061)	Autoclave 20 min	Room temperature (RT)
TYG	As described in (Whitaker et al., 2017)	0.22- μ m filter	4 °C
GAM	Commercially available (HiMedia M1801)	Autoclave 20 min	RT
YCFA	As described in (Duncan et al., 2002). Vitamins and cysteine added 48 h before experiment	0.22- μ m filter	4 °C

21 **Table S2: Media used in this study, related to Figure 1.**



23

24 **Figure S1: Robust, high-throughput cultivation of fecal-derived SICs and**

25 **colonization of a germ-free mice by an SIC. Related to Figures 1 and 2.**

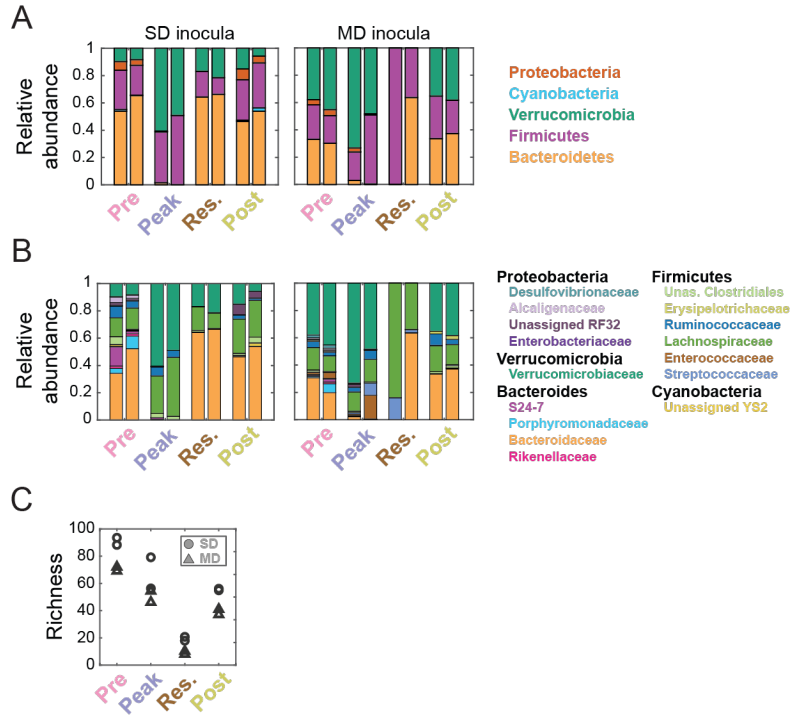
26 A) *In vitro* passaging leads to stable and complex SICs. Family-level composition of
27 three replicate SICs from two fecal samples (Pre MD mice #1 and #2) during
28 passaging in BHI for 16 rounds *in vitro*. Passage 0 is the fecal inoculum.

29 B) *In vitro*-passaged SICs are highly reproducible. Summary of the reproducibility of
30 the three replicates for the mouse 1 SICs in Fig. 1 B-D. *R* and *p* were computed using
31 only ASVs present in both samples. Also shown is the percentage of ASVs that were
32 present in both replicates at >0.1% abundance (“shared”), with the total number of
33 ASVs in parentheses.

34 C,D) SICs can be grown in larger volumes and can be frozen and revived without
35 affecting composition. ASV-level relative abundance values for the SIC in Fig. 1 B-D
36 after 7 passages were compared with the same SIC grown for one passage in a larger
37 vessel without shaking (C), and after freezing with 25% glycerol at -80 °C and
38 reviving for one passage (D). *R* and *p* were computed using only ASVs present in
39 both samples. ASVs with relative abundance < 10⁻⁴ were set to 10⁻⁴ for visualization.

40 E) Families that are overrepresented *in vitro* recede *in vivo*. Shown is a comparison of
41 family-level relative abundance of an SIC and the humanized-mouse fecal inoculum
42 from which it was derived (left), or ex-germ-free mice colonized with the SIC (right,
43 SIC *n*=1, inoculum *n*=1, SIC-colonized *n*=3). Families with relative abundance < 10⁻⁴
44 were set to 10⁻⁴ for visualization. Mouse data are the same as in Fig. 2B.

45 F) The secreted proteome of mice colonized with SIC is similar to the proteome of
46 humanized mice. *M. musculus* proteins present at levels 10-fold higher in mice than
47 in SICs were normalized by their mean abundance in humanized mice.
48 Dendrograms resulting from hierarchical clustering of normalized relative
49 abundance of proteins in germ free (GF), SIC-colonized (SIC col.), or humanized
50 mice (Hum.) housed in three cages (A, B, and C).



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52 **Figure S2: Diet and antibiotics have large, distinct, interacting effects on the**

53 **composition of the microbiota, related to Figure 3.**

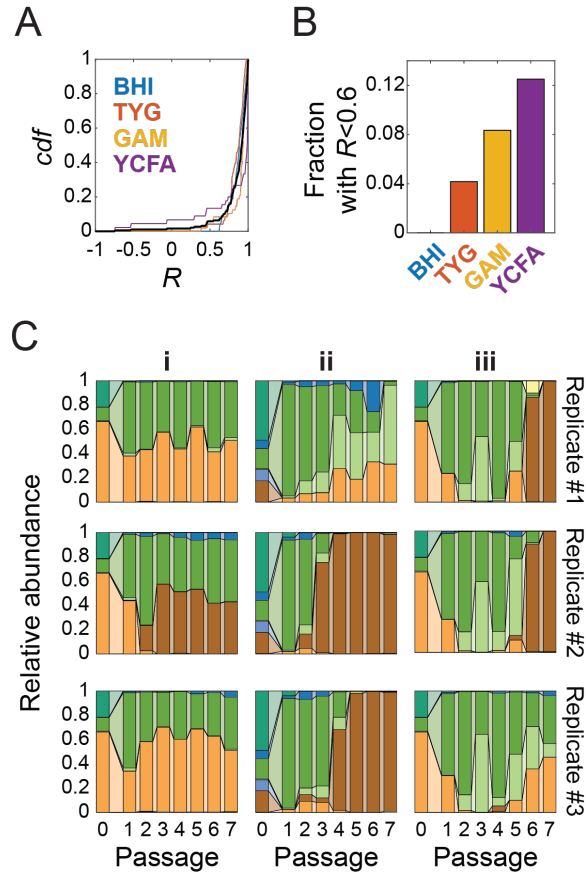
54 A,B) Ciprofloxacin elicits large changes in microbiota composition at the phylum (A)

55 and family (B) level *in vivo*. Each time point has two bars corresponding to the two

56 mice in each group.

57 C) Diversity *in vivo* decreases during ciprofloxacin treatment and does not fully recover

58 after treatment. Richness (number of ASVs in rarefied data) of fecal inocula.



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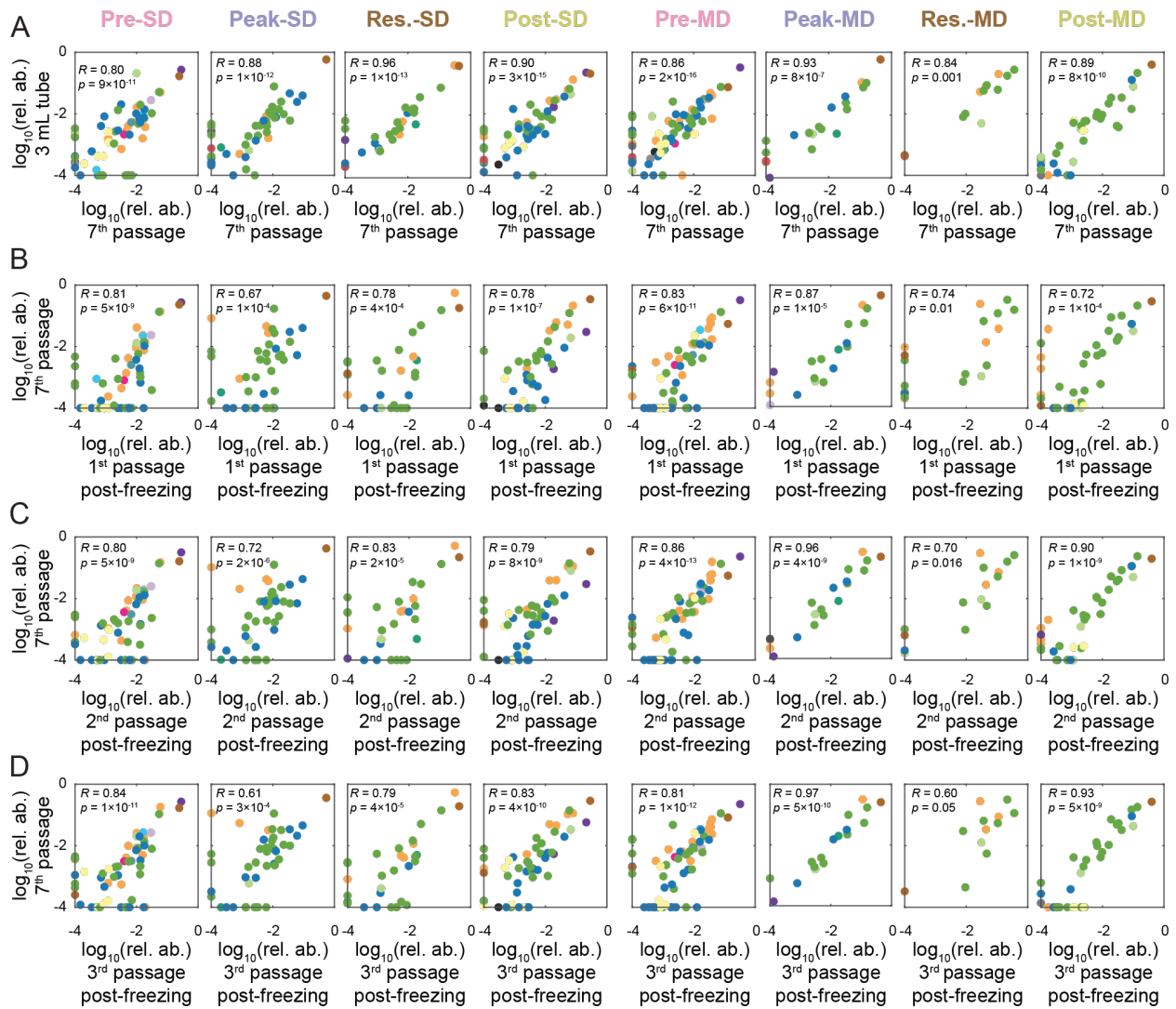
60 **Figure S3: SICs are more reproducible in BHI than in other media, related to Figure 3.**

61 A) Technical replicates are largely reproducible. Cumulative density function of
 62 Pearson correlation coefficient (R) for all pairwise comparisons between technical
 63 replicates.

64 B) Proportion of technical replicate pairwise correlations $R < 0.6$ for the 4 growth media.

65 C) Non-reproducible technical replicates share similar dynamics during early passages.
 66 Family-level composition during *in vitro* passaging for 7 rounds of three SICs with
 67 low correlation coefficients after 7 passages. (i) Technical replicates of SIC
 68 originating from SD mouse #2 during pre-treatment, grown in GAM. (ii) Technical

69 replicates of SIC originating from MD mouse #2 during peak of treatment, grown in
70 YCFA. (iii) Technical replicates of SIC originating from SD mouse #2 during residual
71 treatment, grown in YCFA.

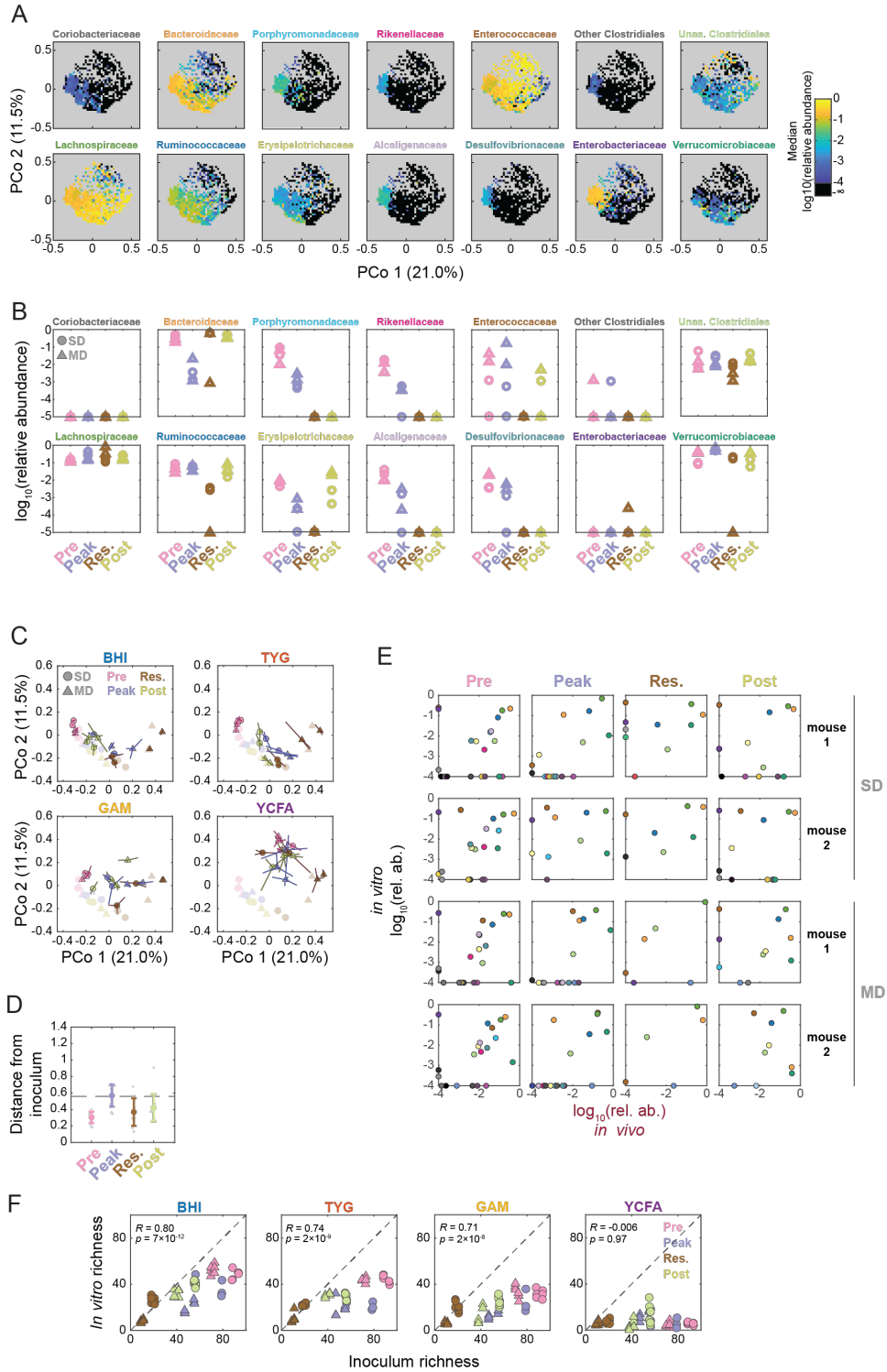


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73 **Figure S4: SICs maintain composition after growth in larger volumes without**
 74 **shaking and during revival after freezing, related to Figure 3.**

75 A-D) Correlation plots of \log_{10} (relative abundance) at the ASV level for 8 SICs grown in
 76 BHI after 7 passages against the same SIC grown for one passage in a larger volume
 77 (3 mL) without shaking (A), and after freezing and reviving for one (B), two (C), or
 78 three (D) passages. Pearson coefficients (R) and their p -values were computed using

79 only data points present in both samples. ASVs with relative abundance $<10^{-4}$ were
80 set to 10^{-4} for visualization purposes.

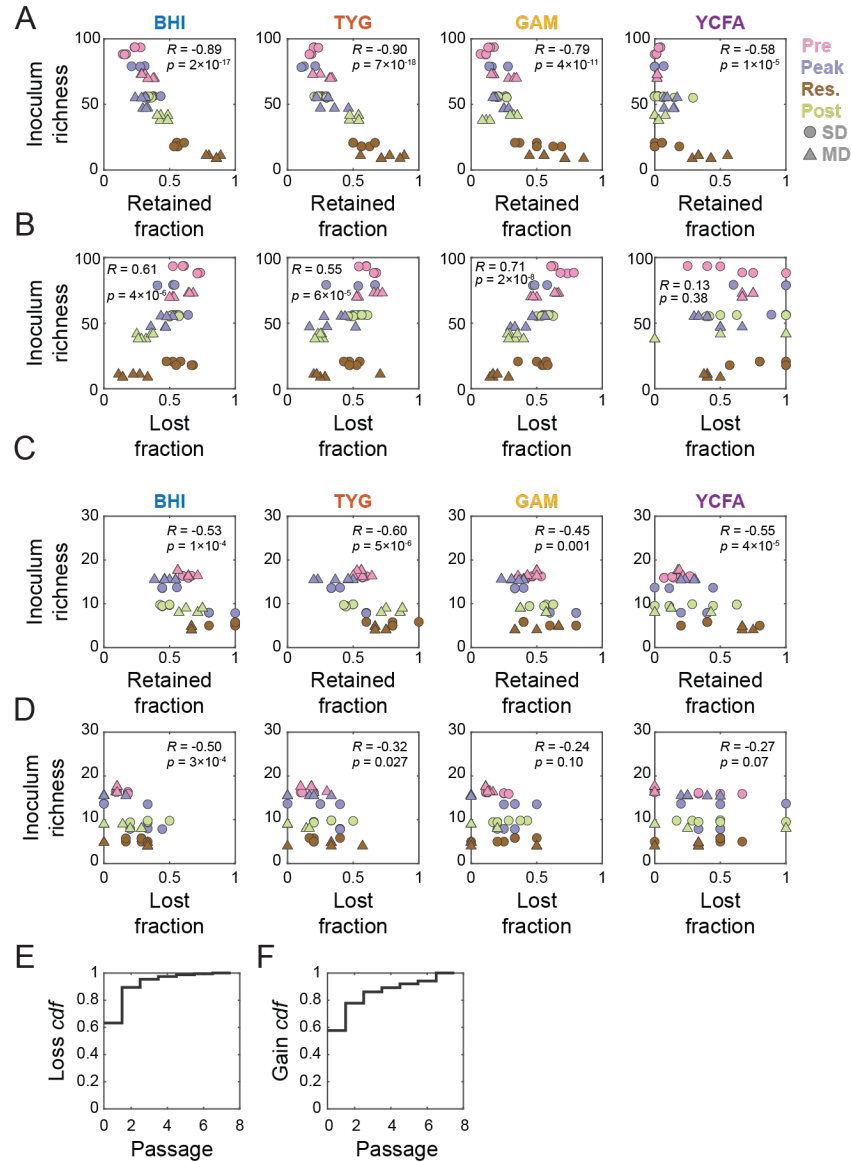


81

82 **Figure S5: Certain families generally co-occur in SICs, related to Figure 3.**

- 83 A) Median \log_{10} (relative abundance) of families across all samples ($n=1728$) binned by
84 first two principal components. Only families present in >10% of the samples are
85 shown.
- 86 B) \log_{10} (relative abundance) of families in (A) in each of the inocula. Families with
87 relative abundance $<10^{-4}$ were set to 10^{-4} for visualization purposes.
- 88 C) Medium and inoculum determine the final composition of passaged SICs. The 7th
89 passage of all 192 SICs in a PCoA of SIC composition using unweighted Unifrac
90 distance computed on all *in vivo* and *in vitro* samples at the ASV level. Samples are
91 separated by media, with colors and shapes representing the timepoint during
92 ciprofloxacin treatment and diet, respectively, in the mice from which the inocula
93 were taken. Symbols are the centroid of three replicates, with lines connecting the
94 replicates to the centroid. Original fecal inocula are plotted in light colors. BHI data
95 is the same as in Fig. 2C.
- 96 D) SICs derived from feces at the peak of treatment *in vivo* are most distinct from the
97 composition of their inoculum suggesting low viability of detected strains. Weighted
98 Unifrac distance of the 7th passage in BHI of each SIC to their fecal inoculum.
99 Colored circles, mean distance for each medium; individual SICs in gray. Error bars,
100 standard deviations; $n=12$. Dashed line, mean distance between fecal samples.
- 101 E) *In vitro* passaging in BHI can produce an SIC that retain abundant families in the
102 fecal inoculum. Mean family-level relative abundances (from mean of passages 4-7)

103 for three SIC triplicates compared with the fecal inoculum from which it originated.
104 Families with relative abundance $<10^{-4}$ were set to 10^{-4} for visualization purposes.
105 F) SIC yield scales with diversity. Mean OD of SICs after 20 h of growth in BHI
106 passages 3-7 increases with increasing richness (number of ASVs in rarefied data) in
107 passage 7. R , Pearson correlation coefficient; $n=48$. BHI data is the same as in Fig. 2E.



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109 **Figure S6: Taxa loss and emergence are correlated with inoculum diversity, related to**

110 **Figure 3.**

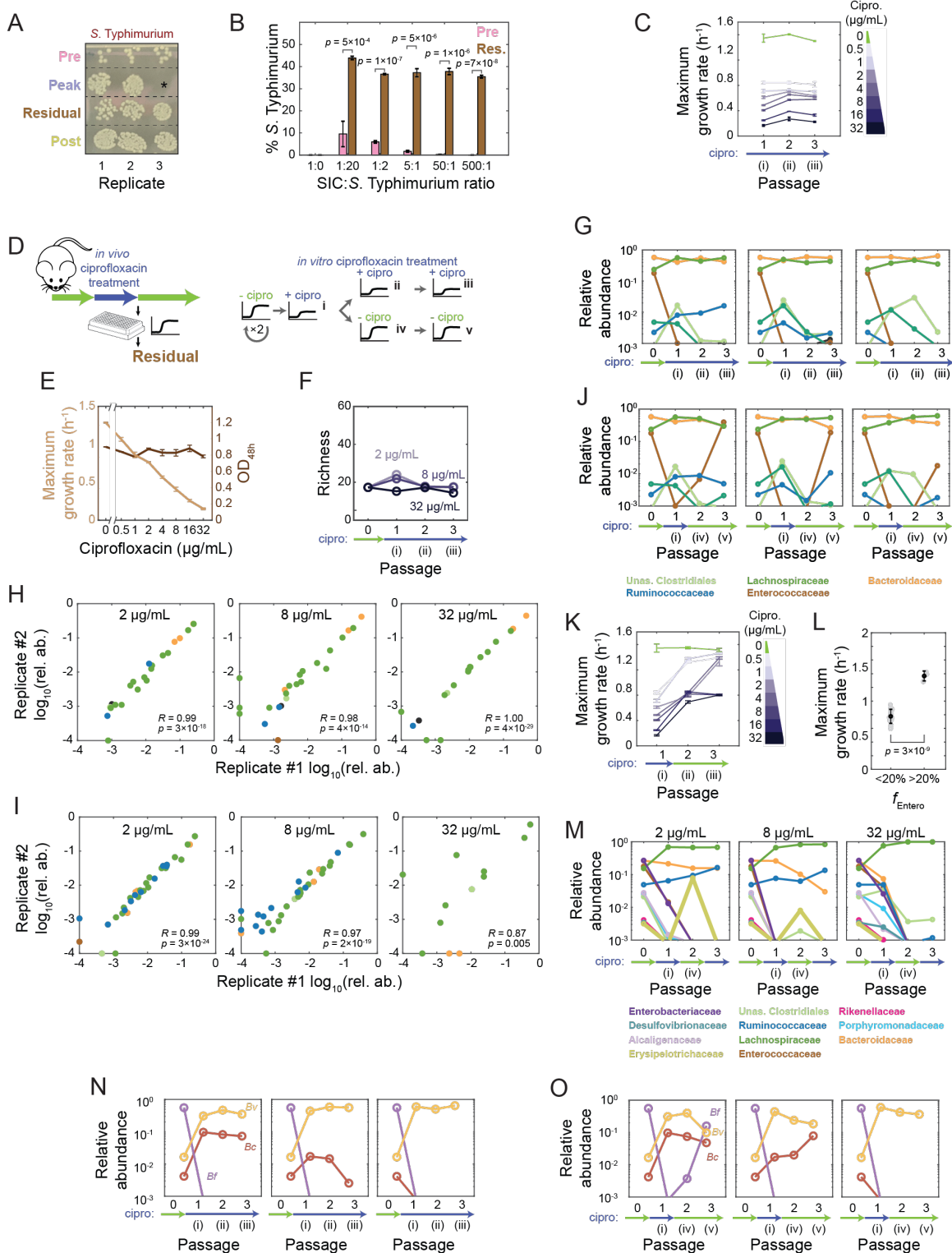
111 A-D) Correlation plots for inoculum diversity (number of ASVs in (A) and (B), and

112 families in (C) and (D), in rarefied data) with fraction of taxa lost in (A) and (C) and

113 emerged in (B) and (D), for all 192 passaged SICs. Pearson correlation coefficients (R)

114 and their p -values were computed from all data points in each plot ($n=48$).

115 E,F) Most ASV loss or emergence occurred within the first three passages. Cumulative
116 density function for the passage at which an ASV was lost (E) or emerged (F).



117

118 **Figure S7: Effects and dynamics of antibiotic treatment on SICs, related to Figures 4-**

119 **6.**

120 A,B) *In vivo* ciprofloxacin treatment makes SICs more susceptible to *S. Typhimurium*
121 invasion. (A) Colonies of *S. Typhimurium* SL1344 after 48 h of growth with SICs
122 spotted on LB+streptomycin in aerobic conditions after a 1:10⁴ dilution. These data
123 are a biological replicate (SICs derived from mice housed in a different cage) of Fig.
124 4B. *: missing replicate. (B) Single-cell quantification of mCherry-tagged *S.*
125 *Typhimurium* 14028s after 48 h of co-culturing with SICs derived from pre- and
126 residual-treatment mice fecal inocula. *p*-values are from a Student's two-sided *t*-test
127 between each pairwise comparison, *n*=3.

128 C) SICs show little adaptation to continuous ciprofloxacin treatment. Maximum growth
129 rate decreases or remains approximately constant across three passages in
130 ciprofloxacin.

131 D-J) SICs derived from pre-exposed inocula show increased resilience to ciprofloxacin
132 treatment. (D) Experimental setup for *in vitro* antibiotic treatment of a pre-exposed
133 SIC. An SIC passaged in BHI from residual treatment humanized mouse fecal
134 inoculum (Res-SD) was revived after freezing and passaged twice in BHI. The SIC
135 was passaged in ciprofloxacin three times (i,ii,iii) or in ciprofloxacin once and then
136 twice without the drug (i,iv,v). (E) In contrast to the Pre-SD SIC (Fig. 3C), the yield
137 of Res-SD SIC was virtually unaffected by ciprofloxacin, although the growth rate
138 decreased with concentration. OD was measured after 48 h of growth with
139 ciprofloxacin. Lines, means of triplicate growth curves; error bars, standard

140 deviations. (F) Richness (number of ASVs in rarefied data) of Pre-SD SIC remained
141 constant during continuous ciprofloxacin treatment. Data are means of two technical
142 replicates. (G) Bacteroidaceae and Lachnospiraceae dominated during three rounds
143 of ciprofloxacin treatment of the Res-SD SIC. Data are the mean of family-level
144 abundances across two technical replicates. (H) Treatment of the residual treatment
145 humanized mouse fecal inoculum (Res-SD) SIC led to highly reproducible outcomes.
146 Shown are comparisons of relative abundance at the ASV level between replicates
147 after 3 passages of growth in BHI with ciprofloxacin. Pearson coefficient (R) and its
148 p -value were computed only from data points present in both samples. ASVs with
149 relative abundance $<10^{-4}$ were set to 10^{-4} for visualization. (I) Treatment outcome of
150 Pre-SD SICs was less reproducible than Res-SD SICs at high concentrations. Shown
151 are comparisons of relative abundance at the ASV level between replicates after 3
152 passages of growth in BHI with ciprofloxacin. Pearson coefficient (R) and its p -value
153 were computed only from data points present in both samples. ASVs with relative
154 abundance $<10^{-4}$ were set to 10^{-4} for visualization. (J) Enterococcaceae can recover
155 after one round of ciprofloxacin treatment in the Res-SD SIC. Data are the mean of
156 two replicates.

157 K-M) SICs derived from pre-exposed inocula show increased resilience to ciprofloxacin
158 treatment. (K) Pre-SD SIC growth rate can recover to levels similar to values before
159 treatment at low concentrations. Maximum growth rate was calculated across

160 ciprofloxacin concentrations during one round of antibiotic treatment and two
161 rounds of recovery. Lines, mean of three technical replicates; error bars, standard
162 deviations. (L) Growth rate recovery after transient ciprofloxacin treatment is linked
163 to the recovery of fast-growing species. Maximum growth rate of SICs grown in BHI
164 after one round of treatment with ciprofloxacin and two rounds without drug. SICs
165 were classified by their summed relative abundances of Enterococcaceae and
166 Enterobacteriaceae ($f_{\text{Enterococcaceae}}$). Black circles are the mean maximum growth rate for each
167 group ($n=21$ for $f_{\text{Enterococcaceae}} < 20\%$ and $n=3$ for $f_{\text{Enterococcaceae}} > 20\%$), error bars are standard
168 deviations. Individual data points are plotted in gray. p -value is from a Student's
169 two-sided t -test between the two groups. (M) Erysipelotrichaceae recovery is
170 reversed by a second ciprofloxacin treatment. Data are the family-level mean
171 \log_{10} (relative abundance) of two replicates during one round of ciprofloxacin
172 treatment followed by one round of recovery and a second treatment.

173 N,O) Bacteroidaceae are remodeled in the Res-SD SIC during and after ciprofloxacin
174 treatment, with *B. vulgatus* as the only member to generally survive. Relative
175 abundances of *Bacteroides* species in the Res-SD SIC during continuous ciprofloxacin
176 treatment (N) or after one round of treatment followed by two rounds of recovery
177 (O).