

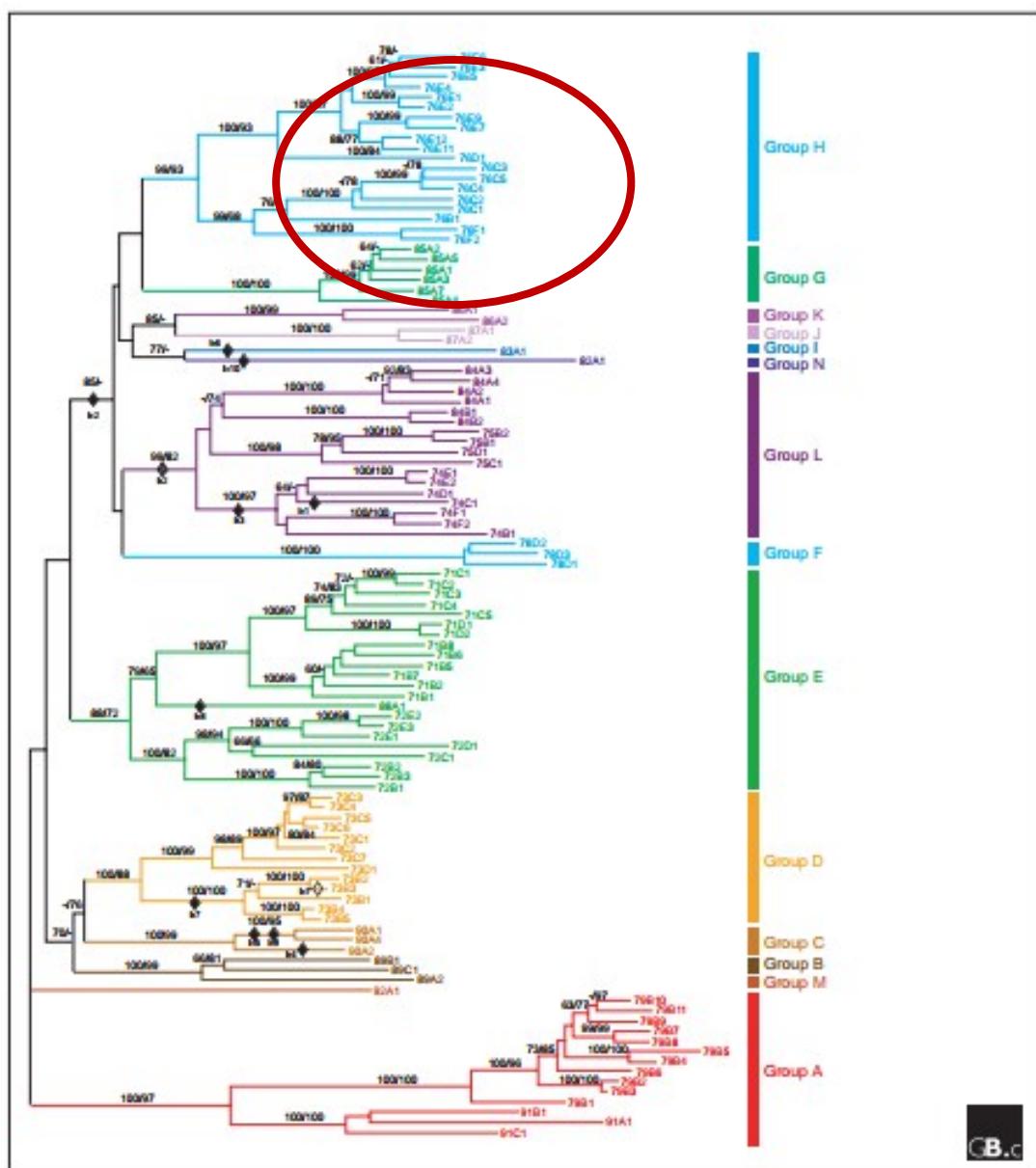
# High Throughput Mass Spectrometry-based Characterisation of *Arabidopsis thaliana* GROUP H Glycosyltransferases

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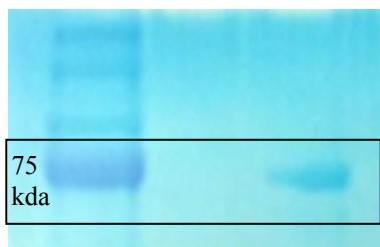
## Phylogenetic classification of *Arabidopsis thaliana*



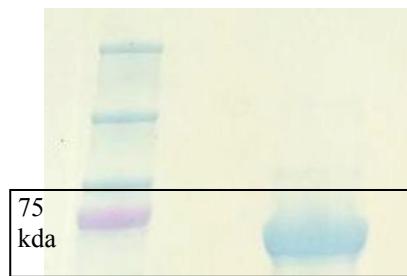
**Fig 1: Phylogenetic grouping of *Arabidopsis thaliana* UGTs (Ross et al., 2001)**

## RESULTS AND DISCUSSION

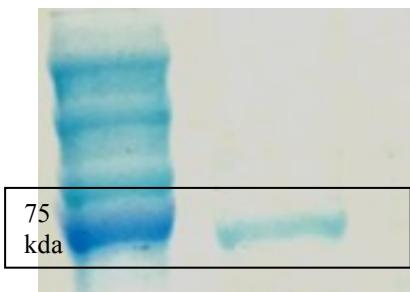
### Protein purification; SDS PAGE



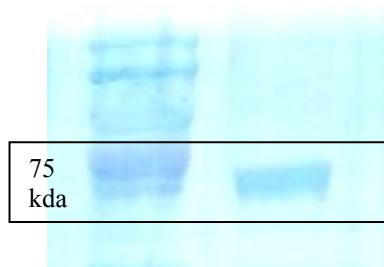
**Fig 2a:** Lane 1: Protein ladder; Lane 3:Purified 76E5 MW ( 80kda)



**Fig 2b:** Lane 1: Protein ladder Lane 3:Purified 76E1 (MW 65kda)



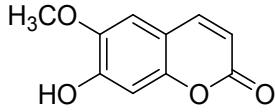
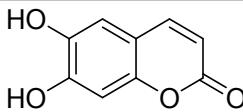
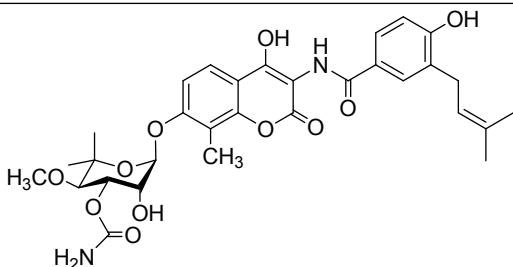
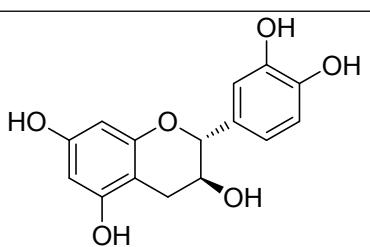
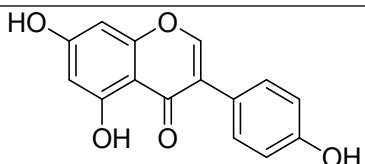
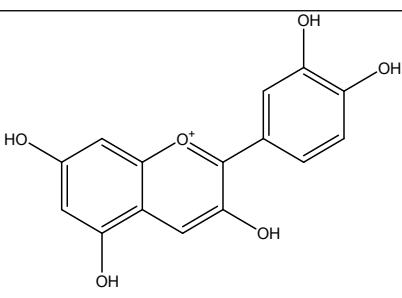
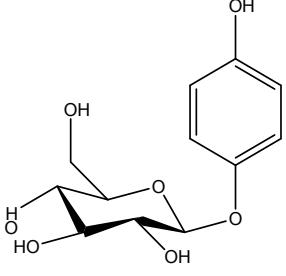
**Fig 2c:** Lane 1: Protein ladder  
Lane 3:Purified 76D1 (MW 65kda)

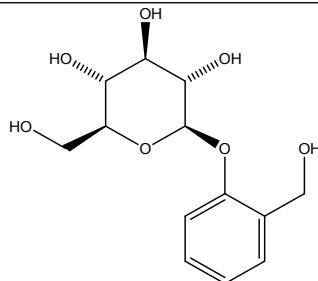
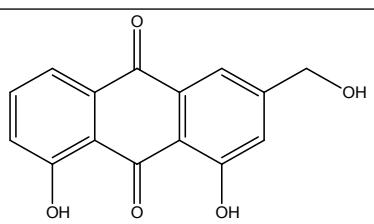
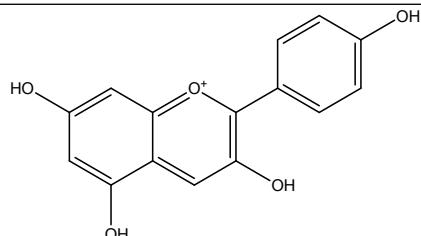
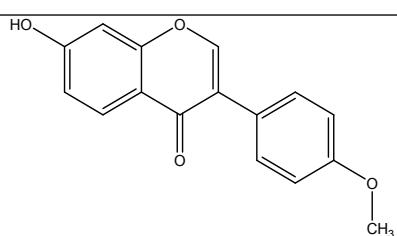
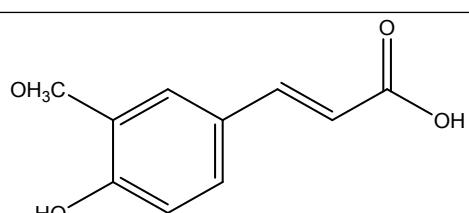
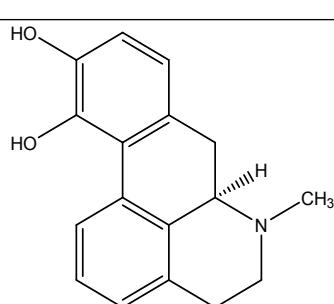
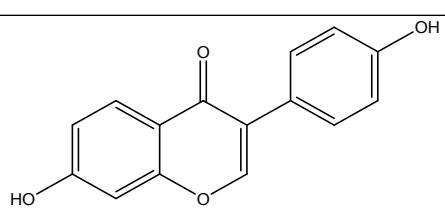


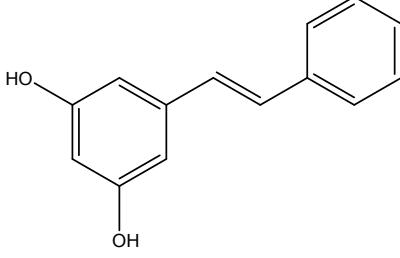
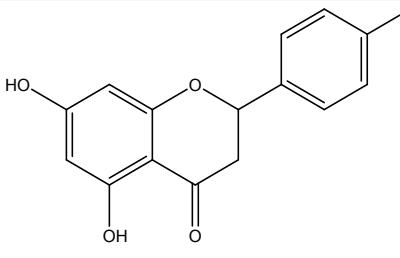
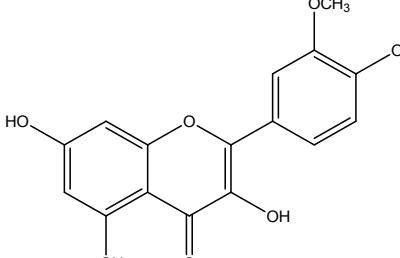
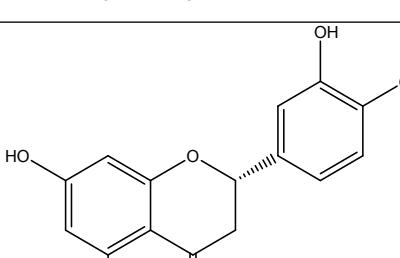
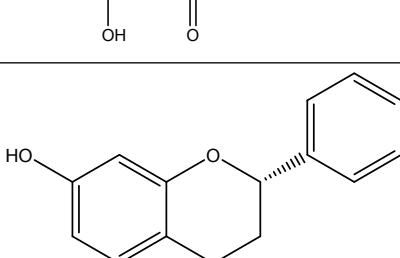
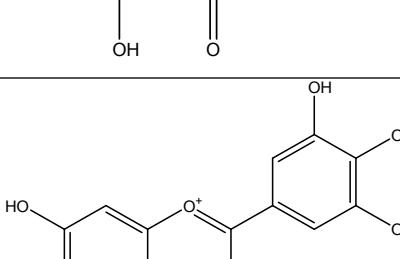
**Fig 2d:** Lane 1: Protein ladder Lane 3:Purified 76E2 (MW 65kda)

### ACCEPTOR LIBRARY USED

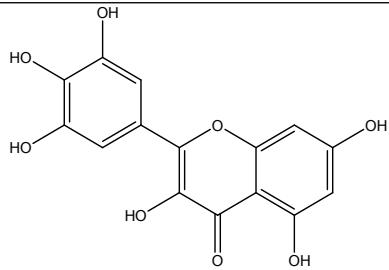
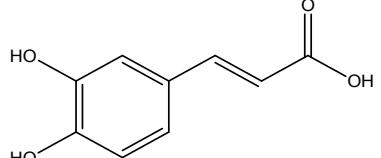
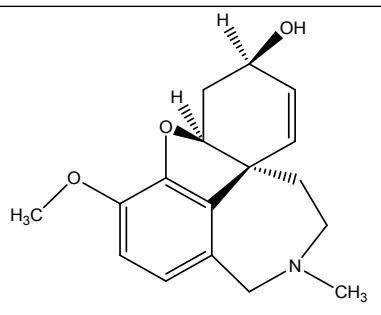
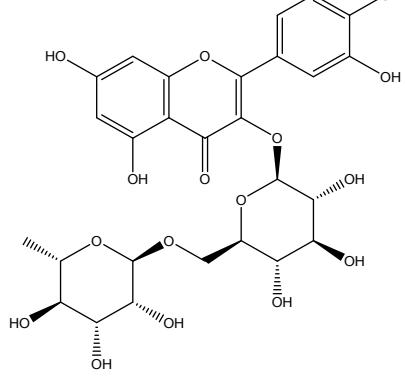
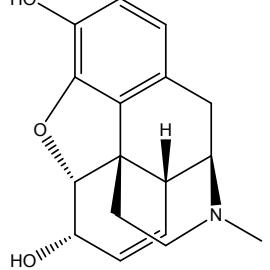
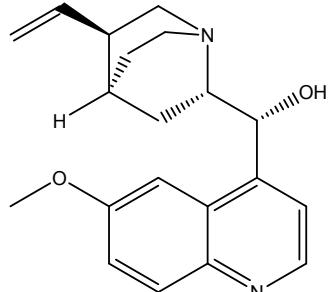
	Name	Structure	Molecular weight
1.	Umbelliferone		<b>162.0</b>
2.	Sinapic acid		<b>224.0</b>
3.	3,4-dihydroxybenzoic acid		<b>154.0</b>
4.	2,5-dihydroxybenzoic acid		<b>154.0</b>
5.	Abscisic acid		<b>264.0</b>
6.	Zeatin		<b>219.0</b>
7.	Quercetin		<b>302.0</b>
8.	Fisetin		<b>286.0</b>
9.	Kaempferol		<b>286.0</b>

10.	Scopoletin		<b>192.0</b>
11.	Esculetin		<b>178.0</b>
12.	Novobiocin		<b>613.0</b>
13	Catechin		<b>290.0</b>
14	Genistein		<b>270.0</b>
15	Cyanidin		<b>286.0</b>
16	Arbutin		<b>272.0</b>

17	Salicin		<b>286.0</b>
18	Aloe-emodin		<b>270.0</b>
19	Pelargonidin		<b>271.0</b>
20	Formononetin		<b>268.0</b>
21	Ferulic acid		<b>194.0</b>
22	Apomorphine		<b>267.0</b>
23	Daidzein		<b>254.0</b>

24	Resveratrol		<b>228.0</b>
25	Apigenin		<b>270.0</b>
26	Isorhamnetin		<b>316.0</b>
27	Hesperetin		<b>302.0</b>
28	Naringenin		<b>272.0</b>
29	Delphinidin		<b>303.0</b>

30	Galangin		<b>270.0</b>
31	Morin		<b>302.0</b>
32	Biochanin A		<b>284.0</b>
33	Eriodictyol		<b>288.0</b>
34	Diosmetin		<b>300.0</b>
35	Chrysin		<b>254.0</b>
36	Dihydromyricetin		<b>320.0</b>

37	Myricetin		<b>318.0</b>
38	Caffeic acid		<b>180.0</b>
39	Galantamine		<b>287.0</b>
40	Rutin		<b>611.0</b>
41	Morphine		<b>285.0</b>
42	Quinine		<b>324.0</b>

**Table 1: Library of acceptor compounds screened**

## MASS SPECTRA SUMMARISED IN GAR SCREEN

### A) 76E1 Acceptor screening result

MS spectra; UDP Glucose (MW 162) was added to the test aglycones. m/z of target peaks (glycosylated products) are shown in bold in the red boxes below.

#### i) Apigenin

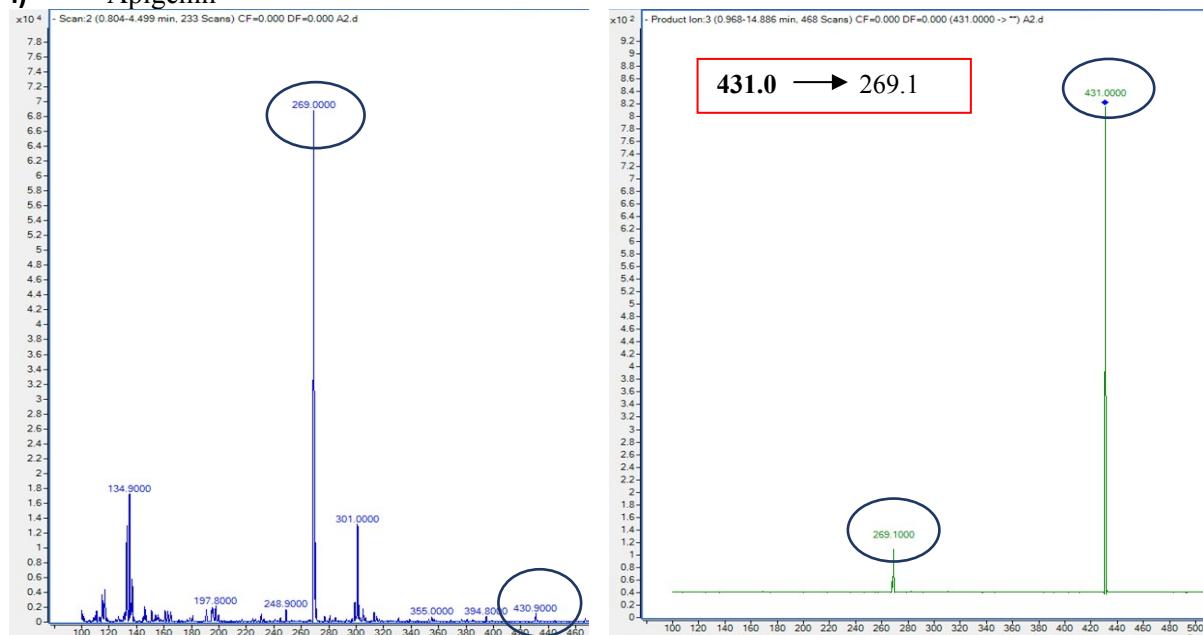


Fig 3a: MS spectrum of 76E1 glycosylation of apigenin (Full scan)

Fig 3b: MS spectrum showing fragmentation of apigenin glucoside (Product ion scan)

MS/MS was used to fragment apigenin glucoside (MW 431). The observation of apigenin (MW 269) confirms the glucoside formation, and hence the glycosylation process.

#### ii) Esculetin

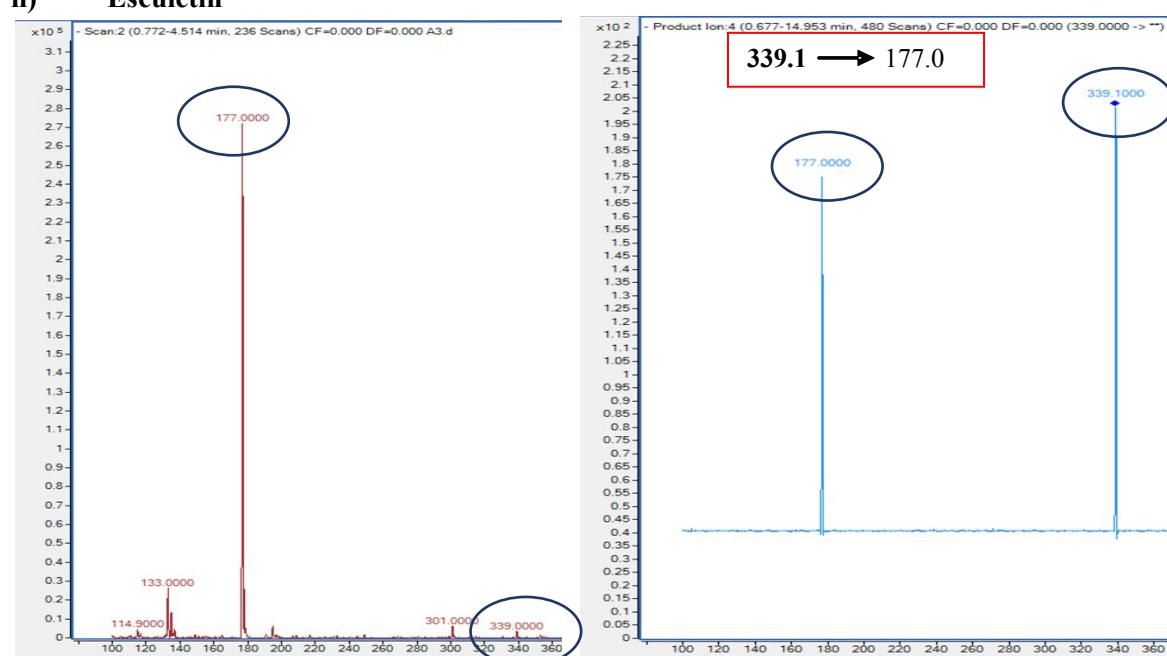


Fig 3c: MS spectrum of 76E1 glycosylation of esculetin (Full scan) 3d: MS spectrum showing fragmentation of esculetin glucoside (Product ion scan)

iii) Quercetin

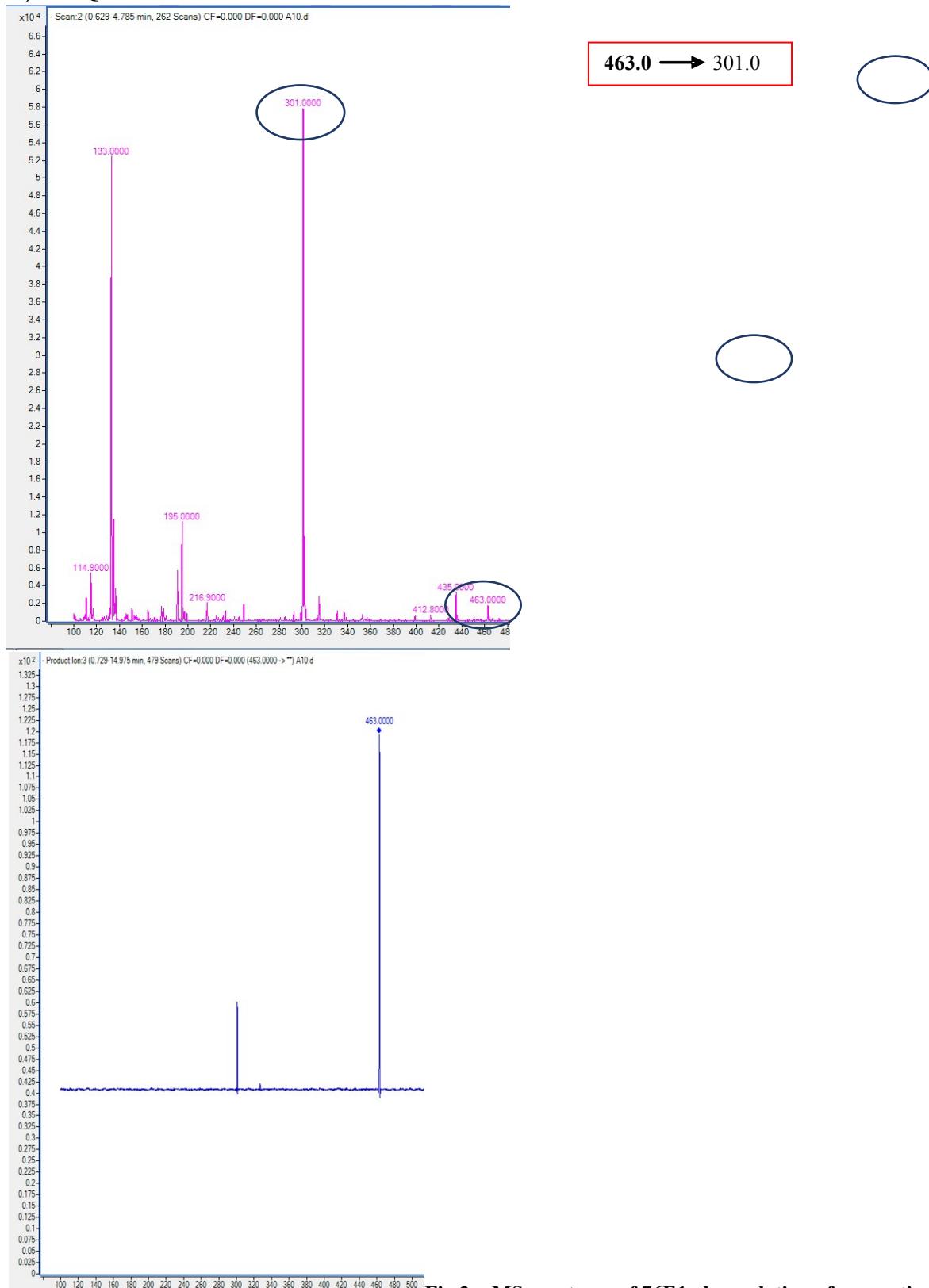
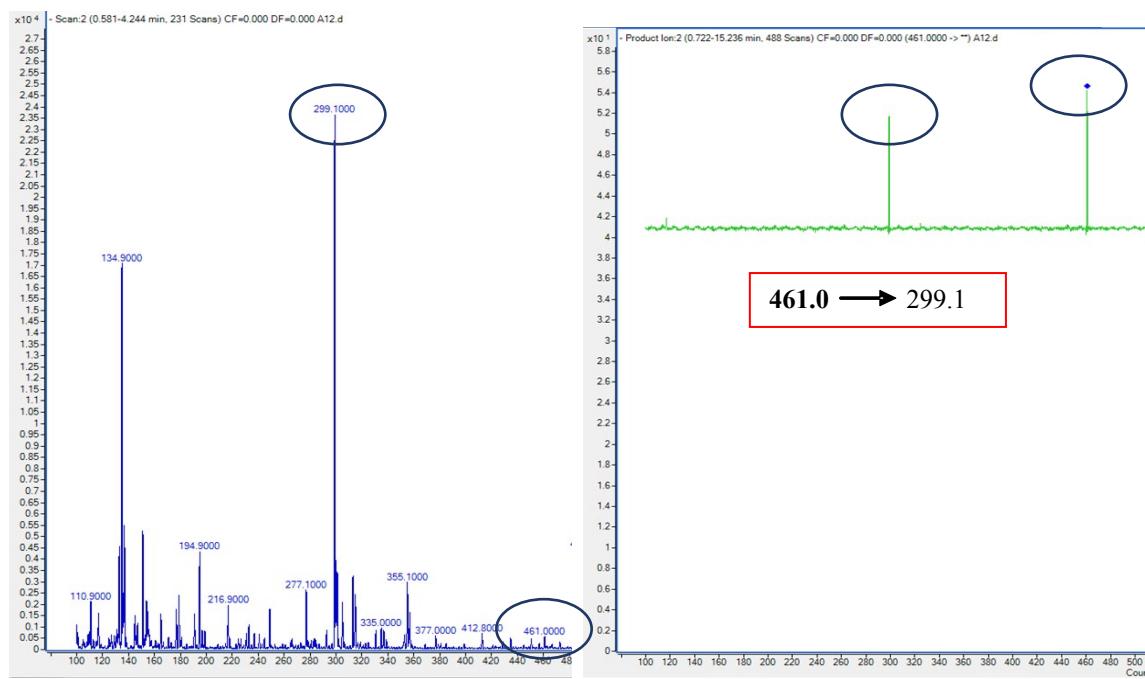


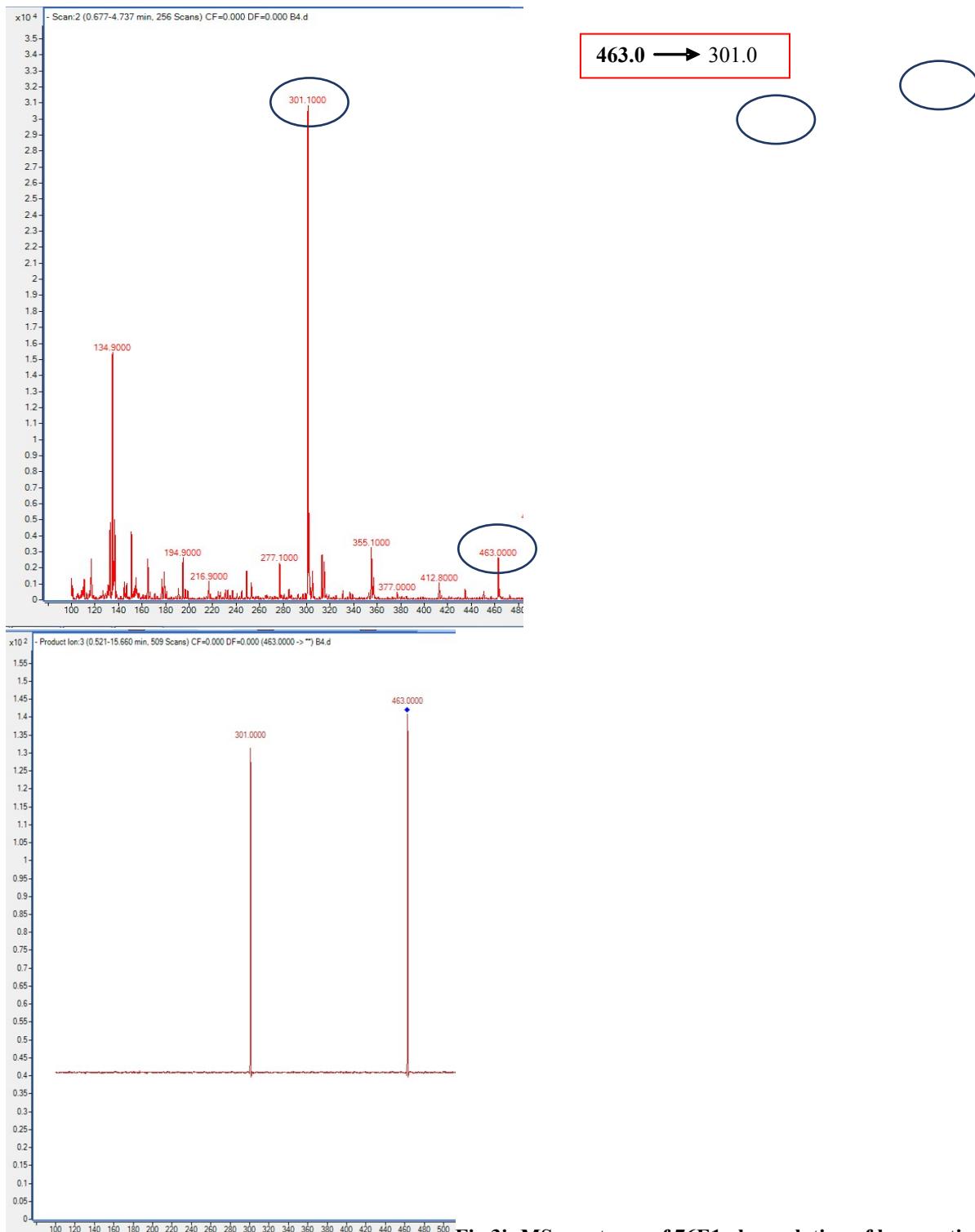
Fig 3e: MS spectrum of 76E1 glycosylation of quercetin  
(Full scan) 3f: MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)

iv) Diosmetin

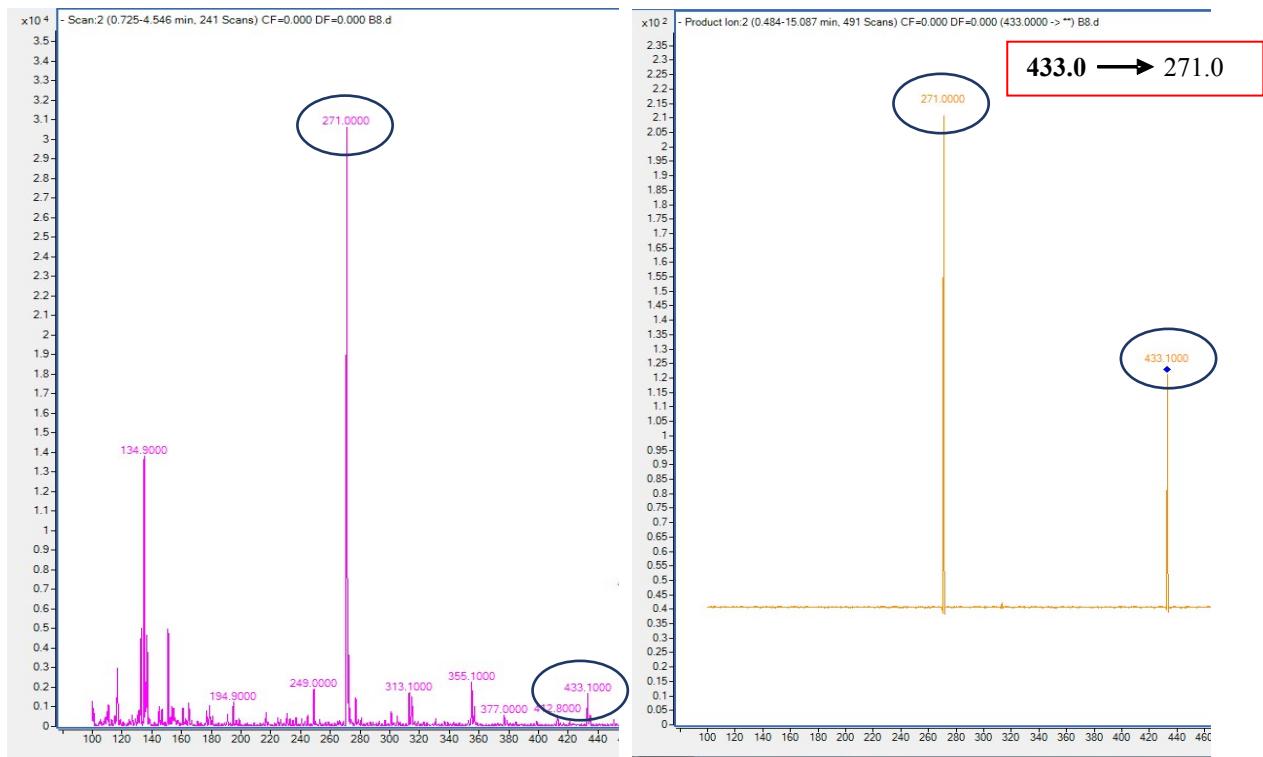


**Fig 3g: MS spectrum of 76E1 glycosylation of diosmetin (Full scan) 3h: MS spectrum showing fragmentation of diosmetin glucoside (Product ion scan)**

v) **Hesperetin**

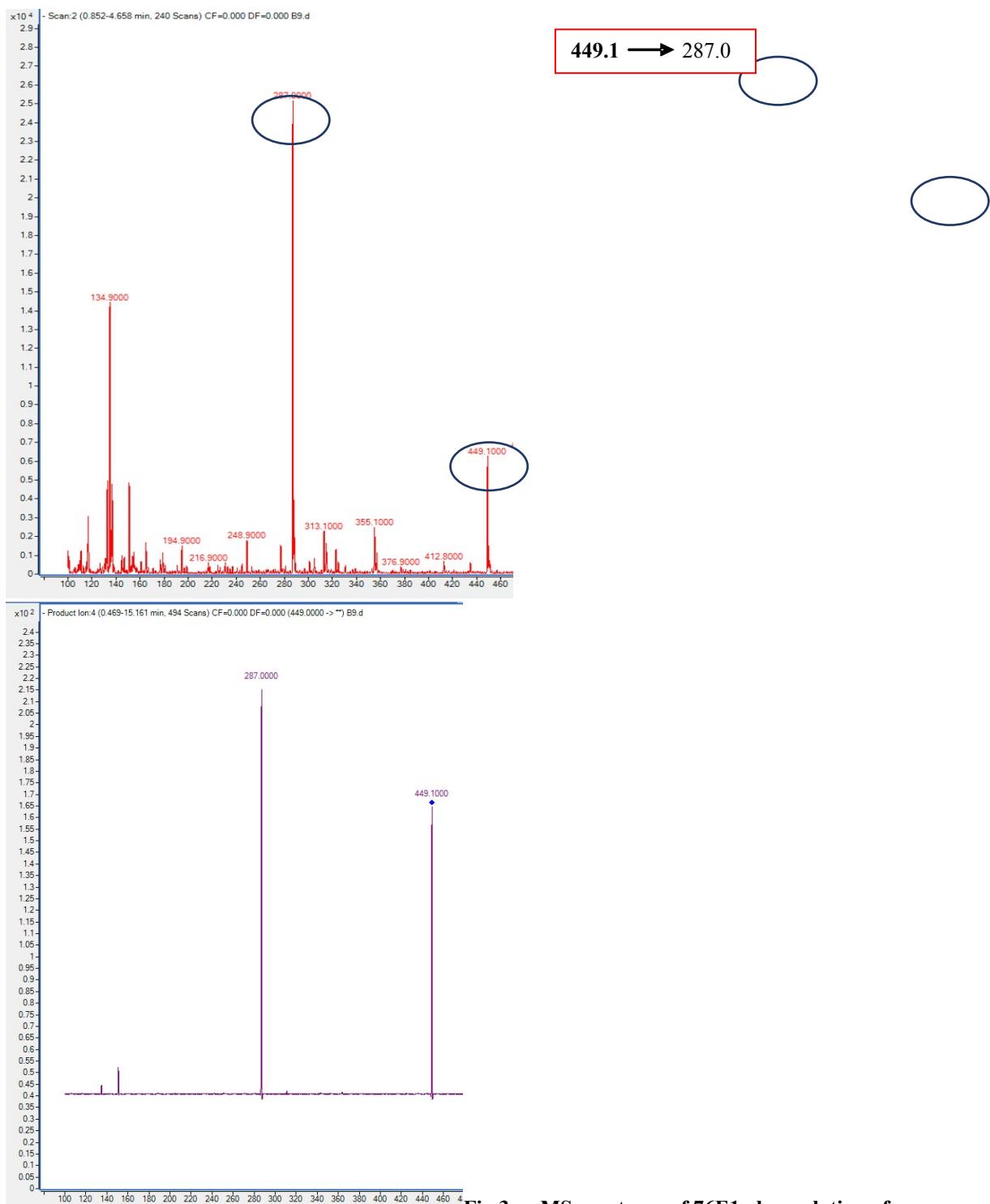


**vi) Naringenin**



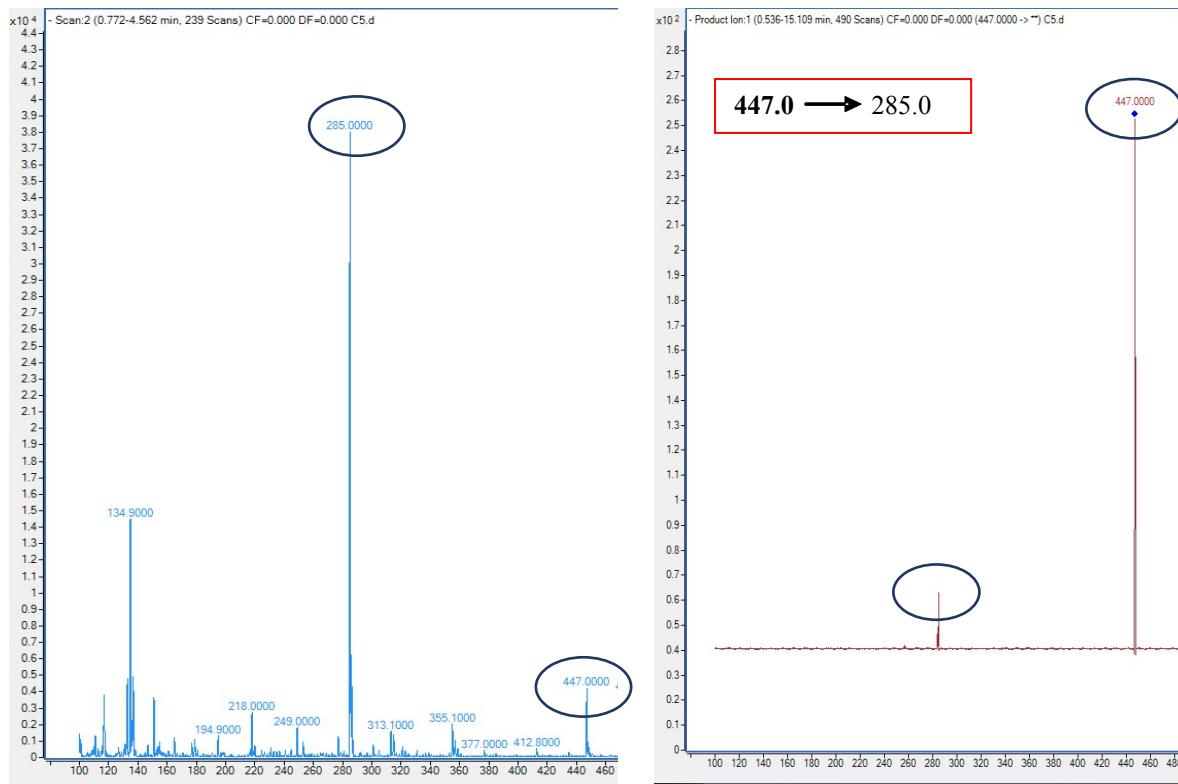
**Fig 3k: MS spectrum of 76E1 glycosylation of naringenin (Full scan) 3l: MS spectrum showing fragmentation of naringenin glucoside (Product ion scan)**

vii) Eriodictyol



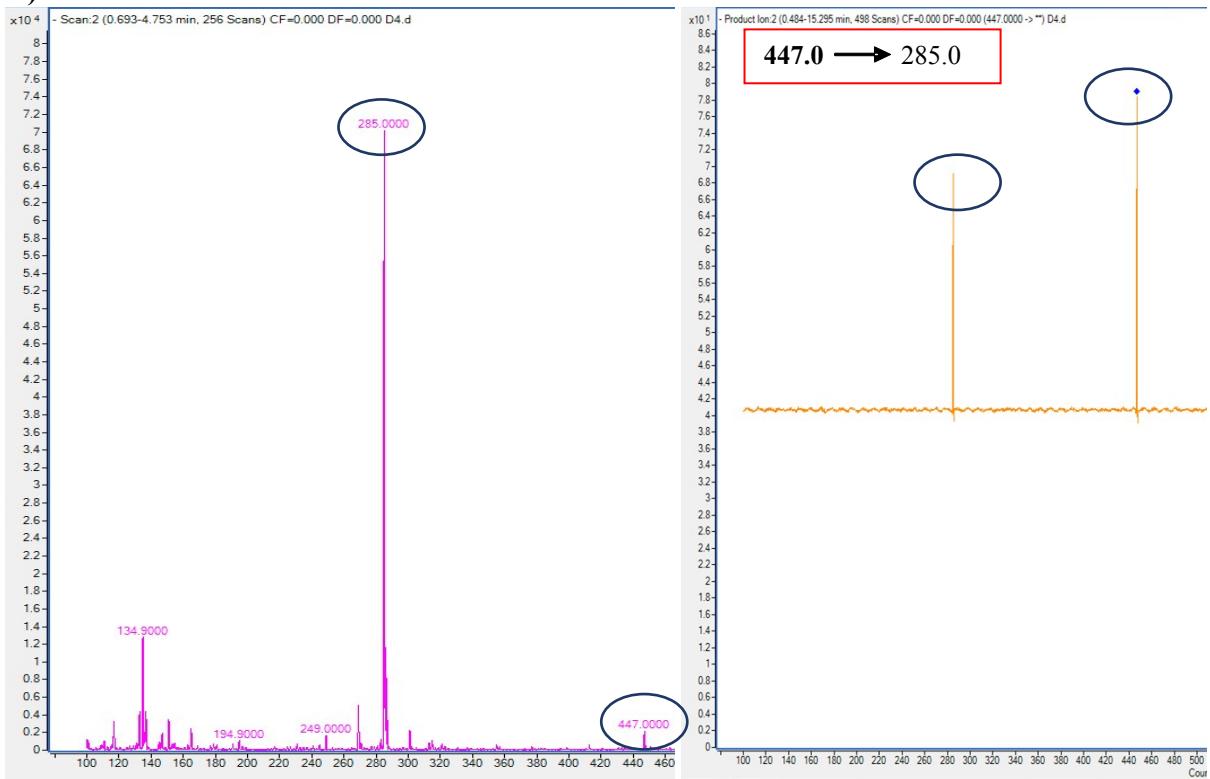
**Fig 3m:** MS spectrum of 76E1 glycosylation of eriodictyol (Full scan) **3n:** MS spectrum showing fragmentation of eriodictyol glucoside (Product ion scan)

### viii) Kaempferol



**Fig 3o:** MS spectrum of 76E1 glycosylation of kaempferol (Full scan) 3p: MS spectrum showing fragmentation of kaempferol glucoside (Product ion scan)

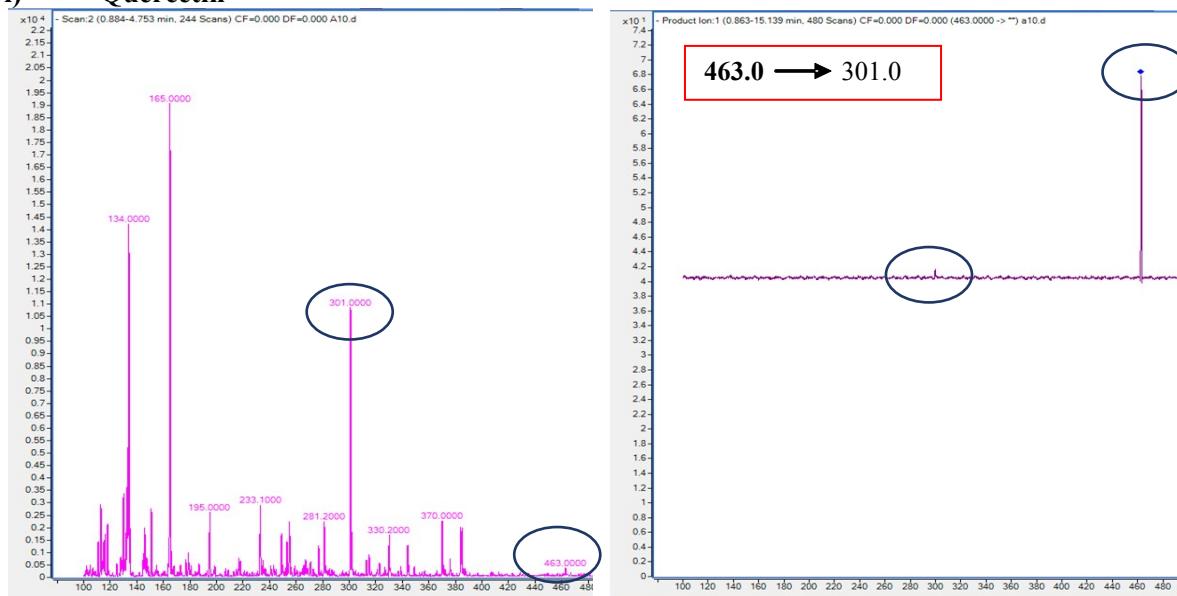
### ix) Fisetin



**Fig 3q:** MS spectrum of 76E1 glycosylation of fisetin (Full scan) 3r: MS spectrum showing fragmentation of fisetin glucoside (Product ion scan)

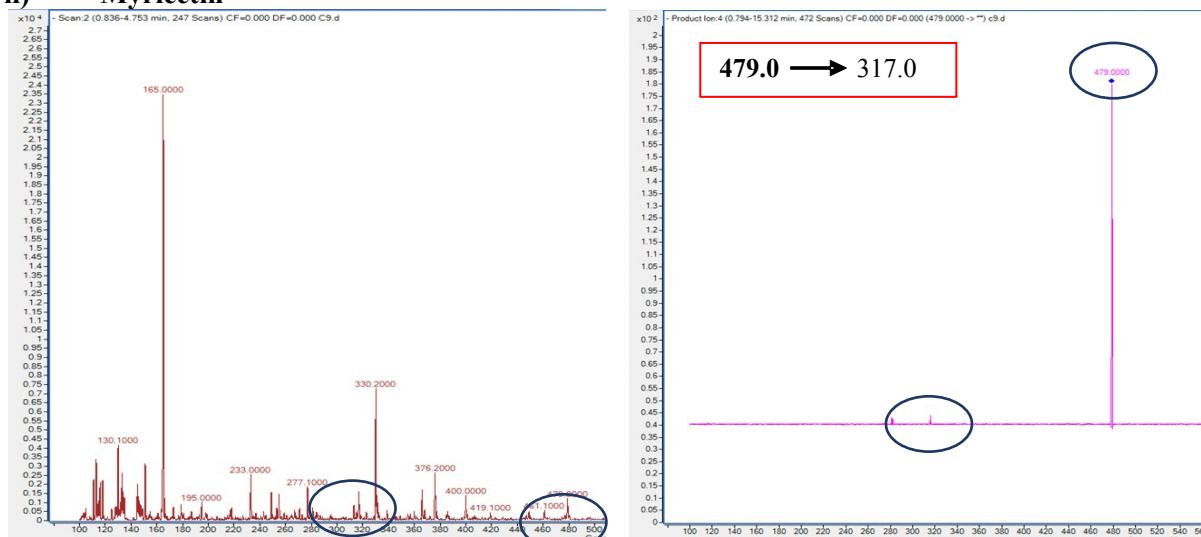
### B) 76E2 Acceptor screening

i) **Quercetin**



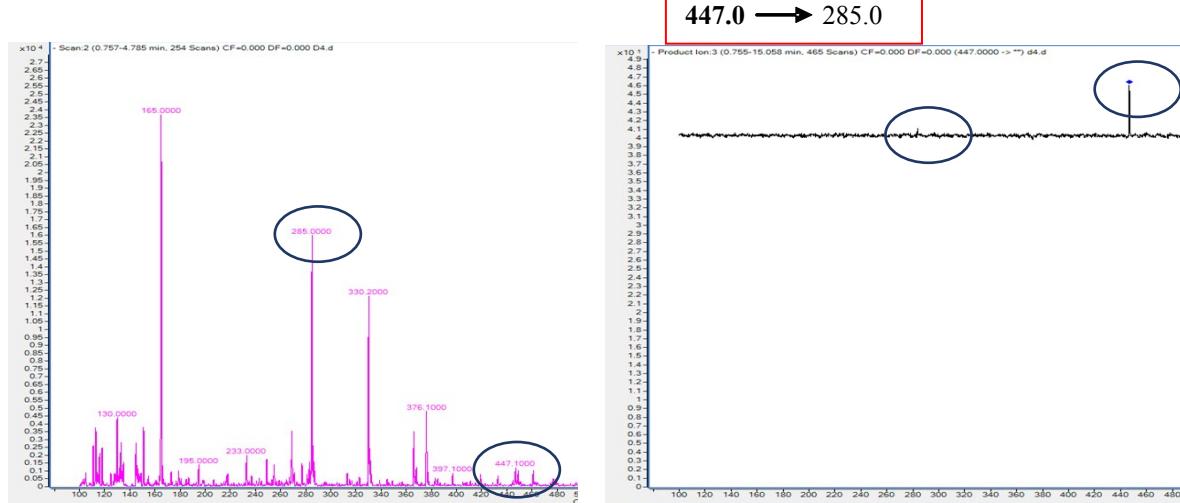
**Fig 4a:** MS spectrum of 76E2 glycosylation of quercetin (Full scan) **4b:** MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)

ii) **Myricetin**

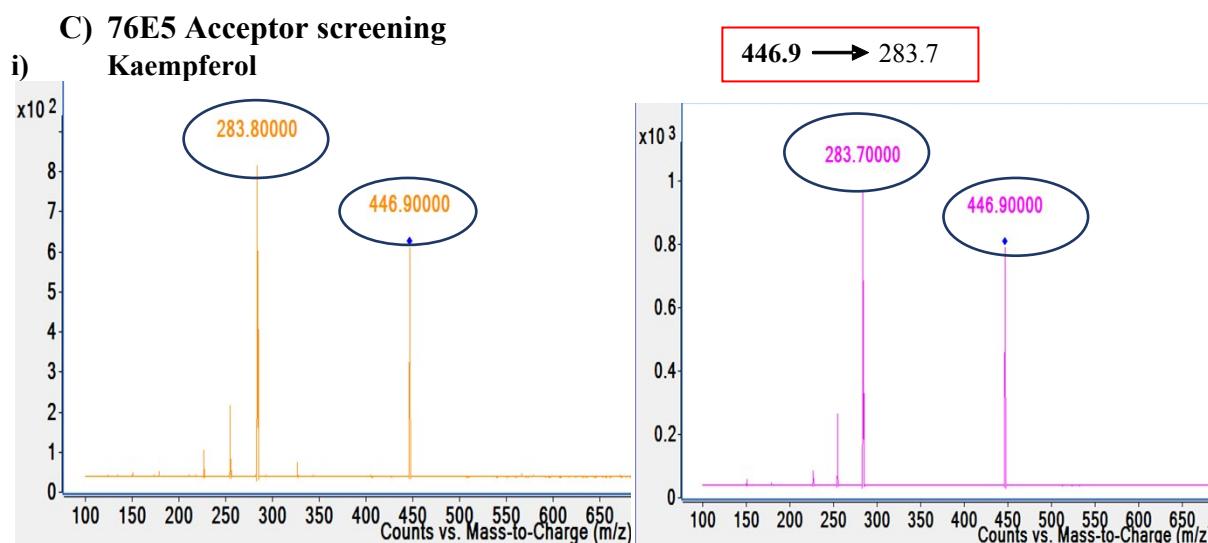


**Fig 4c:** MS spectrum of 76E2 glycosylation of myricetin (Full scan) **4d:** MS spectrum showing fragmentation of myricetin glucoside (Product ion scan)

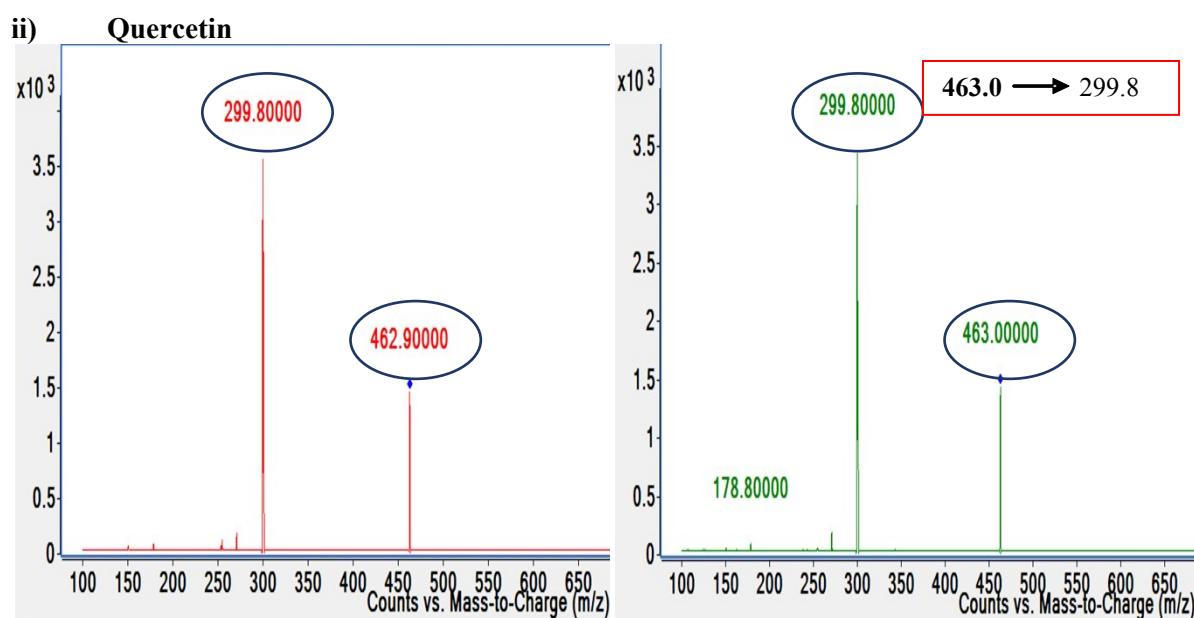
iii) **Fisetin**



**Fig 4e:** MS spectrum of 76E2 glycosylation of fisetin (Full scan) **4f:** MS spectrum showing fragmentation of fisetin glucoside (Product ion scan)

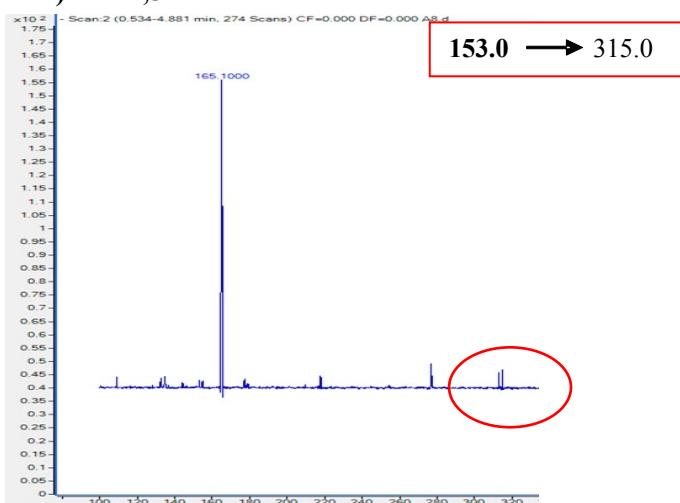


**Fig 5a:** MS spectrum of 76E5 glycosylation of kaempferol (Full scan) **5b:** MS spectrum showing fragmentation of kaempferol glucoside (Product ion scan)

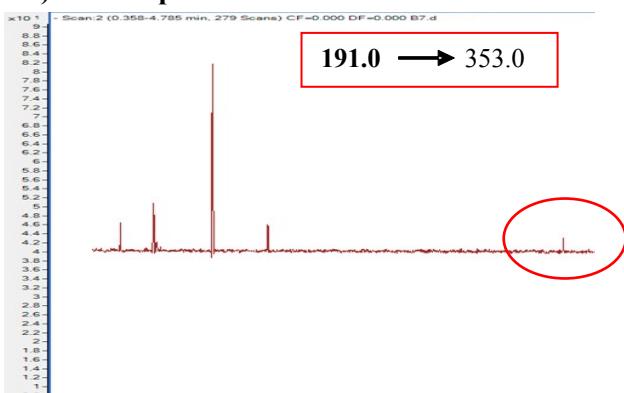


**Fig 5c: MS spectrum of 76E5 glycosylation of quercetin (Full scan) 5d: MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)**

iii) 2,5DHBA

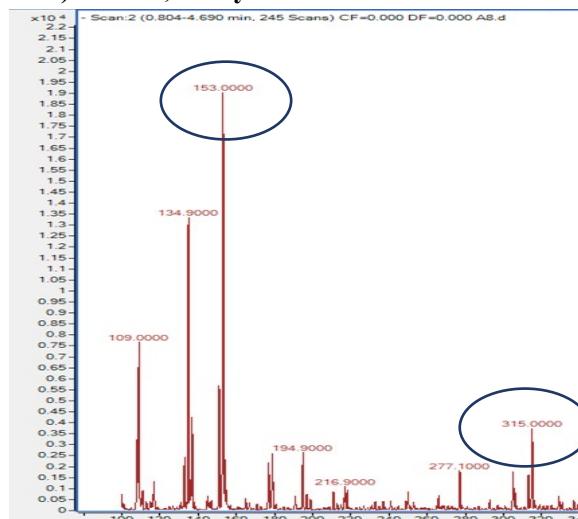


iv) Scopoletin



D) 7

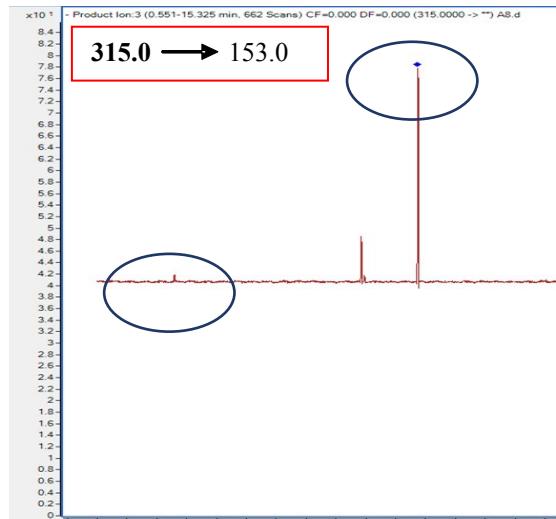
i) 2,5 dihydrobenzoic acid



**Fig 5e: MS spectrum of 76E5 glycosylation of 2,5-dihydroxybenzoic acid (Full scan)**

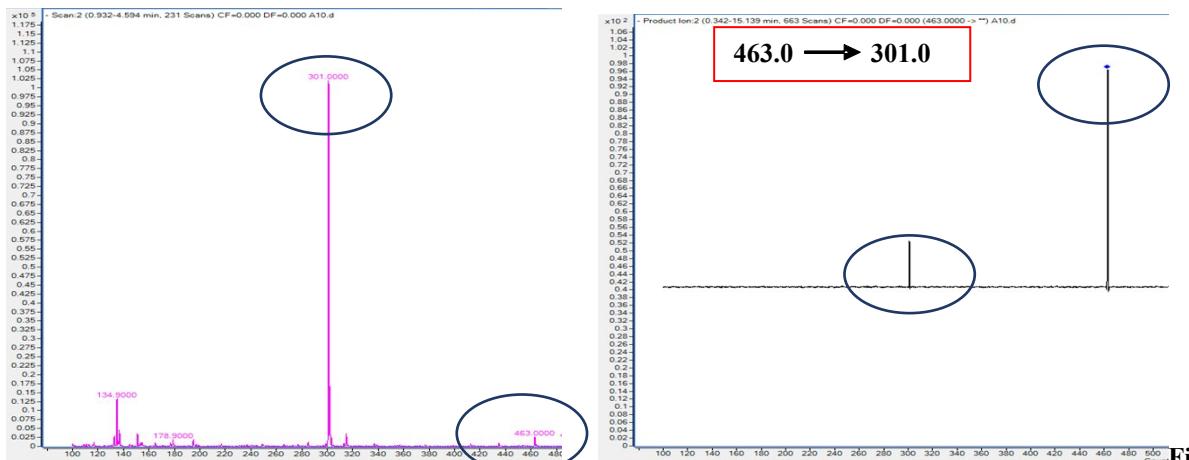
**Fig 5f: MS spectrum of 76E5 glycosylation of scopoletin (Full scan)**

315.0 → 153.0



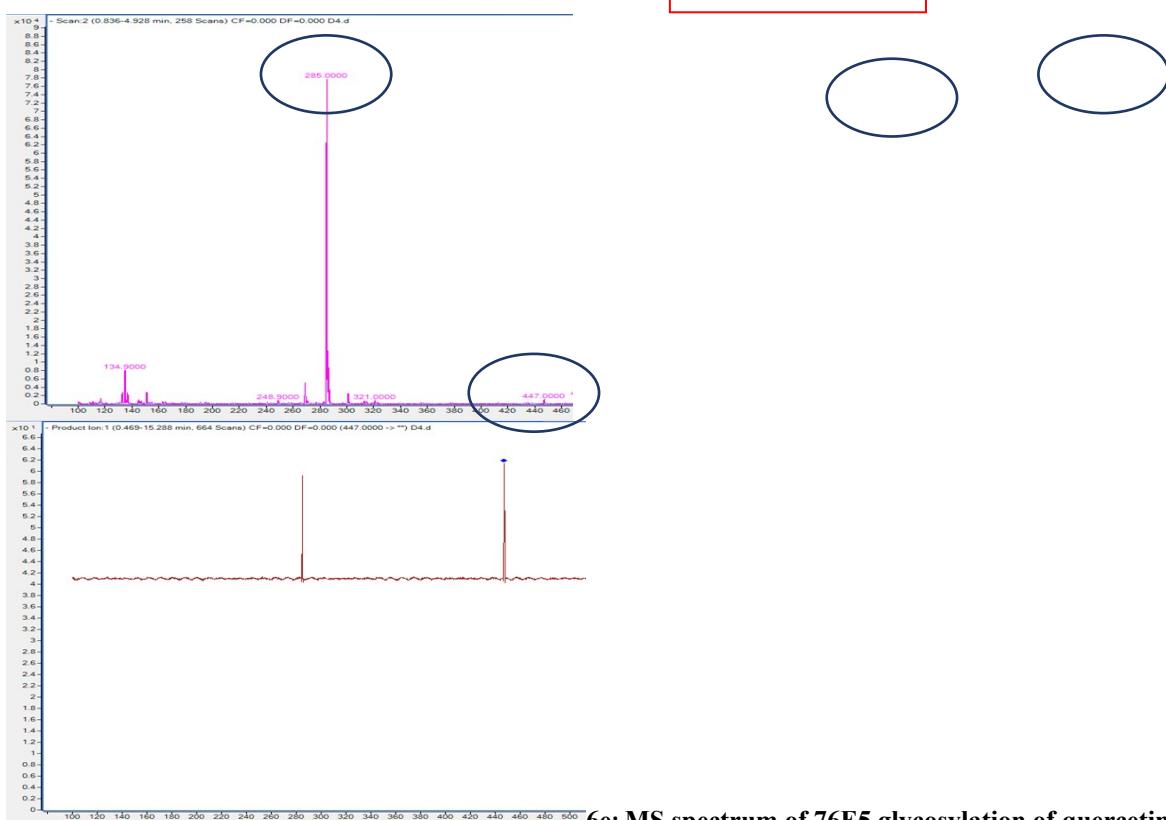
**Fig 6a: MS spectrum of 76E5 glycosylation of 2,5 dihydrobenzoic acid (Full scan) 6b: MS spectrum showing fragmentation of 2,5 dihydrobenzoic acid glucoside (Product ion scan)**

ii) Quercetin



**6c:** MS spectrum of 76E5 glycosylation of quercetin (Full scan) **6d:** MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)

### iii) Fisetin

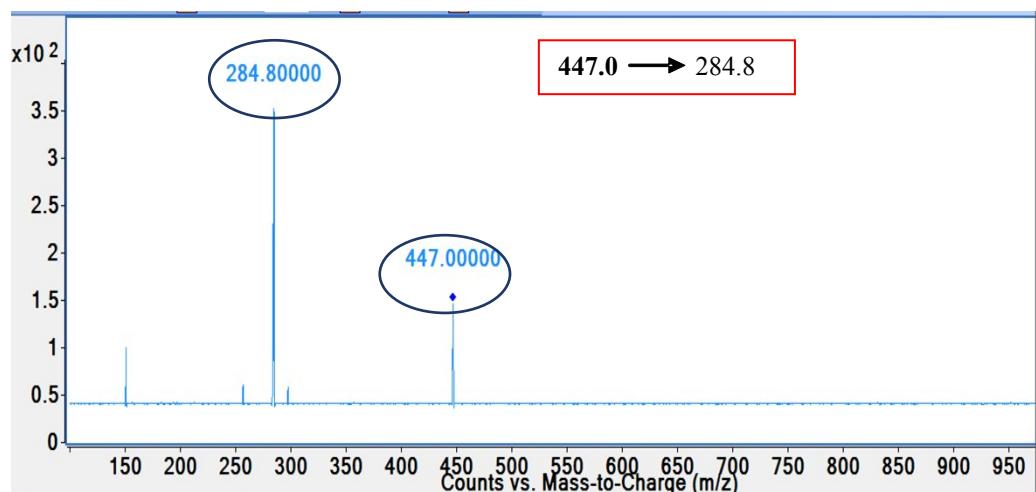


**6e:** MS spectrum of 76E5 glycosylation of quercetin (Full scan) **6f:** MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)

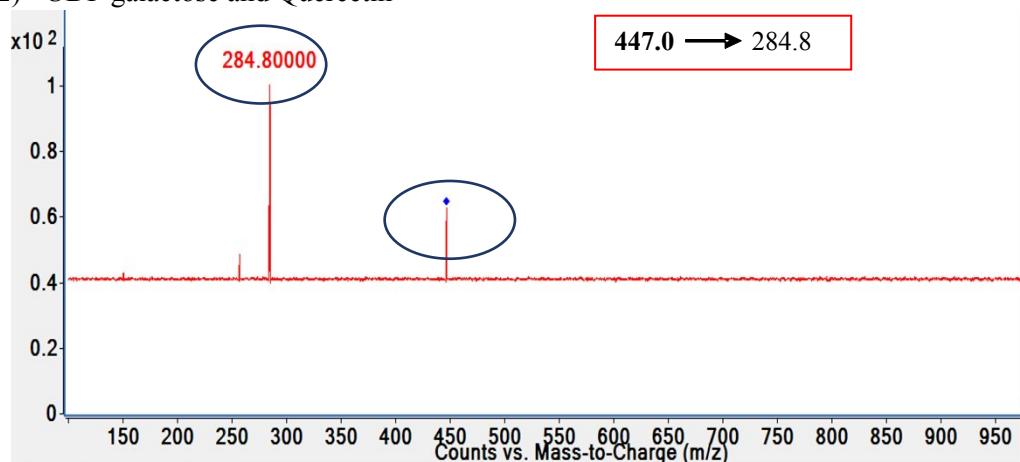
## DONOR SCREENING RESULT AND GLYCOSYLATION POSITION

### A) 76E1 Donor screening results

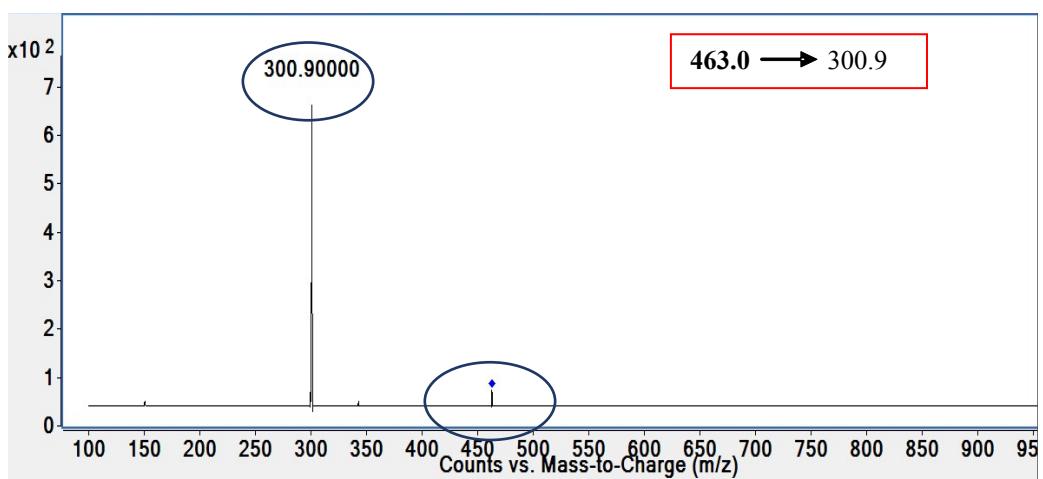
- 1) UDP glucose and Kaempferol



2) UDP galactose and Quercetin



3) UDP glucose and Quercetin



4) UDP glcnac and Quercetin

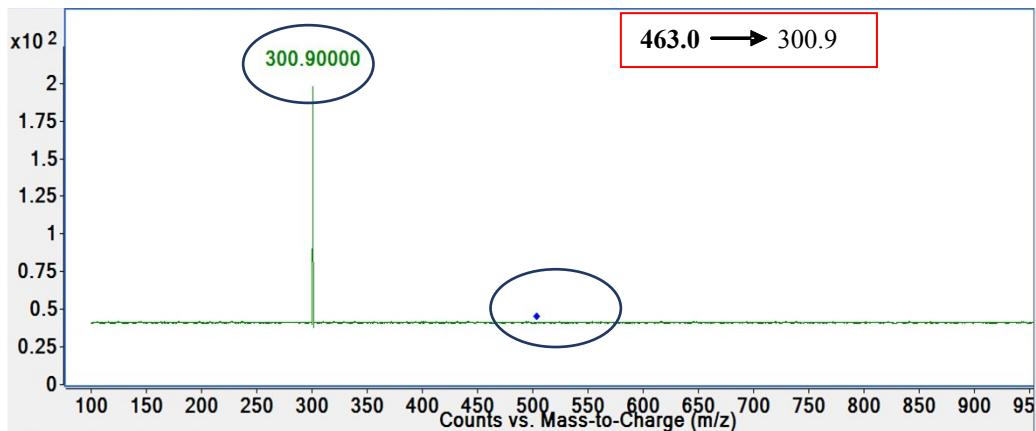


Fig 7: MS spectrum of 76E1 recognition of UDP glc, gal and glenac

**B) 76E2 Donor screening  
UDP glucose and Quercetin**

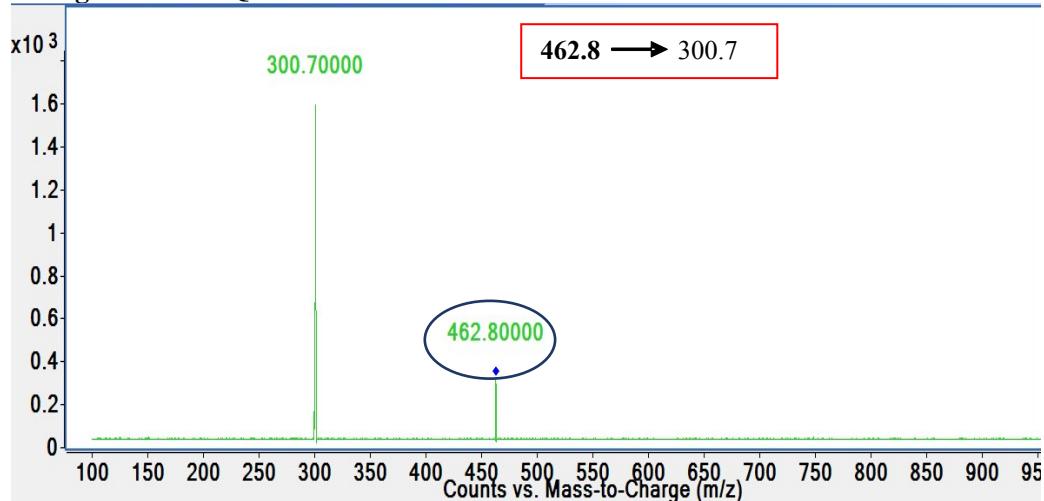
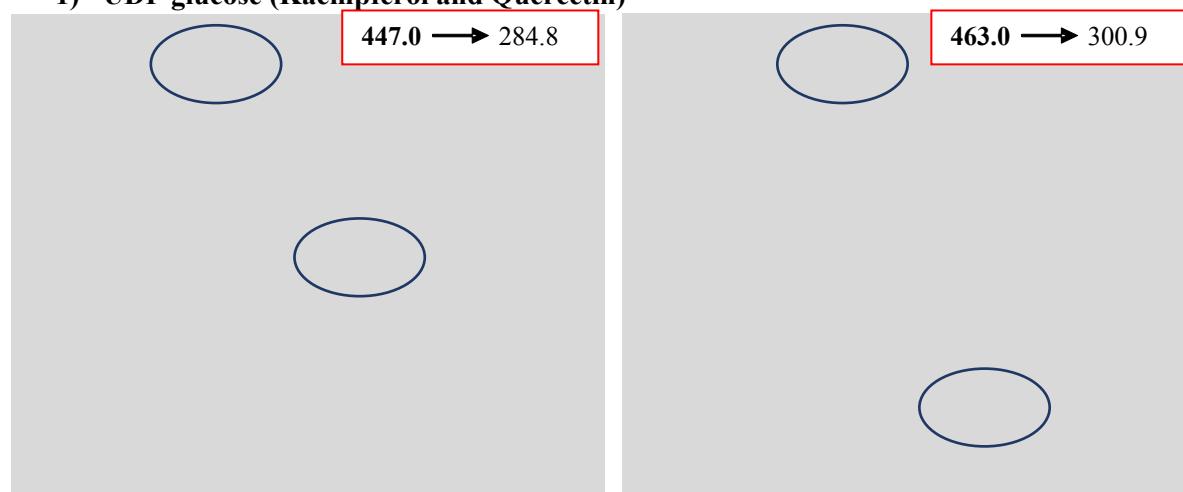


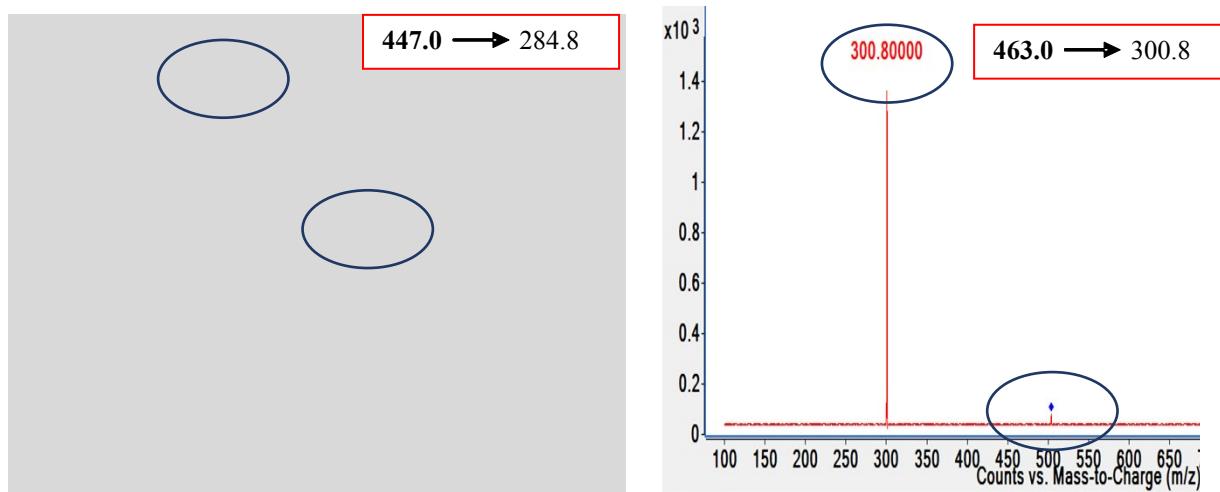
Fig 8: MS spectrum of 76E2 recognition of UDP glc

**C) 76E5 Donor screening**

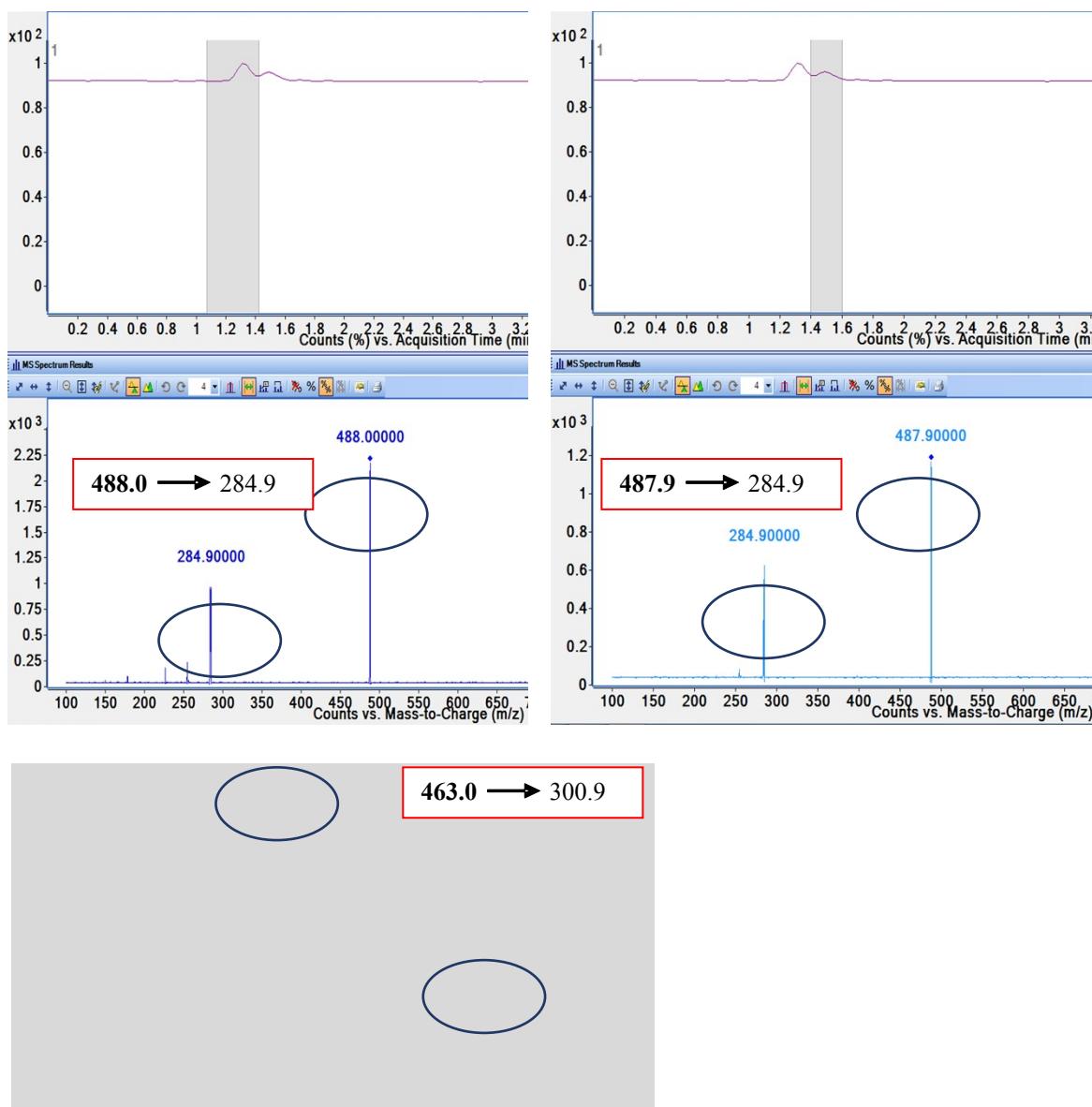
**1) UDP glucose (Kaempferol and Quercetin)**



**2) UDP galactose (Kaempferol and Quercetin)**

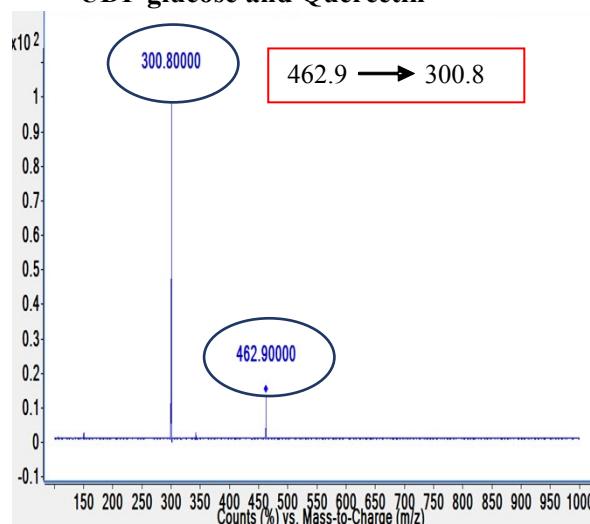


### 3) UDP glcnac (Kaempferol and Quercetin)



**Fig 9: MS spectrum of 76E5 recognition of UDP glc, gal and glcnac**  
**D) 76D1 Donor screening**

i) UDP glucose and Quercetin



**Fig 10:** MS spectrum of 76E5 recognition of UDP glc

**Table showing m/z of precursor ion and product ion for MS/MS data**

Enzyme	Reaction	Precursor ion (Glycoside peaks)	Product ion
76E1	UDP Glc/Gal + Kaempferol	<b>447</b>	<b>285</b>
	UDP Glc + Quercetin	<b>463</b>	<b>301</b>
	UDP GlcNAc + Quercetin	<b>504</b>	<b>301</b>
76E2	UDP Glc + Quercetin	<b>463</b>	<b>301</b>
76D1	UDP Glc + Quercetin	<b>463</b>	<b>301</b>
76E5	UDP Glc/Gal + Kaempferol	<b>447</b>	<b>285</b>
	UDP Glc/Gal + Quercetin	<b>463</b>	<b>301</b>
	UDP GlcNAc + Kaempferol	<b>488</b>	<b>285</b>
	UDP GlcNAc + Quercetin	<b>504</b>	<b>301</b>
76E2 N320S	UDP Glc/Gal + Kaempferol	<b>447</b>	<b>285</b>
	UDP Glc/Gal + Quercetin	<b>463</b>	<b>301</b>
	UDP GlcNAc + Kaempferol	<b>488</b>	<b>285</b>
	UDP GlcNAc + Quercetin	<b>504</b>	<b>301</b>

**Table 2:** m/z of precursor and product ions

## Sanger Sequencing

The sequencing of mutant plasmids was carried out by Source Bioscience to confirm the incorporation of the mutation by template DNAs. The sequence was compared with wild type DNA sequences using NCBI BLAST.

### 76E1 S318N

Score	Expect	Identities	Gaps	Strand	Frame
2002 bits(1084)	0.0()	1092/1098(99%)	0/1098(0%)	Plus/Minus	
<b>Features:</b>					
Query 72	CTACATGAAATTCACTCATTTTAAGGAATTGACAAAGTTGTCTAATGAGCTGAATGAGGA				131
Sbjct 1362	CTACATGAAATTCACTCATTTTAAGGAATTGACAAAGTTGTCTAATGAGCTGAATGAGGA				1303
Query 132	ACCTCTACTCTTGACAGAGGCTTGAAGCTTCTCTTCAAGATTGATAACTCTCTCCAT				191
Sbjct 1302	ACCTCTACTCTTGACAGAGGCTTGAAGCTTCTCTTCAAGATTGATAACTCTCTCCAT				1243
Query 192	TTCTGCTCCTTCTCATCCATAATCAATCTCTACAGCTCTCTCCACTGTTCCATTATC				251
Sbjct 1242	TTCTGCTCCTTCTCATCCATAATCAATCTCTACAGCTCTCTCCACTGTTCCATTATC				1183
Query 252	CAGCTCTCCTTCCATTGAACCCCCATTCTCAAACCTCTCTAAGTAACCTCGCATTGAC				311
Sbjct 1182	CAGCTCTCCTTCCATTGAACCCCCATTCTCAAACCTCTCTAAGTAACCTCGCATTGAC				1123
Query 312	TTTCTGATCTCCGTAAAGGCCAACAGATCATCGAACCTCTTCCCCATGCTCTCTAG				371
Sbjct 1122	TTTCTGATCTCCGTAAAGGCCAACAGATCATCGAACCTCTTCCCCATGCTCTCTAG				1063
Query 372	GGTCGAGTTCCATCCGCAGTGACTCCAAAAACCCCTCCACTGCAGGATGCTGAGAACCTC				431
Sbjct 1062	GGTCGAGTTCCATCCGCAGTGACTCCAAAAACCCCTCCACTGCAGGATGCTGAGAACCTC				1003
Query 432	TATCTGTGGTGCCTATTCAAAATGTAACCTCTTCTGAAACCAACCAATTAAATTCCCTC				491
Sbjct 1002	TATCTGTGGTGCCTATTCAAAATGTAACCTCTTCTGAAACCAACCAACTAAATTCCCTC				943
Query 492	CGGTAAGACTCTGCCATTCCGAGCCGGAAATGAAACCCGGTCGGATCACCCATAAGAA				551
Sbjct 942	CGGTAAGACTCTGCCATTCCGAGCCGGAAATGAAACCCGGTCGGATCACCCATAAGAA				883
Query 552	AGGTGGTTGCTATTACGTAACCCCAGCCATCTCCACATGTCTTAGTTCCATTAG				611
Sbjct 882	AGGTGGTTGCTATTACGTAACCCCAGCCATCTCCACATGTCTTAGTTCCATTAG				823
Query 612	AGCCAAGCTTCCCAAACCTATGTAATCACTGAGCCTATTTTGCTGTCAACCAACTC				671
Sbjct 822	AGCCAAGCTTCCCAAACCTATGTAATCACTGAGCCTATTTTGCTGTCAACCAACTC				763
Query 672	AAGGCAACTCCTGTCCCTTCAAGTAAACAGTAGAAGGCGCTGAAGCTGCAATGTGAAGTGG				731
Sbjct 762	AAGGCAACTCCTGTCCCTTCAAGTAAACAGTAGAAGGCGCTGAAGCTGCAATGTGAAGTGG				703
Query 732	GCCTATAGGATACACTGGAACTTGCAGTTGTTTGTAAACCATGCCAAGATGAGCTCTC				791
Sbjct 702	GCCTATAGGATACACTGGAACTTGCAGTTGTTTGTAAACCATGCCAAGATGAGCTCTC				643
Query 792	TAGACAGCTTGTGAGTTGATGATAACTGCCGAAGCTGTCGAATGTTGACAGTCTCACT				851
Sbjct 642	TAGACAGCTTGTGAGTTGATGATAACTGCCGAAGCTGTCGAATGTTGACAGTCTCACT				583
Query 852	GTAAACCTTGAGTATACTCTCTAATGGCCCAAATGCTGAAGTTGGCAGGTCTTGACCT				911
Sbjct 582	GTAAACCTTGAGTATACTCTCTAATGGCCCAAATGCTGAAGTTGGCAGGTCTTGACCT				523

**Fig 11: Query and subject sequence; 76E1 S318N and 76E1 wild type respectively. Codon ATT (reverse AAT) in query is asparagine while ACT (reverse AGT) in subject is serine.**

## 76E5 S311N

Score	Expect	Identities	Gaps	Strand	Frame
2002 bits(1084)	0.0()	1092/1098(99%)	0/1098(0%)	Plus/Minus	
<b>Features:</b>					
Query 72	CTACATGAAATTCATCATTAAAGGAATTGACAAAGTTGTCTAATGAGCTGAATGAGGA				131
Sbjct 1362	CTACATGAAATTCATCATTAAAGGAATTGACAAAGTTGTCTAATGAGCTGAATGAGGA				1303
Query 132	ACCTCTACTCTTGACAGAGGGCTTGAAGCTCTCTTCAAGTTGATAACTCTCTCCAT				191
Sbjct 1302	ACCTCTACTCTTGACAGAGGGCTTGAAGCTCTCTTCAAGTTGATAACTCTCTCCAT				1243
Query 192	TTCTGCTCCTCTTCATCCATAATCAATCTCTACAGCTCTCCACTGTTCCATTATC				251
Sbjct 1242	TTCTGCTCCTCTTCATCCATAATCAATCTCTACAGCTCTCCACTGTTCCATTATC				1183
Query 252	CAGCTCTCCTCCAATTGAACCCCCAATTCTCCAAACTCTCTTAAGTACCTCGCATTGAC				311
Sbjct 1182	CAGCTCTCCTCCAATTGAACCCCCAATTCTCCAAACTCTCTTAAGTACCTCGCATTGAC				1123
Query 312	TTTCTGATCTCCCGTAAAAGGCCTACAGATCATCGAACCTCTCCCCGATGCTCTAG				371
Sbjct 1122	TTTCTGATCTCCCGTAAAAGGCCTACAGATCATCGAACCTCTCCCCGATGCTCTAG				1063
Query 372	GGTCGAGTTCCATCCGGAGTGAACCTCAAAACCCCTCCACTGCAGGATGTCTGAGAACCTC				431
Sbjct 1062	GGTCGAGTTCCATCCGGAGTGAACCTCAAAACCCCTCCACTGCAGGATGTCTGAGAACCTC				1003
Query 432	TATCTGTGGTGCCATTTCACAATGTAACCTCTTCTGAAACCAACATTAAATTCCCTC				491
Sbjct 1002	TATCTGTGGTGCCATTTCACAATGTAACCTCTTCTGAAACCAACATTAAATTCCCTC				943
Query 492	CGGTAAAGACTCTGTCCATTCCGAGCCGGGAATAGAACCCGGTCGGATCACCCATAAGAA				551
Sbjct 942	CGGTAAAGACTCTGTCCATTCCGAGCCGGGAATAGAACCCGGTCGGATCACCCATAAGAA				883
Query 552	AGGTTGGTTGCTATTACGTAACCCCAAGCCATCTCAACATGTCTTAGTTCCATTAG				611
Sbjct 882	AGGTTGGTTGCTATTACGTAACCCCAAGCCATCTCAACATGTCTTAGTTCCATTAG				823
Query 612	AGCCAAGCTCCCAAACCTATGTAACACTGAGCCTATTTTGCTTGTCAACCACTC				671
Sbjct 822	AGCCAAGCTCCCAAACCTATGTAACACTGAGCCTATTTTGCTTGTCAACCACTC				763
Query 672	AAGGCAACTCTGTCCCTCTCAAGTAAACTAGAAGGCGCTGAAGCTGCAATGTGAAGTGG				731
Sbjct 762	AAGGCAACTCTGTCCCTCTCAAGTAAACTAGAAGGCGCTGAAGCTGCAATGTGAAGTGG				703
Query 732	GCCTATAGGATAACTGGAACTTGCAGTTGTTTGTAAACCATGCCAAGATGAGCTCTC				791
Sbjct 702	GCCTATAGGATAACTGGAACTTGCAGTTGTTTGTAAACCATGCCAAGATGAGCTCTC				643

**Fig 12: Query and subject sequence; 76E5 S311N and 76E5 wild type respectively. Codon ATT (reverse AAT) in query is asparagine while ACT (reverse AGT) in subject is serine.**

## 76E2 N320S

Score	Expect	Identities	Gaps	Strand	Frame
2023 bits(1095)	0.0()	1097/1098(99%)	0/1098(0%)	Plus/Minus	
<b>Features:</b>					
Query 42	TCACATGGAATTAAACAAAGTCGTCTAGTGAGCTGCATGAGGAACCTCACAATTCAACAGA				101
Sbjct 1350	TCACATGGAATTAAACAAAGTCGTCTAGTGAGCTGCATGAGGAACCTCACAATTCAACAGA				1291
Query 102	GGTTTCAATCTTCTTTCAAGTCATGGCTCTTCCATTTCTGTCCTTCTTCATC				161
Sbjct 1290	GGTTTCAATCTTCTTTCAAGTCATGGCTCTTCCATTTCTGTCCTTCTTCATC				1231
Query 162	CACAAGCAACCACCTCTACAGCTCTCTCCACAGTTCTTATCCAGATCTCCCTCCAATTG				221
Sbjct 1230	CACAAGCAACCACCTCTACAGCTCTCTCCACAGTTCTTATCCAGATCTCCCTCCAATTG				1171
Query 222	AACCCCATTCTCCAAACTCTCTTAAGTACCTCGCATTGACTTTCTGATCCCCGGTGAA				281
Sbjct 1170	AACCCCATTCTCCAAACTCTCTTAAGTACCTCGCATTGACTTTCTGATCCCCGGTGAA				1111
Query 282	AGGCCTACATATCATCGGAACCTCTTCCCGATGCTCTACTGTTGAGTTCCATCCACA				341
Sbjct 1110	AGGCCTACATATCATCGGAACCTCTTCCCGATGCTCTACTGTTGAGTTCCATCCACA				1051
Query 342	GTGACTCCAAAACCCCTCTACTGCAGGATGTCGTGAGAACCTCCATCTGGAGGCCATT				401
Sbjct 1050	GTGACTCCAAAACCCCTCTACTGCAGGATGTCGTGAGAACCTCCATCTGGAGGCCATT				991
Query 402	CACAATGTAACCTTTCTGAAACCAACCTTAAATTCTCTGGTAAGGACTCTGTCCA				461
Sbjct 990	CACAATGTAACCTTTCTGAAACCAACCTTAAATTCTCTGGTAAGGACTCTGTCCA				931
Query 462	TTCTGACCCCGGAATAGAGCCGGTCTGACCACCCATAAGAAAGGTTGGTGTATTACT				521
Sbjct 930	TTCTGACCCCGGAATAGAGCCGGTCTGACCACCCATAAGAAAGGTTGGTGTATTACT				871
Query 522	TAATCCCCAAGCCATCTCCACATGTCTTGTTGTCATTAGGCCAAGCTTCCAAAGCT				581
Sbjct 870	TAATCCCCAAGCCATCTCCACATGTCTTGTTGTCATTAGGCCAAGCTTCCAAAGCT				811
Query 582	TATGTAAAATAACTGAATTGATTTTGCTGTGTTCAACCACTCAACGCAACTCTGTCTC				641
Sbjct 810	TATGTAAAATAACTGAATTGATTTTGCTGTGTTCAACCACTCAACGCAACTCTGTCTC				751
Query 642	TTCTAGTAAAATAGAAGGCCTGAAGCTGTAATATGAAGTGGGCTATAGGATAACACCGG				701
Sbjct 750	TTCTAGTAAAATAGAAGGCCTGAAGCTGTAATATGAAGTGGGCTATAGGATAACACCGG				691
Query 702	AACTTGCAAGTTGTTGCAACCTTGCAAAGATGAGCTCTAAACAGCTTGCTGAGTT				761
Sbjct 690	AACTTGCAAGTTGTTGCAACCTTGCAAAGATGAGCTCTAAACAGCTTGCTGAGTT				631
Query 762	GATGATAACACCGGAAGCTGTCGAGTGTACAGTCACGTCAACTGTAACACCTTGAGCGTACT				821
Sbjct 630	GATGATAACACCGGAAGCTGTCGAGTGTACAGTCACGTCAACTGTAACACCTTGAGCGTACT				571
Query 822	CTCTATTGGCCAAATACTGAAGTTGGTAGATCCTGTAACCTCAGAGGATGCAACCTGG				881
Sbjct 570	CTCTATTGGCCAAATACTGAAGTTGGTAGATCCTGTAACCTCAGAGGATGCAACCTGG				511
Query 882	AAATACTTTGTTGTTGAGGATCTTCATGTCATCAAGAACGACTCTGCGTGAC				941
Sbjct 510	AAATACTTTGTTGTTGAGGATCTTCATGTCATCAAGAACGACTCTGCGTGAC				451
Query 942	TCTAGACAAAACAGAGCGACAGACAAAAGCAGTAGCAGCTTGCGTGTCAAAGACGACACT				1001
Sbjct 450	TCTAGACAAAACAGAGCGACAGACAAAAGCAGTAGCAGCTTGCGTGTCAAAGACGACACT				391
Query 1002	AGGAAGTTGAAACTCTTTACTGCAGCATGAGAGAAGTACATGACTCATCGTAGACGAC				1061
Sbjct 390	AGGAAGTTGAAACTCTTTACTGCAGCATGAGAGAAGTACATGACTCATCGTAGACGAC				331

**Fig 13: Query and subject sequence; 76E2 N320S and 76E2 wild type respectively. Codon ACT (reverse AGT) in query is serine while ATT (reverse AAT) in subject is asparagine.**

## WILD TYPE SEQUENCING

### 76E5

	Score 1539 bits(833)	Expect 0.0	Identities 833/833(100%)	Gaps 0/833(0%)	Strand Plus/Minus
Query	31	TCAAGTATTCTATACTCTGCCTCCAAGTAATGGACGAGCTCATCCAATGCATTGTATGA		90	
Sbjct	1344	TCAAGTATTCTATACTCTGCCTCCAAGTAATGGACGAGCTCATCCAATGCATTGTATGA		1285	
Query	91	AGAGCCTCCACTCTTACAGAGGCATTGAGCTCTCTTTAAAACAAGGGCTCTCCCT		150	
Sbjct	1284	AGAGCCTCCACTCTTACAGAGGCATTGAGCTCTCTTTAAAACAAGGGCTCTCCCT		1225	
Query	151	CATTCTACACCTTCATCATCCACAATCAACCTTTCACAGCTCTCTACACATCCTCT		210	
Sbjct	1224	CATTCTACACCTTCATCATCCACAATCAACCTTTCACAGCTCTCTACACATCCTCT		1165	
Query	211	CTCCACTTCTCCTTGAAGCAGAACCCCTACTCTCAAACACTTTCTATATACATCGCGTT		270	
Sbjct	1164	CTCCACTTCTCCTTGAAGCAGAACCCCTACTCTCAAACACTTTCTATATACATCGCGTT		1105	
Query	271	TAACTTCTGCTCACCGTTAACCGGTCTGCAAATCATTGAAACTCCTCCACAATACTCTC		330	
Sbjct	1104	TAACTTCTGCTCACCGTTAACCGGTCTGCAAATCATTGAAACTCCTCCACAATACTCTC		1045	
Query	331	GAGTGTGAGTCCATCCACAGTGGCTCCAGAACCTCCACTGCAGGATGCACAAGTAC		390	
Sbjct	1044	GAGTGTGAGTCCATCCACAGTGGCTCCAGAACCTCCACTGCAGGATGCACAAGTAC		985	
Query	391	TTCATTCTGTGGCGCCCATTCAATGCATCCTCTTCCGAGACAATCTTACTGACTTC		450	
Sbjct	984	TTCATTCTGTGGCGCCCATTCAATGCATCCTCTTCCGAGACAATCTTACTGACTTC		925	
Query	451	CACTGGCATTGACTCTGTACCGGGTCGGATTACCCATAGAAAAGGTGGTTGCTATTATA		510	
Sbjct	924	CACTGGCATTGACTCTGTACCGGGTCGGATTACCCATAGAAAAGGTGGTTGCTATTATA		865	
Query	511	CAATCCCCAAGCCATCTCAAATCTCCTGGTTCCATGTGAGCTATGCTTCTACGCT		570	
Sbjct	864	CAATCCCCAAGCCATCTCAAATCTCCTGGTTCCATGTGAGCTATGCTTCTACGCT		805	
Query	571	TATGTATATAACTGACCTCAGTTCTGCTTGTCAGCCATTCAATGCAGCTCCTGCTTC		630	
Sbjct	804	TATGTATATAACTGACCTCAGTTCTGCTTGTCAGCCATTCAATGCAGCTCCTGCTTC		745	
Query	631	TTCAAGTAAACTAAAATTGCTGAAGTTGAATGTGAAGAGGGCCTAATGGATACACTGG		690	
Sbjct	744	TTCAAGTAAACTAAAATTGCTGAAGTTGAATGTGAAGAGGGCCTAATGGATACACTGG		685	
Query	691	AATACTGAGTTCTGTTCAGCCATGACAGAGACGAGCTCTAGACAACCTGACGTATT		750	
Sbjct	684	AATACTGAGTTCTGTTCAGCCATGACAGAGACGAGCTCTAGACAACCTGACGTATT		625	
Query	751	GATTATAACAGCGGAAGCTGTTCTTGACAACTTCGGCACAAATCTCCAAAAATCG		810	
Sbjct	624	GATTATAACAGCGGAAGCTGTTCTTGACAACTTCGGCACAAATCTCCAAAAATCG		565	
Query	811	CTCTAGCGGCCCATTCCTGAAGTTGGTAAGTCTTTGTATTTAGTGGATGTA		863	
Sbjct	564	CTCTAGCGGCCCATTCCTGAAGTTGGTAAGTCTTTGTATTTAGTGGATGTA		512	

**Fig 14: Query and subject sequence; 76E5 recombinant plasmid and 76E5 sequence on database respectively.**

## 76E2

	Score 1934 bits(1047)	Expect 0.0	Identities 1047/1047(100%)	Gaps 0/1047(0%)	Strand Plus/Minus
Query	50	TCACATGGAATTAAACAAAGTCGTAGTGAGCTGCATGAGGAACCTCCACTTCAACAGA		109	
Sbjct	1350	TCACATGGAATTAAACAAAGTCGTAGTGAGCTGCATGAGGAACCTCCACTTCAACAGA		1291	
Query	110	GGTTTCAATCTTCTTCAAGTCATGGCTCTTCCTCATTTCTGCTCCTCTTCATC		169	
Sbjct	1290	GGTTTCAATCTTCTTCAAGTCATGGCTCTTCCTCATTTCTGCTCCTCTTCATC		1231	
Query	170	CACAAGCAACCCTACAGCTCTCCACAGTTCTTATCCAGATCTCCCTCCAATTG		229	
Sbjct	1230	CACAAGCAACCCTACAGCTCTCCACAGTTCTTATCCAGATCTCCCTCCAATTG		1171	
Query	230	AACCCCATTCTCCAAACTCTCTTAAGTACCTCGCATTGACTTCTGATCCCCGGTGAA		289	
Sbjct	1170	AACCCCATTCTCCAAACTCTCTTAAGTACCTCGCATTGACTTCTGATCCCCGGTGAA		1111	
Query	290	AGGCCTACATATCATCGGAACTCCTCCCCGATGCTCTACTGTTGAGTTCCATCCACA		349	
Sbjct	1110	AGGCCTACATATCATCGGAACTCCTCCCCGATGCTCTACTGTTGAGTTCCATCCACA		1051	
Query	350	GTGACTCCAAAACCCCTCCTACTGCAGGATGTCAGAACCTCCATCTGCGGAGCCCATT		409	
Sbjct	1050	GTGACTCCAAAACCCCTCCTACTGCAGGATGTCAGAACCTCCATCTGCGGAGCCCATT		991	
Query	410	CACAATGTAACCTCTTCTGAAACCAACCTATTGAATTCTCTGTAAGGACTCTGTCCA		469	
Sbjct	990	CACAATGTAACCTCTTCTGAAACCAACCTATTGAATTCTCTGTAAGGACTCTGTCCA		931	
Query	470	TTCTGACCCCGGAATAGAGCCCGGTCTGACCACCCATAAGAAAGGTTGGTTGCTATTACT		529	
Sbjct	930	TTCTGACCCCGGAATAGAGCCCGGTCTGACCACCCATAAGAAAGGTTGGTTGCTATTACT		871	
Query	530	TAATCCCCAAGCCATCTCAAACATGTCTTGGTGTCCATTAGAGCCAAGCTTCCAAAGCT		589	
Sbjct	870	TAATCCCCAAGCCATCTCAAACATGTCTTGGTGTCCATTAGAGCCAAGCTTCCAAAGCT		811	
Query	590	TATGTAATAACTGAATTGATTTGTTGTTCAACCCTCAACGCAACTCCTGTCTTC		649	
Sbjct	810	TATGTAATAACTGAATTGATTTGTTGTTCAACCCTCAACGCAACTCCTGTCTTC		751	
Query	650	TTCTAGTAAACTAGAAGGCCGTGAAGCTGTAATATGAAGTGGGCTATAGGATACACCGG		709	
Sbjct	750	TTCTAGTAAACTAGAAGGCCGTGAAGCTGTAATATGAAGTGGGCTATAGGATACACCGG		691	
Query	710	AACTTGCAGTTGTTGCAACCTTGCCAAAGATGAGCTCTCTAAACAGCTTGCTGAGTT		769	
Sbjct	690	AACTTGCAGTTGTTGCAACCTTGCCAAAGATGAGCTCTCTAAACAGCTTGCTGAGTT		631	
Query	770	GATGATAACAGCGGAAGCTGTTGAGTGTTCACAGTCTCACTGTAACCTTGAGCGTACT		829	
Sbjct	630	GATGATAACAGCGGAAGCTGTTGAGTGTTCACAGTCTCACTGTAACCTTGAGCGTACT		571	
Query	830	CTCTATTGGCCCAAATACTGAAGTTGGTAGATCCTGTACCTCAGAGGATGCAACCCCTGG		889	
Sbjct	570	CTCTATTGGCCCAAATACTGAAGTTGGTAGATCCTGTACCTCAGAGGATGCAACCCCTGG		511	

**Fig 15: Query and subject sequence; 76E2 recombinant plasmid and 76E2 sequence on database respectively.**

## 76D1

Score 1157 bits(626)	Expect 0.0	Identities 626/626(100%)	Gaps 0/626(0%)	Strand Plus/Minus
Query 13	TCATTGTCGTCAATTGCATCATTATTGCATGGACCAAATTGTTAAAGAATTGTGAGA	72		
Sbjct 1359	TCATTGTCGTCAATTGCATCATTATTGCATGGACCAAATTGTTAAAGAATTGTGAGA	1300		
Query 73	AGAGCCTTCGGTTGTGACAGAGGCTTCAACCTCTCCTCAATATGGTGGCTCTCATTC	132		
Sbjct 1299	AGAGCCTTCGGTTGTGACAGAGGCTTCAACCTCTCCTCAATATGGTGGCTCTCATTC	1240		
Query 133	CATCTCTGACCTTCTTGATCCACAATGAGCCTCTCACGGCCATCTCAACAGCTCTCT	192		
Sbjct 1239	CATCTCTGACCTTCTTGATCCACAATGAGCCTCTCACGGCCATCTCAACAGCTCTCT	1180		
Query 193	TTCCAATTCACCTCGATCTCATACGCGGTTGCCAACATGTGACATAAGTCGAGTATT	252		
Sbjct 1179	TTCCAATTCACCTCGATCTCATACGCGGTTGCCAACATGTGACATAAGTCGAGTATT	1120		
Query 253	CACCCCTCTGATCACCAGAATACGGCTACAAATATTGGTACACCAACTGCTTATGCTCTC	312		
Sbjct 1119	CACCCCTCTGATCACCAGAATACGGCTACAAATATTGGTACACCAACTGCTTATGCTCTC	1060		
Query 313	CAAGCAGAGTTCCATCCACCATGGTCCAAAACCTCCACTGCTCTATGCCTTAATAC	372		
Sbjct 1059	CAAGCAGAGTTCCATCCACCATGGTCCAAAACCTCCACTGCTCTATGCCTTAATAC	1000		
Query 373	CTCTTTTGTTGGGCCATTTCACAACAAAACCTCTCCATCGGTACCGTTGGTGAA	432		
Sbjct 999	CTCTTTTGTTGGGCCATTTCACAACAAAACCTCTCCATCGGTACCGTTGGTGAA	940		
Query 433	CTGTTCCGTAAGAAGTCTAAAGATTCTGTCGTTATAGAGCCTGGATCACCCA	492		
Sbjct 939	CTGTTCCGTAAGAAGTCTAAAGATTCTGTCGTTATAGAGCCTGGATCACCCA	880		
Query 493	CAAGAAGGGTTGATTACTCTGGACAAATCCCATGCCATCTCACAGCCTCTATCTTG	552		
Sbjct 879	CAAGAAGGGTTGATTACTCTGGACAAATCCCATGCCATCTCACAGCCTCTATCTTG	820		
Query 553	TGTCATGCCAAGCTCCCCATGCTTATGTAGATCACTGAGCTGTTCTGCTTCAAG	612		
Sbjct 819	TGTCATGCCAAGCTCCCCATGCTTATGTAGATCACTGAGCTGTTCTGCTTCAAG	760		
Query 613	CCATTCAAGACAGTTCTTCTTCTT 638			
Sbjct 759	CCATTCAAGACAGTTCTTCTTCTT 734			

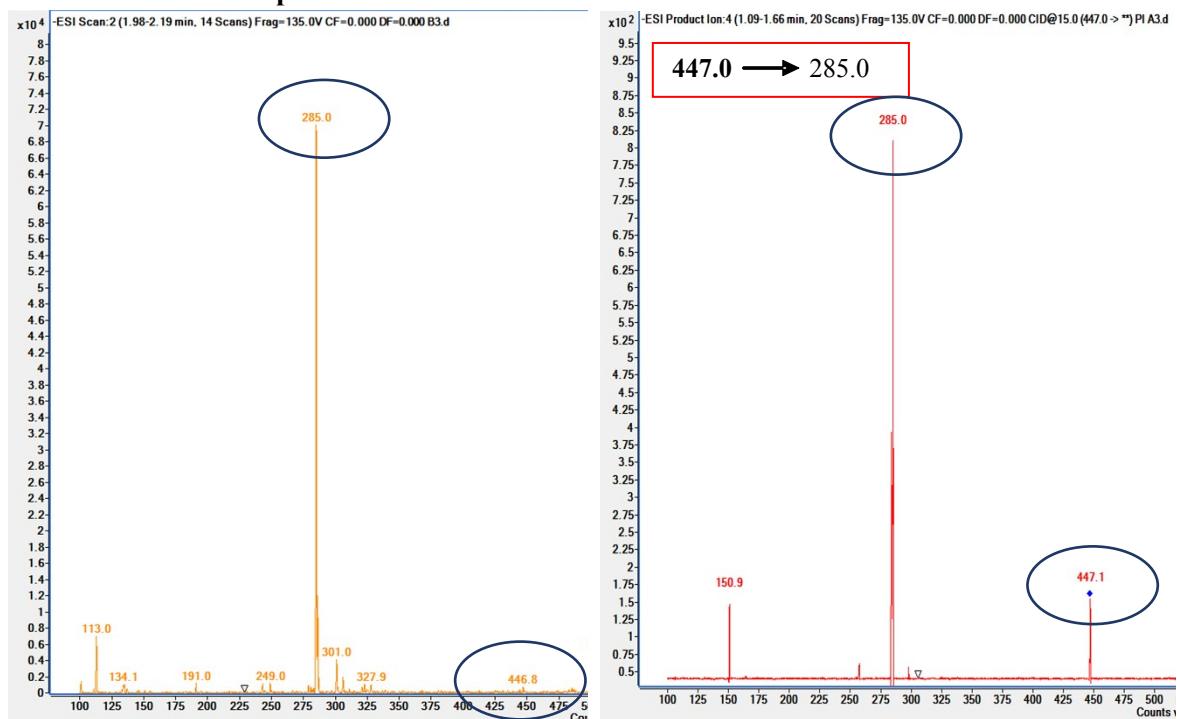
**Fig 16: Query and subject sequence; 76D1 recombinant plasmid and 76D1 sequence on database respectively.**

## 76E1

Query	71	CTACATGAAATTCACTCATTTAAGGAATTGACAAAGTGTCTAATGAGCTGAATGAGGA	130
Sbjct	1362	CTACATGAAATTCACTCATTTAAGGAATTGACAAAGTGTCTAATGAGCTGAATGAGGA	1303
Query	131	ACCTCTACTCTTGACAGAGGCTGAAGCTTCCTCAAGTGTAACTCTCCAT	190
Sbjct	1302	ACCTCTACTCTTGACAGAGGCTGAAGCTTCCTCAAGTGTAACTCTCCAT	1243
Query	191	TCTGCTCTCTCATCCATAATCAATCTTACAGCTCTCCACTGTTCTTATC	250
Sbjct	1242	TCTGCTCTCTCATCCATAATCAATCTTACAGCTCTCCACTGTTCTTATC	1183
Query	251	CAGCTCTCTTCCAATTGAACCCCAATTCTCAAACCTCTCTAAGTAACCTCGCATTGAC	310
Sbjct	1182	CAGCTCTCTTCCAATTGAACCCCAATTCTCAAACCTCTCTAAGTAACCTCGCATTGAC	1123
Query	311	TTTCTGATCTCCCGTAAAAGGCCAACAGATCATCGGAACCTCTCCCCGATGCTCTTAG	370
Sbjct	1122	TTTCTGATCTCCCGTAAAAGGCCAACAGATCATCGGAACCTCTCCCCGATGCTCTTAG	1063
Query	371	GGTCGAGTTCATCCGAGTGACTCCAAAACCTCCCACTGCAGGATGCTGAGAACTTC	430
Sbjct	1062	GGTCGAGTTCATCCGAGTGACTCCAAAACCTCCCACTGCAGGATGCTGAGAACTTC	1003
Query	431	TATCTGTGGTGCCTTACAAATGTAACCTCTGAAACCAACCTACTGAATTCCC	490
Sbjct	1002	TATCTGTGGTGCCTTACAAATGTAACCTCTGAAACCAACCTACTGAATTCCC	943
Query	491	CGGTAAAGACTCTGTCCTTCCGAGCCGGGAATAGAACCCGGTCGGATCACCCATAAGAA	550
Sbjct	942	CGGTAAAGACTCTGTCCTTCCGAGCCGGGAATAGAACCCGGTCGGATCACCCATAAGAA	883
Query	551	AGGTTGGTTGCTTACGTAACCCCCAACCATCTCAACATGTCCTTAGTTCATTAG	610
Sbjct	882	AGGTTGGTTGCTTACGTAACCCCCAACCATCTCAACATGTCCTTAGTTCATTAG	823
Query	611	AGCCAAGCTCCAAACTATGTAANTCACTGAGCCTATTTGCTTGTCAACCACTC	670
Sbjct	822	AGCCAAGCTCCAAACTATGTAANTCACTGAGCCTATTTGCTTGTCAACCACTC	763
Query	671	AAGGCAACTCTGTCCTTCAAGTAACAGAACGCTGAAGCTGCAATGTGAAGTGG	730
Sbjct	762	AAGGCAACTCTGTCCTTCAAGTAACAGAACGCTGAAGCTGCAATGTGAAGTGG	703
Query	731	GCTTATAGGATACACTGGAACCTGCAGTTGTTGTAAACCATGCCAAAGATGANCTTC	790
Sbjct	702	GCTTATAGGATACACTGGAACCTGCAGTTGTTGTAAACCATGCCAAAGATGAGCTTC	643
Query	791	TAGACAGCTGNTGAGTTGATGATAACTGCCGAANCTGNTCNAATGNTGACAGTCTCACT	850
Sbjct	642	TAGACAGCTGNTGAGTTGATGATAACTGCCGAAGCTGTCGAATGTTGACAGTCTCACT	583

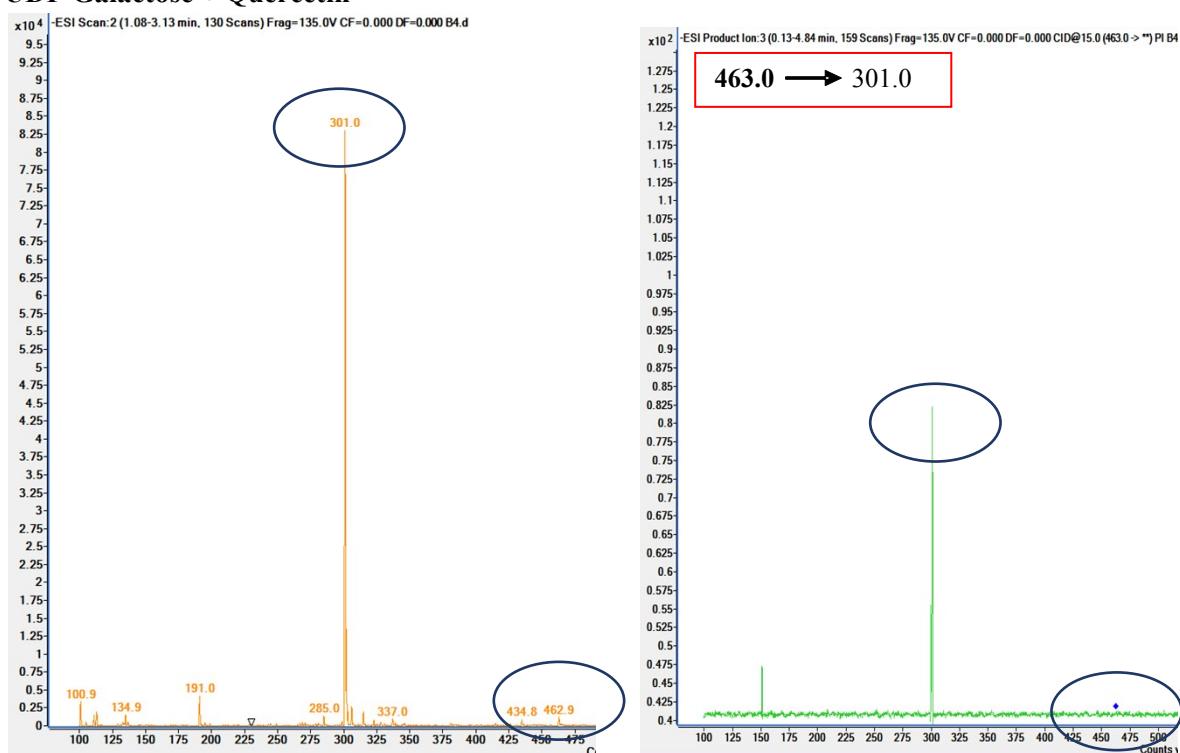
**Fig 17: Query and subject sequence; 76E1 recombinant plasmid and 76E1 sequence on database respectively.**

**MUTAGENESIS EXPERIMENTS - MASS SPECTRA**  
**76E2 N320S DONOR SCREEN (New activity)**  
**UDP Galactose + Kaempferol**



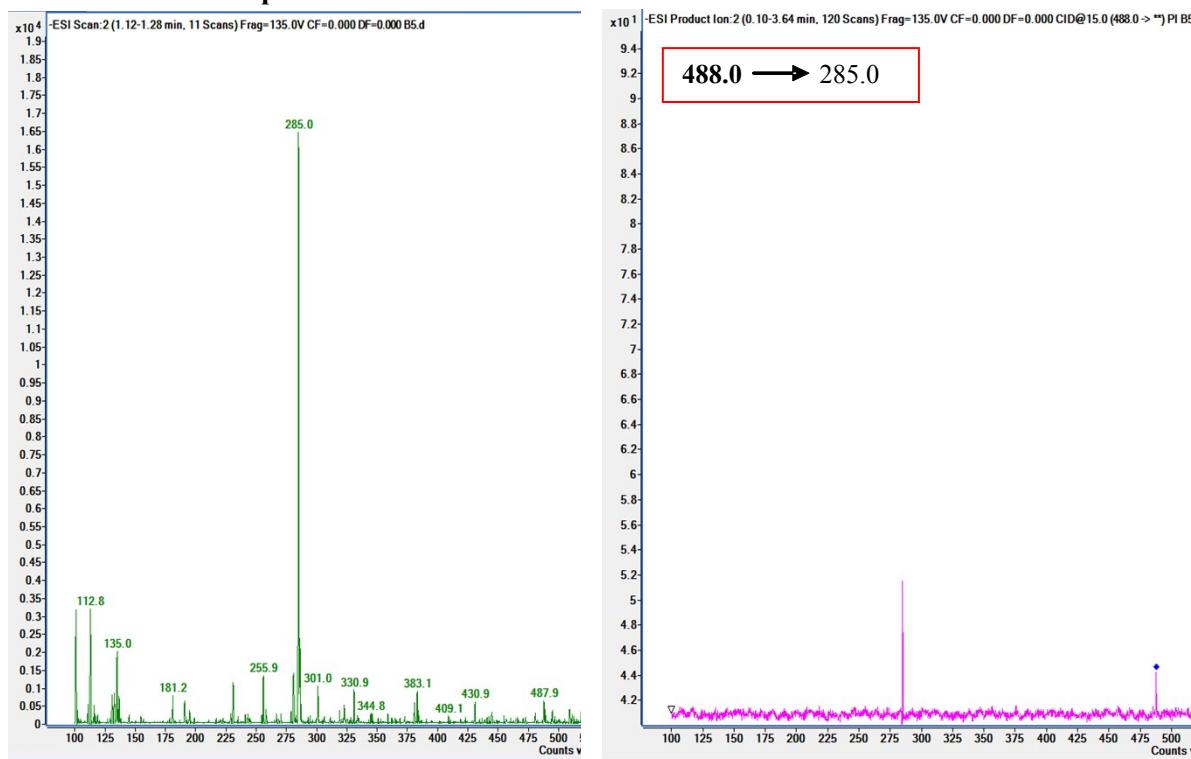
**Fig 18a:** MS spectrum of 76E2 N320S glycosylation of kaempferol (Full scan) **18b:** MS spectrum showing fragmentation of kaempferol galactoside (Product ion scan)

**UDP Galactose + Quercetin**



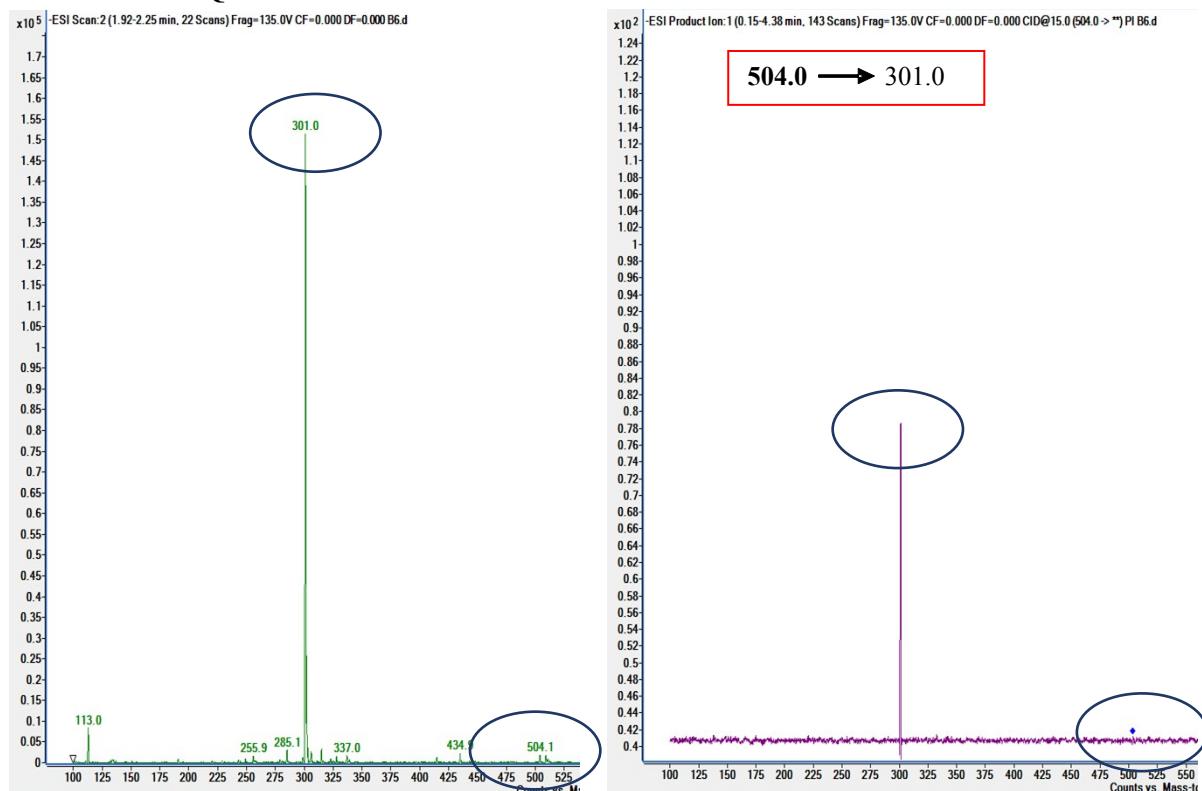
**Fig 19a:** MS spectrum of 76E2 N320S glycosylation of quercetin (Full scan) **19b:** MS spectrum showing fragmentation of quercetin galactoside (Product ion scan)

### UDP GlcNAc + Kaempferol



**Fig 20a:** MS spectrum of 76E2 N320S glycosylation of kaempferol (Full scan) **20b:** MS spectrum showing fragmentation of kaempferol glycoside (Product ion scan)

### UDP GlcNAc + Quercetin



**Fig 21a:** MS spectrum of 76E2 N320S glycosylation of quercetin (Full scan) **21b:** MS spectrum showing fragmentation of quercetin glycoside (Product ion scan)

### UDP Glucose + Kaempferol

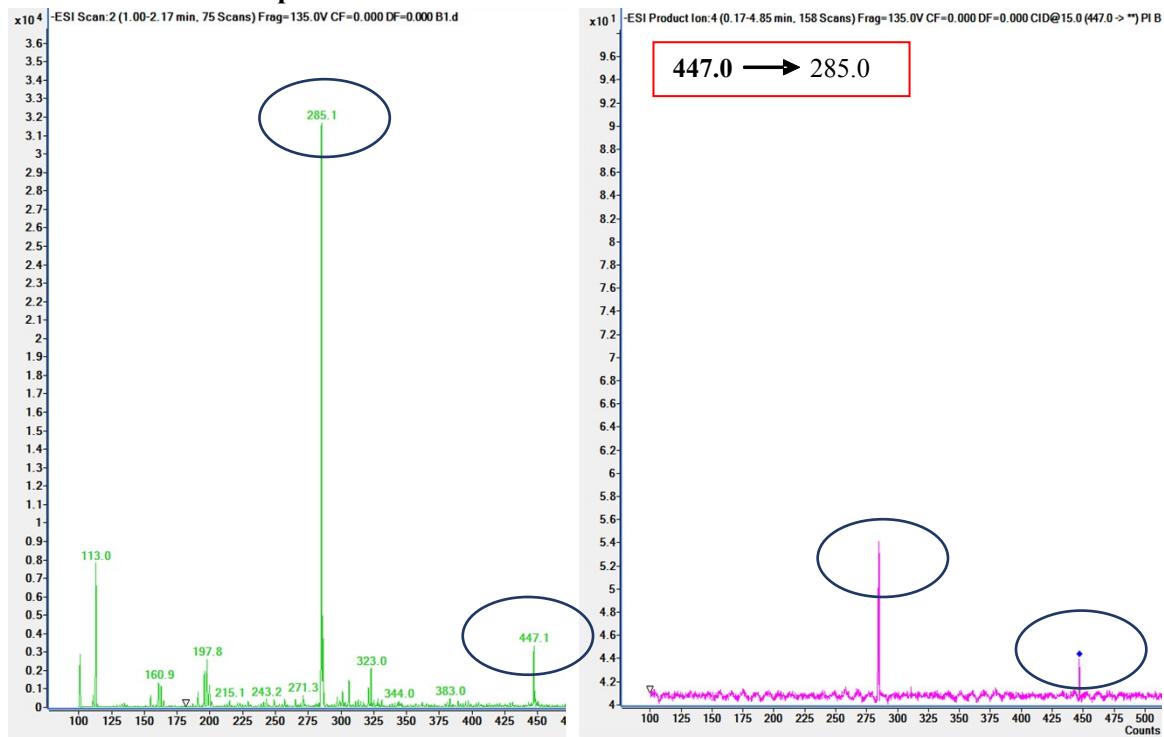


Fig 22a: MS spectrum of 76E2 N320S glycosylation of kaempferol (Full scan) 22b: MS spectrum showing fragmentation of kaempferol glucoside (Product ion scan)

### UDP Glucose + Quercetin

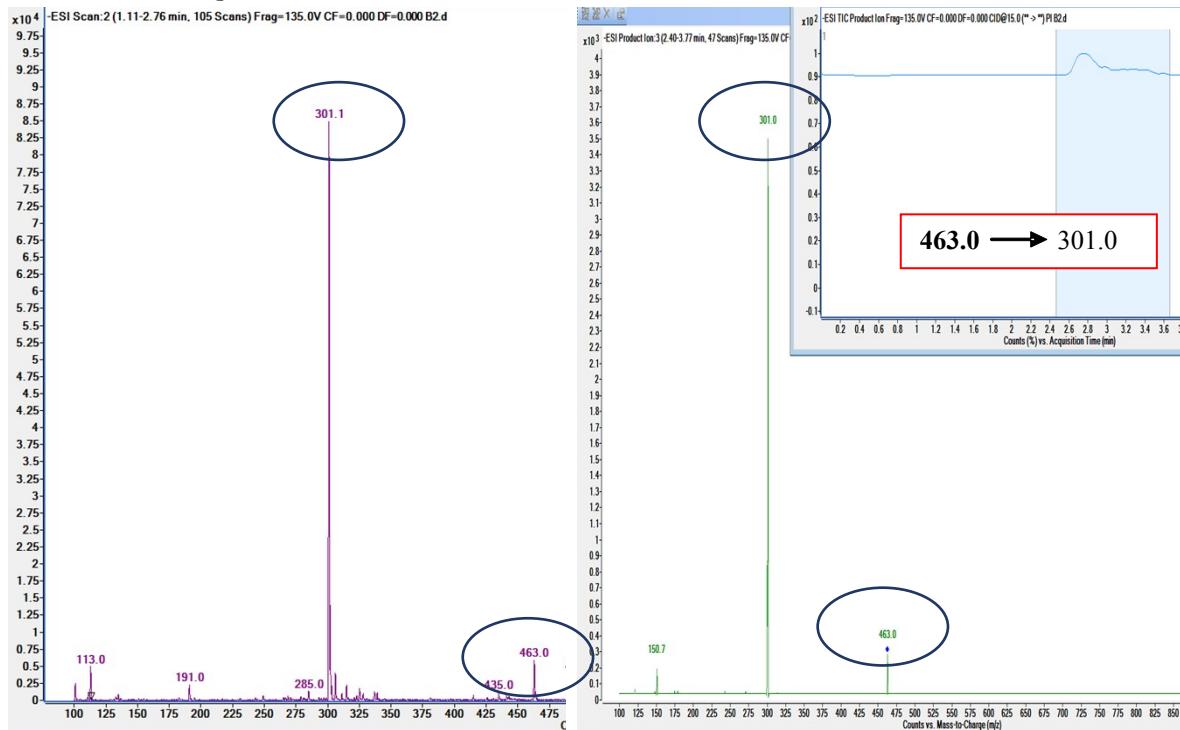


Fig 23a: MS spectrum of 76E2 N320S glycosylation of quercetin (Full scan) 23b: MS spectrum showing fragmentation of quercetin glucoside (Product ion scan)

## 76E1 S318N and 76E5 S311N DONOR SCREEN (Loss of activity)

### UDP Glucose + Kaempferol

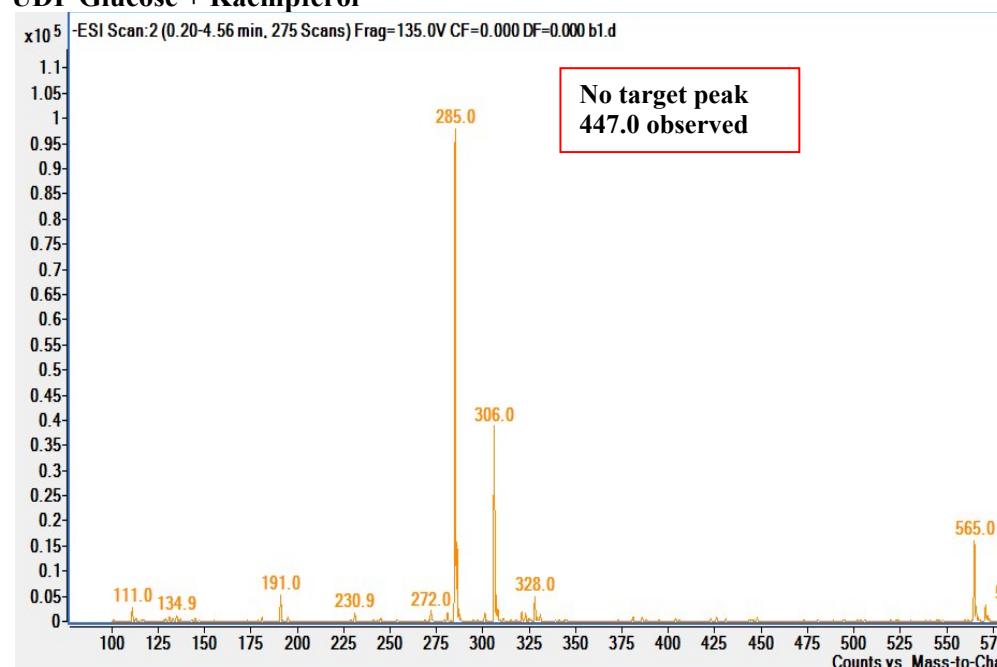


Fig 24a: MS spectrum of asparagine mutants showing no glycosylation with kaempferol (Product ion scan)

### UDP Glucose + Quercetin

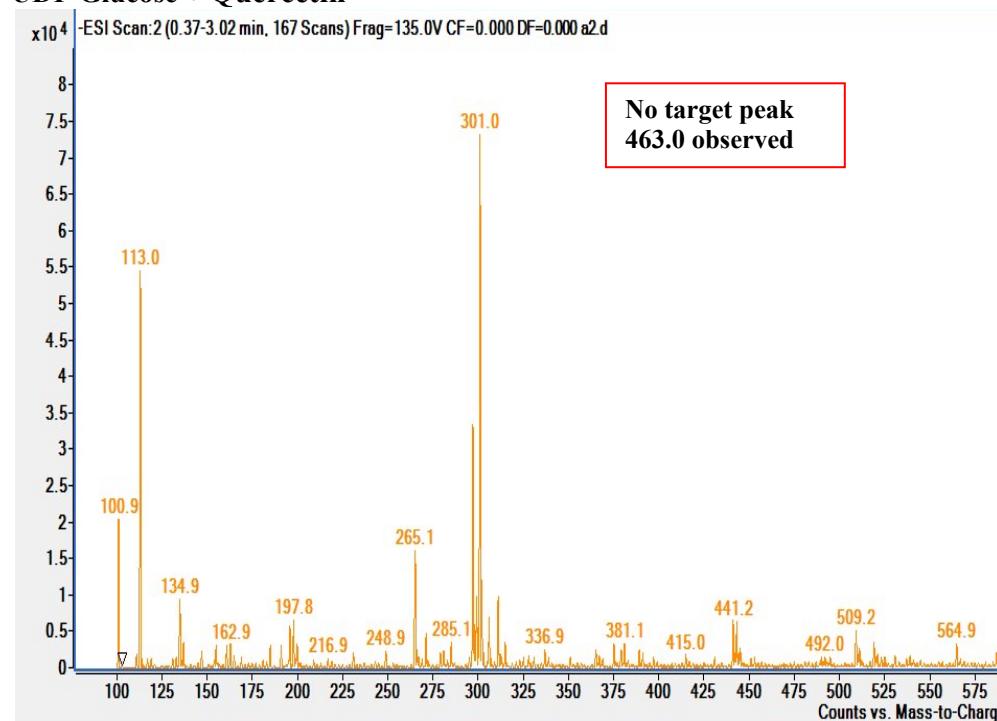


Fig 24b: MS spectrum of asparagine mutants showing no glycosylation with quercetin (Product ion scan)

### UDP Galactose + Quercetin

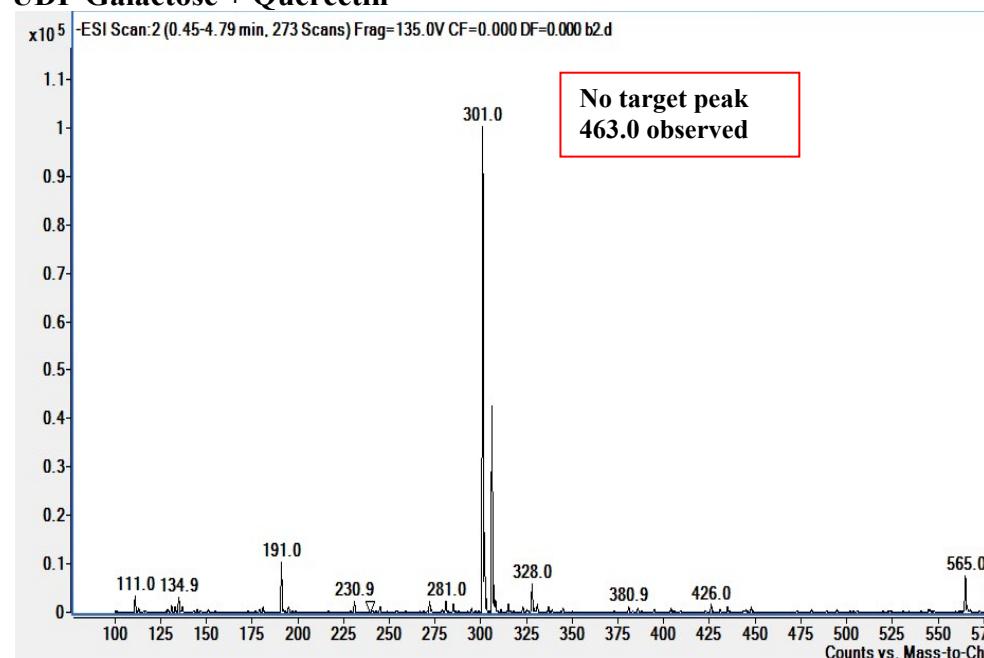


Fig 24c: MS spectrum of asparagine mutants showing no glycosylation with quercetin (Product ion scan)

### UDP Galactose + Kaempferol

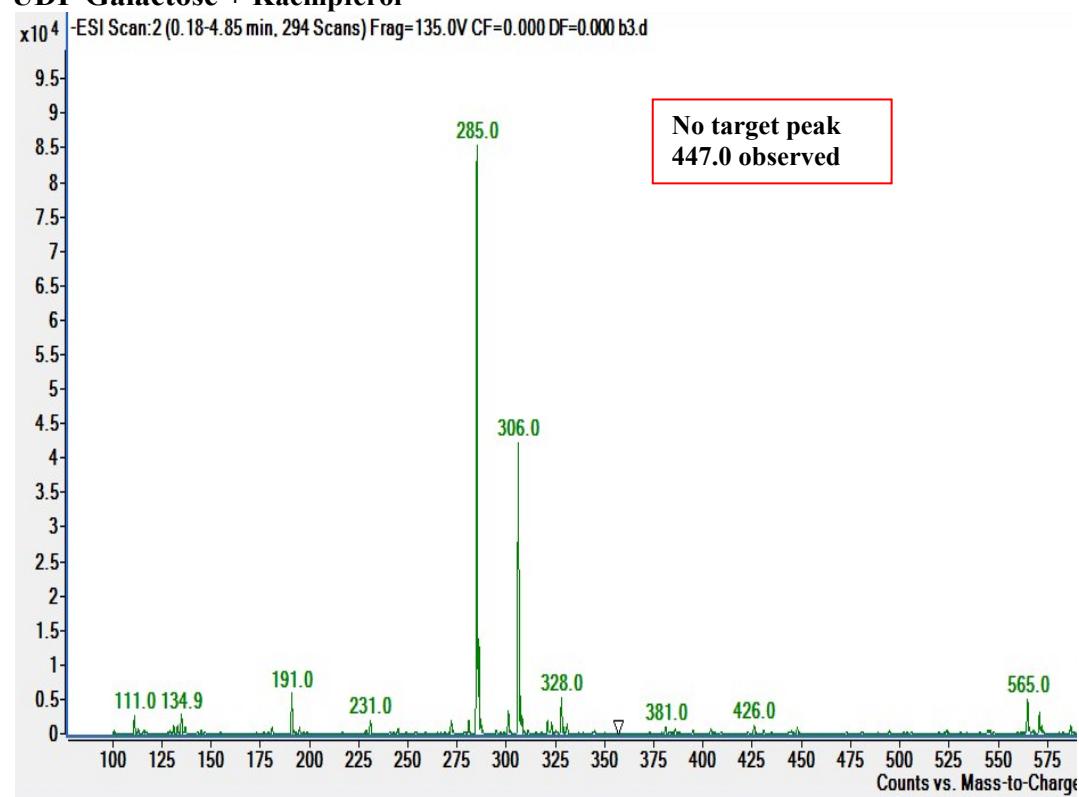


Fig 24d: MS spectrum of asparagine mutants showing no glycosylation with kaempferol (Product ion scan)

### UDP GlcNAc + Kaempferol

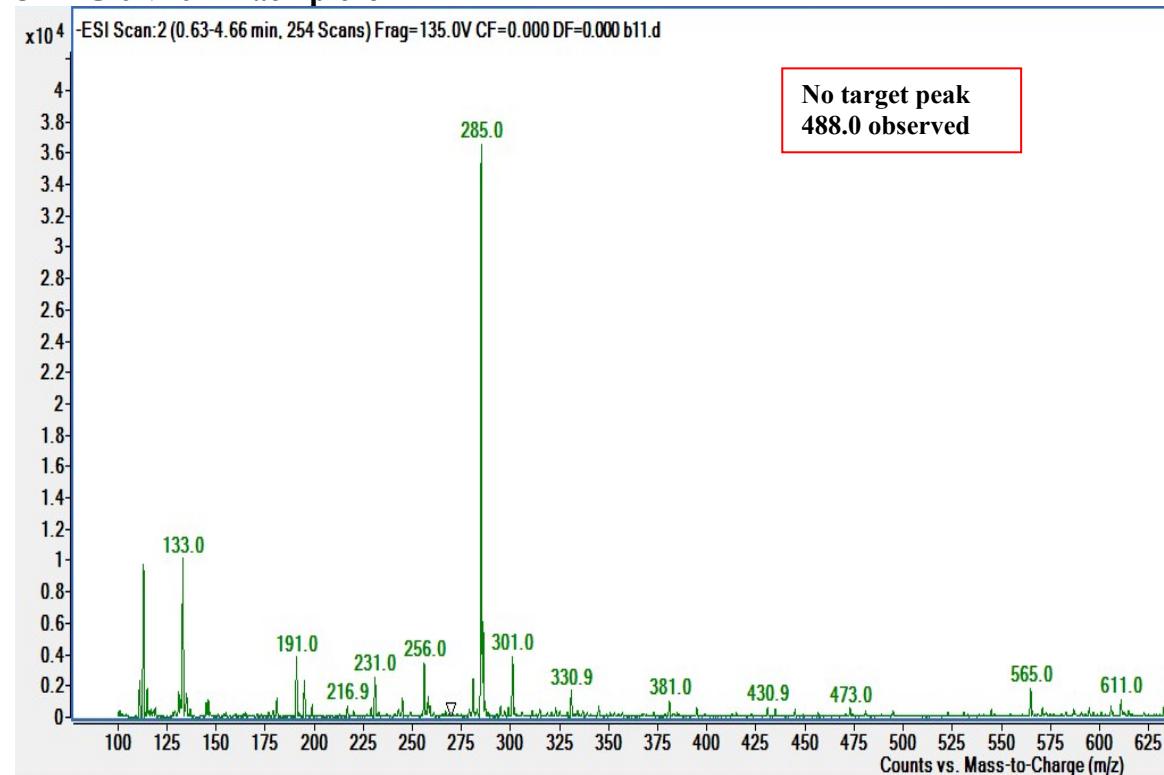


Fig 24e: MS spectrum of asparagine mutants showing no glycosylation with kaempferol (Product ion scan)

### UDP GlcNAc + Quercetin

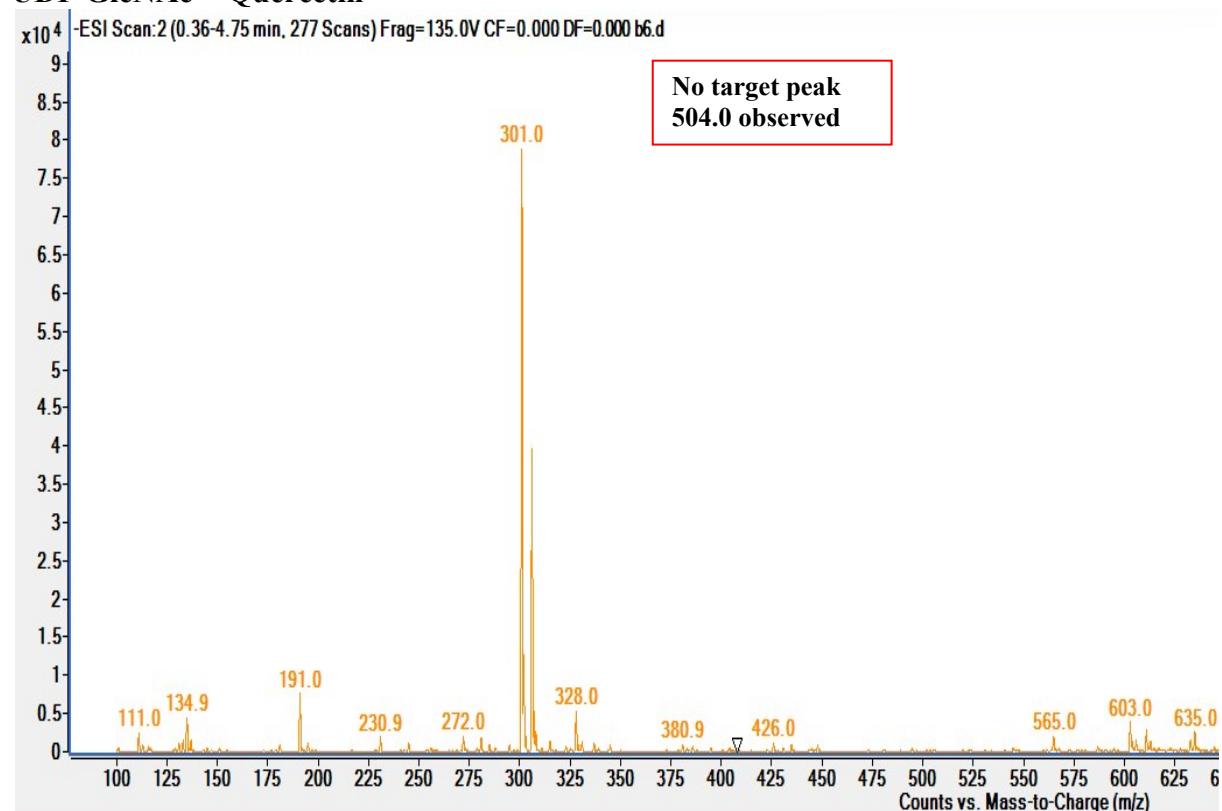
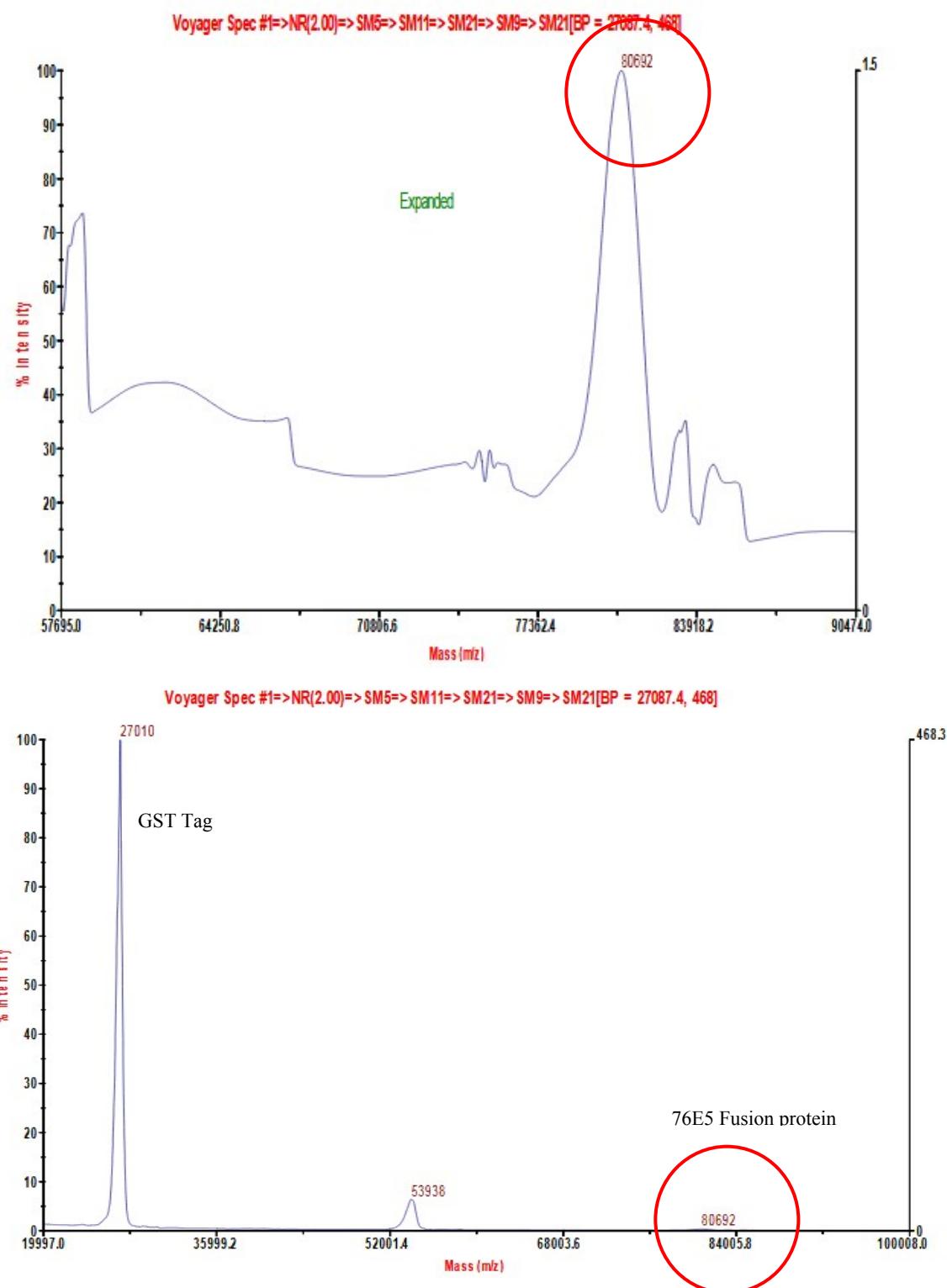


Fig 24f: MS spectrum of asparagine mutants showing no glycosylation with quercetin (Product ion scan)

## MALDI-TOF CONFIRMATION OF PROTEIN EXPRESSION

### 1. 76E5



1	27010	24999	28185	0	468	100	559778
2	53938	51124	55511	0	30	6	44166
3	80692	79919	81429	0	2	0	595

Fig 25: MALDI spectrum to confirm protein expression

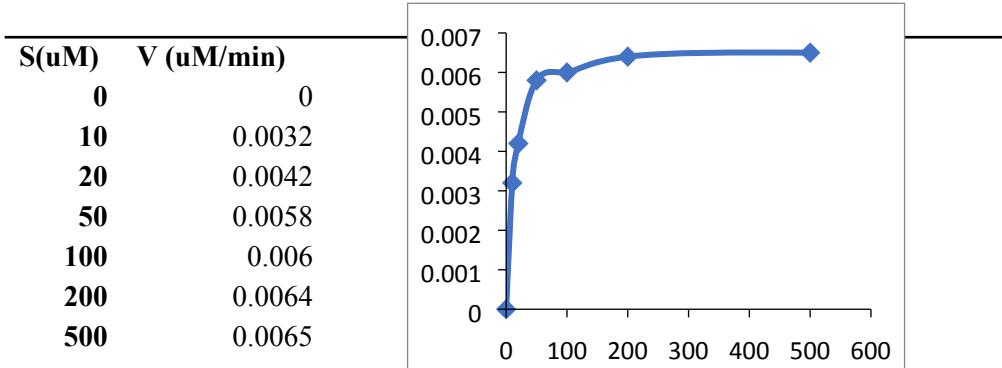
The empirical determination of 76E5 molecular weight (using our sequenced DNA data, was converted into protein sequence and lastly used to calculate the weight) was determined. This was confirmed by the MALDI-TOF data, 80.69kda.

<https://web.expasy.org/translate/> (DNA sequence conversion to protein sequence)  
[https://www.bioinformatics.org/sms/prot\\_mw.html](https://www.bioinformatics.org/sms/prot_mw.html) (protein sequence changed to molecular weight)

## KINETICS DATA

### 1) 76E1 UDP GLUCOSE

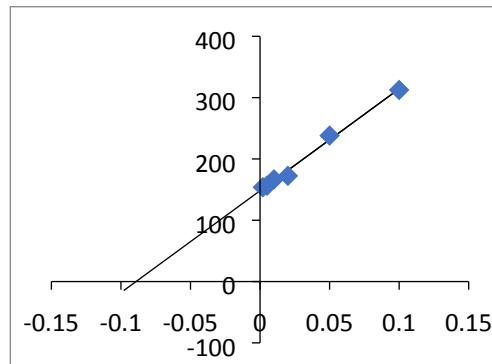
#### Michaelis menten plot



#### Lineweaver Burk

##### $1/S$

$1/S$	$1/V$
0.1	312.5
0.05	238.0952
0.02	172.4138
0.01	166.6667
0.005	156.25
0.002	153.8462



$$K_m = 10.5 \pm 1.38 \mu M$$

$$k_{cat}/K_m = 0.79 s^{-1} mM^{-1}$$

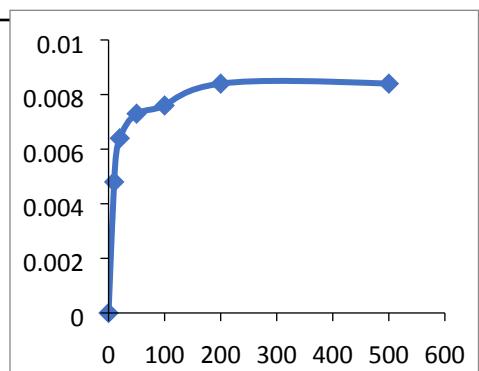
$$k_{cat} = 0.00832 s^{-1}$$

Fig 26: Michaelis menten and Lineweaver-Burk plots for 76E1

## 2) 76E2 UDP GLUCOSE

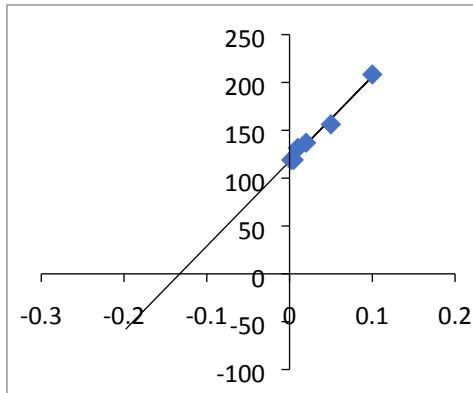
**Michaelis menten plot**

S (uM)	V (uM/min)
0	0
10	0.0048
20	0.0064
50	0.0073
100	0.0076
200	0.0084
500	0.0084



**Lineweaver-Burk**

1/S	1/V
0.1	208.3333
0.05	156.25
0.02	136.9863
0.01	131.5789
0.005	119.0476
0.002	119.0476



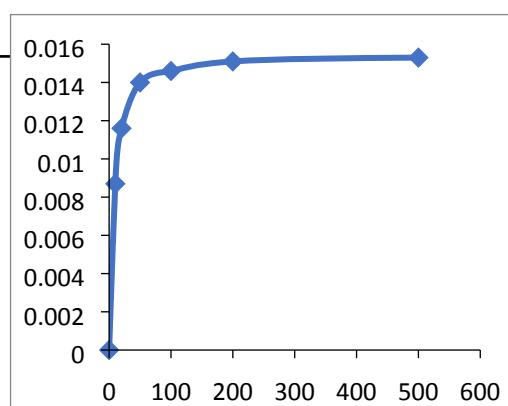
$$\begin{aligned} K_m &= 8.75 \pm 1.38 \mu M \\ k_{cat}/K_m &= 1.23 \text{ s}^{-1} \text{ mM}^{-1} \\ k_{cat} &= 0.0107 \text{ s}^{-1} \end{aligned}$$

Fig 27: Michaelis menten and Lineweaver-Burk plots for 76E2

### 3) a) 76E5 UDP Glucose

**Michaelis menten plot**

S (uM)	V (uM/min)
0	0
10	0.0087
20	0.0116
50	0.014
100	0.0146
200	0.0151
500	0.0153

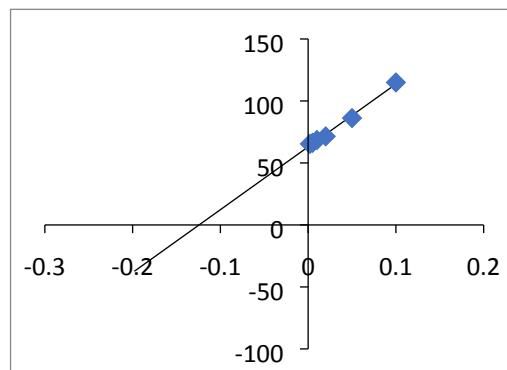


**Lineweaver Burk**

$1/S$

$1/V$

<b>0.1</b>	114.9425
<b>0.05</b>	86.2069
<b>0.02</b>	71.42857
<b>0.01</b>	68.49315
<b>0.005</b>	66.22517
<b>0.002</b>	65.35948



**K<sub>m</sub>**                     $8.79 \pm 1.64 \text{ } \mu\text{M}$

**k<sub>cat</sub>/K<sub>M</sub>**             $2.21 \text{ } \text{s}^{-1}\text{mM}^{-1}$

**k<sub>cat</sub>**                     $0.0194 \text{ } \text{s}^{-1}$

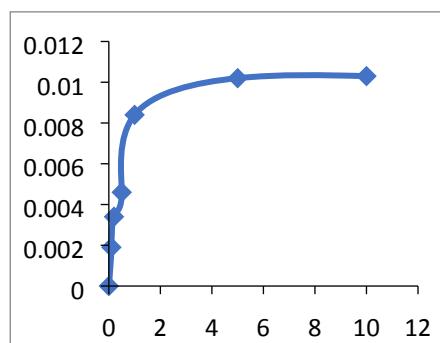
Fig 28a: Michaelis menten and Lineweaver-Burk plots for 76E5

**b) 76E5 UDP Galactose**

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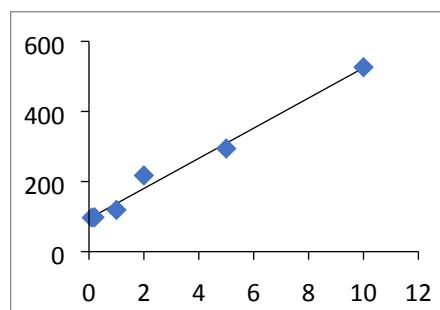
**Michaelis menten plot**

S (uM)	V (uM/min)
0	0
0.1	0.0019
0.2	0.0034
0.5	0.0046
1	0.0084
5	0.0102
10	0.0103



**Lineweaver Burk**

1/S	1/V
10	526.3158
5	294.1176
2	217.3913
1	119.0476
0.2	98.03922
0.1	97.08738



<b>K<sub>m</sub></b>	$0.57 \pm 0.19 \mu\text{M}$
<b>k<sub>cat</sub>/K<sub>M</sub></b>	$22.8 \text{ s}^{-1}\text{mM}^{-1}$
<b>k<sub>cat</sub></b>	$0.0130 \text{ s}^{-1}$

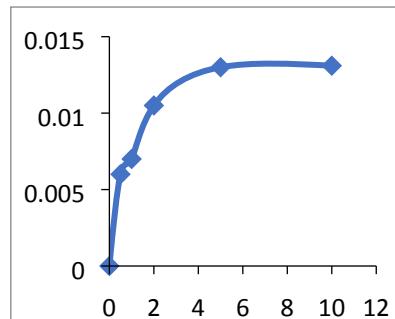
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Fig 28b: Michaelis menten and Lineweaver-Burk plots for 76E5

**c) 76E5 UDP N-acetyl glucosamine**

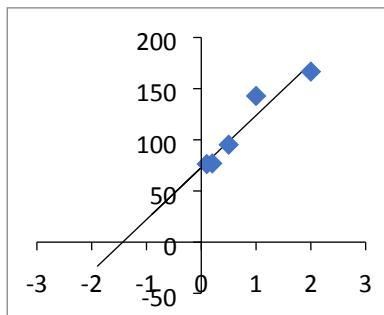
**Michaelis menten plot**

S (uM)	V (uM/min)
0	0
0.5	0.006
1	0.007
2	0.0105
5	0.013
10	0.0131



**Lineweaver Burk**

1/S	1/V
2	166.6667
1	142.8571
0.5	95.2381
0.2	76.92308
0.1	76.33588



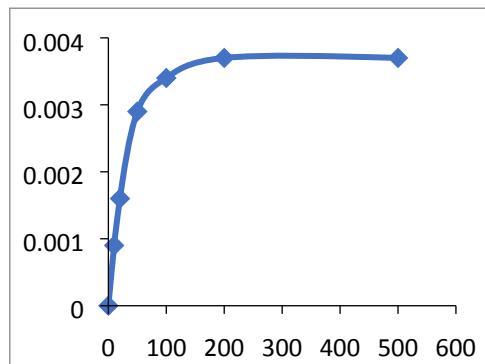
<b>K<sub>m</sub></b>	$0.78 \pm 0.28 \mu\text{M}$
<b>k<sub>cat</sub>/K<sub>M</sub></b>	$21.3 \text{ s}^{-1}\text{mM}^{-1}$
<b>k<sub>cat</sub></b>	$0.0166 \text{ s}^{-1}$

Fig 28c: Michaelis menten and Lineweaver-Burk plots for 76E5

#### 4) 76D1 UDP GLUCOSE

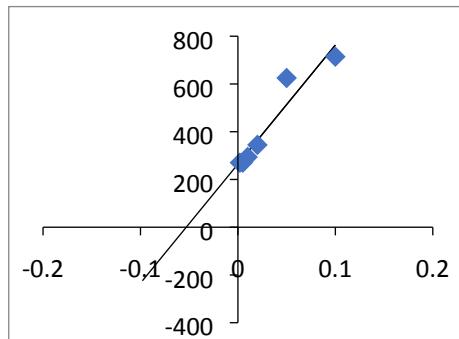
##### Michaelis menten plot

S ( $\mu\text{M}$ )	V ( $\mu\text{M}/\text{min}$ )
0	0
10	0.0009
20	0.0016
50	0.0029
100	0.0034
200	0.0037
500	0.0037



##### Lineweaver Burk

1/S	1/V
0.1	714.2857
0.05	625
0.02	344.8276
0.01	294.1176
0.005	270.2703
0.002	270.2703



<b>K<sub>m</sub></b>	$25.77 \pm 8.82 \mu\text{M}$
<b>k<sub>cat</sub>/K<sub>M</sub></b>	$0.185 \text{ s}^{-1}\text{mM}^{-1}$
<b>k<sub>cat</sub></b>	$0.00476 \text{ s}^{-1}$

Fig 29: Michaelis menten and Lineweaver-Burk plots for 76D1

