

Supplemental Figure 1. Validation of ASC-Citrine system in tissue imaging. (A) Representative images of ASC speck formation detected with ASC-Citrine and ASC antibody signals. Live spleen tissue culture slices from naïve WT and ASC-Citrine mice were used with NLRP3 inflammasome stimulation. Scale bar is 50  $\mu$ m. (B) Quantification of ASC specks in the iLNs and cLNs of ASC-Citrine mice during EAE. Each datapoint represents a value of an average value from two cross-sections of LNs (25  $\mu$ m thickness) from one mouse. One-way ANOVA, *p*=0.0021 (iLN), *p*=0.3235 (cLN), with Dunnett's multiple comparisons test. (C and D) Comparison of ASC speck images and numbers in SC between Type-A and Type-B EAE. Representative images (C) and quantification (D) of ASC specks in the SC of ASC-Citrine mice at 30-dpi for Type A (*n*=5) and Type B (*n*=8) EAE. Each datapoint represents a value from one mouse. Mann-Whitney test was used (*D*). Scale bar is 200  $\mu$ m. (*B*, *D*) *ns*; not significant (*p*>0.05), \**p*<0.05, \*\**p*<0.01. Error bars denote mean ± SEM (*B*, *D*).



Supplemental Figure 2. Validation of Bone Marrow Chimeras. (A) BM chimera were created by transferring WT BM cells to irradiated WT or Pycard<sup>-/-</sup> recipients (n=7 for each group). Reconstitution efficiency of BM chimeras determined by flow cytometry, quantified as % of total CD45<sup>+</sup> cells in peripheral blood for congenic markers of CD45.1 (donor) or CD45.2 (recipient). Each datapoint represents a value from one mouse. Mann-Whitney test used. (B and C) BM chimera were created by transferring ASC-Citrine BM cells irradiated WT recipients (ASC-Citrine  $\rightarrow$  WT, n=6) and vice versa (WT  $\rightarrow$  ASC-Citrine mice, n=8). Reconstitution efficiency (B) and EAE disease score (C) of indicated BM chimera. Each datapoint represents a value from one mouse (B). Each datapoint denotes mean EAE score per group, and Mann-Whitney test of total AUC was used for statistical evaluation (C). (D) Representative images of SC from WT recipients reconstituted with ASC-Citrine BM cells at indicated time points during EAE. No apparent ASC specks were observed. Scale bar is 500 µm. (E) Representative image of ALDH1L1 counterstaining of astrocytes in ASC-Citrine mice at 30-dpi EAE. Scale bar is 10 µm. (F) Representative images of ASC specks and strings counter-stained with antibodies against NG2 (for OPCs) and MBP (for mature oligodendrocytes) in SC from naïve versus 30-dpi EAE ASC-Citrine mice. Scale bar is 20 µm. (G) Quantification of ASC specks in OPCs and mature oligodendrocytes of SC from naïve versus 30dpi EAE ASC-Citrine mice. Each datapoint represents a value from one mouse. Two-way repeated measures (RM) ANOVA was used (main effect of cell type: nsp<0.7807). (H) Percentages of ASC specks detected in OPC or mature oligodendrocytes out of total ASC specks per section. L5 spinal cords at 30-dpi EAE were used for the analysis. (I) Percentage of ChAT<sup>+</sup> and ChAT<sup>-</sup> VH neurons containing ASC specks in SC from naïve vs. 30-dpi EAE ASC-Citrine mice. Each datapoint represents a value from one mouse (n=5). Two-way RM ANOVA was used (main effect of cell type: p < 0.001) with Sidak's multiple comparisons test post hoc. ns; not significant (p > 0.05), \*\*p < 0.01. Error bars denote mean ± SEM (A, B, G-I).



Supplemental Figure 3. Validation of EAE mice with cell type-specific ASC-Citrine expression. (A) Flow cytometry histograms showing tamoxifen-mediated expression of ASC-citrine reporter expression in microglia and splenic monocytes from  $Cx3cr1^{CreERT2}$ ; Asc-Citrine<sup>LSL</sup> mice with or without tamoxifen (TAM) treatment. (B) Flow cytometry gating strategy for identifying microglia. (C) EAE disease score of  $Cx3cr1^{CreERT2}$ ; Asc-Citrine<sup>LSL</sup> (*n*=5) vs.  $Cx3cr1^{CreERT2}$ ; Asc-Citrine<sup>LSL</sup> (*n*=5) mice. Both groups were treated with TAM. (D and E) EAE disease score of Asc-Citrine<sup>LSL</sup> (*n*=10) vs. Gfap<sup>Cre</sup>; Asc-Citrine<sup>LSL</sup> (*n*=13) (D) and Asc-Citrine<sup>LSL</sup> (*n*=13) vs. Syn1<sup>Cre</sup>; Asc-Citrine<sup>LSL</sup> (*n*=10) (E). Mann-Whitney test of total AUC of disease score was used for statistical analysis (C, D, E). ns; not significant (*p*>0.05). Each datapoint denotes mean EAE score per group with an error bar of mean ± SEM (C-E).



Supplemental Figure 4. Expression of inflammasome components and cell death markers in astrocytes during EAE. (A) Gene-set enrichment analysis of inflammasome-associated genes in bulk SC lysates and astrocytes (with astrocyte-specific Ribotag-HA enriched RNA) in naïve and 30-dpi EAE mice. Data represented as raw transcript counts derived from publicly available data (GEO Accession #: GSE100329). (B-G) Representative images (*B-D*) and quantification (*E-F*) of caspase-1 (*B*, *E*), IL-1 $\beta$  (*C*, *F*), and GSDMD (*D*, *G*) expression in spleen and SC astrocytes from naïve versus 30-dpi EAE ASC-Citrine mice. Scale bar is 20 µm. Each datapoint represents a value from one mouse. Individual astrocytes were identified using the Imaris software and the mean intensity per cell was quantified for caspase-1 (*E*), IL-1 $\beta$  (*F*) and GSDMD (*G*). Mann-Whitney test was used. (*E-G*) *ns*; not significant (*p*>0.05). Error bars denote mean ± SEM (*E-G*).



Supplemental Figure 5. Expression of inflammasome components in primary cortical astrocyte cell line (A and B) WB quantitative evaluation of culture supernatant samples of mature caspase-1 (*A*) and IL-1 $\beta$  (*B*). (C-F) WB quantitative evaluation of cell lysate samples of procaspase-1 (*C*), pro-IL-1 $\beta$  (*D*), GSDMD-FL (*E*), and GSDMD-NT (*F*). In (*A*-*F*), Cells in group 1 were unstimulated. Cells in group 2 were treated with Ultrapure LPS alone. Cells in groups 3 and 4 were pre-treated with Ultrapure LPS, and were further stimulated with nigericin and poly(dA:dT)/liposome to activate the NLRP3 and AIM2 inflammasomes, respectively. (G) Quantification of active caspase-3 (CC3) in spinal cord astrocytes comparing astrocytes with or without ASC specks in *Gfap*<sup>Cre</sup>;*Asc-Citrine*<sup>LSL</sup> (*n*=6) mice at 30-dpi EAE. Each datapoint represents a value from one mouse. Individual astrocytes were identified using the Imaris software and were quantified by CC3 puncta staining. Mann-Whitney test was used. \*\**p*<0.01. Error bars denote mean ± SEM.



Fig. S6. Validation of EAE phenotype of *NIrp3*-<sup>*I*</sup>;ASC-Citrine mice and immune phenotype of *Aim2*-<sup>*I*</sup> mice with EAE. (A) EAE disease score of ASC-Citrine (*n*=7) vs. *NIrp3*-<sup>*I*</sup>;ASC-Citrine (*n*=8) mice with Type B-EAE. Mann-Whitney test of total AUC for disease score was used. Each datapoint denotes mean EAE score per group with an error bar of mean ± SEM. (**B**–**D**) Leukocyte counts in SC (*B*), iLN (*C*) and spleen (*D*) at 16-dpi EAE in WT vs. *Aim2*-<sup>*I*</sup> mice induced with Type-A EAE. Each datapoint represents a value from one mouse Error bars denote mean ± SEM. Two-way RM ANOVA was used with Sidak's multiple comparisons test post hoc. (**E**) Representative image of GFAP staining in SC from WT versus *Aim2*-<sup>*I*</sup> mice at 30-dpi of EAE. Scale bar is 200 µm. *ns*; not significant (*p*>0.05) (*A-D*).