

Supplementary Materials for

IDH2-mediated regulation of the biogenesis of the oxidative phosphorylation system

Anjaneyulu Murari, Naga S. V. Goparaju, Shauna-Kay Rhooms, Kaniz F. B. Hossain, Felix G. Liang, Christian J. Garcia, Cindy Osei, Tong Liu, Hong Li, Richard N. Kitsis, Rajesh Patel, Edward Owusu-Ansah*

*Corresponding author. Email: eo2364@cumc.columbia.edu

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Figs. S1 to S18 Legends for tables S1 to S8

Other Supplementary Material for this manuscript includes the following:

Tables S1 to S8

Figure S1: Replicates of immunoblots shown in Figures 1E-1G

The figure legend is the same as what is described for figure panels 1E-1G.

Figure S2: Additional replicates of immunoblots shown in Figures 1E-1G

The figure legend is the same as what is described for figure panels 1E-1G.

Figure S3: Quantification of immunoblots shown in Figures 1E-1G

The antibodies quantified for 1E-1G are listed above each graph. In all instances except the 1F panels, p values are based on one-way ANOVAs followed by the Dunnett's test. In the 1F panels, p values are based on the student's t-test for unpaired two-tailed samples. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 15 flies per replicate.

Figure S4: NADPH or ferroptosis inhibitors fail to rescue the OXPHOS assembly defects observed when dIDH2 is knocked down.

(A-F) Mitochondrial preparations from thoraces isolated from Dmef2-Gal4/w1118 (wild type) and dIDH2-KD1 flies raised on a diet supplemented with 5mM and 10mM NADPH 4 days after eclosure. The mitochondrial preparations were analyzed by BN-PAGE, followed by immunoblotting with the antibodies indicated. The blots were imaged following a short exposure to detect the holoenzyme and supercomplexes, after which the region corresponding to the holoenzyme and supercomplexes was cut off, and the rest of the blot re-imaged after a longer exposure to detect the assembly intermediates (denoted as *). The *#* refers to the initiating AI of the Q-module. The antibodies used were anti-NDUFS3 which detects dNDUFS3 (A), anti-dNDUFS7 (B), anti-dNDUFS8 (C), anti-dNDUFA9 (D), anti-dND1 (E), and anti-dND3 (F). Anti-ATP synthase, subunit B (ATP5F1B) which detects the *Drosophila* ortholog, dATP-Synβ, was used as a loading control.

(G-J) Mitochondrial preparations from thoraces isolated from Dmef2-Gal4/w1118 (wild type) and dIDH2-KD2 flies raised on a diet supplemented with 1mg/ml each of ferrostatin-1 and liproxstatin-1 (FL-1) 4 days after eclosure. The antibodies used were anti-NDUFS3 which detects dNDUFS3

(G), anti-dNDUFA9 (H), anti-dNDUFV3 (I) and anti-dNDUFB5 (J). Anti-ATP synthase, subunit B (ATP5F1B) which detects the *Drosophila* ortholog, dATP-Syn β , was used as a loading control.

Figure S5: Replicates of immunoblots shown in Figure 4

The figure legend is the same as what is described for Figure 4.

Figure S6: Additional replicates of immunoblots shown in Figure 4

The figure legend is the same as what is described for Figure 4.

Figure S7: Quantification of immunoblots shown in Figure 4

The antibody quantified for each figure panel is listed above each graph. In all instances, p values are based on one-way ANOVAs followed by the Dunnett's test. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 40 flies per replicate.

Figure S8: Replicates of immunoblots shown in Figure 5

The figure legend is the same as what is described for Figure 5.

Figure S9: Additional replicates of immunoblots shown in Figure 5

The figure legend is the same as what is described for Figure 5.

Figure S10: Quantification of immunoblots shown in Figure 5

The antibody quantified for each figure panel is listed above each graph. In all instances, p values are based on one-way ANOVAs followed by the Dunnett's test. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 40 flies per replicate.

Figure S11: Replicates of immunoblots shown in Figure 8

The figure legend is the same as what is described for Figure 8.

Figure S12: Additional replicates of immunoblots shown in Figure 8

The figure legend is the same as what is described for Figure 8.

Figure S13: Quantification of immunoblots shown in Figures 8A-8C

The antibodies quantified for figure panels are listed above each graph. In 8C, p values are based on one-way ANOVAs followed by Dunnett's multiple comparisons test. In 8A and 8B, one-way ANOVAs followed by Tukey's multiple comparisons test was used. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 15 flies per replicate.

Figure S14: Quantification of immunoblots shown in Figures 8D and 8E

The antibodies quantified for the figure panels are listed above each graph. Due to recurrent phosphorylation-dephosphorylation oscillations, we quantified each kinase experiment separately in Figure 8D. The p values in 8E are based on one-way ANOVAs followed by Dunnett's multiple comparisons test. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 15 flies per replicate.

Figure S15: Quantification of immunoblots shown in Figure 8F

The antibody quantified for each figure panel is listed above each graph. Due to recurrent phosphorylation-dephosphorylation oscillations, we quantified each kinase experiment separately. For HSP60 and cytochrome C, one-way ANOVAs followed by Tukey's multiple comparisons test was used. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 15 flies per replicate.

Figure S16: Replicates of immunoblots shown in Figure 9

The figure legend is the same as what is described for Figure 9.

Figure S17: Additional replicates of immunoblots shown in Figure 9

The figure legend is the same as what is described for Figure 9.

Figure S18: Quantification of immunoblots shown in Figure 9

The antibodies quantified for the figure panels are listed above each graph. In all instances, p values are based on the student's t-test for unpaired two-tailed samples. The fold change shown refers to the mean \pm s.e.m (standard error of the mean); and n.s. denotes p > 0.05, * = p<0.05, ** = p<0.01 and *** = p<0.001. The number of replicates (n) = 3 biological replicates, with 40 flies per replicate.

Table S1: Comparison of identified proteins and their relative quantitation among four A samples using the spectra counting method (sorted by protein name)

Table S2: Comparison of identified proteins and their relative quantitation among four B samples using the spectra counting method (sorted by protein name)

Table S3: Comparison of identified proteins and their relative quantitation among four C samples using the spectra counting method (sorted by protein name)

Table S4: Comparison of identified proteins and their relative quantitation among four Dsamples using the spectra counting method (sorted by protein name)

Table S5: Comparison of identified proteins and their relative quantitation among four E samples using the spectra counting method (sorted by protein name)

Table S6: Comparison of identified proteins and their relative quantitation among four F samples using the spectra counting method (sorted by protein name)

Table S7: RNA-seq profile when dIDH2 is disrupted in flight muscles

Table S8: A list of the peptide antigens and their target proteins.



















1E. a-dUQCRC2



























5L. α-dNDUFB1

5M. α-dNDUFB5

5N. α-dNDUFB6

5O. α-dNDUFB8

























8C. α-dHSP10A















25

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GRA

25[.]

0

w1118

Since; GFP

annu.re

25

0

JIDHD PROF. CFP ann Person Conta

wills

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w1118 DH2. POWL GFP

ann. ann.

HDHL.PONS,

