Supporting Information

In vivo neutralization of Myotoxin II, a phospholipase A_2 homolog from *Bothrops asper* venom, using peptides discovered via phage display technology

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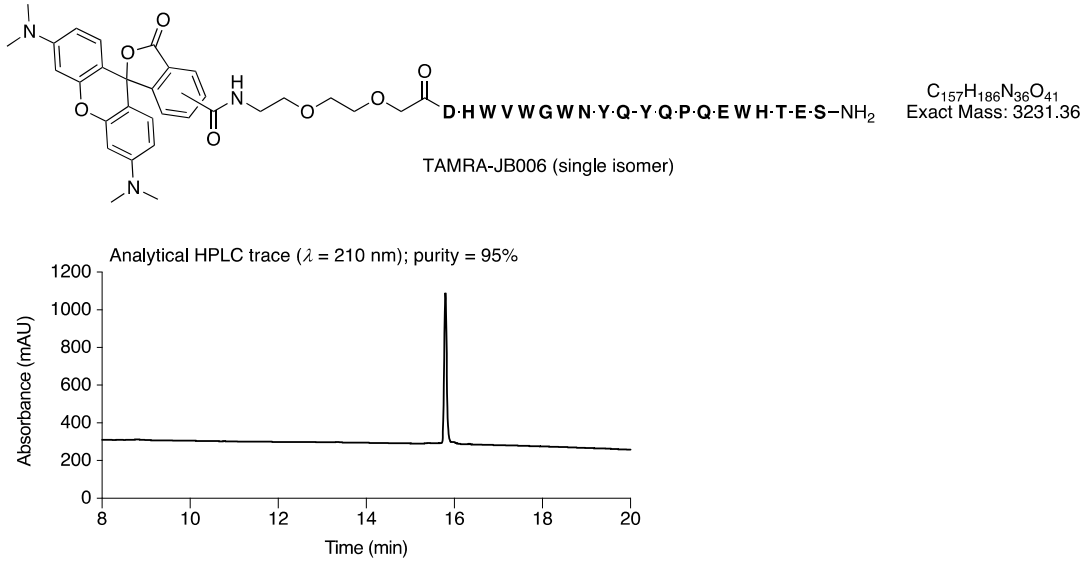

Figure S1. Structure of TAMRA-JB006 and analytical HPLC trace of purified TAMRA-JB006. The peptide was isolated as single TAMRA isomer but the identity of the isomer was not resolved.

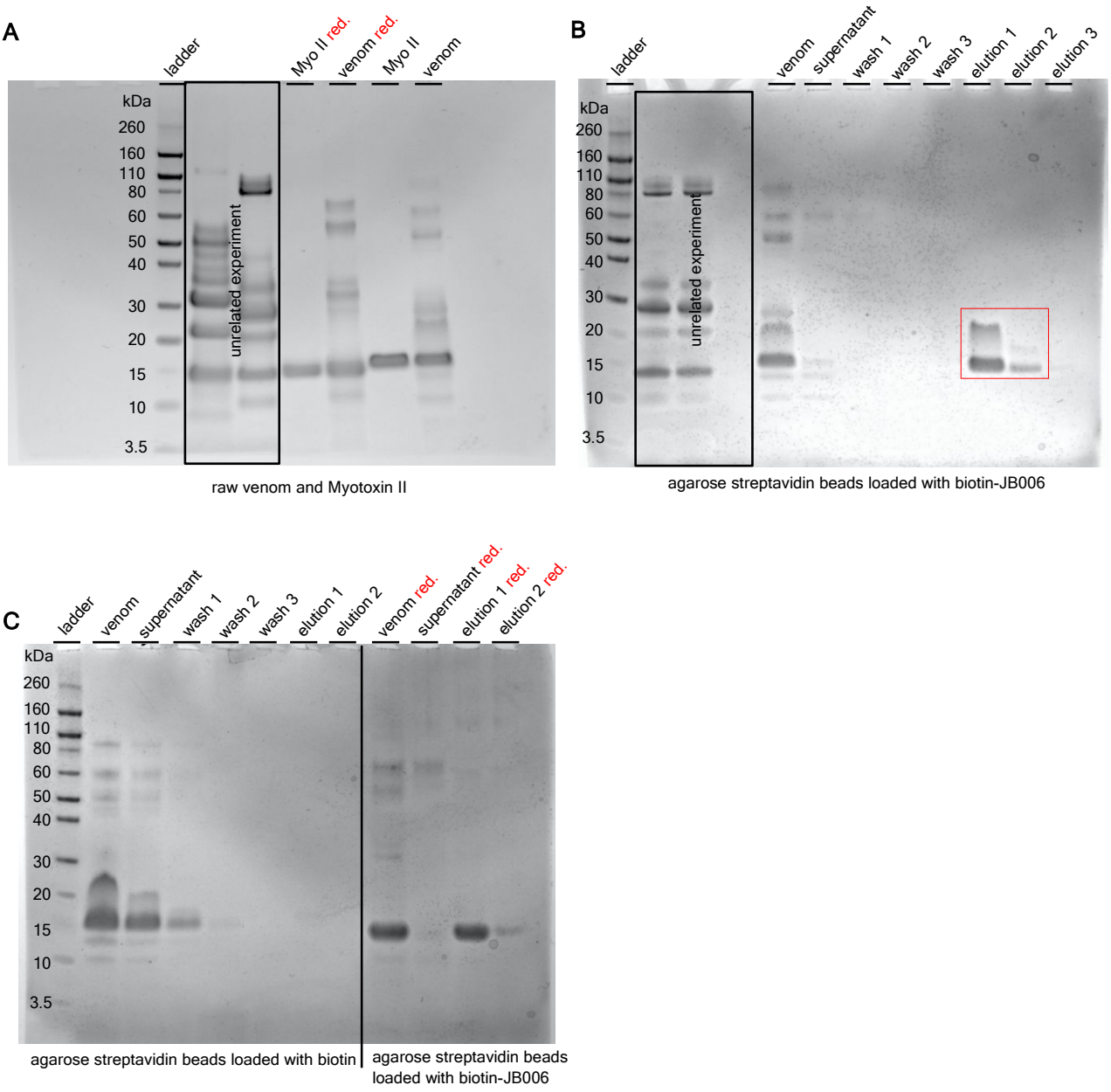

Figure S2. SDS-page gels stained with Coomassie blue. **A)** Myotoxin II and *B. asper* crude venom reduced and non-reduced. **B)** Pulldown experiment with agarose streptavidin beads loaded with biotinylated JB006 peptide; lane 5: *B. asper* crude venom (1.0 mg/mL) before incubation, lane 6: supernatant after overnight incubation of *B. asper* crude venom at 4 °C, lane 7–9: washing of beads with PBS buffer, lane 10–12: elution from beads with glycine HCl buffer (0.1 M, pH 2.8). **C)** Pulldown experiment with agarose streptavidin beads loaded with biotin; lane 2: *B. asper* crude venom (1.0 mg/mL) before incubation, lane 3: supernatant after overnight incubation of *B. asper* crude venom at 4 °C, lane 4–6: washing of beads with PBS buffer, lane 7–8: elution from beads with glycine HCl buffer (0.1 M, pH 2.8). Pulldown experiment with agarose streptavidin beads loaded with biotinylated JB006; lane 9: *B. asper* crude venom (1.0 mg/mL) before incubation, reduced, lane 10: supernatant after overnight incubation of *B. asper* crude venom at 4 °C, reduced, lane 11–12: elution from beads with glycine HCl buffer (0.1 M, pH 2.8) after 3 washing steps with PBS buffer, reduced.