

1 **Supplementary materials**

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3 **Methylation-dependent and -independent roles of EZH2 synergize in CDCA8**
4 **activation in prostate cancer**

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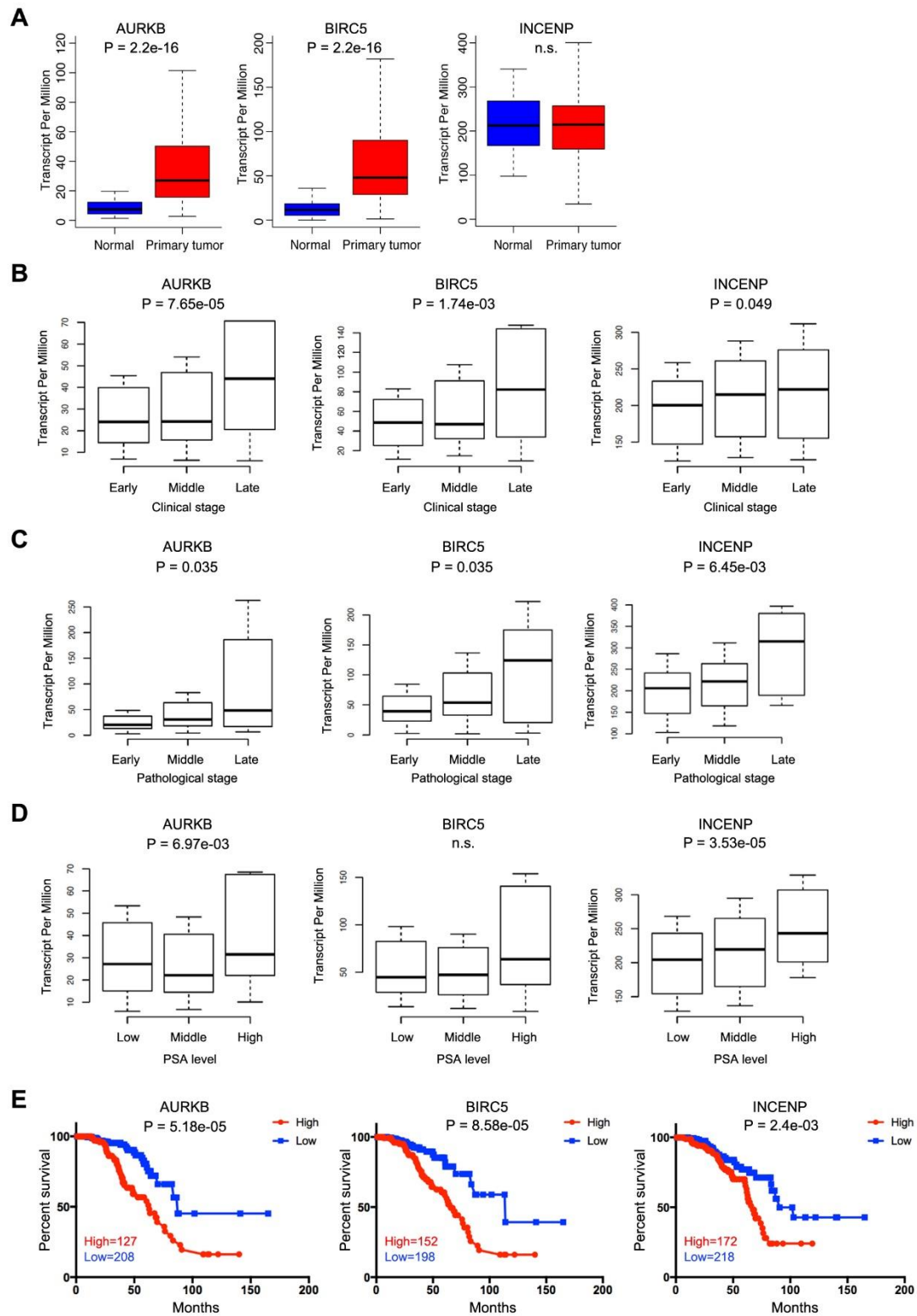
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9 **Contents**

10 **Supplementary Figures 1-6**

11 **Supplementary Table 1, 4 and 5**

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14 **Supplementary Figure 1. Dysregulation of CPC members other than CDCA8 in**
 15 **PCa**

16 (A) Box plots showing the mRNA levels of AURKB, BIRC5 and INCENP in normal
 17 (n=52) and PCa (n=497) specimens using data from TCGA.

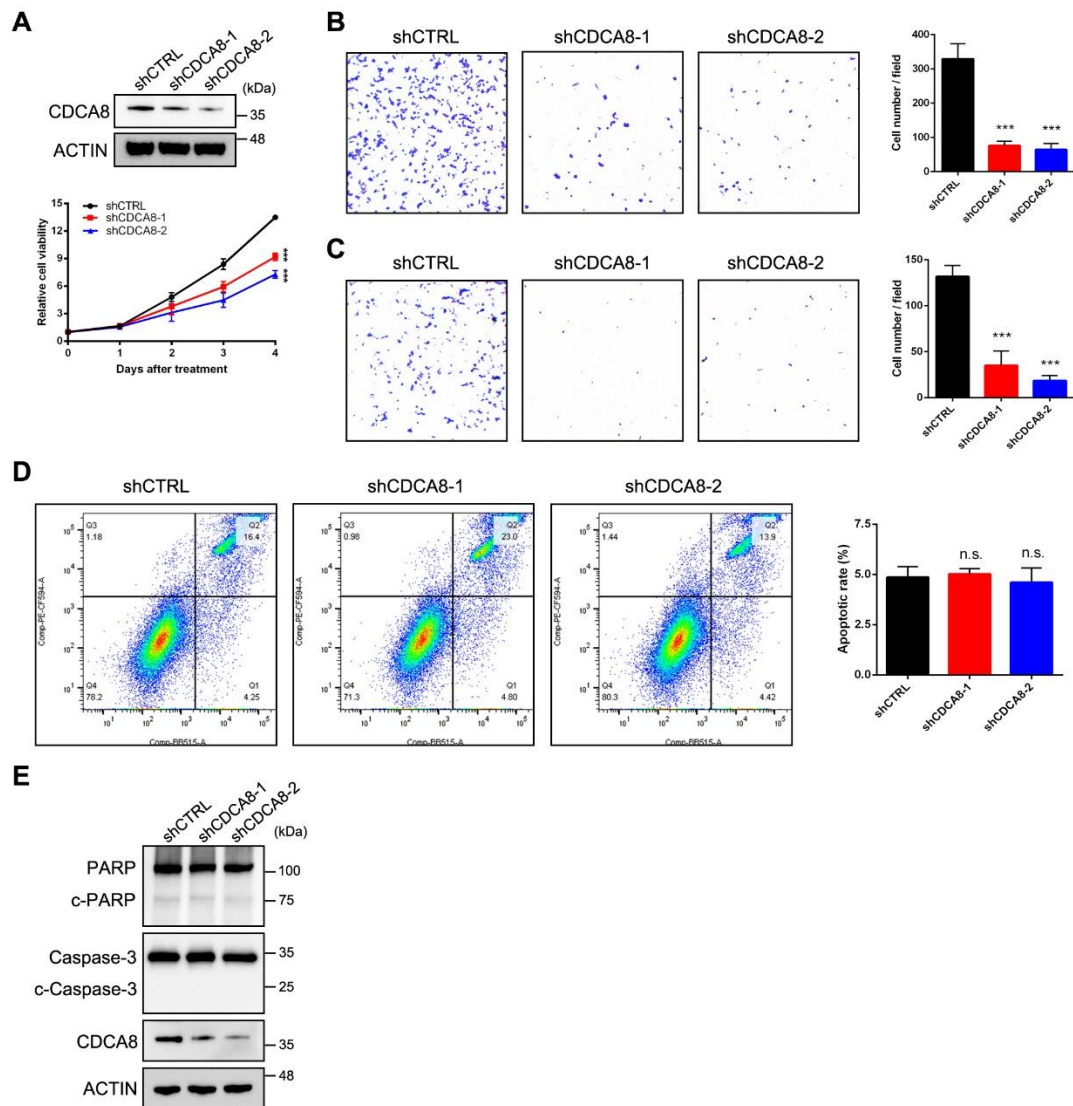
18 **(B)** Box plots showing the mRNA levels of AURKB, BIRC5 and INCENP in PCa
19 patients with different clinical stages ($n_{\text{early}}=177$, $n_{\text{middle}}=173$, $n_{\text{late}}=53$) using data
20 from TCGA.

21 **(C)** Box plots showing the mRNA levels of AURKB, BIRC5 and INCENP in PCa
22 patients with different pathological stages ($n_{\text{early}}=186$, $n_{\text{middle}}=293$, $n_{\text{late}}=10$) using
23 data from TCGA.

24 **(D)** Box plots showing the mRNA levels of AURKB, BIRC5 and INCENP in PCa
25 patients with different PSA levels ($n_{\text{low}}=181$, $n_{\text{middle}}=209$, $n_{\text{high}}=48$) using data from
26 TCGA.

27 **(E)** The association between AURKB, BIRC5 or INCENP expression and Disease-free
28 survival time of PCa patients was analyzed by Kaplan-Meier analysis using data from
29 TCGA.

30



31

32 **Supplementary Figure 2. CDCA8 maintains the aggressiveness of PCa cells**

33 **(A)** Cell viability assay was used to assess the proliferative capacity of control and
 34 CDCA8-deficient PC-3 cells. The knockdown efficiency of CDCA8 was validated by
 35 western blot.

36 **(B)** Boyden chamber migration assay was performed to determine the migratory
 37 capability of PC-3 cells after CDCA8 depletion. Graph showing the number of migrated
 38 cells in the lower surface of filter at 24 h. Data represent Mean \pm SD from n=5 random
 39 fields per filter.

40 **(C)** Boyden chamber invasion assay was performed to determine the invasive capability
 41 of PC-3 cells after CDCA8 depletion. Graph showing the number of migrated cells
 42 passing through Matrigel at 24 h. Data represent Mean \pm SD from n=5 random fields

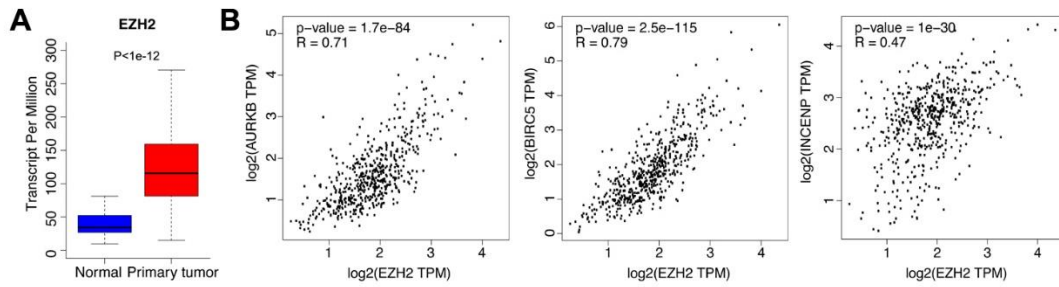
43 per filter.

44 **(D)** Flow cytometry assays were performed to analyze the apoptotic rate in control and
45 CDCA8-deficient C4-2 cells. Graph showing the percentage of apoptotic cells at each
46 group.

47 **(E)** Western blot analysis of the protein levels of full-length and cleaved PARP or
48 Caspase-3 in C4-2 cells upon CDCA8 knockdown.

49 *, $P < 0.05$, **, $P < 0.01$, ***, $P < 0.001$ is based on the student's t-test unless otherwise
50 stated. Values are mean \pm SD of three independent experiments.

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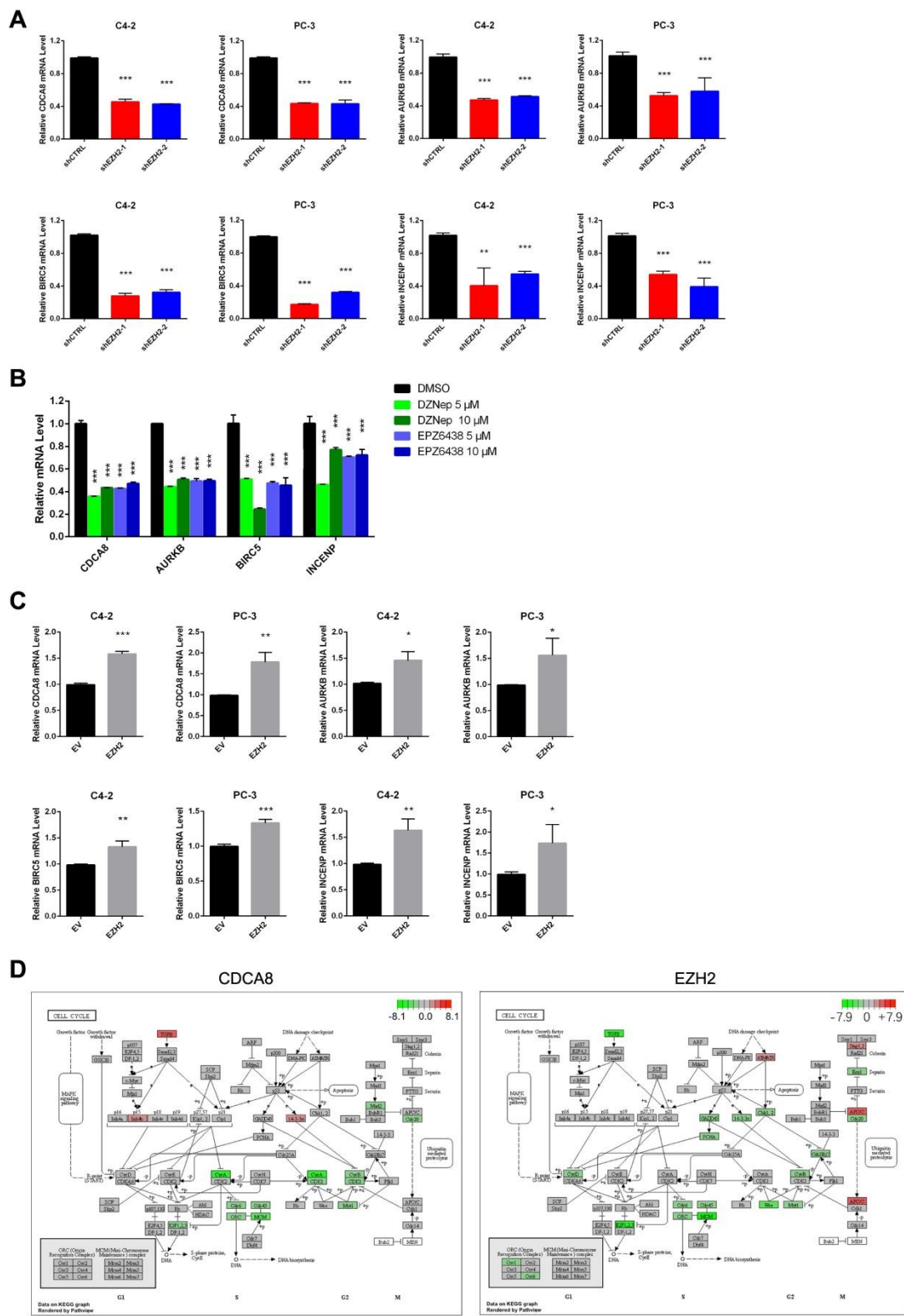
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53 **Supplementary Figure 3. CPC members other than CDCA8 are positively**
 54 **correlated with EZH2 in PCa**

55 **(A)** Box plot showing the mRNA level of EZH2 in normal (n=52) and PCa (n=497)
 56 specimens using data from TCGA.

57 **(B)** Scatter plots showing the relationship between AURKB/BIRC5/INCENP and
 58 EZH2 expressions using data from TCGA, with Spearman correlation coefficient (R)
 59 and P values as indicated. TPM, transcript per million.

60



61

62 **Supplementary Figure 4. EZH2 regulates transcription of CDCA8 and other CPC**
 63 **members**

64 (A) RT-qPCR analysis of the mRNA levels of CDCA8 and other CPC members in PCa
 65 cell lines upon EZH2 knockdown.

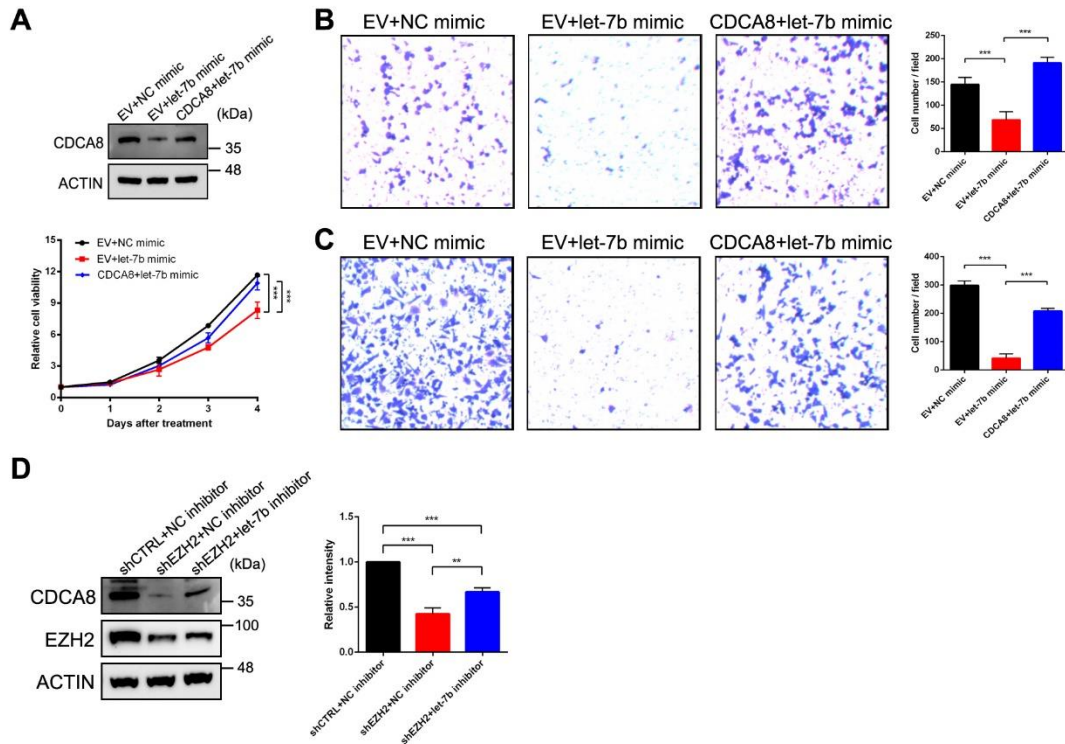
66 **(B)** RT-qPCR analysis of the mRNA levels of CDCA8 and other CPC members in C4-
67 2 cells after treatment of EZH2 inhibitors as indicated.

68 **(C)** RT-qPCR analysis of the mRNA levels of CDCA8 and other CPC members in PCa
69 cell lines upon EZH2 overexpression.

70 **(D)** KEGG graphs showing the dysregulated genes in cell cycle pathway upon CDCA8
71 knockdown or EZH2 knockdown in C4-2 cells.

72 *, P < 0.05, **, P < 0.01, ***, P < 0.001 is based on the student's t-test unless otherwise
73 stated. Values are mean \pm SD of three independent experiments.

74



75

76 **Supplementary Figure 5. Repression of let-7b is partially responsible for the**
 77 **EZH2-mediated CDCA8 upregulation in PCa**

78 **(A)** Cell viability assay showing that overexpression of CDCA8 could rescue the
 79 decreased proliferation rate in C4-2 cells treated with let-7b mimics. The protein level
 80 of CDCA8 in each group was detected by western blot. EV, empty vector.

81 **(B)** Boyden chamber migration assay showing that overexpression of CDCA8 could
 82 rescue the decreased migration rate in C4-2 cells treated with let-7b mimics. Graph
 83 showing the number of migrated cells in the lower surface of filter at 24 h. Data
 84 represent Mean \pm SD from n=5 random fields per filter.

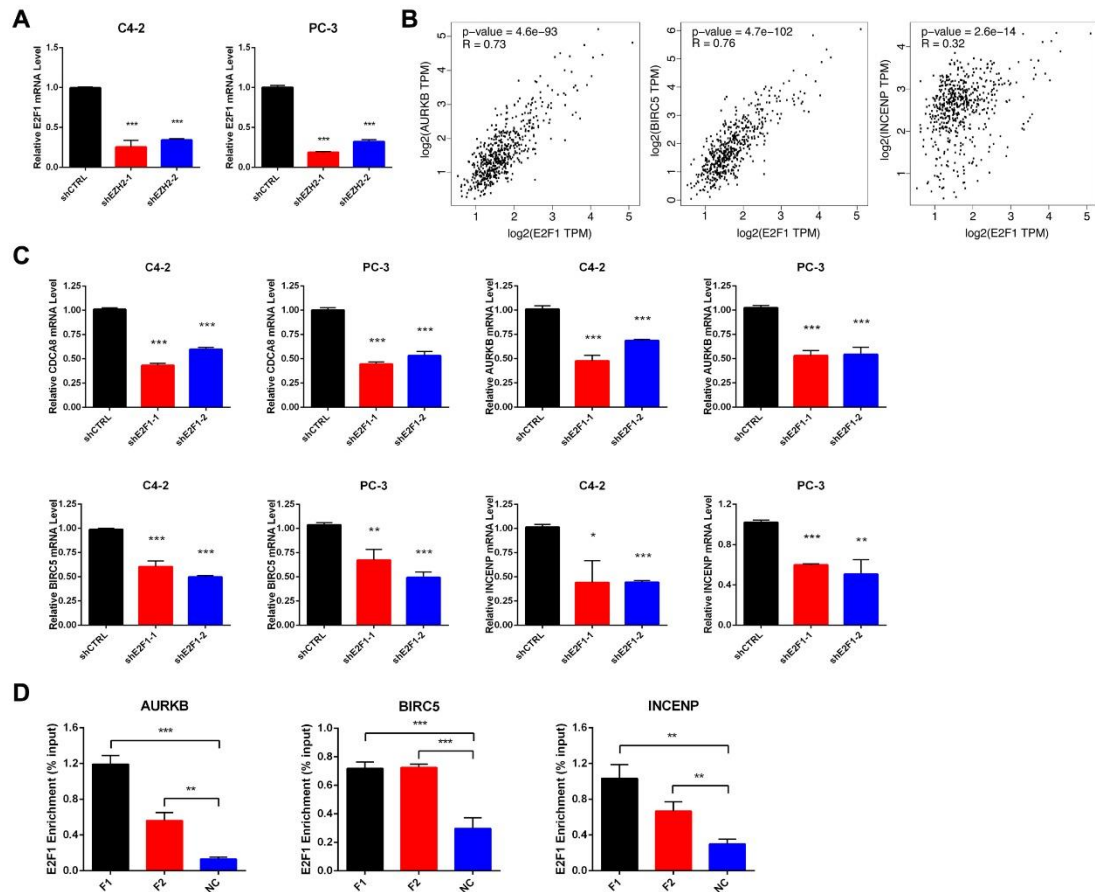
85 **(C)** Boyden chamber invasion assay showing that overexpression of CDCA8 could
 86 rescue the decreased invasion rate in C4-2 cells treated with let-7b mimics. Graph
 87 showing the number of migrated cells passing through Matrigel at 24 h. Data represent
 88 Mean \pm SD from n=5 random fields per filter.

89 **(D)** Rescue assay showing that treatment of let-7b inhibitor could not fully restore the
 90 downregulation of CDCA8 in EZH2-deficient PC-3 cells, as measured by western blot.
 91 Graph represents the relative CDCA8 protein level in each group.

92 *, P < 0.05, **, P < 0.01, ***, P < 0.001 is based on the student's t-test unless otherwise

93 stated. Values are mean \pm SD of three independent experiments.

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Supplementary Figure 6. E2F1 activates transcription of CPC members other than CDCA8 in PCa

97

98 **(A)** RT-qPCR analysis of the mRNA level of E2F1 in PCa cell lines upon EZH2
99 knockdown.

100 **(B)** Scatter plot showing the relationship between AURKB/BIRC5/INCENP and E2F1
101 expressions using data from TCGA, with Spearman correlation coefficient (R) and P
102 value as indicated.

103 **(C)** RT-qPCR analysis of the mRNA levels of all CPC members in PCa cell lines upon
104 E2F1 knockdown.

105 **(D)** ChIP-qPCR assay to monitor the enrichment of E2F1 at the promoter region of
106 AURKB/BIRC5/INCENP in C4-2 cells. Two pairs of primers (F1 and F2) were used to
107 amplify fragments inside AURKB/BIRC5/INCENP promoter region while another pair
108 of primers (NC) targeting nearby region was used as negative control.
109

110 **Supplementary Table 1. PCa TMA IHC data revealed correlation between**
 111 **expressions of CDCA8 and EZH2**

Pearson r Correlation	
CDCA8 vs. EZH2	
Pearson r	
r	0.8234
95% confidence interval	0.7569 to 0.8730
R squared	0.6779
P value	
P (two-tailed)	< 0.0001
P value summary	****
Significant? (alpha = 0.05)	Yes
Number of XY Pairs	124

112

113

Supplementary Table 4. Antibodies used in this study

Target	Source	Application (Dilution)
CDCA8	Abcam, Cat#ab74473	IHC (1:50), WB (1:1000)
AURKB	Sigma, Cat#MABE627	WB (1:1000)
BIRC5	Cell signaling, Cat#2808	WB (1:1000)
INCENP	Abcam, Cat#ab12183	WB (1:1000)
EZH2	Cell signaling, Cat#5246	IHC (1:50), WB (1:1000), ChIP-qPCR (1:100)
E2F1	Cell signaling, Cat#3742	WB (1:1000), ChIP-qPCR (1:100)
ACTIN	Cell signaling, Cat#3700	WB (1:1000)
GAPDH	Cell signaling, Cat#5174	WB (1:1000)
Histone H3	Cell signaling, Cat#4499	WB (1:1000)
H3K27me3	Cell signaling, Cat#9733	WB (1:1000), ChIP-qPCR (1:100)
LC3A/B	Cell signaling, Cat#12741	WB (1:1000)
p62/SQSTM1	Cell signaling, Cat#88588	WB (1:1000)
PARP	Cell signaling, Cat# 9532	WB (1:1000)
Caspase-3	Cell signaling, Cat#9662	WB (1:1000)

Supplementary Table 5. Primers used in this study

Name	Sequence (5' to 3')	Application
GAPDH	F: GGAGCGAGATCCCTCCAAAAT	RT-qPCR
	R: GGCTGTTGTCATACTTCTCATGG	
CDCA8	F: GAAGGGCAGTAGTCGGGTG	RT-qPCR
	R: TCACGGTCGAAGTCTTTCAGA	
AURKB	F: CAGTGGGACACCCGACATC	RT-qPCR
	R: GTACACGTTTCCAACTTGCC	
BIRC5	F: AGGACCACCGCATCTCTACAT	RT-qPCR
	R: AAGTCTGGCTCGTTCTCAGTG	
INCENP	F: AAGTCATGGAGTTTCTCTGC	RT-qPCR
	R: CGTCTCTTCTCCGTCGGTTC	
E2F1	F: ACGCTATGAGACCTCACTGAA	RT-qPCR
	R: TCCTGGGTCAACCCCTCAAG	
let-7b promoter -F1	F: CCCCAGGAAGGTGGTAGCC	ChIP-qPCR
	R: GGACAGAGTGTAGCATGAGGATGA	
let-7b promoter -F2	F: GTGACAGCGTCGCAAATG	ChIP-qPCR
	R: GCAGGAAACCACCAACCAG	
let-7b promoter -NC	F: ACGGGGCAGCCACCAA	ChIP-qPCR
	R: TGAGGGAGGACGGAAGGAC	
CDCA8 promoter -F1	F: AGCAGAATCCTACAGCCGACC	ChIP-qPCR
	R: GCGAACTATACAACTACTACTCCCG	
CDCA8 promoter -F1	F: CGCATTGGGCGGAAGA	ChIP-qPCR
	R: CCCGAGACAAGGGCTGAG	
CDCA8 promoter -NC	F: CAAGACCCGTGTGCGAGC	ChIP-qPCR
	R: TGTTGATTACATCAGGGAGCGT	
AURKB promoter -F1	F: TCAGAGGGTCCGTTGGGC	ChIP-qPCR
	R: CCGTGAGAAGCAGAGAAAAAGAG	
AURKB promoter	F: CTAAACTGGAAGCCAAGCGTG	ChIP-qPCR

-F2	R: TCCAAGGCACTGCTACTCTCC	
AURKB promoter	F: TGCCTCCCAGGTTCAAGC	ChIP-qPCR
-NC	R: GCCAACACAGTGAAACCCC	
BIRC5 promoter	F: ACTCAAGTGATGCTCCTGCCT	ChIP-qPCR
-F1	R: CGACTGCTTTCAAAGAACGC	
BIRC5 promoter	F: AGGACTTACTGTTGGTGGGACG	ChIP-qPCR
-F2	R: TGACTGCACGACCTGGGTTT	
BIRC5 promoter	F: TGGCAGCCCCACACAGA	ChIP-qPCR
-NC	R: AAGGAAAGCATGAGCACGACT	
INCENP promoter	F: TTGGTTGGGAAGTGTGGACTT	ChIP-qPCR
-F1	R: GGGGTCTGGGGCTTTCTC	
INCENP promoter	F: GTGTTTTCTCTTGCTCAATGCTT	ChIP-qPCR
-F2	R: CCTCCCACAACCCTCTCCC	
INCENP promoter	F: CCTCATCTGGCGATTGTGTC	ChIP-qPCR
-NC	R: TGTAAGCAAGTAGGTTGGGTCA	
E2F1 promoter	F: CGCCGTTGTTCCCGTCA	ChIP-qPCR
-F1	R: CGCCGCTGCCTGCAAAGT	
E2F1 promoter	F: GCCTATGTTCCGGTGTCCC	ChIP-qPCR
-F2	R: GCGGCGGTTCCCTATTGG	
E2F1 promoter	F: AGACGGGGAGCATCACAGG	ChIP-qPCR
-NC	R: CCCTCATCCCTCACCACAGA	