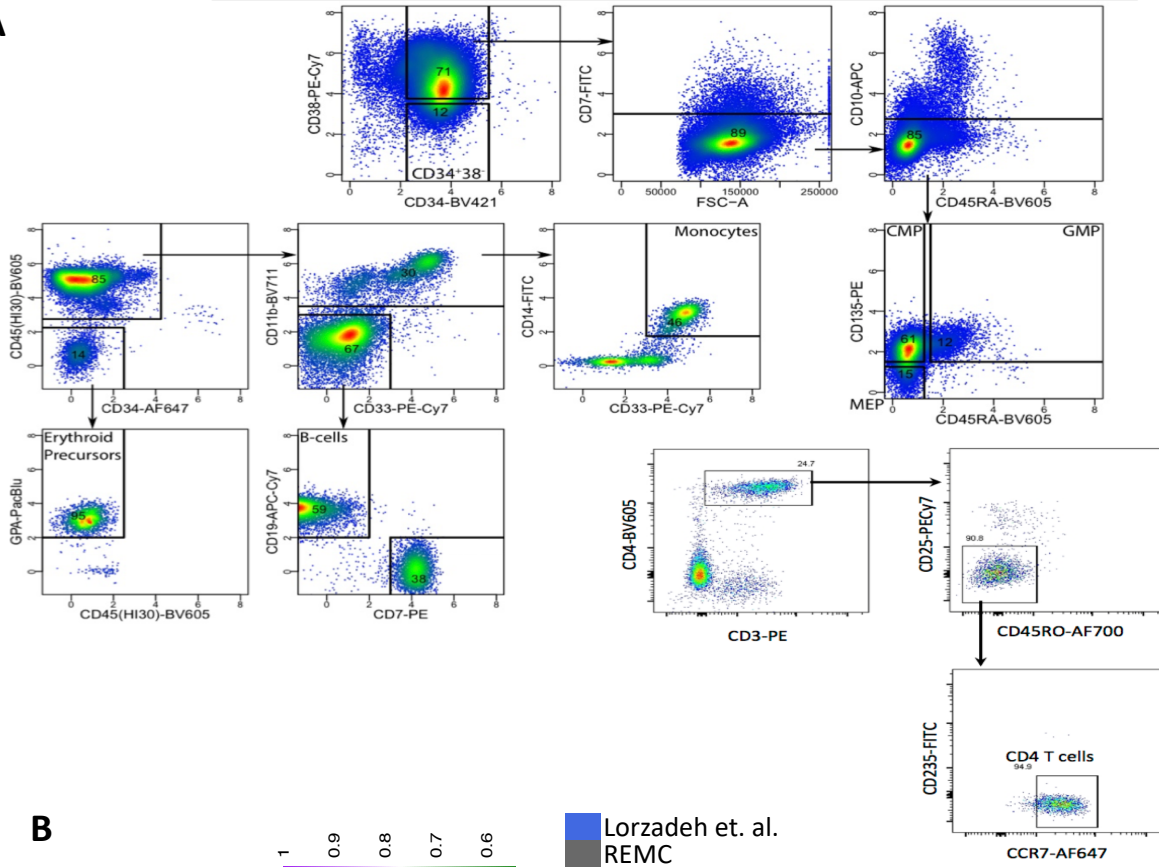


Supplemental Figures and Legends

A



B



Figure S1. Sorting strategy for hematopoietic populations profiled in this study. A) Representative examples of sorting strategies for CD34+CD38-, CMP, GMP, MEP, monocytes, erythroblasts, B and T cell isolated from cord blood pools. **B)** Unsupervised hierarchical clustering and heatmap of pairwise spearman correlations for protein coding gene RPKM values across blood cell types profiled in this study in the context of all cell types profiled by NIH Epigenome RoadMap Consortium. The cluster of blood cell types is indicated by the shaded box.

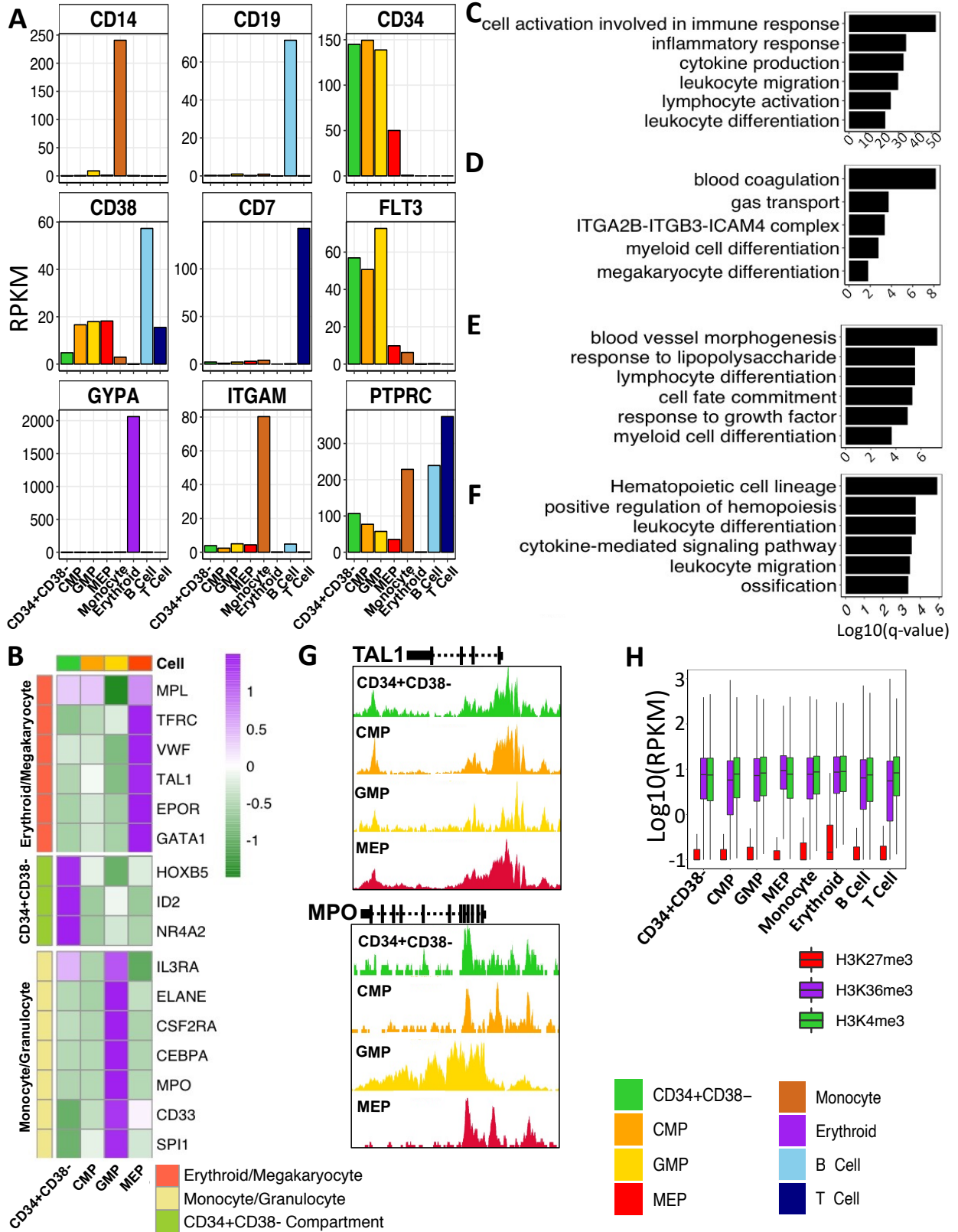


Figure S2. Progenitor populations possess unique expression profiles. **A)** Expression (RPKM) of cell type specific cell surface marker genes across cell types as indicated by the colour legend on the bottom right. **B)** Expression of previously identified progenitor population specific genes across CD34+CD38-, CMP, GMP and MEP. Pathway enrichment analysis of GMP (**C**), MEP (**D**), CD34+CD38- (**E**), and CMP (**F**) differentially expressed gene sets identified by DEFine (FDR > 0.01). **G)** Genome browser view of H3K4me3 density in progenitor populations at the *TALI* and *MPO* locus across CD34+CD38-, CMP, GMP and MEP. **H)** Expression of genes marked with H3K4me3, H3K27me3 or H3K36me3 across each cell type.

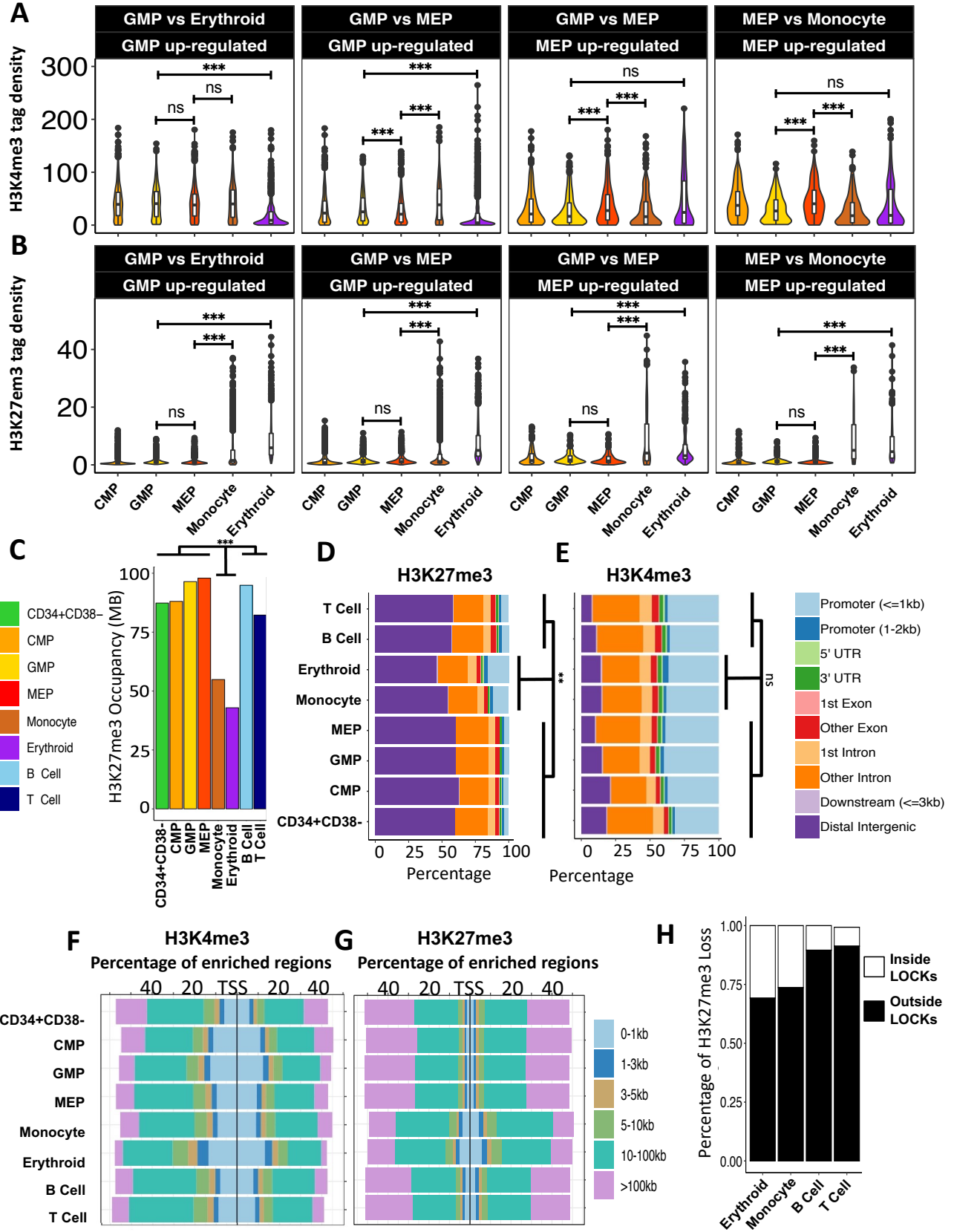


Figure S3. H3K27me3 promoter density signatures are conserved across progenitor populations. H3K4me3 (A) and H3K27me3 (B) tag density ± 2 Kb of transcription start sites of genes up-regulated in GMP and MEP across CMP, GMP, MEP, monocyte and erythroid precursor as indicated by the colour legend. C) Plot of the cumulative number of base pairs marked by H3K27me3 across cell types indicated by colour legend on the bottom right. Percentage of H3K27me3- (D) and H3K4me3- (E) enriched regions at the genomic features indicated. Percentage of H3K4me3 (F) and H3K27me3 (G) enriched regions with respect to their binned distance to the TSS of coding genes across cell types indicated by colours. Distance from TSS for each bin indicated in the bottom panel. H) Fraction of H3K27me3 loss in erythroid and monocyte within or outside the LOCKs (** $P < 0.01$, *** $P < 0.001$).

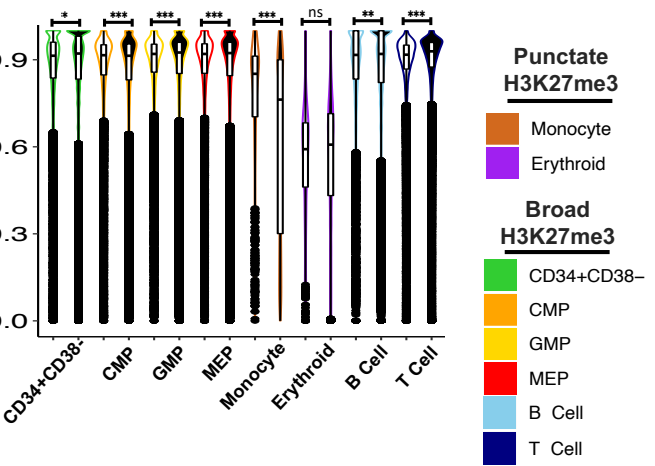
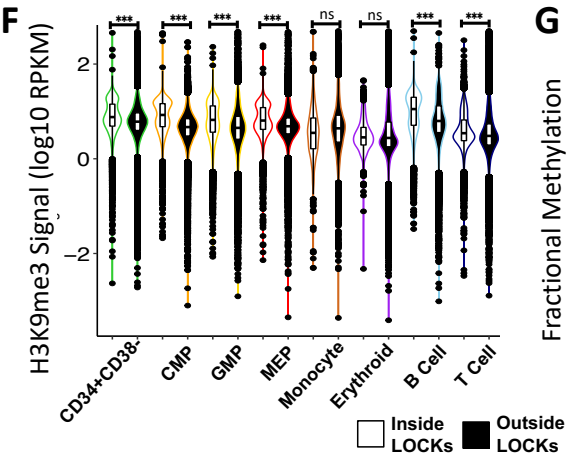
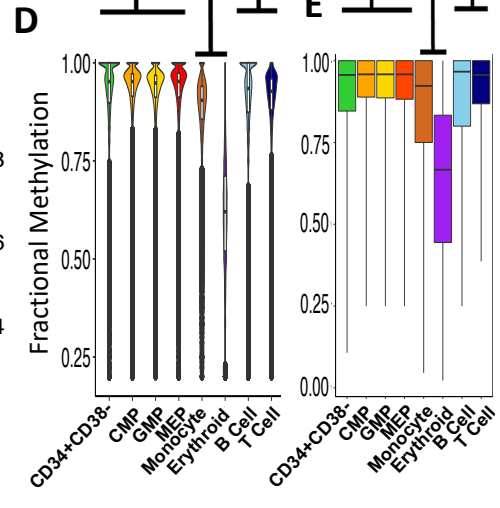
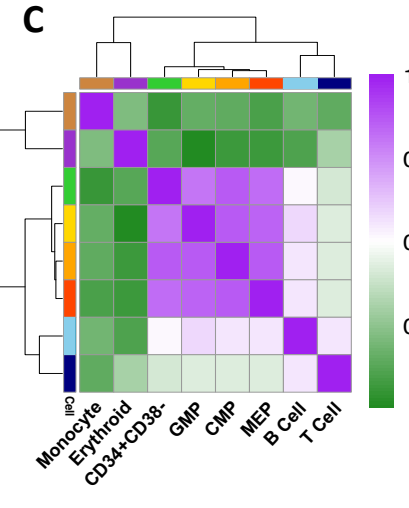
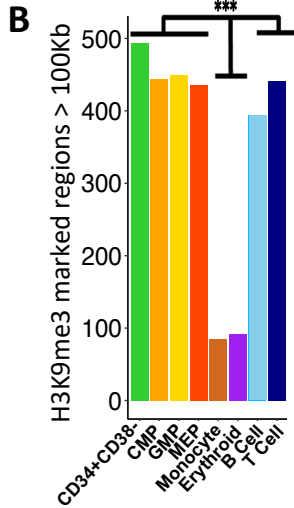
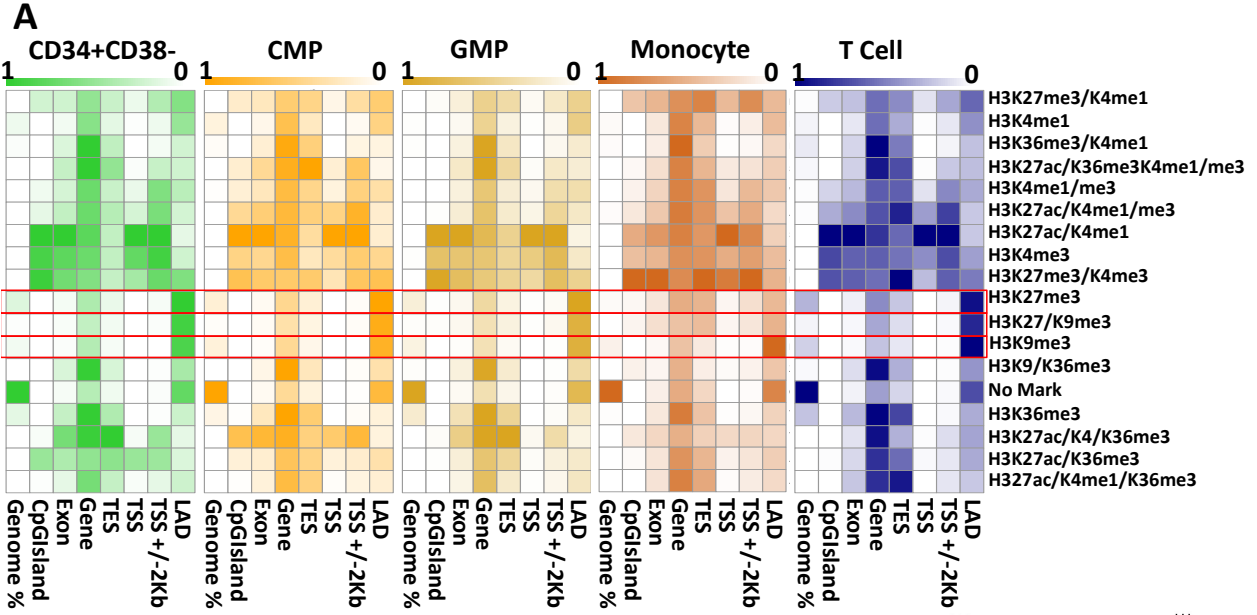


Figure 4. H3K27me3 LOCKs lost during myeloid differentiation are associated with LADs in CD34+ progenitor cells. **A)** Emission probabilities by histone mark for the 18 states of the ChromHMM chromatin state model across all cell types (upper panel). Emission probabilities are based on ChIP-seq data presented in this study. Enrichment of chromatin states within genomic features for MEPs (red), erythroblasts (purple) and B cells (blue). **B)** Cumulative number of bases marked by indicated histone modification within progenitor population LOCKs as indicated by the colour legend (bottom right). **C)** Genome browser view of H3K27me3 and H3K9me3 density on chromosome 20 across cell types indicated by the colour legend (bottom left). The shaded box indicates a H3K27me3 LOCK (FDR < 0.05) present in progenitor cells but lost in monocytes and erythroblasts. ChromHMM derived heatmap of z-score enrichment of identified polycomb repressed (**D**) and dually repressed (**E**) chromatin states at genomic features indicated. **F)** Barplot showing percentage of LOCKs overlapping with laminB binding sites. **G)** Barplot showing percentage overlap of LOCKs lost in monocyte or erythroblasts with laminB binding site. **H)** Violin plot showing occupancy of H3K9me3 (log10(bps)) at LOCKs across all populations as indicated by the colour legend (bottom right). Violin plot of fractional CpG methylation levels within progenitors' LOCKs (**I**), H3K27me3 enriched regions (**J**) and regions containing both H3K27me3 and H3K9me3 (**K**) across all populations as indicated by the colour legend (bottom right) (***) $p < 0.001$.

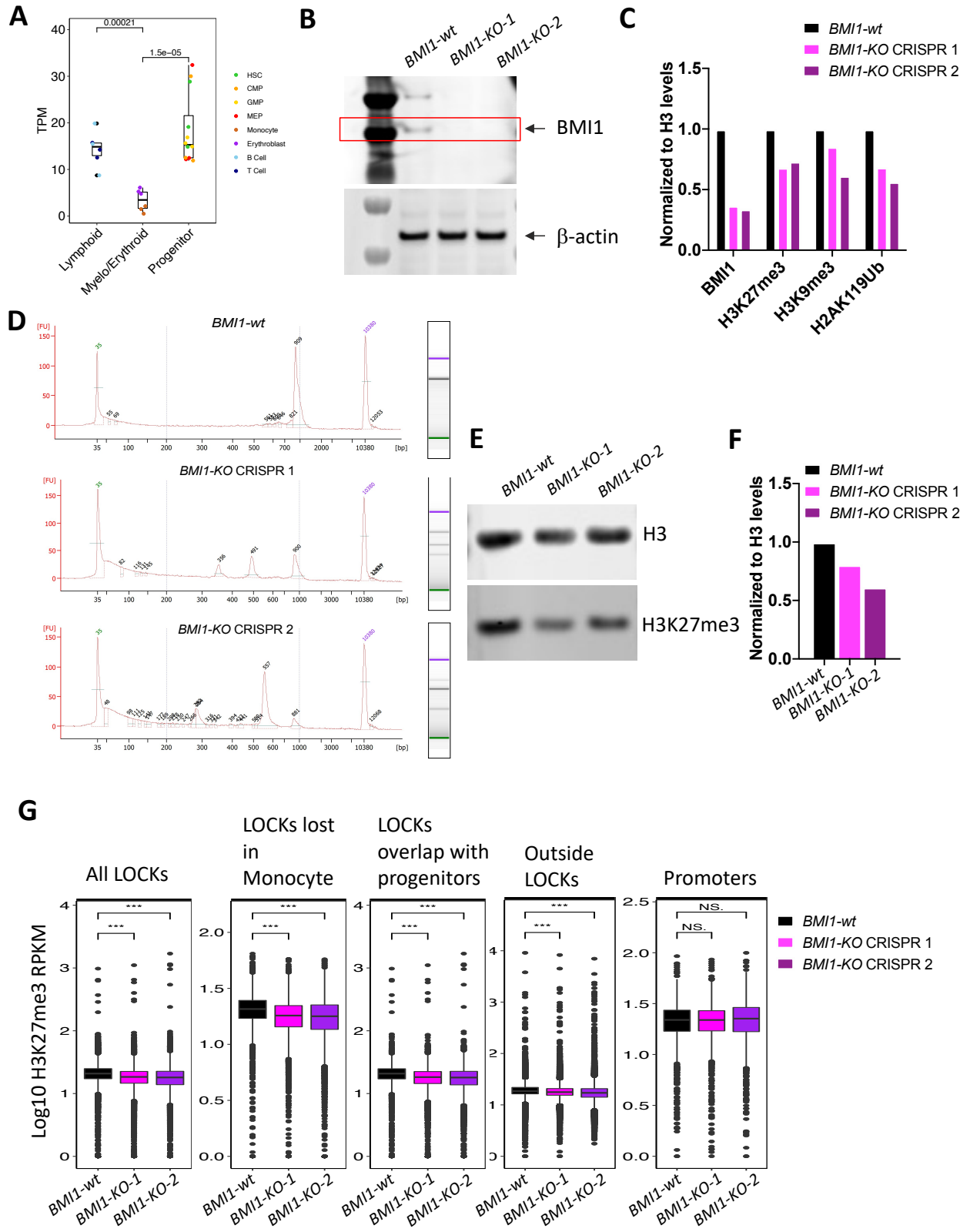


Figure S5. BMI1 loss reduces genome-wide levels of H3K27me3. **A)** BMI1 expression across hematopoietic cells. RNAseq acquired from Blueprint consortium. **B)** Global measurement of BMI1 and β -actin by western blot across *BMI1-wt* and *KO* (N=2). **C)** Quantitative measurement of western blot intensity shown in **Figure 5C** and **S5A** (N=2). **D)** Agilent bioanalyzer profile of PCR amplicon after T7E1 mutation cleavage assay in CRISPR Control (*BMI1-wt*) and *BMI1-KO* replicates. **E)** Global measurement of H3 and H3K27me3 by western blot in *BMI1-wt* and *KO* (N=2). **F)** Quantitative measurement of western blot intensity shown in **S5D** (N=2). **G)** H3K27me3 density at LOCKs identified in HL60 , those overlapped with primary CB progenitor LOCKs, those that are lost in monocyte across *BMI1-wt* and *KO*, at H3K27me3 enriched regions outside the LOCKs and at promoters (\pm 2kb TSS). (* $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$).

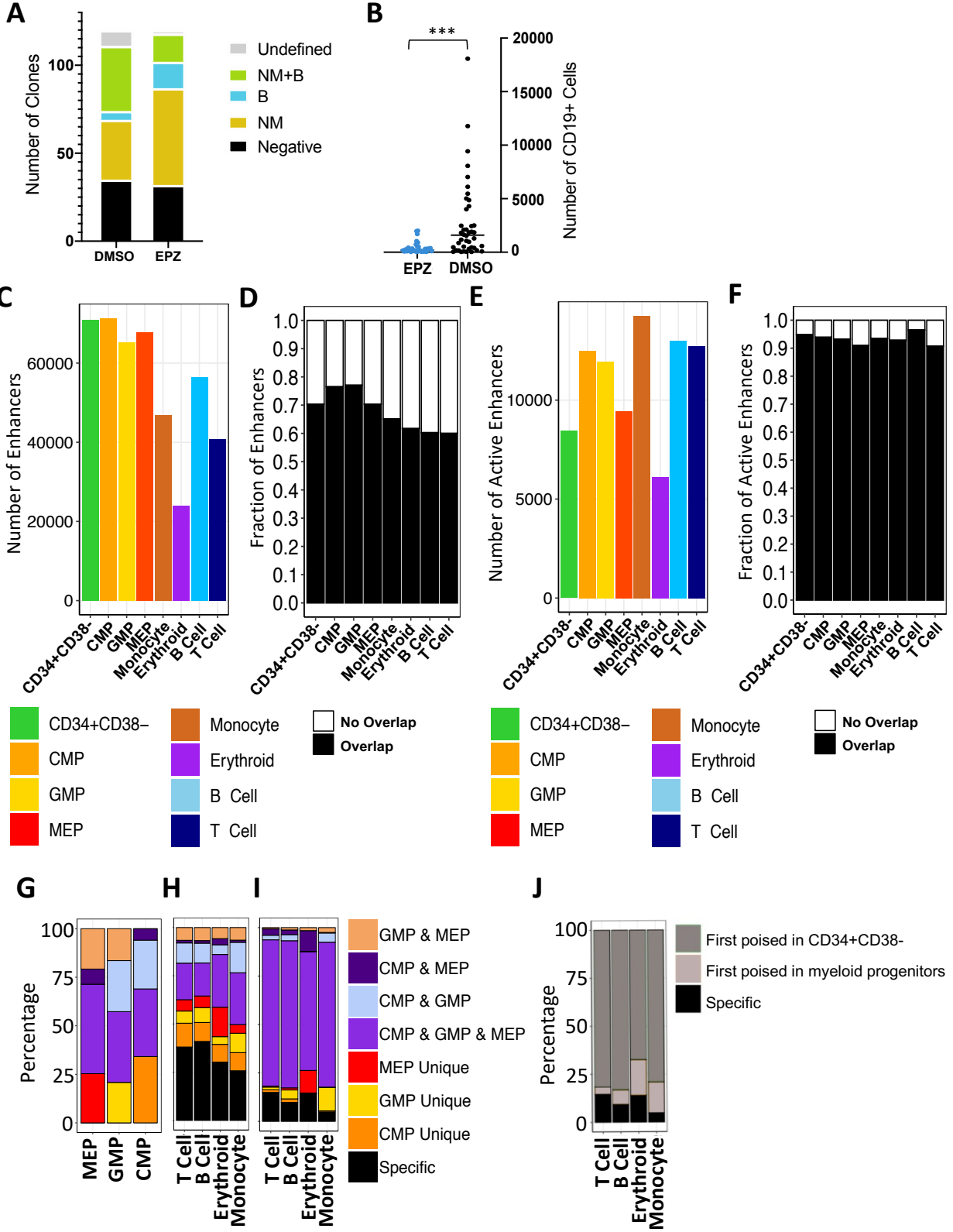


Figure S6. Enhancers dynamics in erythroid precursor, monocyte and differentiated lymphoid cells. **A)** Bar plot showing the number of clones with different contents. M = CD14+ monocytes, N= CD15+ neutrophils, B = CD19+ B cells, negative = <10 CD45+ events, undefined = no detectable mature cells. **B)** Total number of cells expressing CD19+. **C)** Number of enhancers (active and primed enhancers) across cell type. **D)** Fraction of Enhancers (active and primed enhancers) that overlap with open chromatin regions. **E)** Total number of active enhancers across cell types as indicated by the colour legend. **F)** Fraction of active Enhancers that overlap with open chromatin regions. **G)** Percentage of active enhancers that are shared between or unique to each progenitor populations. **H)** Percentage of active enhancers that are present in progenitor populations or *de novo* (specific) in differentiated cells as indicated by the colour legend. **I)** Percentage of active enhancers that are poised in progenitor populations or *de novo* in differentiated cells as indicated by the colour legends. **J)** Percentage of active enhancers present in differentiated cells that are first poised in CD34+CD38- cells, progenitor cells, or appear *de novo* in erythroid precursors, monocytes or lymphoid cells. Cell types are indicated by the colour legend. (* $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$)

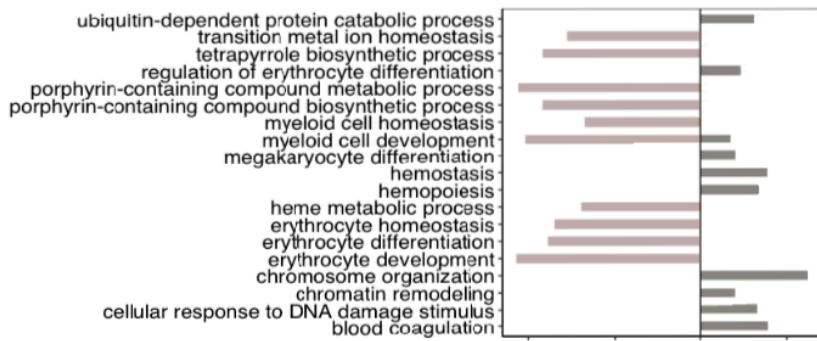
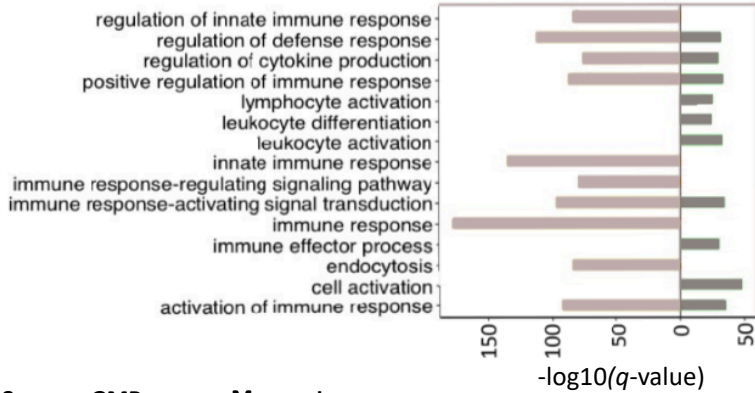
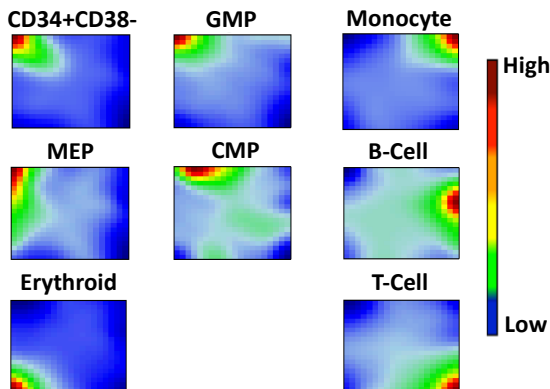
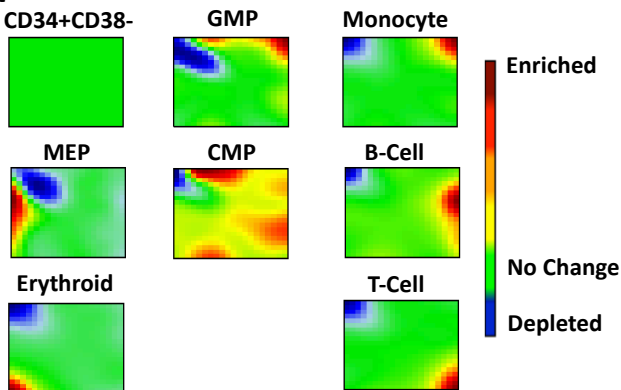
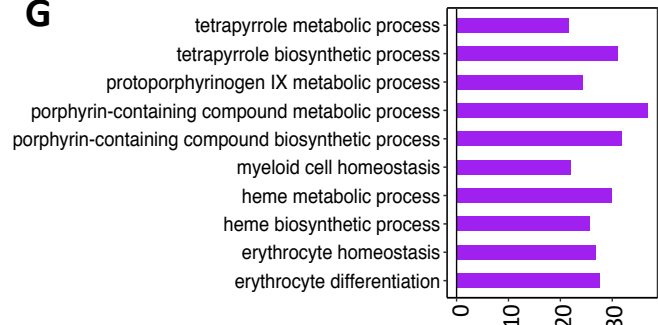
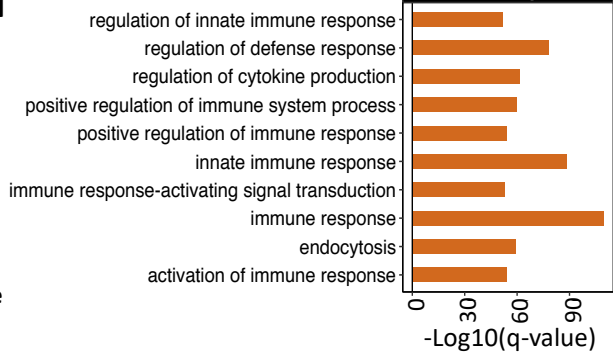
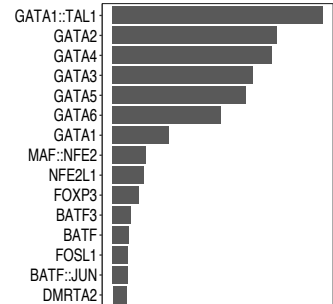
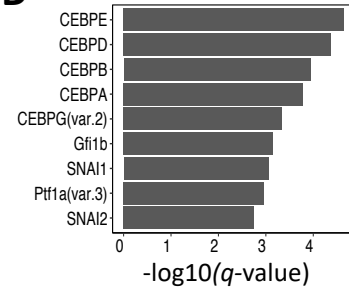
A**B****E****F****G****H****C****D**

Figure S7. Lineage-specific enhancers are marked by H3K27ac in hematopoietic progenitor subsets. GREAT pathway enrichment analysis of active enhancers in erythroid precursor (**A**) and monocytes (**B**) that are first apparent in CD34⁺CD38⁻ or other progenitor subsets. Transcription binding site enrichment at active enhancers in erythroblasts (**C**) and monocytes (**D**) that first appear in their myeloid progenitor populations with respect to those that first appear in CD34⁺CD38⁻. **E**) SOM plot of rank normalized H3K27ac signal contained within the hematopoietic enhancer catalogue across cell types profiled in this study. **F**) SOM plot of rank normalized H3K27ac signal within hematopoietic enhancers with respect to CD34⁺CD38⁻ for each cell type. Gene ontology analysis of super enhancers in erythroid precursor (**G**) and monocyte (**H**).

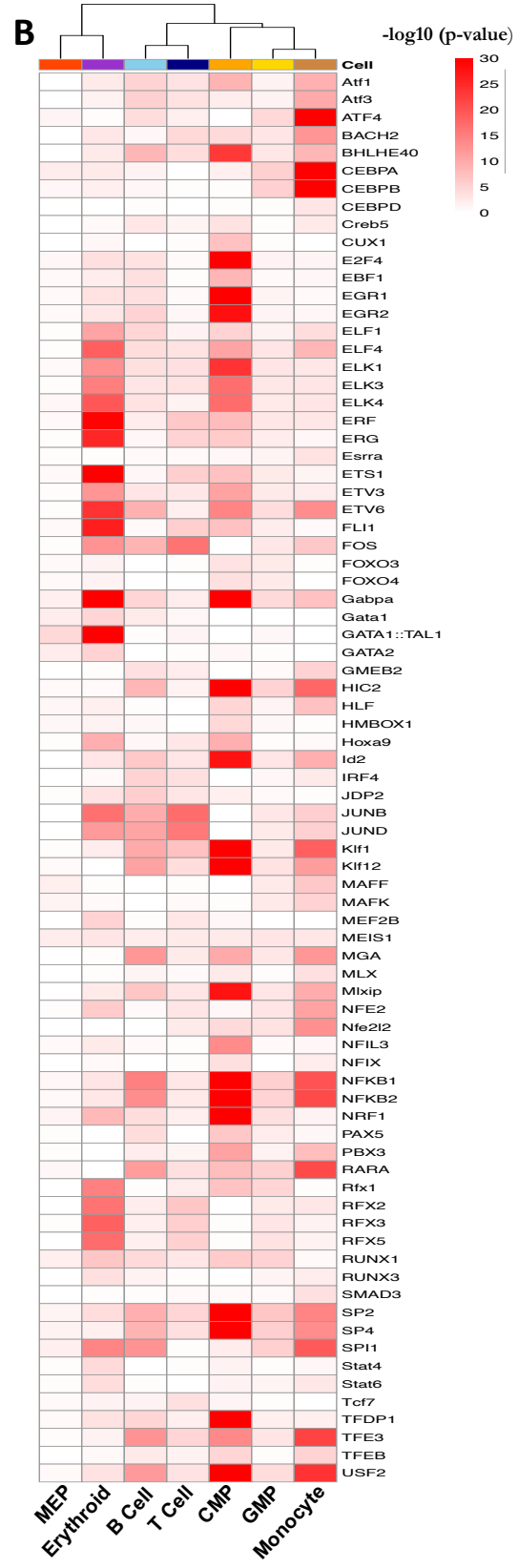
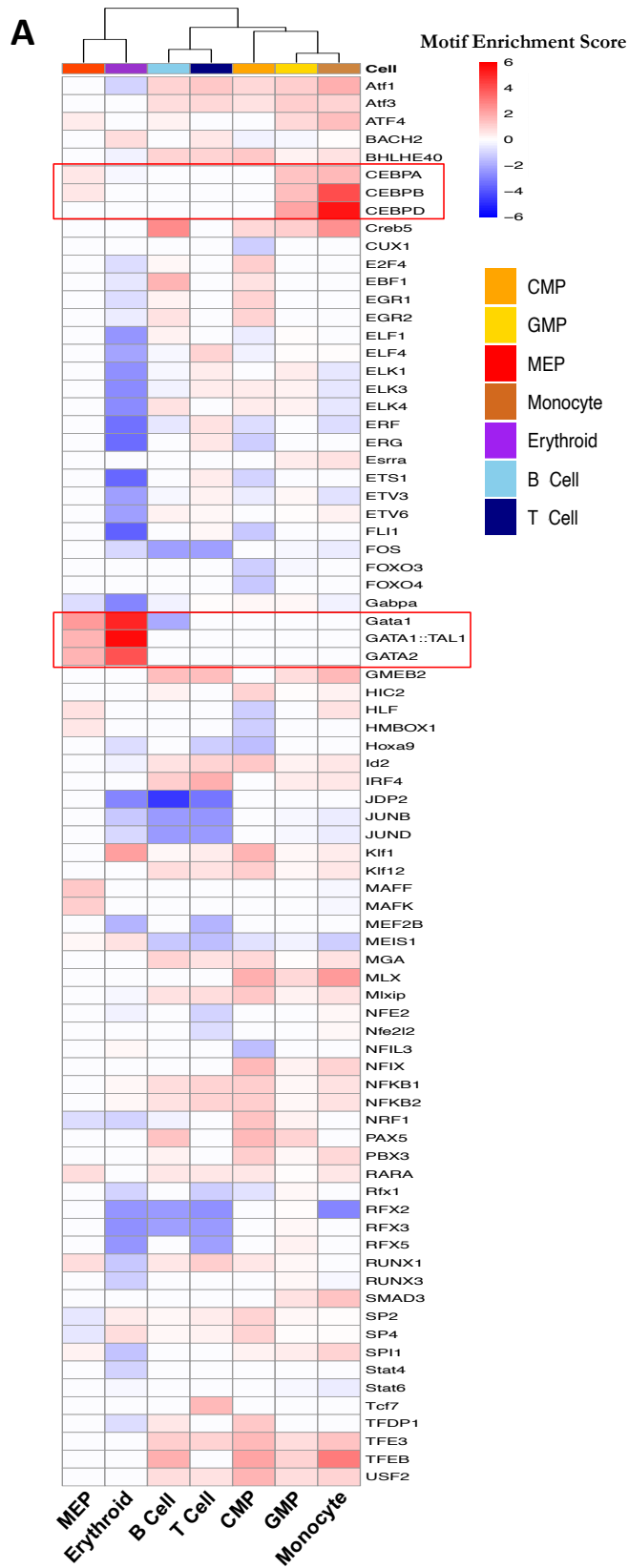


Figure S8. Lineage specific enhancers harbour lineage specific TF binding site are marked with H3K27ac in progenitor population. A) Enrichment of Transcription factor binding site at H3K27ac enriched regions with respect to CD34⁺CD38⁻ compartment. **B)** Significant value for enrichment of TF binding site shown in **(A)**.