Supplementary Information for

3 Yin-Feng Kang^{1#}, Cong Sun^{1#}, Jing Sun^{2#}, Chu Xie¹, Zhen Zhuang², Hui-Qin Xu³, 4 Zheng Liu³, Yi-Hao Liu^{4,5}, Sui Peng⁴, Run-Yu Yuan^{6 ∞}, Jin-Cun Zhao^{2,7 ∞}, Mu-Sheng 5 $Zeng^{1.8}$

 State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Guangdong Key Laboratory of Nasopharyngeal Carcinoma Diagnosis and Therapy, Department of Experimental Research, Sun Yat-sen University Cancer Center (SYSUCC), Sun Yat-sen University, Guangzhou, 510060, P. R. China. ² State Key Laboratory of Respiratory Disease, National Clinical Research Center for

- Respiratory Disease, Guangzhou Institute of Respiratory Health, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, 510182, P. R. China.
- ³ Cryo-electron Microscopy Center, Southern University of Science and Technology,
- Shenzhen, 518000, P. R. China.

⁴ Institute of Precision Medicine, Clinical Trials Unit, The First Affiliated Hospital of

- Sun Yat-sen University, Guangzhou, 510080, P. R. China.
- ⁵ Department of Endocrinology, The First Affiliated Hospital of Sun Yat-sen University, Guangzhou, 510080, P. R. China.
- ⁶ Guangdong Provincial Institution of Public Health, Guangdong Provincial Center
- for Disease Control and Prevention, Guangzhou, 511430, P. R. China.
- ⁷ Guangzhou Laboratory, Bio-island, Guangzhou, 510320, P. R. China.
- ⁸ Guangdong-Hong Kong Joint Laboratory for RNA Medicine, Guangzhou, 510120, P.
- R. China.
- 26 # These authors contributed equally: Yin-Feng Kang, Cong Sun, Jing Sun.
- 27 Email: cecilia yry@hotmail.com; zhaojincun@gird.cn; zengmsh@sysucc.org.cn
-
-

Supplementary Figures

 Supplementary Fig. 1. Binding curves of SARS-CoV-2 Spike-specific neutralization antibodies against HexaPro-based nanoparticle immunogens measured by ELISA. The data are presented as means ± SD in duplicate from three independent experiments SARS-CoV spike-specific CR3022 antibody and EBV gH/gL-specific AMMO1 antibody were used as controls. Source data are provided as

a Source Data file.

Supplementary Fig. 2. Thermostability analysis ofHexaPro-based immunogens.

 a. Thermostability parameter table of SARS-CoV-2 HexaPro or Hexapro-based nanoparticles determined by DSF from three replicate experiments. Tm: melting temperature; Tagg 266: aggregation temperature identified by the static light scattering at 266nm.

 b. Barycentric mean (BCM) of the intrinsic protein fluorescence from 300-430nm and static light scattering intensity at 266nm of the HexaPro-based immunogens. The curves presented the lined mean data of each temperature point from three replicate experiments. Source data are provided as a Source Data file.

 Supplementary Fig. 3. Cross-neutralization of bat and human coronaviruses by sera elicited by HexaPro-based nanoparticle immunogens. Sera were collected from two weeks after the second booster dose (n=4 cynomolgus macaques in each group) and used to measure the cross-neutralization antibody titers using the pseudoviruses assay. The data were expressed as means ±SD. Comparison between the two groups were performed using a two-tailed Mann-Whitney U test. Mosaic NP 59 vs WT NP in SARS-CoV pseudovirus neutralization titers ${}^{*}p = 0.0286$, Mosaic NP vs 60 WT NP in HCoV-NL63 pseudovirus neutralization titers *p = 0.0286. *p < 0.05; ns, no significant.Source data are provided as a Source Data file.

65 **Supplementary Fig. 4. Expression of immune-related cytokines and chemokines** 66 **and viral burden in infected lung tissues at 2 days post B.1.351 variant strain** 67 **infection.** a and b. The expression levels of viral burden (a) and cytokines and 68 chemokines (b) in the lungs at 2 days post infection was measured by $qRT-PCR$. The 69 data were expressed as means \pm SD. Comparisons between the two groups were 70 performed using a Kruskal-Wallis ANOVA with Dunn's correction. $\frac{1}{2}p < 0.05$, $\frac{1}{2}p <$ 71 0.01; ns, no significant. (a) Mosaic NP vs PBS in *ORF1ab* and *N* transcript copies, 72 **p = 0.0082. (b) WT NP and Cocktail NP vs PBS in *CCL2* fold change, *p = 0.0175, 73 *p = 0.0210, respectively. Cocktail NP and Mosaic NP vs PBS in *IL6* fold change, *p $74 = 0.0250$, **p = 0.0055, respectively. WT NP and Cocktail NP vs PBS in *IFIT1* fold 75 change, ${}^*p = 0.0354$, ${}^*p = 0.0298$, respectively. WT NP and Mosaic NP vs PBS in 76 *MX2* fold change, ${}^*p = 0.0175$, ${}^*p = 0.0145$, respectively. WT NP and Cocktail NP vs 77 PBS in*CXCL10* fold change, *p = 0.0250. WT HexaPro and WT NP vs PBS in*IL10* 78 fold change, ${}^*p = 0.0298$, ${}^*p = 0.0419$, respectively. Cocktail NP and Mosaic NP vs 79 PBS in *IFIT3* fold change, ${}^*p = 0.0419$, ${}^{**}p = 0.0067$, respectively. Mosaic NP vs 80 PBS in*ISG15* fold change, **p = 0.0082. Source data are provided as a Source Data

file.

 Supplementary Fig. 5. Protective efficacy of HexaPro-bearing immunogens in mice following challenge with authentic ancestral SARS-CoV-2 virus *in vivo***.** a-c, six-week-old male BALB/c mice were subcutaneously immunized with an equivalent amounts of HexaPro-based immunogens (equal to 5 µg HexaPro) at weeks 0 and 3. At 3 weeks after the second vaccination, the mice were transduced intranasally with 2.5×10^8 FFU of Ad5-hACE2, and after 5 days of transduction, the mice were 92 intranasally inoculated with 2×10^6 PFU/ml of authentic ancestral SARS-CoV-2 virus.

 Lung tissues were collected for virus titer quantification and clinicopathological analysis.

 a. Body weight change of mice (n=3 in PBS-treated and WT NP-vaccinated mice, n=4 mice in WT HexaPro, Cocktail NP and Mosaic NP-vaccinated group) after infection with ancestral SARS-CoV-2. The body weight of each mouse was recorded daily for 8 days.

- 99 b. Virus titers of lung tissues from mice challenged with ancestral SARS-CoV-2 ($n=4$) 100 mice in each group) at 2 days post challenge. LOD, limit of detection. Statistical significance was determined by two-tailed unpaired t test. WT HexaPro, WT NP, 102 Cocktail NP and Mosaic NP vs PBS, $***p = 0.0001$.
- c. Immunohistological analysis of lung tissues from mice challenged with ancestral SARS-CoV-2 at 4 days post challenge (n=2 mice in each experimental group). Hematoxylin and eosin staining (HE, left) and immunohistochemistry (IHC, right) microscopic images are shown in the figure at magnification. Scale bar for H&E, 250 μm (left); 50 μm (middle); 25 μm (right), Scale bar for IHC, 50 μm. In a and b, the 108 data were expressed and plotted as means \pm SD. Source data are provided as a Source Data file.

 Supplementary Fig. 6. Expression of immune-related cytokines and chemokines and viral burden in infected lung tissues at 2 days post infection with the ancestral SARS-CoV-2 strain. a and b. The expression levels of viral burden (a) and cytokines and chemokines (b) in lungs at 2 days post infection were measured by 117 qRT-PCR assay. The data were expressed as means \pm SD. Comparison between the two groups was performed using a Kruskal-Wallis ANOVA with Dunn's correction. $*_{p} < 0.05$; $**_{p} < 0.01$; ns, no significant. (a) Cocktail NP and Mosaic NP vs PBS in *ORF1ab* transcript copies, ${}^*p = 0.0419$ and ${}^*p = 0.0175$, respectively, Mosaic NP vs 121 PBS in *N* transcript copies, $*p = 0.0120$. (b) Cocktail NP vs PBS in *IL6* fold change, $*_{p} = 0.0298$. WT NP vs PBS in *IFIT1* fold change, $*_{p} = 0.0354$. Cocktail NP vs PBS 123 in $MX2$ fold change, *p = 0.0495. WT NP vs PBS in *CXCL10* fold change, **p = 124 0.0098. WT HexaPro vs PBS in $IL10$ fold change, $**p = 0.0210$. WT NP and Cocktail 125 NP vs PBS in *IFIT3* fold change, ${}^*p = 0.0250$, ${}^*p = 0.0145$, respectively. WT NP and Cocktail NP vs PBS in*ISG15* fold change, *p = 0.0495. Source data are provided as a Source Data file.

129 **Supplementary Tables**

- 130 **Supplementary Table 1: Kinetic analysis of SARS-CoV-2 mAbs and ACE2**
- 131 **receptor to HexaPro and HexaPro-based nanoparticle immunogens of**
- 132 **SARS-CoV-2 prototype and variants.**

133

134 For antibody or receptor binding, SARS-CoV-2 RBD-directed mAbs (REGN10933 and P2B-2F6),

135 NTD-directed mAb (4A8) and ACE2 receptor was immobilized onto the biosensors, and then 136 twofold diluted HexaPro or HexaPro-based nanoparticle immunogens injected into the wells. 137 Binding kinetics were analyzed with a 1:1 model in Octet Analysis Studio. Affinity constant 138 (K_D), kinetic constants of associations (K_{on}) and dissociations (K_{off}) were summarized in this table. 139 K_D is calculated from K_{off}/K_{on} .

140

141

143 **Supplementary Table 2: Quantitative RT-PCR primer used in this study.**

144