

Supplementary Information

MYC drives aggressive prostate cancer by disrupting transcriptional pause release at androgen receptor targets

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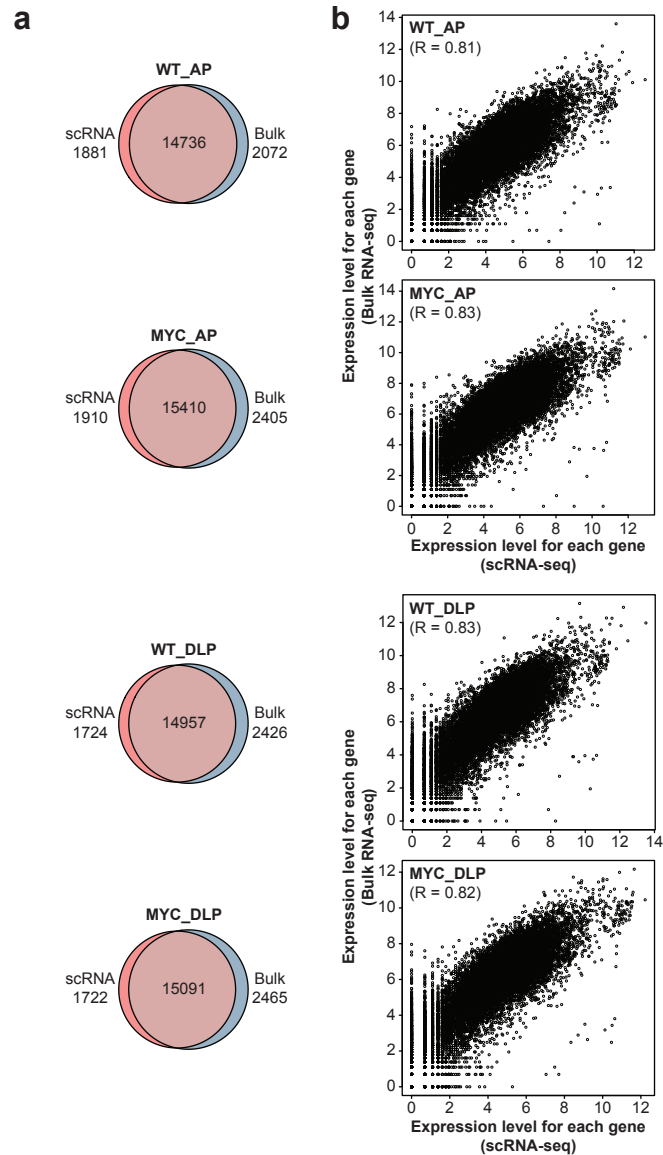
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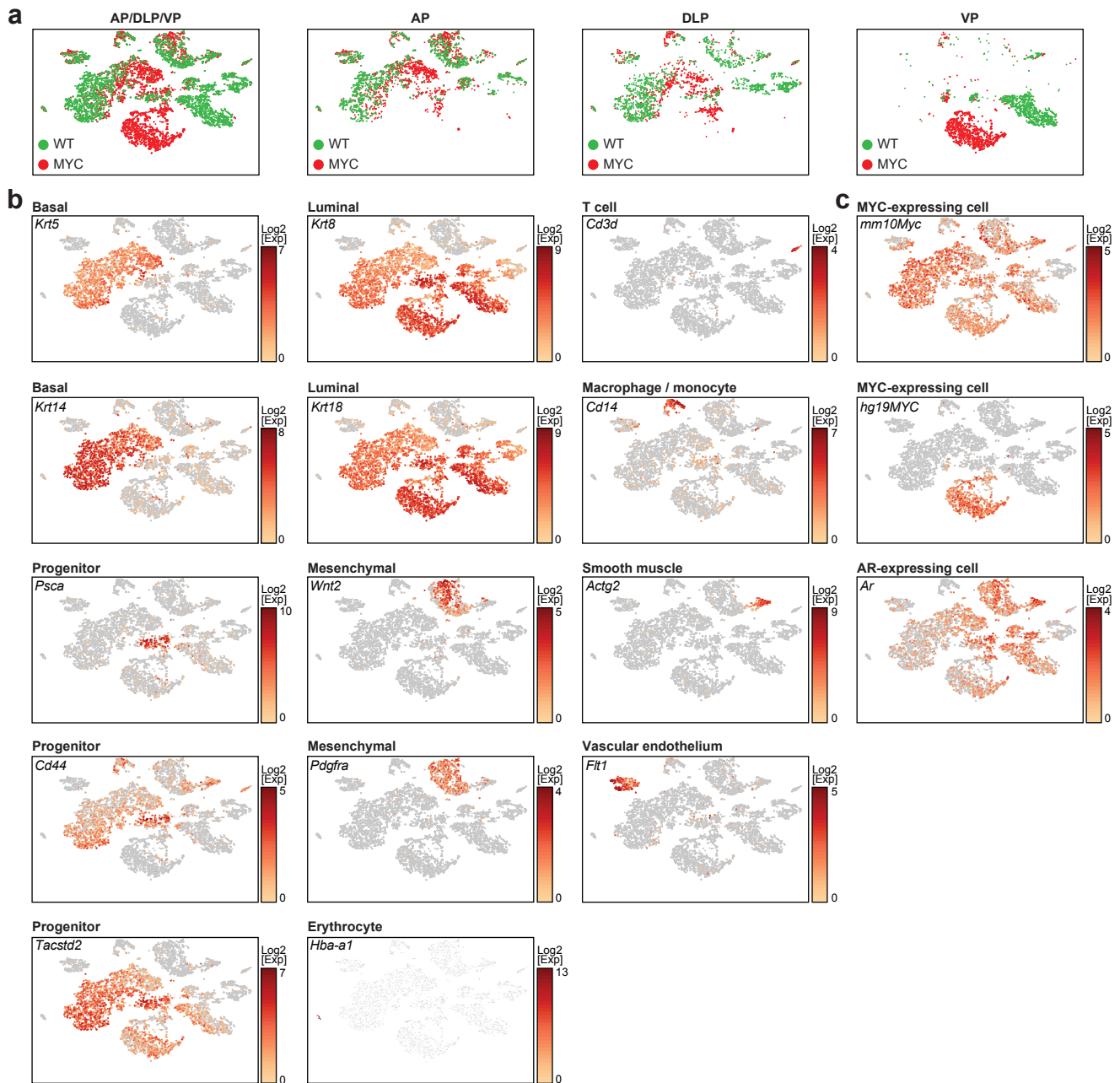
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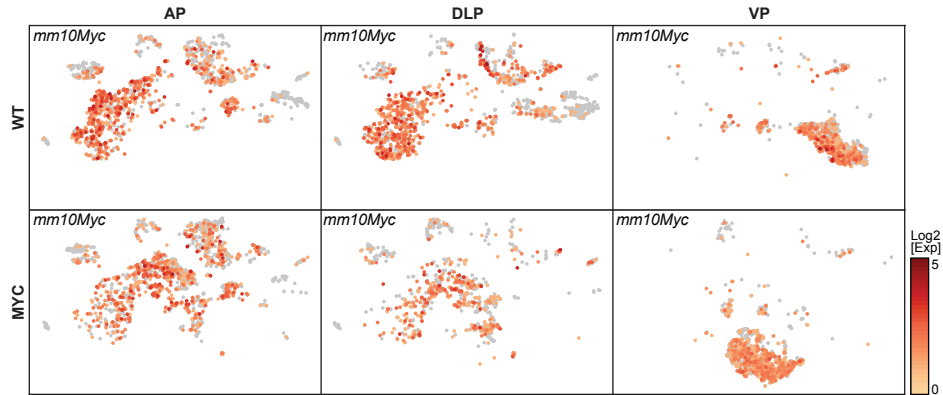
*E-Mail: henry_long@dfci.harvard.edu (H.W.L.), david.labbe@mcgill.ca (D.P.L.)



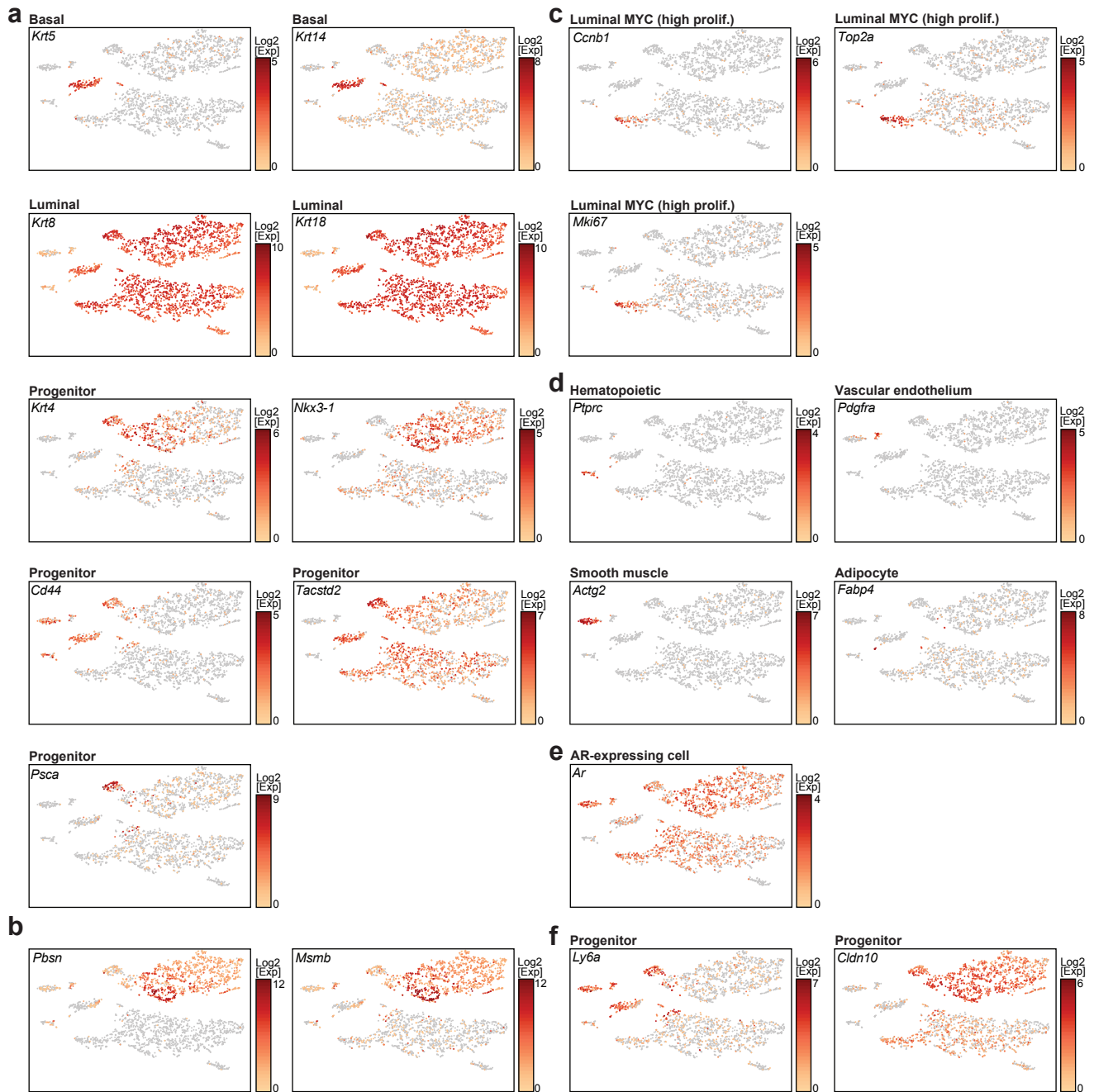
Supplementary Figure 1: Single-cell transcriptome is highly correlated with bulk gene expression in AP and DLP lobes. (a, b) Transcriptional profiling of WT and MYC-transformed AP and DLP lobes reveals high concordance for the total number of genes detected **(a)** and their expression levels **(b)** between bulk and single-cell RNA-seq (AP, DLP; matched bulk and single-cell RNA-seq; $n = 1$ per genotype). WT: wild-type; AP: anterior prostate; DLP: dorsolateral prostate.



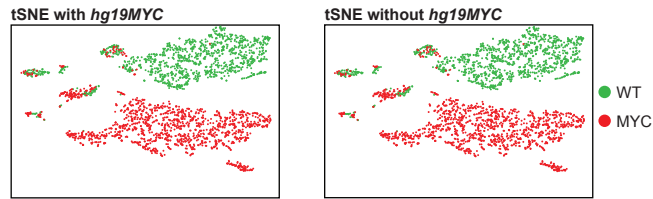
Supplementary Figure 2: Molecular characterization of murine WT and MYC-transformed prostate lobes. (a) Single-cell census of WT and MYC-transformed AP, DLP and VP. tSNE of scRNA-seq profiles (as in **Figure 2a**), colored by genotype (AP, DLP, VP; $n = 1$ per genotype). (b, c) Expression of selected markers of different subsets (b; AP, DLP, VP; $n = 1$ per genotype) as well as murine *Myc* (*mm10Myc*), human *MYC* (*hg19MYC*) and the *Ar* (c; AP, DLP, VP; $n = 1$ per genotype). WT: wild-type; VP: ventral prostate; DLP: dorsolateral prostate; AP: anterior prostate.



Supplementary Figure 3: Murine Myc is expressed across cell subpopulations and prostate lobes. Expression of murine *Myc* (*mm10Myc*) in WT and MYC-transformed AP, DLP and VP (AP, DLP, VP; $n = 1$ per genotype). WT: wild-type; AP: anterior prostate; DLP: dorsolateral prostate; VP: ventral prostate.




Supplementary Figure 4: Molecular characterization of murine WT and MYC-transformed VP. (a-f) Expression of selected markers of different subsets (VP; $n = 1$ per genotype). WT: wild-type; VP: ventral prostate.

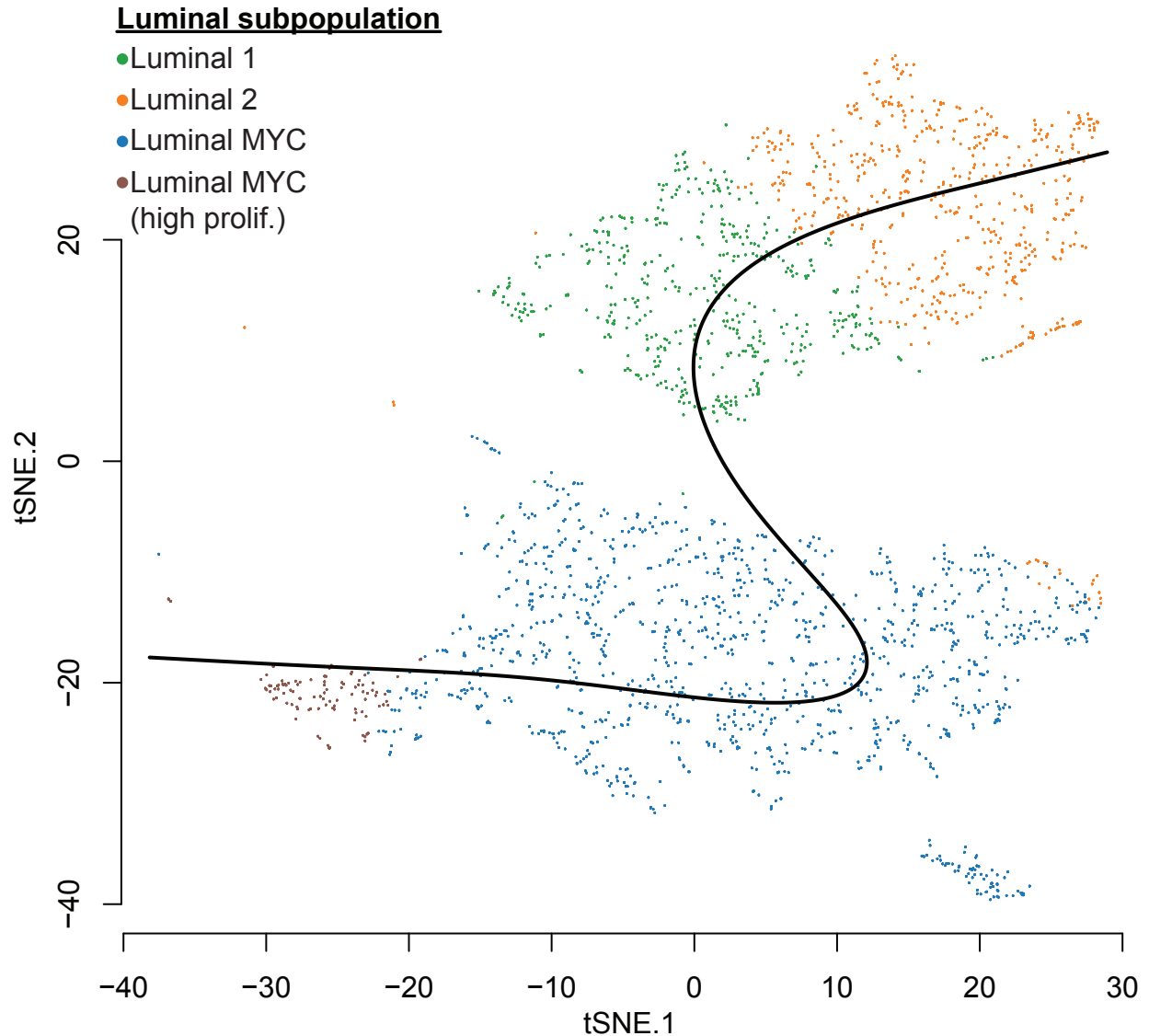


Supplementary Figure 5: tSNE of scRNA-seq profiles is not affected by the inclusion of human *MYC* transcript. tSNE of VP generated with (*left*) or without (*right*) the inclusion of human *MYC* (*hg19MYC*; VP; $n = 1$ per genotype). WT: wild-type; VP: ventral prostate.

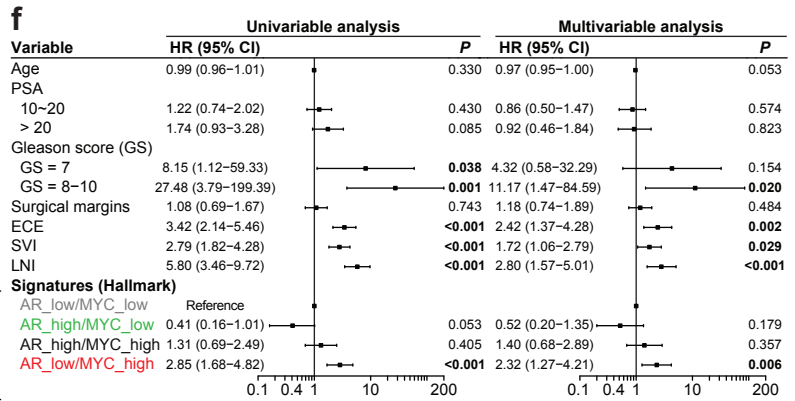
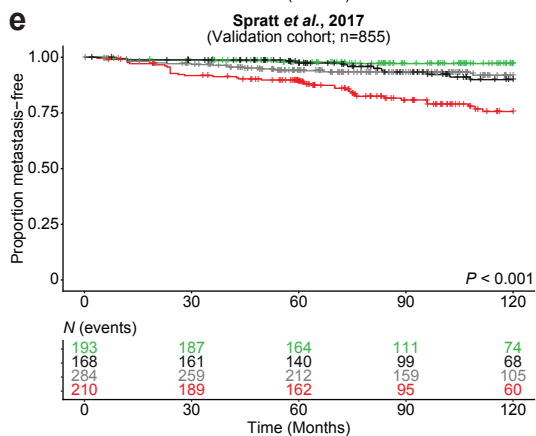
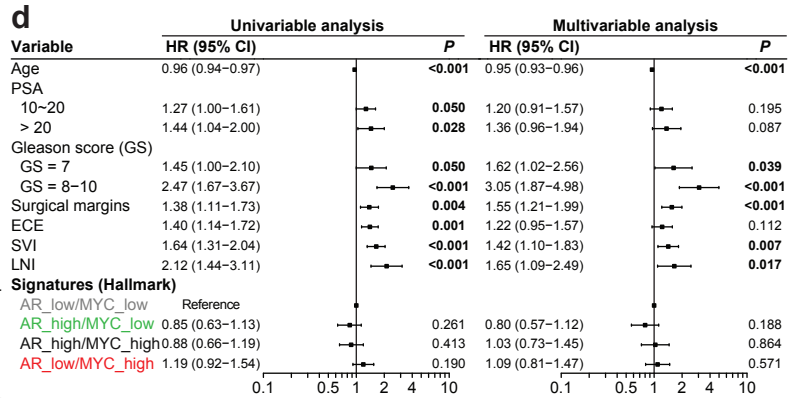
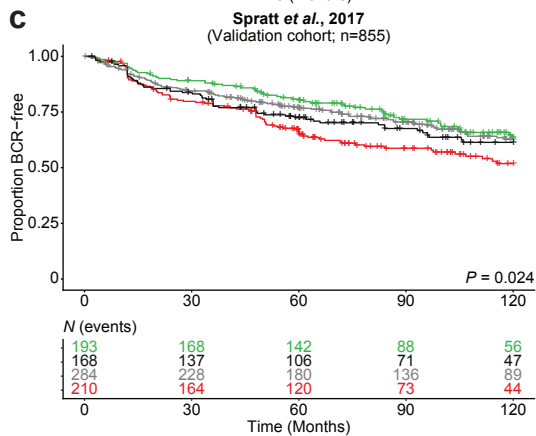
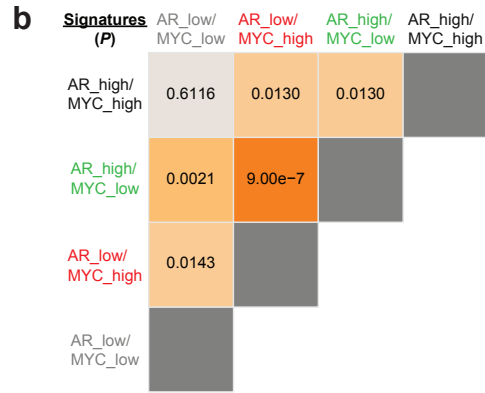
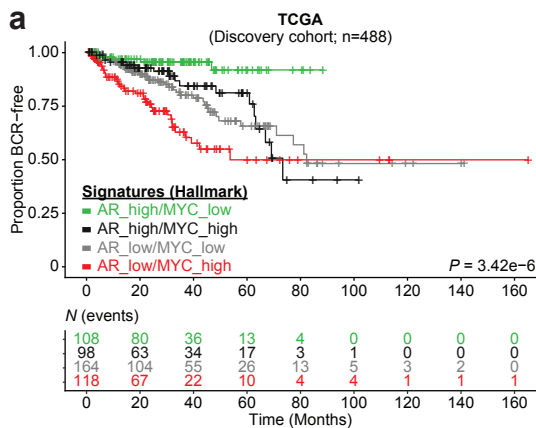
a

FOXA1 ChIP-seq binding motif
FOXA1(Forkhead)/LNCAP-FOXA1-ChIP-seq(GSE27824)/Homer

WT_1: $P = 1e-690$
WT_2: $P = 1e-738$
MYC_1: $P = 1e-769$
MYC_2: $P = 1e-823$

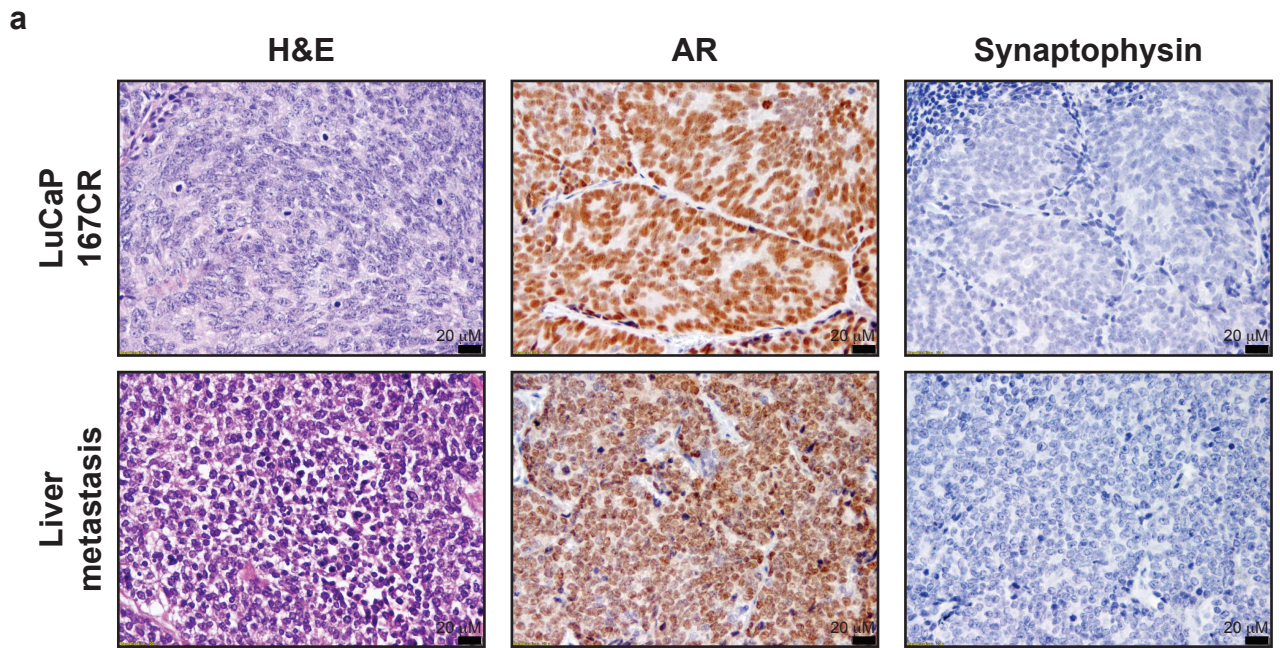
b



Supplementary Figure 6: Integration of ChIP-seq with scRNA-seq. (a) FOXA1 ChIP-seq identifies FHRE as the top FOXA1 binding motif in WT and MYC-transformed VP (VP; $n = 2$ pools of biological replicates ($n = 8-13$) per genotype). (b) Slingshot pseudotime inference used to order luminal cells in **Figure 4g** (VP; $n = 1$ per genotype). WT: wild-type; VP: ventral prostate; FHRE: forkhead response element.



Supplementary Figure 7: Divergent MYC and AR transcriptional programs dictate disease progression. (a, b) Kaplan-Meier curves (a) and log-rank tests (b) reveal that patients bearing a primary tumor characterized by low AR transcriptional signature (Hallmark) and concurrent high MYC transcriptional signature (Hallmark) have a shorter time to biochemical recurrence (BCR) within the discovery cohort (TCGA). (c, d) Kaplan-Meier curves (c) but not univariable and multivariable analysis (d; Cox proportional hazards model) confirms that tumors with concurrent low AR and high MYC transcriptional signatures have a significant more rapid development of BCR than tumors with low AR transcriptional signature without an active MYC transcriptional program in the validation cohort (Spratt *et al.*, 2017¹; $n = 855$; HR \pm 95% CI). (e, f) Kaplan-Meier curves (e), univariable and multivariable analyses (f; Cox proportional hazards model) reveal that tumors with concurrent low AR and high MYC transcriptional signatures are more likely to develop metastatic disease ($n = 855$; HR \pm 95% CI). PSA: prostate-specific antigen; HR: hazard ratio; CI: confidence interval; GS: Gleason score; ECE: extracapsular extension; SVI: seminal vesicles invasion; LNI: lymph node involvement.



b AR ChIP-seq binding motif
 ARE(NR)/LNCAP-AR-ChIP-seq(GSE27824)/Homer

AGAACAATGTTCTT

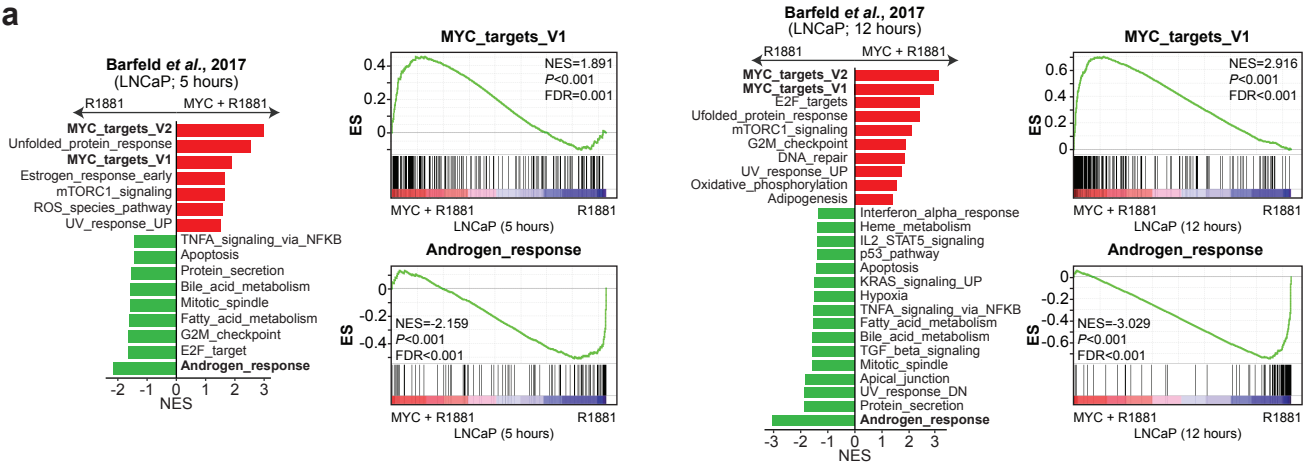
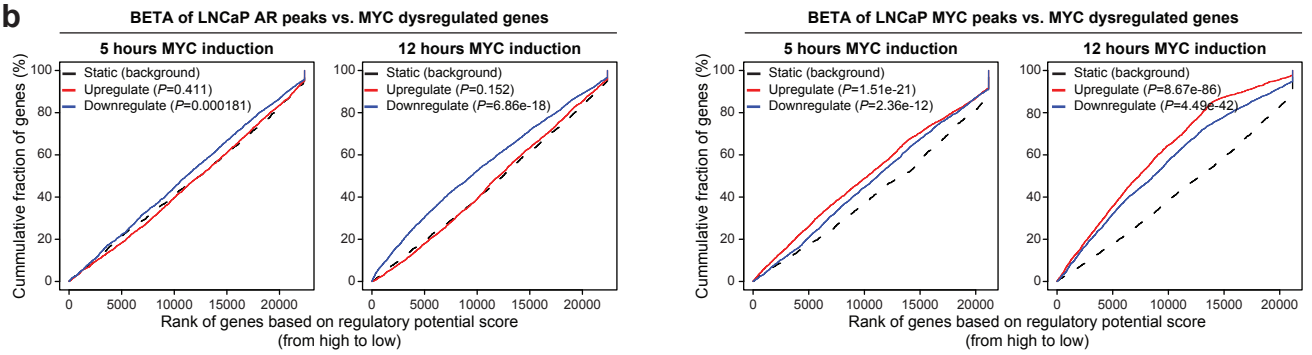
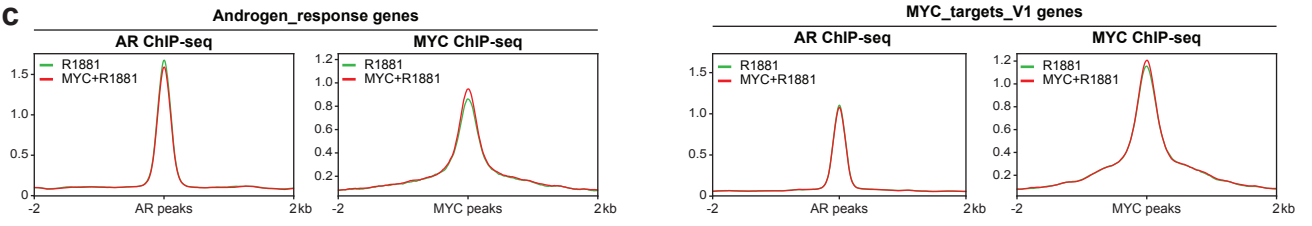
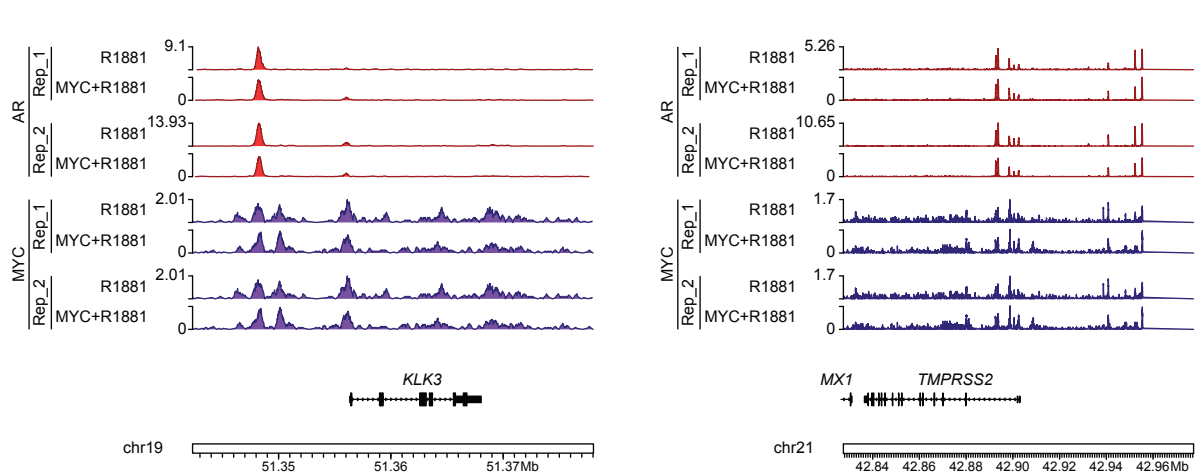
MYC-low		MYC-high	
PDX	P	PDX	P
105CR	1e-673	167CR	1e-1322
136CR	1e-628	70CR	1e-706
147CR	1e-318	78CR	1e-666
96CR	1e-327	81CR	1e-641

c FOXA1 ChIP-seq binding motif
 FOXA1(Forkhead)/LNCAP-FOXA1-ChIP-seq
 (GSE27824)/Homer

AAAGTAAACA

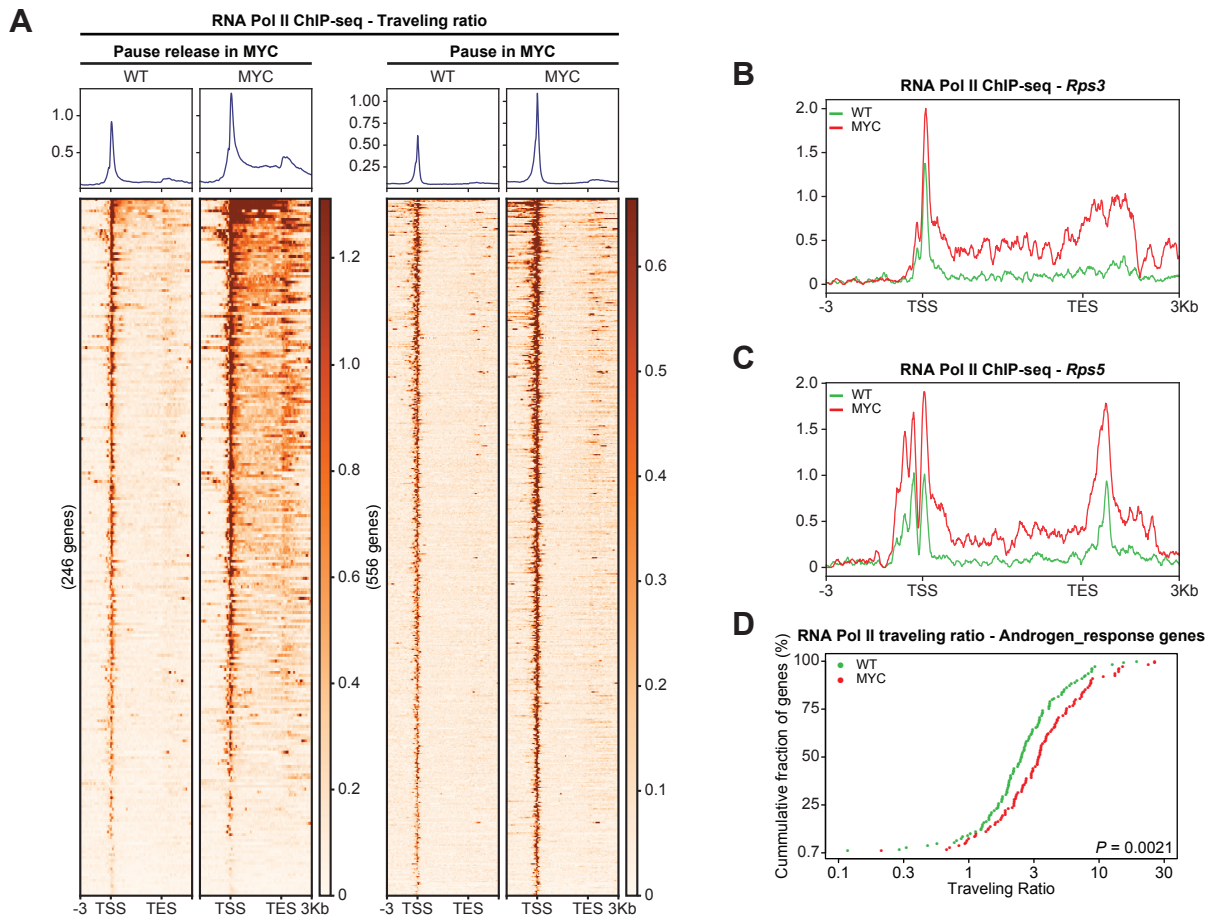
MYC-low		MYC-high	
PDX	P	PDX	P
105CR	1e-729	167CR	1e-693
136CR	1e-801	70CR	1e-869
147CR	1e-916	78CR	1e-558
96CR	1e-764	81CR	1e-658

Supplementary Figure 8: mCRPC LuCaP patient-derived xenograft (PDX) 167CR characterization and top binding motifs in LuCaP PDXs AR and FOXA1 ChIP-seq. (a) LuCaP PDX 167CR was established from a liver metastasis of a male who died of abiraterone-, carboplatin- and docetaxel-resistant CRPC. LuCaP 167CR expresses AR, responds to castration and is negative for synaptophysin. Morphology of the PDX was concordant with the original liver metastasis (b, c) AR and FOXA1 ChIP-seq identifies ARE (b) and FHRE (c), respectively as the top binding motif in mCRPC LuCaP PDXs. ARE: androgen response element; FHRE: forkhead response element.

a**b****c****d**

Supplementary Figure 9

Supplementary Figure 9: MYC overexpression disrupts the AR transcriptional program in LNCaP cells. Reanalysis of transcriptomics and epigenetics data from Barfeld and colleagues ². **(a)** Gene Set Enrichment Analysis (GSEA, Hallmark, $P < 0.05$ and $FDR < 0.1$) revealed an enriched MYC transcriptional program and a depleted AR response following 5 or 12 hours of MYC induction ($P < 0.001$ and $FDR < 0.001$; Source data are provided as a Source Data file). **(b)** BETA analysis revealed that AR binding sites are associated with gene downregulation while MYC binding sites are associated with gene upregulation following MYC induction. **(c)** AR and MYC binding nearby *Androgen_response* and *MYC_targets_V1* genes is unchanged following MYC induction despite a dampened AR and a heightened MYC transcriptional program. **(d)** Unchanged AR and MYC binding at *KLK3* and *TMPRSS2* loci, AR-dependent genes downregulated by MYC overexpression. NES: normalized enrichment score; ES: enrichment score.



Supplementary Figure 10: RNA Pol II promoter-proximal pausing. (a) RNA Pol II occupancy at pause release (*left*) and pause genes (*right*) following MYC overexpression (VP; $n = 2$ pools of biological replicates ($n = 8-13$) per genotype). (b, c) Pause release at *Rps3* (b) and *Rps6* (c) MYC_targets_V1 genes (VP; $n = 2$ pools of biological replicates ($n = 8-13$) per genotype). (d) RNA Pol II traveling ratio reveals greater promoter-proximal pausing at Androgen_response genes (non-smoothed curves. TSS: transcription start site; TES: transcription end site).

SUPPLEMENTARY REFERENCES

- 1 Spratt, D. E. *et al.* Individual Patient-Level Meta-Analysis of the Performance of the Decipher Genomic Classifier in High-Risk Men After Prostatectomy to Predict Development of Metastatic Disease. *J Clin Oncol* **35**, 1991-1998, doi:10.1200/JCO.2016.70.2811 (2017).
- 2 Barfeld, S. J. *et al.* c-Myc Antagonises the Transcriptional Activity of the Androgen Receptor in Prostate Cancer Affecting Key Gene Networks. *EBioMedicine* **18**, 83-93, doi:10.1016/j.ebiom.2017.04.006 (2017).