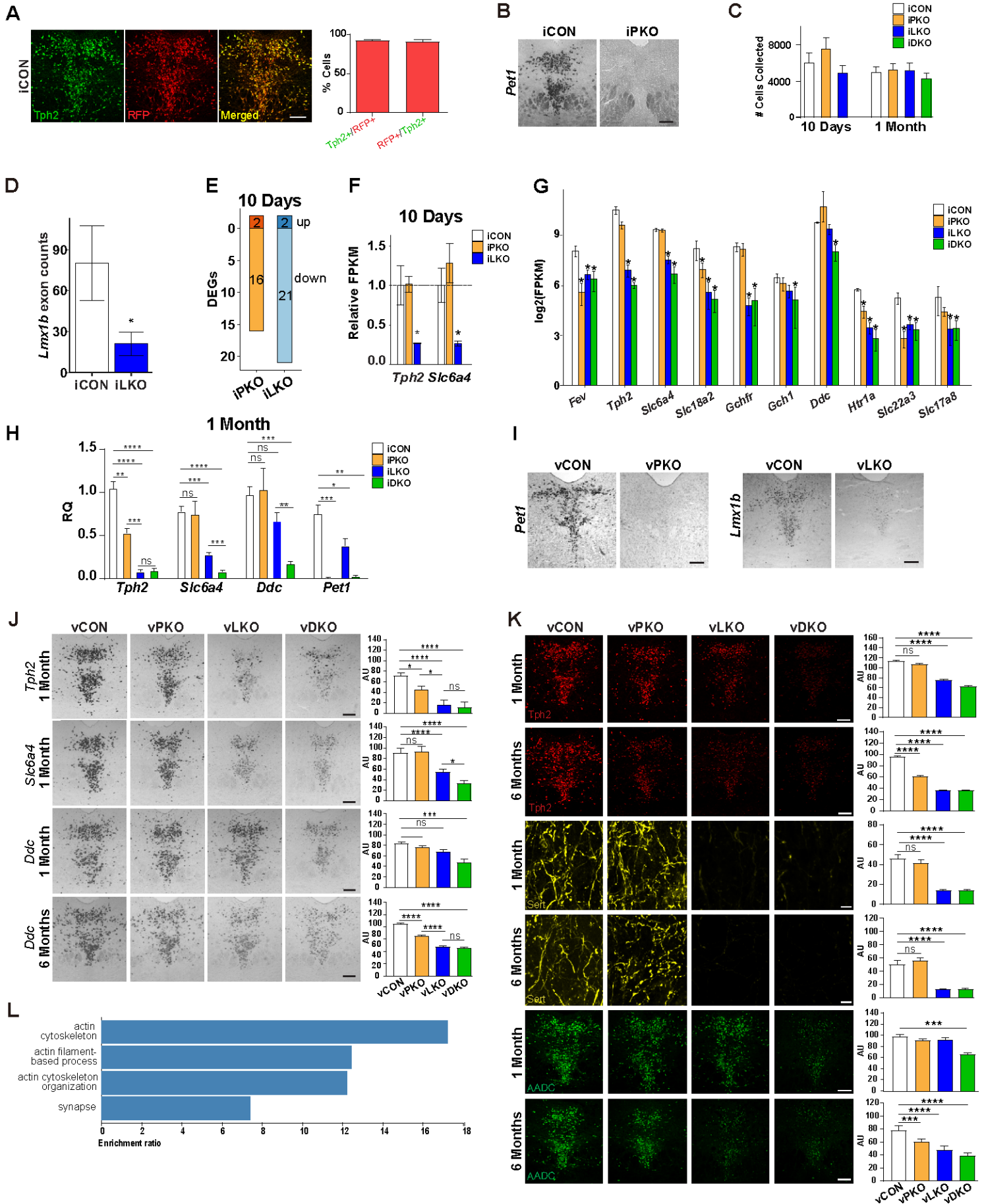


Cell Reports, Volume 39

Supplemental information

**An adult-stage transcriptional program
for survival of serotonergic connectivity**

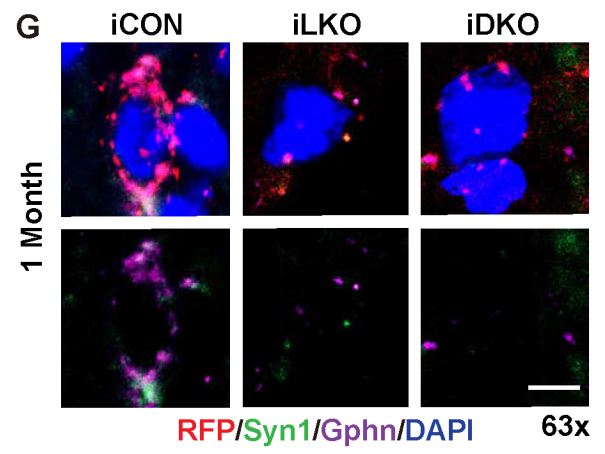
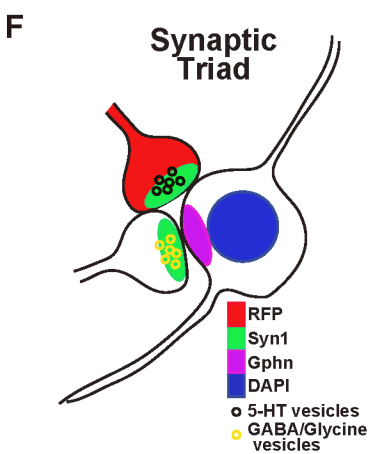
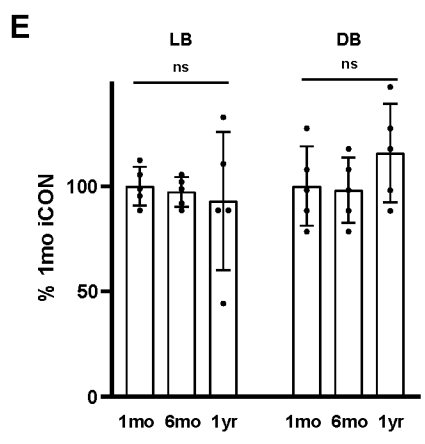
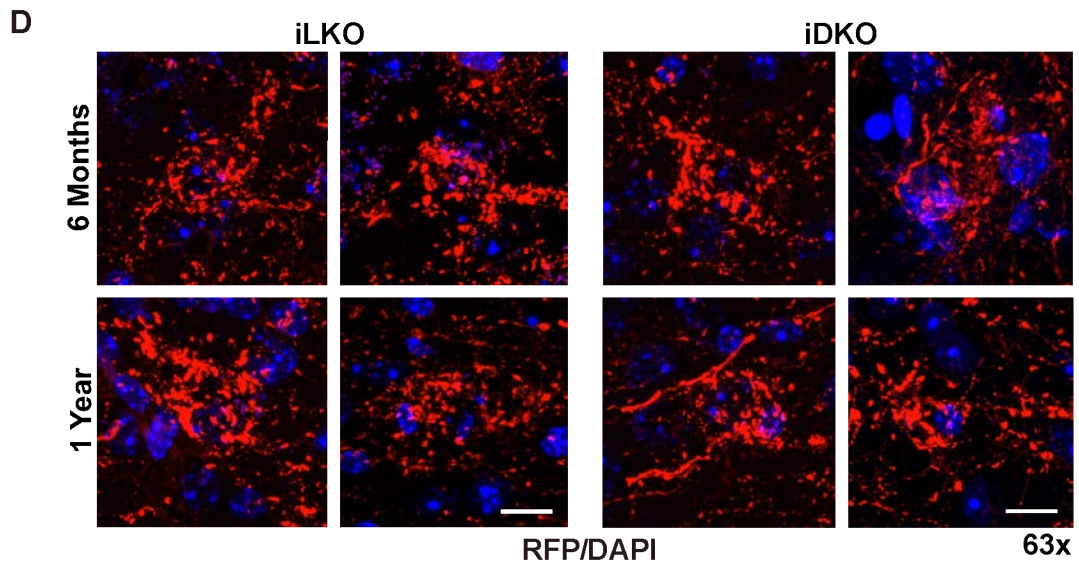
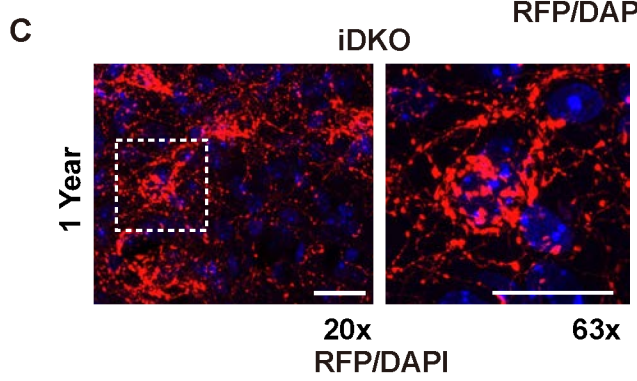
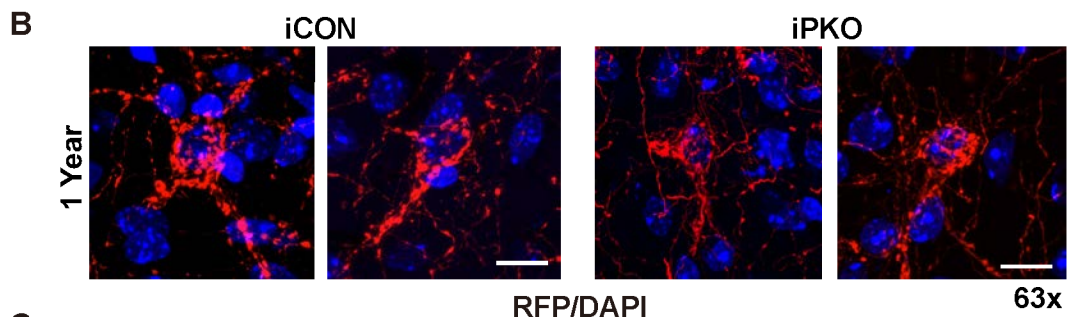
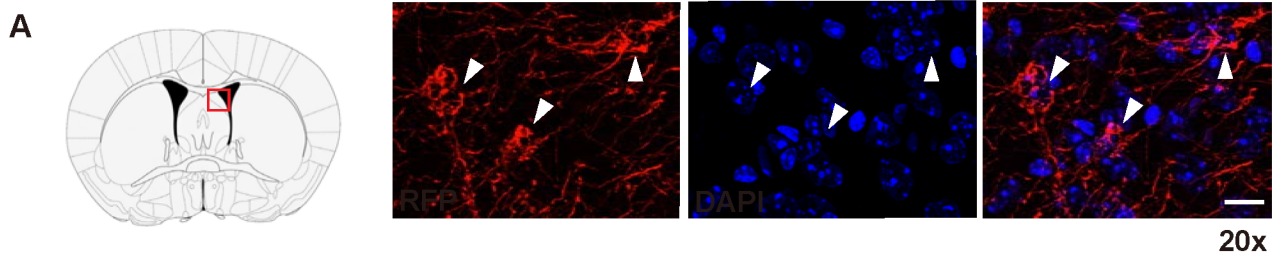
Meagan M. Kitt, Nobuko Tabuchi, W. Clay Spencer, Heath L. Robinson, Xinrui L. Zhang, Brent A. Eastman, Katherine J. Lobur, Jerry Silver, Lin Mei, and Evan S. Deneris



Supplemental Fig 1. Adult stage targeting of 5-HT neuron gene expression, Related to Figure 1.

- A. Tph2-CreER targeting efficiency (Tph2+/RFP+) and specificity (RFP+/Tph2+) assessed at 1 month post-TAM. n=2 iCON mice.
- B. *In situ* hybridization analysis of adult stage *Pet1* targeting at 1 month post-TAM; n= 3 mice. Scale bar: 100µm.
- C. Number of neurons flow sorted for RNA-seq at 10 days and 1 month post-TAM.
- D. Reduced *Lmx1b* exon 4 reads demonstrates effective *Lmx1b* targeting.
- E. iPKO and iLKO DEGs at 10 days post-TAM; FDR≤0.05.
- F. Relative expression of *Tph2* and *Slc6a4* in iCON, iPKO and iLKO neurons 10 days post-TAM.
- G. Relative expression of 5-HT neurotransmission genes and *Slc17a8* at 1 month post-TAM.
- H. qPCR analysis of 5-HT gene expression 1 month post-TAM; 2 mice were dissected per replicate (n), n=3-5 mice per genotype; ± SEM; one-way ANOVA.
- I. ISH analysis of *Pet1* (left) or *Lmx1b* (right) 1 month after AAV-Cre injection into the DRN of vCON and vPKO or vLKO mice. Scale bar: 100µm.
- J. ISH of *Tph2*, *Slc6a4*, and *Ddc* in vCON, vPKO, vLKO and vDKO mice at 1 or 6 months post AAV-Cre injection; ± SEM; n= 1,200 -1,500 cells from 3 sections obtained from 3 mice per genotype; 1-way ANOVA. Scale bar: 100µm.
- K. Immunofluorescence staining 1 month and 6 months post AAV-Cre injections; ± SEM; n= 1,200 -1,500 cells from 3 sections obtained from each mouse per genotype; 1-way ANOVA. Scale bar, 100µm, Tph2 panels; 5µm, Sert panels; 100µm, AADC panels.
- L. Gene Ontology (Biological Process) term enrichment for iLKO DEGs at 10 days post-TAM.

*p<0.05, **p<0.01, ***p<0.001, ****p<0.0001, ns, not significant in G-J.



Supplemental Figure 2. Degeneration of pericellular baskets, Related to Figure 2.

A. Coronal schematic highlighting location of dLS where pericellular baskets can be readily imaged. Arrowheads indicate representative dense baskets (DBs) identified at low magnification.

B. Representative images of DBs in iCON (left) and iPKO (right) mice 1 year post-TAM. Scale bar: 10 μ m.

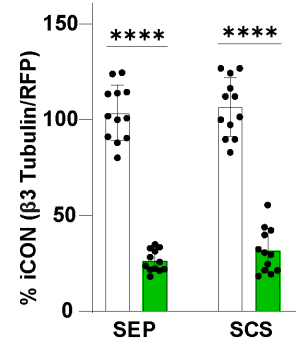
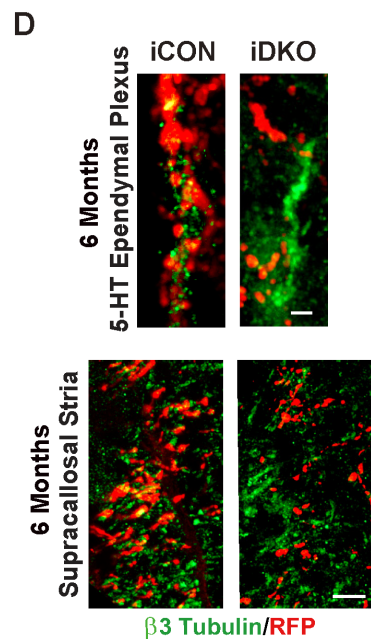
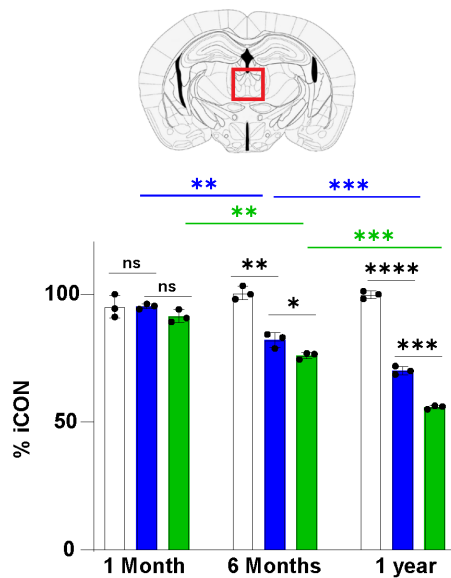
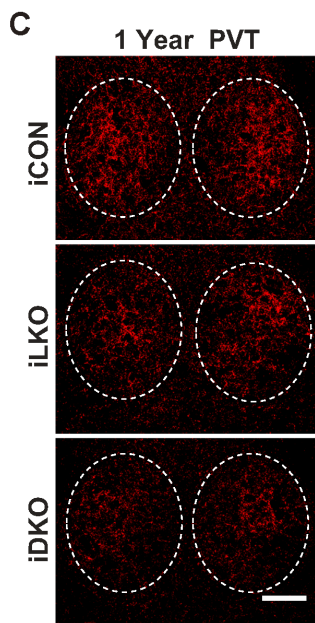
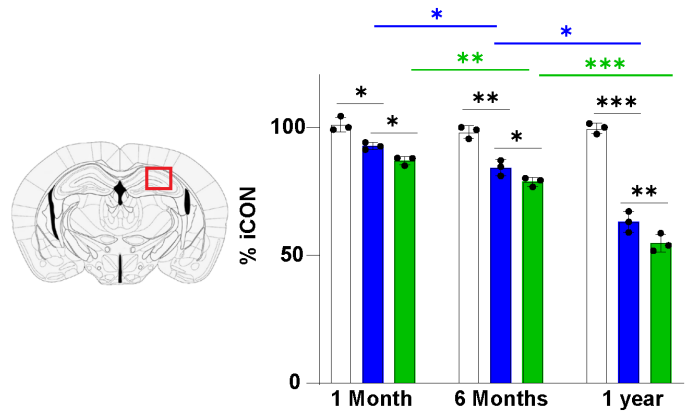
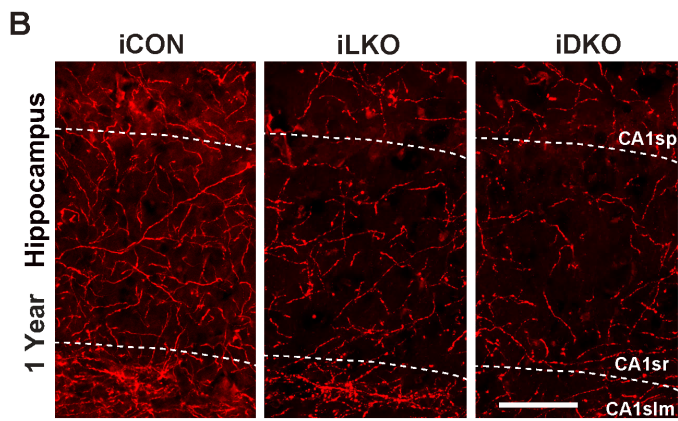
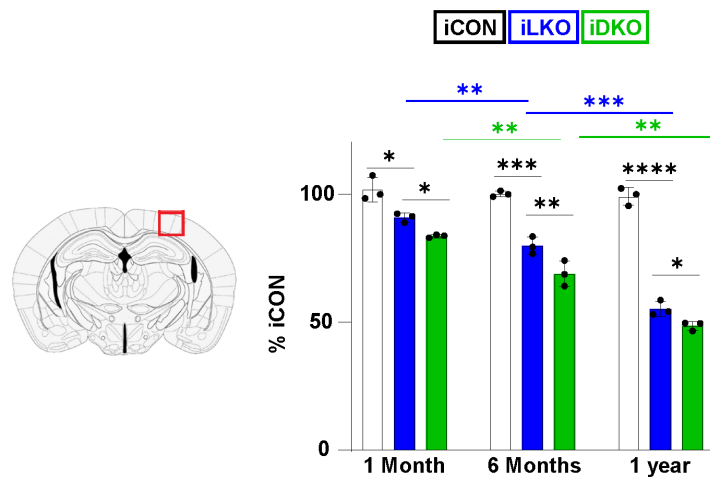
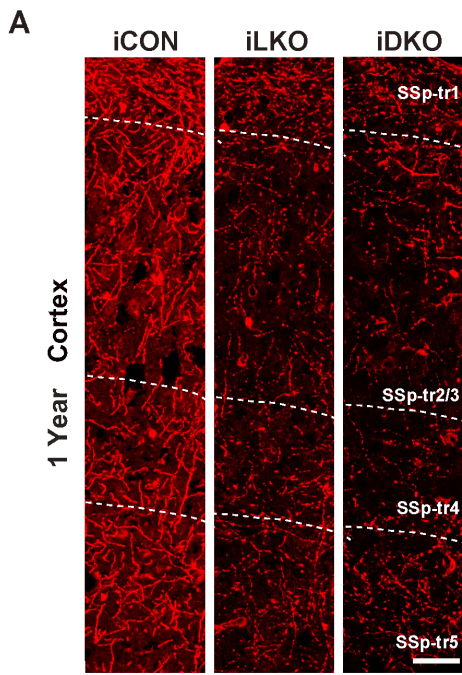
C. Confocal images of readily identifiable DBRs found in iDKO mice 1 year post-TAM. Low magnification (left) indicates that DBRs are easy to identify while higher magnification (right) reveals abnormal morphology. Scale bars: 10 μ m.

D. Representative images of DBRs at 6 months and 1 year post-TAM. Scale bar: 10 μ m.

E. iCON LB and DBs numbers do not change across time points. Data expressed % of iCON numbers at 1 month post-TAM. 1-way ANOVA. n=3 mice per genotype, 2-3 sections per animal were used for counting.

F. Schematic of 5-HT synaptic triad.

G. Representative images of Synapsin1, Gephyrin, and RFP colocalization in iCON, iLKO and iDKO mice at 1 month post-TAM. Scale bar: 5 μ m.



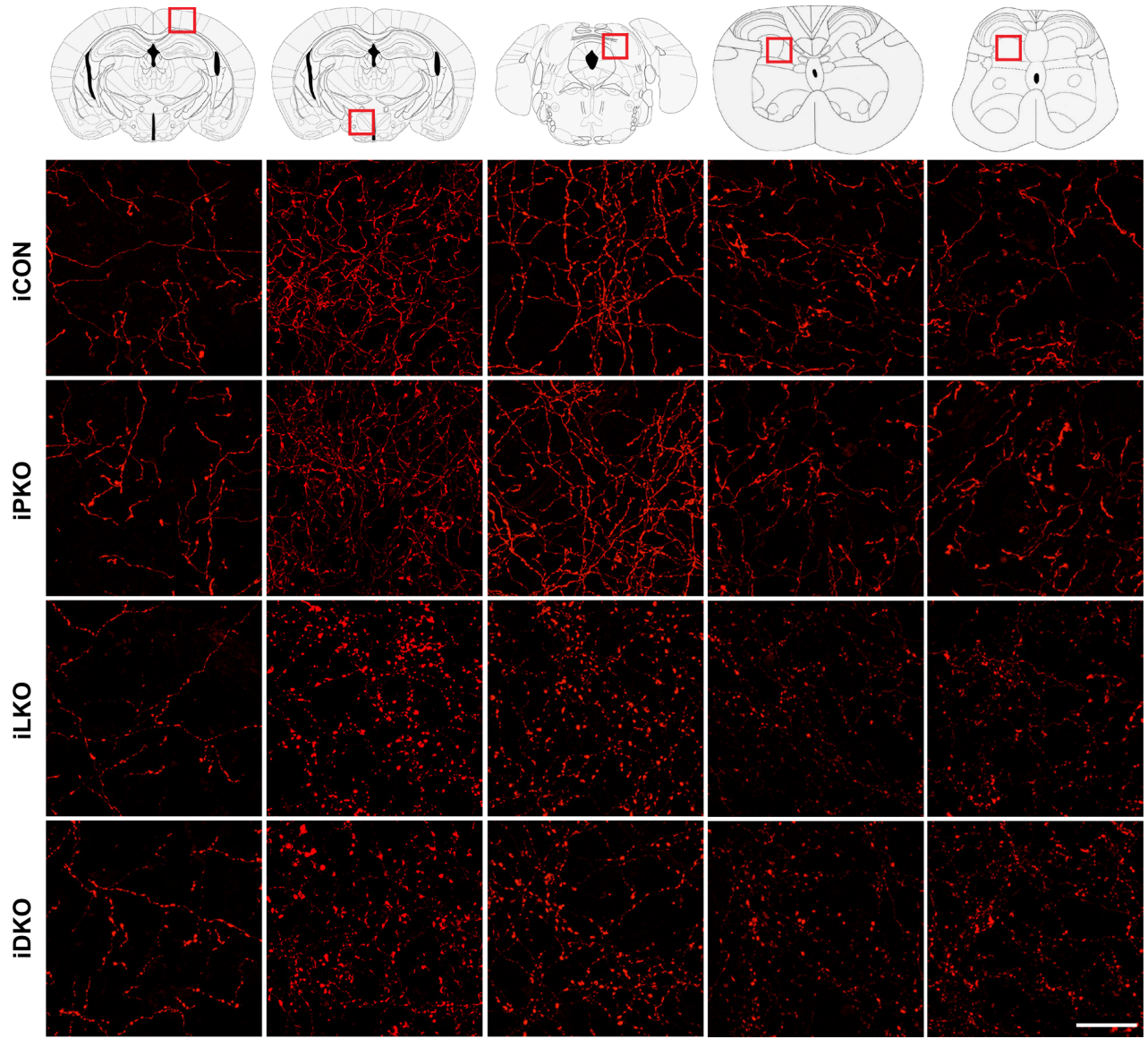
Supplemental Figure 3. Degeneration of serotonergic axons, Related to Figure 3.

A-C, Left, representative images of TdTomato+ (anti-RFP) axons in somatosensory cortex (A), hippocampus (B) and paraventricular nucleus (C) at 1 year post-TAM. Right, quantification of TdTomato+ axon densities; \pm SEM; n=3 mice per genotype; 2-way ANOVA. Scale bars: A, 50 μ m; B, C, 100 μ m.

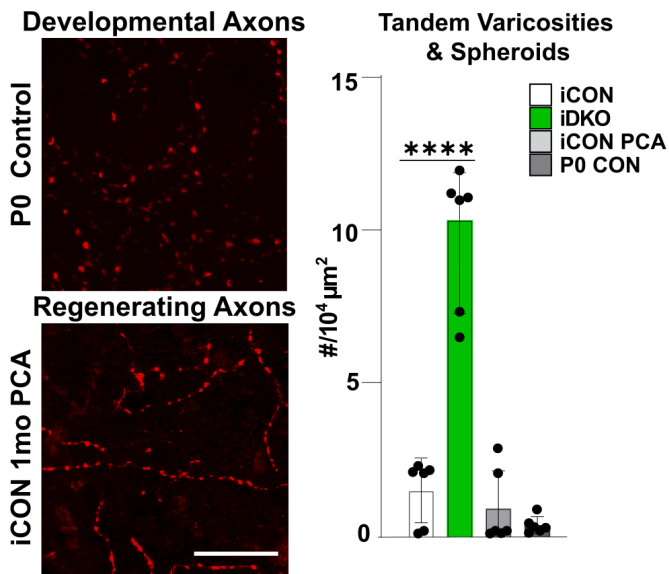
D. Left, representative images of β 3 Tubulin co-immunostaining with RFP at 6 months post-TAM. Scale bars, 2 μ m, 10 μ m. Right, relative ratio of β 3 Tubulin co-immunostaining with RFP at 6 months post-TAM; \pm SEM; n=3 mice per genotype; 1-way ANOVA.

*p<0.05, **p<0.01, ***p<0.001, ****p<0.0001; ns, not significant.

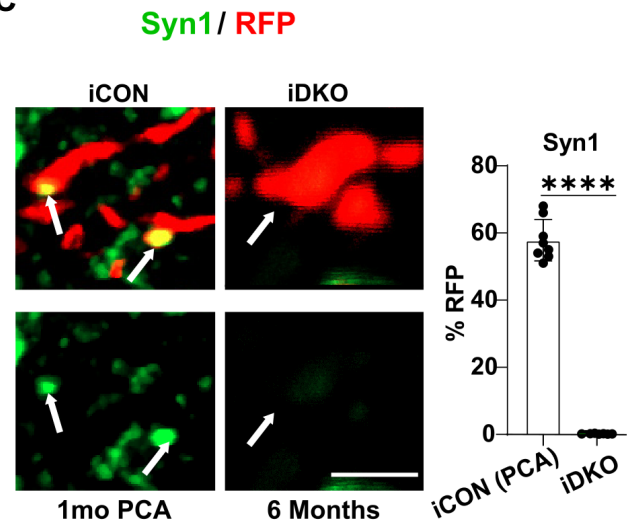
A



B



C



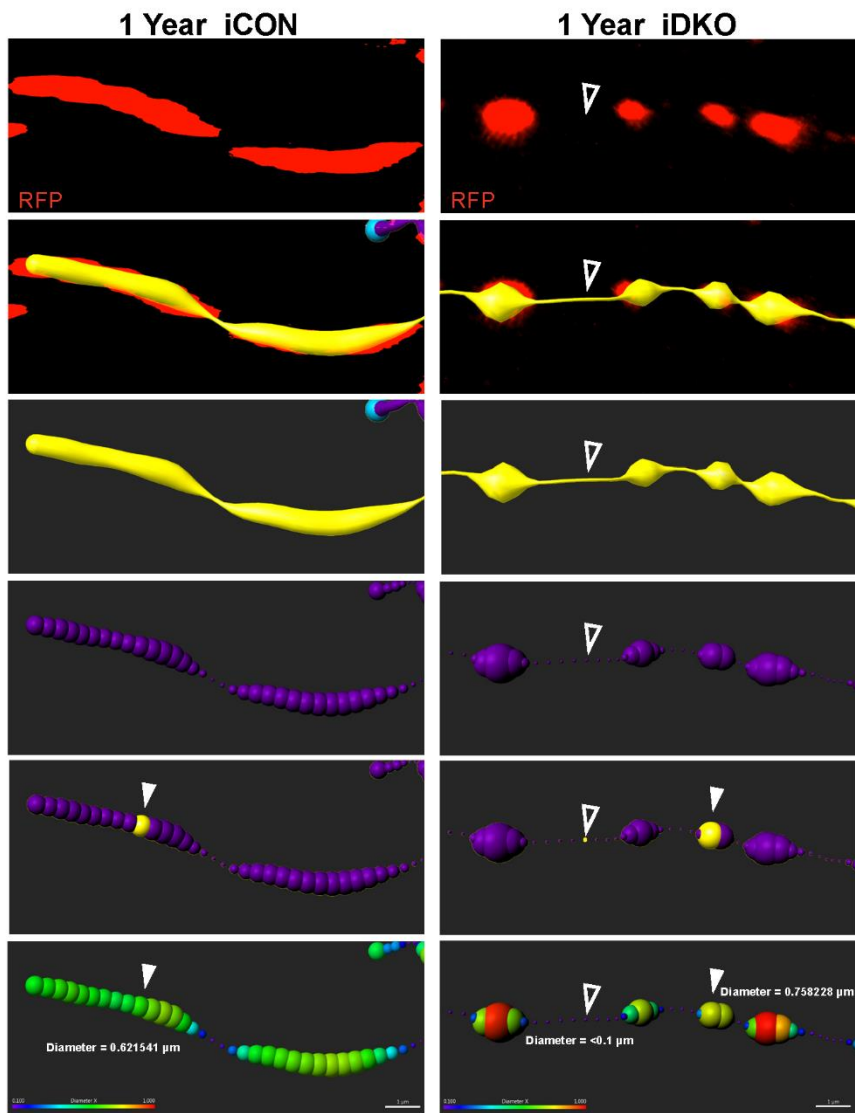
Supplemental Figure 4. Progressive dystrophic morphology, Related to Figure 4.

A. Progressive morphological alterations of TdTomato+ iLKO and iDKO axons at 1 year post-TAM. Left to right: cortex, hypothalamus, inferior colliculus, cervical spinal cord and sacral spinal cord (n=3-5 mice analyzed per genotype).

B. Representative images of control 5-HT axon morphology at birth (upper panel) and adult regenerating axons at 1 month post-PCA administration (lower panel). Right, quantification of tandem varicosities and spheroids; \pm SEM; n=3 mice per genotype.

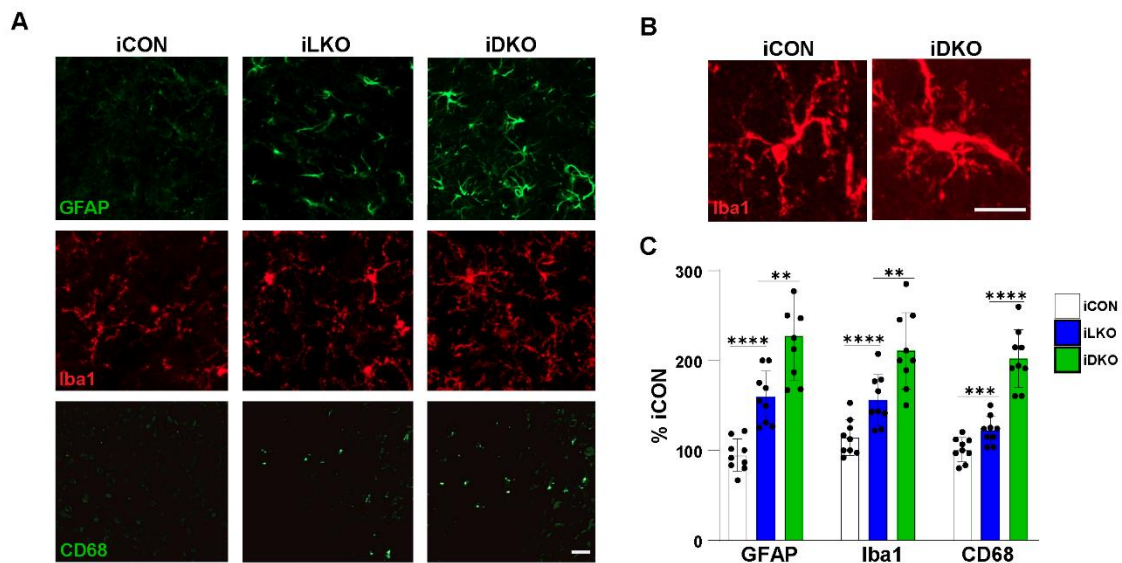
C. Regenerating iCON axons at 1 month post-PCA accumulate Synapsin 1 in varicosities but iDKO fibers do not at 6 month post-TAM. Right, quantification; mean \pm SEM; n=3 mice per genotype.

One-way ANOVA in B, C; ****p<0.0001. Scale bars: 20 μ m in A, B; 2 μ m in C.



Supplemental Figure 5. Axon fragmentation analysis, Related to Figure 4.

Imaris 3D reconstruction modeling and axon segment diameter analysis at 1 year post-TAM. Modeled axon segments representing areas below the detection threshold of $0.1\mu\text{m}$ (open arrowheads) were counted. Fragmentation index was calculated by the number of modeled $<0.1\mu\text{m}$ assignments divided by the sum of $<0.1\mu\text{m}$ and $>0.1\mu\text{m}$ assignments (filled arrowheads).



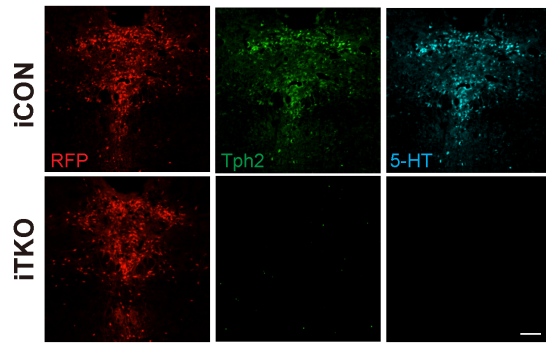
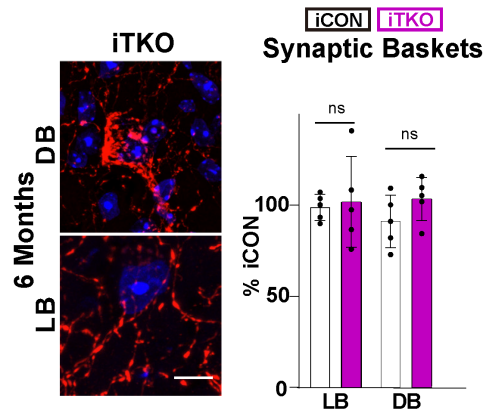
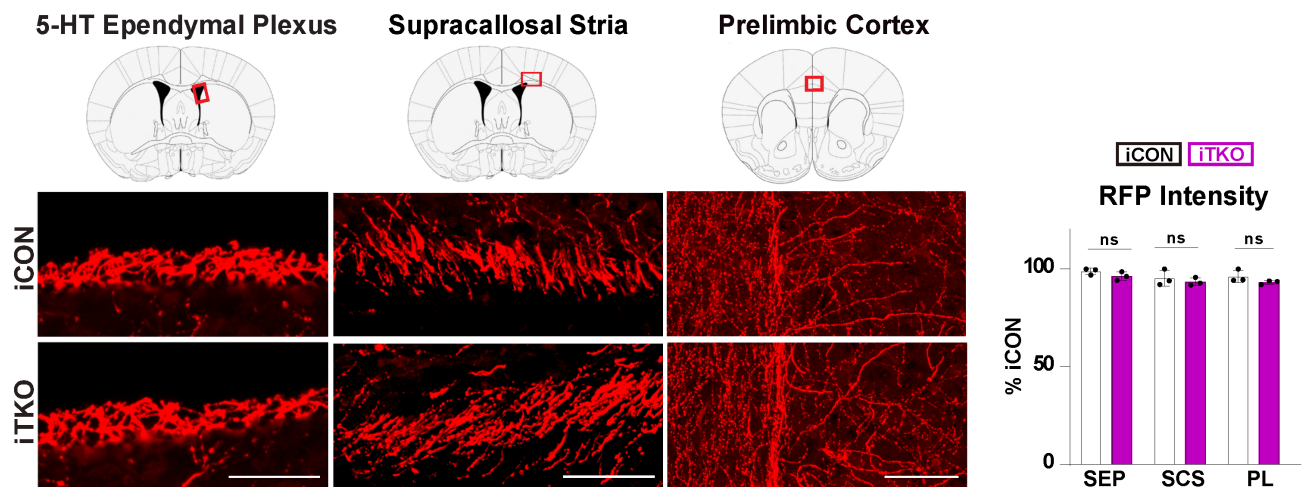
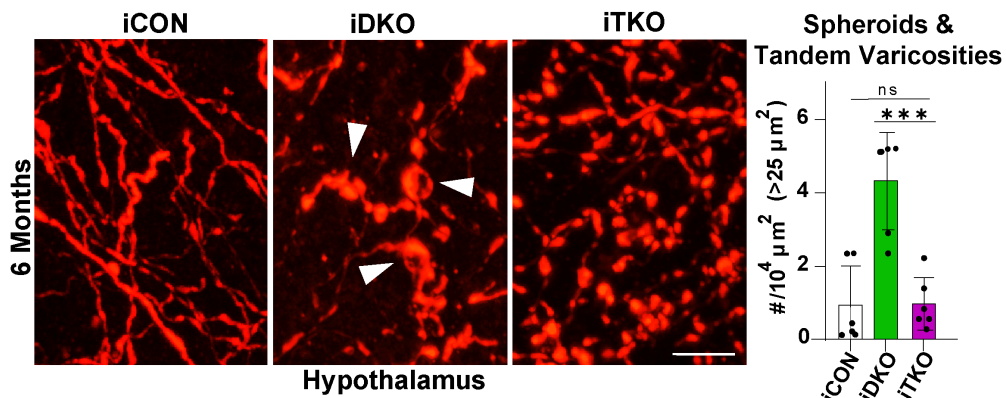
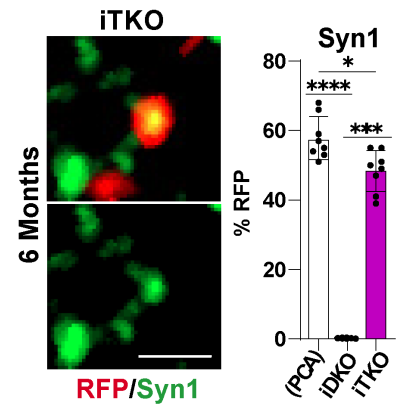
Supplemental Figure 6. Microglia activation and astrogliosis, Related to Figure 4.

A. GFAP, Iba1, and CD68 immunoreactivities are significantly increased in iLKO and iDKO brains compared to iCON at 6 months post-TAM. Scale bar: 20 μ m for each panel.

B. High magnification image of Iba1+ cells in iCON brains exhibiting small cell bodies and ramified processes compared to those in iDKO brains exhibiting an activated morphology. Scale bar: 20 μ m.

C. Quantification of GFAP, Iba1, and CD68 immunofluorescence in iCON, iLKO and iDKO cortex; \pm SEM; n=3 mice using 3 sections per animal per genotype; 1-way ANOVA;

p<0.01, *p<0.001, ****p<0.0001.

A**B****C****D****E**

Supplemental Figure 7. Loss of adult brain 5-HT does not lead to loss of serotonergic connectivity or dystrophic morphology, Related to Figure 4.

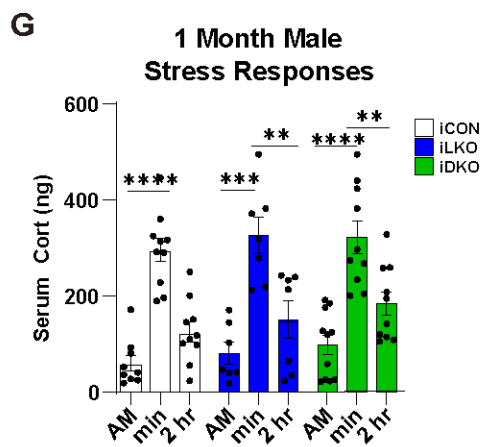
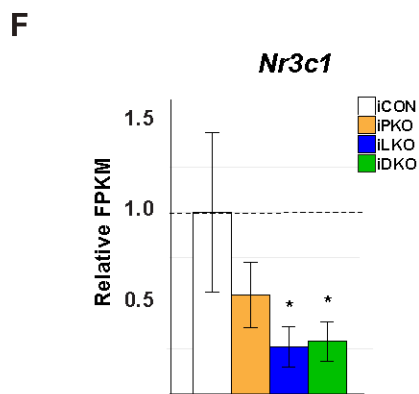
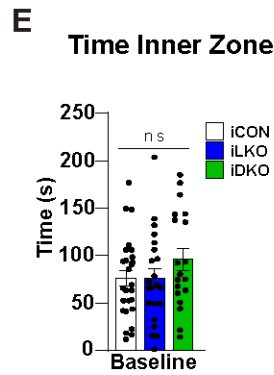
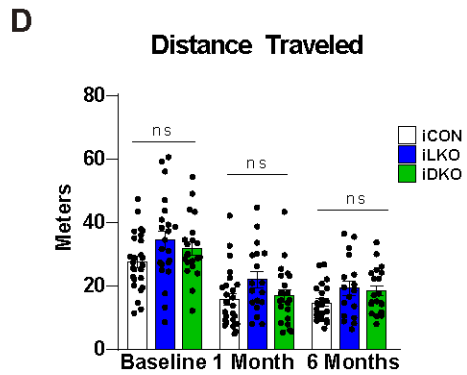
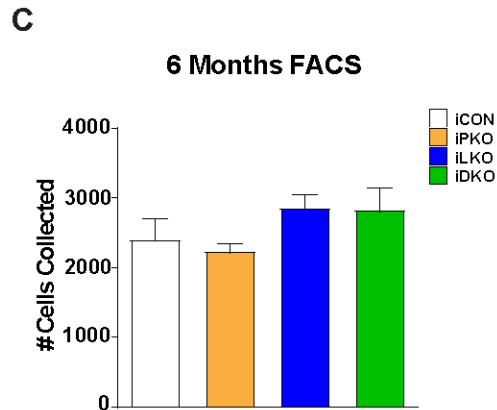
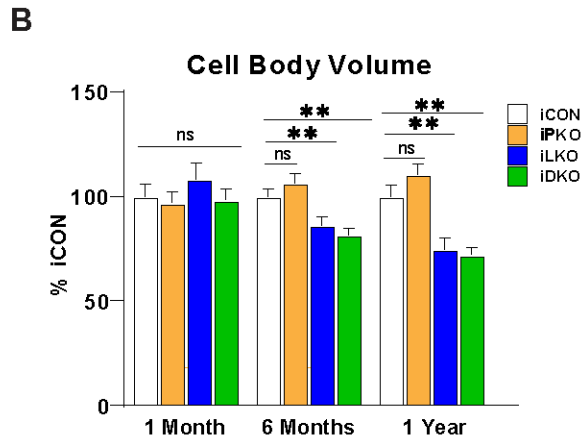
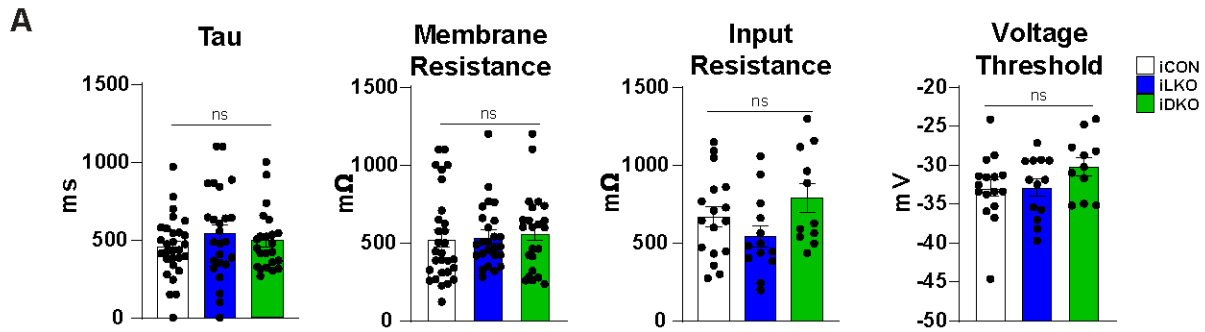
A. Adult stage tamoxifen-inducible *Tph2* targeting.

B. Left, representative images of DBs and LBs in iTKO mice versus iCON at 6 months post-TAM. Right, quantification of pericellular baskets numbers: \pm SEM; n=3 animals per genotype; unpaired t-test with Welch's correction; ns, not significant. Scale bar: 10 μ m.

C. Left, representative images of TdTomato+ (anti-RFP) axons in indicated regions. Right, quantification of RFP fluorescence at 6 months post-TAM: \pm SEM; n=3 mice per genotype; unpaired t-test with Welch's correction; ns, not significant. Scale bars: 50 μ m for SEP and SCS; 100 μ m for PL.

D. Left, TdTomato+ fibers and spheroids (arrowheads) at 6 months post-TAM. Right, quantification of tandem varicosities and spheroids; \pm SEM, n=3 mice per genotype; 1-way ANOVA; ***p<0.001, ns, not significant. Scale bar: 5 μ m.

E. Accumulation of Synapsin1 in iTKO varicosities at 6 months post-TAM; \pm SEM; n=3 mice per genotype; 1-way ANOVA; *p<0.05, ***p<0.001, ****p<0.0001. Scale bar:5 μ m.



Supplemental Figure 8. Passive membrane properties, cell body volumes and behavior assays, Related to Figure 6.

A. Tau, membrane resistance, input resistance, and voltage threshold at 1 month post-TAM; \pm SEM; n=14-18 neurons per genotype; 1-way ANOVA.

B. Cell body volumes at 6 months and 1 year post-TAM; \pm SEM; 150-200 cells were analyzed per mouse, n=2-3 mice per genotype; 1-way ANOVA.

C. Numbers of TdTomato+ neurons flow sorted at 6 months post-TAM.

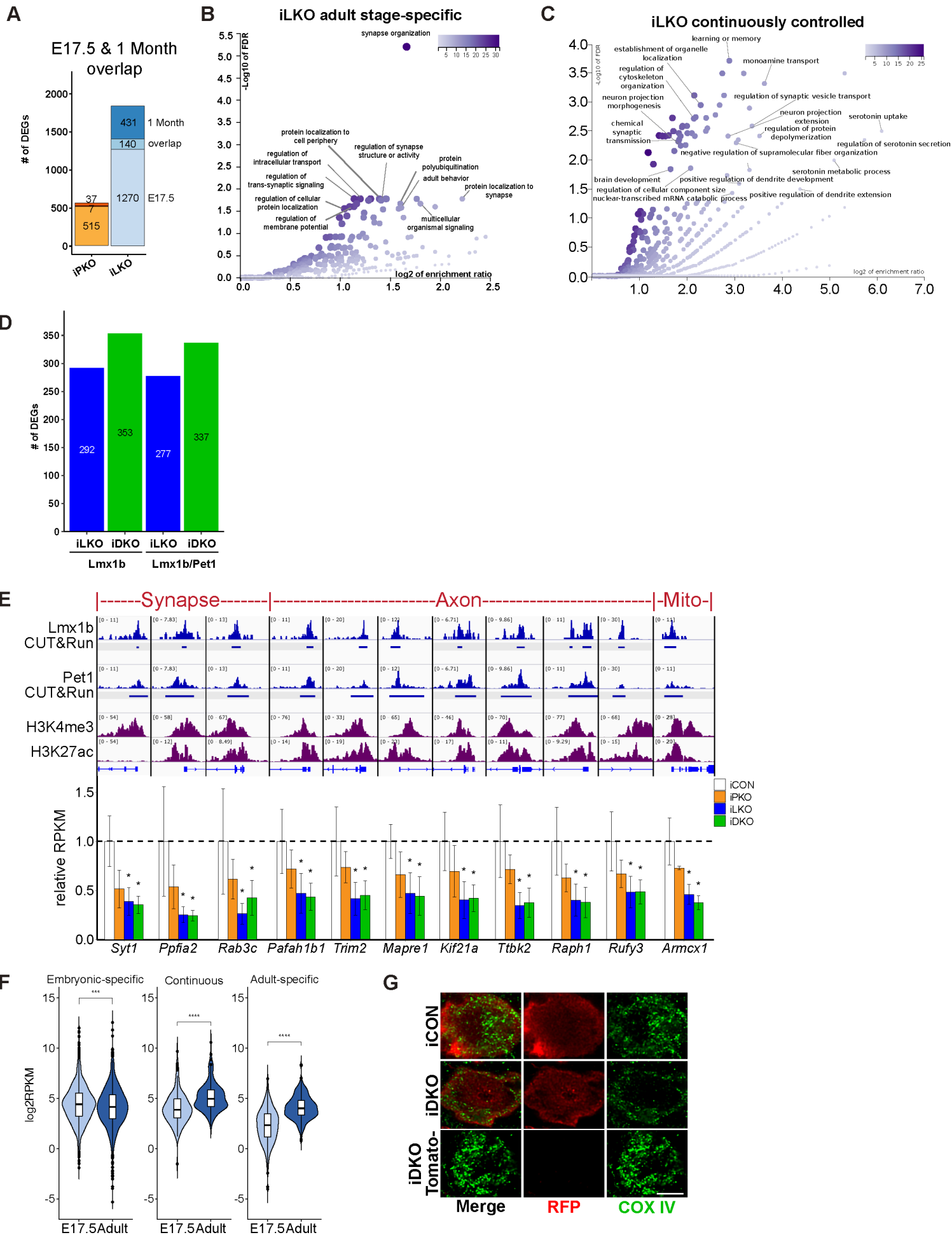
D. Total distance traveled pre-TAM and at 1 and 6 months post-TAM; \pm SEM; n=20-28 mice per genotype; 2-way repeated measures ANOVA.

E. Baseline testing of time in the inner zone of the open field pre-TAM; n=20-28 mice per genotype; 2-way ANOVA and unpaired t-test with Welch's correction.

F. Relative expression (FPKM) of *Nr3c1*. *FDR \leq 0.05.

G. Serum CORT levels after restraint stress in male mice at 1 month post-TAM; \pm SEM; n=8-10 mice per genotype; 2-way repeated measures ANOVA;

p<0.01, *p<0.001, ****p<0.0001, ns, not significant.



Supplemental Figure 9. Adult-stage control of synapse, axon, and mitochondrial genes, Related to Figure 7.

- A. Adult stage-specific DEGs determined by comparison of E17.5 Pet1 and Lmx1b DEGs to 1 month post-TAM Pet1 and Lmx1b DEGs.
- B. Biological Process GO term enrichment of 1 month post-TAM adult stage-specific iLKO down DEGs.
- C. Biological Process GO term enrichment of 1 month post-TAM continuously controlled iLKO down DEGs.
- D. Number of iLKO (blue) or iDKO (green) down DEGs bound by Lmx1b and co-bound by Lmx1b and Pet1 within ± 5 kb of TSSs.
- E. Upper, Genome browser views of Lmx1b and Pet1 co-occupancy (reads in blue) at continuously controlled iLKO and iDKO DEG promoters, defined by enrichment of H3K4me3 and H3K27ac (reads in purple). Lower, bar plots of continuously controlled DEGs; *FDR \leq 0.05.
- F. Violin plots of showing upregulation of adult stage-specific genes. (Wilcoxon rank sum test, ***p \leq 0.001, ****p \leq 0.0001).
- G. Images show mitochondrial fragmentation (COX IV) only in TdTomato+ iDKO cell bodies. Scale bar: 1 μ m.