

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection All data were measured as described in the manuscript without any special software.

Data analysis StepOne software, Zeiss ZEN2, Vilber Bio imaging Evolution Capt , Statcel

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The authors declare that all data supporting the findings of this study are available within this article and its Supplementary Information files. The atomic coordinates and the structure factor files have been deposited in the Protein Data Bank (PDB) under accession number 7X8V. Source data are provided with this paper.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	No sample size calculation was required as used three technical or biological replicates has been historically accepted as valid.
Data exclusions	No exclusion criteria was used, and therefore, no data were excluded.
Replication	All experiments were performed using three biological or three technical replicates
Randomization	Due to the nature of our study design, randomization was not applicable to this study.
Blinding	Due to the nature of the experimental setup, blinding was not applicable .

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

- | n/a | Involved in the study |
|-------------------------------------|--|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Antibodies |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern |

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Antibodies

Antibodies used

anti-PBI1 (Originally made by Tsutomu Kawasaki, dilution 1:1000), anti-PUB44(Originally made by Tsutomu Kawasaki, dilution 1:1000), anti-WRKY45(Originally made by Hiroshi Takatsuji, dilution 1:1000), anti-HA (11867423001 Roche, dilution 1:1000), anti-Myc (MC045 Nakarai tesque, dilution 1:2000), anti-T7 (PM022 Medical Biological Laboratories, dilution 1:500), anti-GFP (50430-2 Proteintech, dilution 1:2000), anti-pMPK (#4370 Cell Signaling, dilution 1:2000), anti-LexA (sc-7544 Santa Cruz Biotechnology, dilution 1:1000), anti-LgBiT (N7100 Promega, dilution 1:500), HRP-linked anti-rabbit (NA934 GE Healthcare, dilution 1:5000), HRP-linked anti-rat (NA935 GE Healthcare, dilution 1:5000), HRP-linked anti-mouse (NA931 GE Healthcare, dilution 1:5000)

Validation

<https://doi.org/10.1038/n.comms6430>, <https://doi.org/10.1073/pnas.1222155110>, https://www.sigmaaldrich.com/catalog/product/roche/roahaha?lang=ja®ion=JP&gclid=EAlaIqobChMIw7OutlL98AIVzF1gCh2wzQSLEAAYASAAEgKllvd_BwE, <https://www.nacalai.co.jp/ss/ec2/ec-srchdetl.cfm?HP=1&l=JP&lc=1&syohin=0436276&syubetsu=3&catalog=&SiireC=&MakerC=&yoro=&mv=1>, <https://ruo.mbl.co.jp/bio/dtl/A/index.html?pcd=PM022>, <https://www.ptglab.co.jp/products/eGFP-Antibody-50430-2-AP.htm>, https://www.cellsignal.jp/products/primary-antibodies/phospho-p44-42-mapk-erk1-2-thr202-tyr204-d13-14-4e-xp-rabbit-mab/4370?_id=1622775538973&Ntt=4370s&thead=true, <https://www.scbt.com/p/lex-a-antibody-2-12>