

**Agonist-dependent coupling of the promiscuous adenosine A<sub>2B</sub> receptor to G $\alpha$  protein subunits**

Jan Hendrik Voss,<sup>1</sup> Andhika B. Mahardhika,<sup>1,2</sup> Asuka Inoue,<sup>3</sup> and Christa E. Müller<sup>1,2\*</sup>

<sup>1</sup>PharmaCenter Bonn, Pharmaceutical Institute, Pharmaceutical & Medicinal Chemistry, University of Bonn, An der Immenburg 4, D-53121 Bonn, Germany

<sup>2</sup>Research Training Group GRK1873, University of Bonn, Germany

<sup>3</sup>Tohoku University, Graduate School of Pharmaceutical Sciences, Sendai, Miyagi, 980-8578 Japan

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**Running title:** A<sub>2B</sub> receptor G $\alpha$  protein coupling

\*To whom correspondence should be addressed: Prof. Dr. Christa E. Müller, Pharmaceutical Institute, Pharmaceutical & Medicinal Chemistry, University of Bonn, An der Immenburg 4, D-53121 Bonn, Germany, Phone: +49-(0)-228-73-2301; Fax: +49-(0)-228-73-2567; E-Mail: [christa.mueller@uni-bonn.de](mailto:christa.mueller@uni-bonn.de)

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**Table S1:** Potency and efficacy of A<sub>2B</sub>AR agonists and CCh at HEK cells recombinantly expressing Gα<sub>q/11</sub> proteins determined in calcium mobilization assays.

<b>Cell line</b>	<b>HEK-Gα<sub>q</sub></b>	<b>HEK-Gα<sub>11</sub></b>	<b>HEK-Gα<sub>14</sub></b>	<b>HEK-Gα<sub>15</sub></b>
<b>Adenosine</b>				
Potency ( <i>p</i> EC <sub>50</sub> )	4.84 ± 0.17 <sup>b</sup>	partial activation	partial activation	5.52 ± 0.29 <sup>c</sup>
Efficacy (AU)	4398 ± 1290 <sup>b</sup>	1800 ± 350 <sup>a</sup>	3570 ± 680 <sup>b</sup>	6480 ± 410 <sup>c</sup>
<b>NECA</b>				
Potency ( <i>p</i> EC <sub>50</sub> )	4.37 ± 0.34 <sup>b</sup>	n.a.	n.a.	5.79 ± 0.21 <sup>c</sup>
Efficacy (AU)	4820 ± 840 <sup>b</sup>	n.a.	n.a.	5450 ± 900 <sup>c</sup>
<b>BAY 60-6583</b>				
Potency ( <i>p</i> EC <sub>50</sub> )	n.a.	n.a.	n.a.	n.a.
Efficacy (AU)	n.a.	n.a.	n.a.	n.a.
<b>CCh</b>				
Potency ( <i>p</i> EC <sub>50</sub> )	6.61 ± 0.24	6.55 ± 0.18	5.12 ± 0.83	5.28 ± 0.32 <sup>a</sup>
Efficacy (AU)	6160 ± 1370	6790 ± 890	8060 ± 2930	4750 ± 900 <sup>a</sup>
Data are presented as means ± SEM from 3 independent experiments performed in duplicates if not otherwise indicated.				
<sup>a</sup> Data from four independent experiments.				
<sup>b</sup> Data from five independent experiments.				
<sup>c</sup> Data from six independent experiments.				
n.a. no activation at the highest tested concentration				

**Table S2:** Potency ( $pEC_{50}$ ) of A<sub>2B</sub>AR agonists at G $\alpha$  proteins determined by TRUPATH BRET2 assays.

TRUPATH Biosensor	Adenosine	NECA -ADA	NECA +ADA	BAY -ADA	BAY 60-6583 + ADA
<b>G<math>\alpha_s</math> subfamily</b>					
G $\alpha_{s-s}$	4.99 ± 0.22	6.73 ± 0.18	6.50 ± 0.19 <sup>c</sup>	7.46 ± 0.55	7.91 ± 0.45 <sup>a</sup>
<b>G<math>\alpha_{i/o}</math> subfamily</b>					
G $\alpha_{i1}$	5.19 ± 0.20	n.d.	7.97 ± 0.30	n.d.	n.a., <5
G $\alpha_{i2}$	5.38 ± 0.09	n.d.	7.43 ± 0.22	n.d.	n.a., <5
G $\alpha_{i3}$	5.58 ± 0.07	n.d.	8.00 ± 0.28	n.d.	n.a., <5
G $\alpha_{oA}$	5.18 ± 0.19 <sup>a</sup>	n.d.	6.36 ± 0.71 <sup>a</sup>	n.d.	n.a., <5
G $\alpha_{oB}$	5.39 ± 0.11 <sup>a</sup>	n.d.	6.29 ± 0.49 <sup>b</sup>	n.d.	n.a., <5
G $\alpha_{gust}$	n.a., <4	n.d.	n.a., <4	n.d.	n.a., <5
G $\alpha_z$	6.19 ± 0.48 <sup>a</sup>	n.d.	7.17 ± 0.28 <sup>a</sup>	n.d.	n.a., <5
<b>G<math>\alpha_q</math> subfamily</b>					
G $\alpha_q$	6.48 ± 0.34 <sup>a</sup>	7.05 ± 0.63 <sup>c</sup>	5.90 ± 0.29 <sup>c</sup>	n.a., <5	n.a., <5
G $\alpha_{11}$	5.85 ± 0.32 <sup>b</sup>	7.89 ± 0.55 <sup>c</sup>	6.90 ± 0.39 <sup>d</sup>	n.a., <5	n.a., <5
G $\alpha_{15}$	6.51 ± 0.32 <sup>d</sup>	7.56 ± 0.25 <sup>d</sup>	8.67 ± 0.23 <sup>b</sup>	7.14 ± 0.69 <sup>c</sup>	9.23 ± 0.08
<b>G<math>\alpha_{12/13}</math> subfamily</b>					
G $\alpha_{12}$	5.66 ± 0.03	n.d.	7.06 ± 0.48 <sup>a</sup>	n.d.	8.61 ± 0.24 <sup>a</sup>
G $\alpha_{13}$	5.66 ± 0.17	n.d.	6.67 ± 0.36 <sup>b</sup>	n.d.	n.a., <5

Data are presented as means ± SEM from three independent experiments (unless otherwise noted) performed in duplicates.  $pEC_{50}$  values calculated from experiments conducted with NECA and BAY 60-6583 were performed in the presence of ADA.

<sup>a</sup> Data from four independent experiments.  
<sup>b</sup> Data from five independent experiments.  
<sup>c</sup> Data from six independent experiments.  
<sup>d</sup> Data from seven independent experiments.

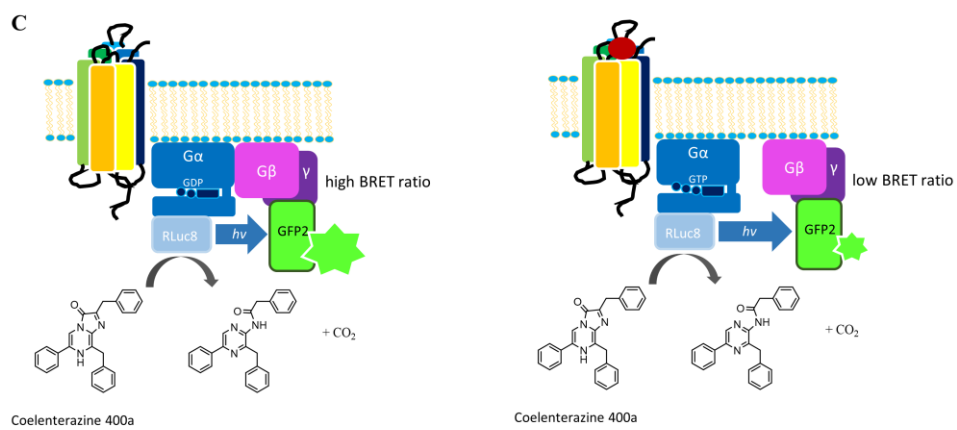
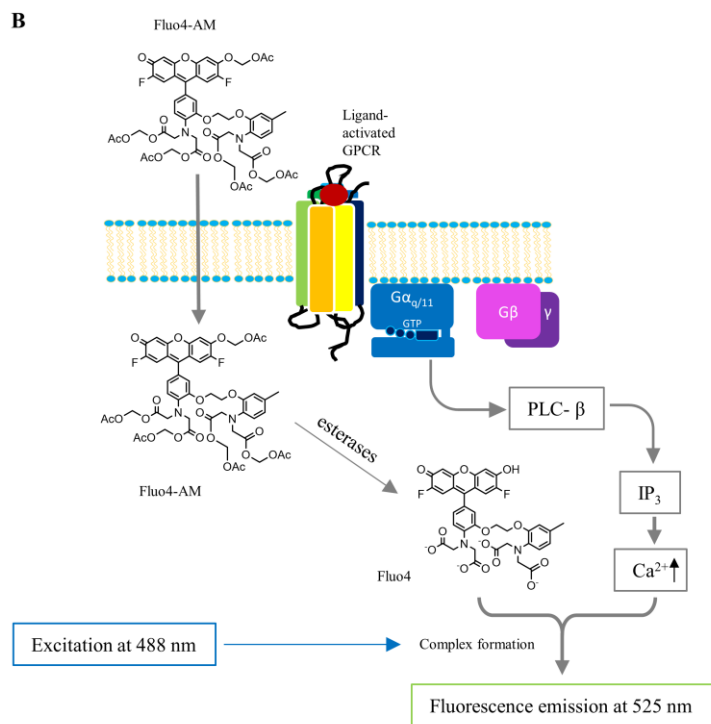
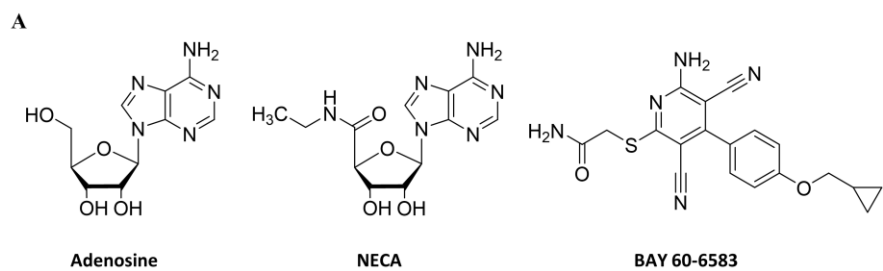
n.a., no activation at the highest tested concentration  
n.d., not determined

**Table S3:** Efficacy ( $E_{\max}$ , % of maximum activation of a model receptor) of A<sub>2B</sub>AR agonists at respective G $\alpha$  proteins determined by TRUPATH BRET2 assays.

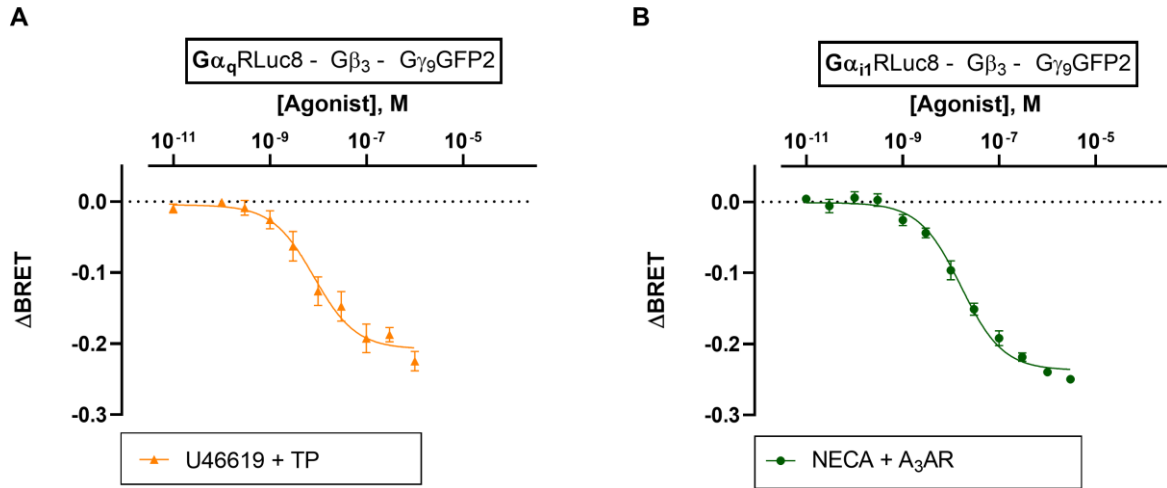
TRUPATH Biosensor	Adenosine	NECA -ADA	NECA +ADA	BAY -ADA	BAY 60-6583 + ADA
<b>G<math>\alpha_s</math> subfamily</b>					
G $\alpha_{s-s}$	120 ± 15	110 ± 23	63 ± 6 <sup>c</sup>	113 ± 38	32 ± 12 <sup>a</sup>
<b>G<math>\alpha_{i/o}</math> subfamily</b>					
G $\alpha_{i1}$	22 ± 5	n.d.	36 ± 6	n.d.	n.a., <10
G $\alpha_{i2}$	49 ± 18	n.d.	76 ± 12	n.d.	n.a., <10
G $\alpha_{i3}$	29 ± 2	n.d.	27 ± 4	n.d.	n.a., <10
G $\alpha_{oA}$	28 ± 4 <sup>a</sup>	n.d.	33 ± 9 <sup>a</sup>	n.d.	n.a., <10
G $\alpha_{oB}$	44 ± 4 <sup>a</sup>	n.d.	37 ± 3 <sup>b</sup>	n.d.	n.a., <10
G $\alpha_{gust}$	n.a., <4	n.d.	n.a., <4	n.d.	n.a., <10
G $\alpha_z$	47 ± 11 <sup>a</sup>	n.d.	44 ± 11 <sup>a</sup>	n.d.	n.a., <10
<b>G<math>\alpha_{q/11}</math> subfamily</b>					
G $\alpha_q$	25 ± 6 <sup>a</sup>	29 ± 6 <sup>c</sup>	22 ± 2 <sup>c</sup>	n.a., <10	n.a., <10
G $\alpha_{11}$	34 ± 4 <sup>b</sup>	28 ± 3 <sup>c</sup>	22 ± 4 <sup>d</sup>	n.a., <10	n.a., <10
G $\alpha_{15}$	88 ± 24 <sup>d</sup>	49 ± 9 <sup>c</sup>	95 ± 18 <sup>b</sup>	64 ± 20 <sup>c</sup>	131 ± 13
<b>G<math>\alpha_{12/13}</math> subfamily</b>					
G $\alpha_{12}$	64 ± 10	n.d.	51 ± 10 <sup>a</sup>	n.d.	23 ± 2 <sup>a</sup>
G $\alpha_{13}$	18 ± 6	n.d.	31 ± 2 <sup>b</sup>	n.d.	n.a., <10

$E_{\max}$  values were calculated by dividing the maximum  $\Delta$ BRET shift measured for the A<sub>2B</sub>AR by the maximum  $\Delta$ BRET shift observed by a control receptor ( $\beta_2$  adrenoceptor for the G $\alpha_s$  family,  $\mu$  opioid receptor for the G $\alpha_{i/o}$  family, neurotensin 1 receptor for the G $\alpha_{q/11}$  and G $\alpha_{12/13}$  family). Means ± SEM from three independent experiments unless otherwise noted.

<sup>a</sup> Data from four independent experiments.  
<sup>b</sup> Data from five independent experiments.  
<sup>c</sup> Data from six independent experiments.  
<sup>d</sup> Data from seven independent experiments.  
n.a., no activation at the highest tested concentration  
n.d., not determined

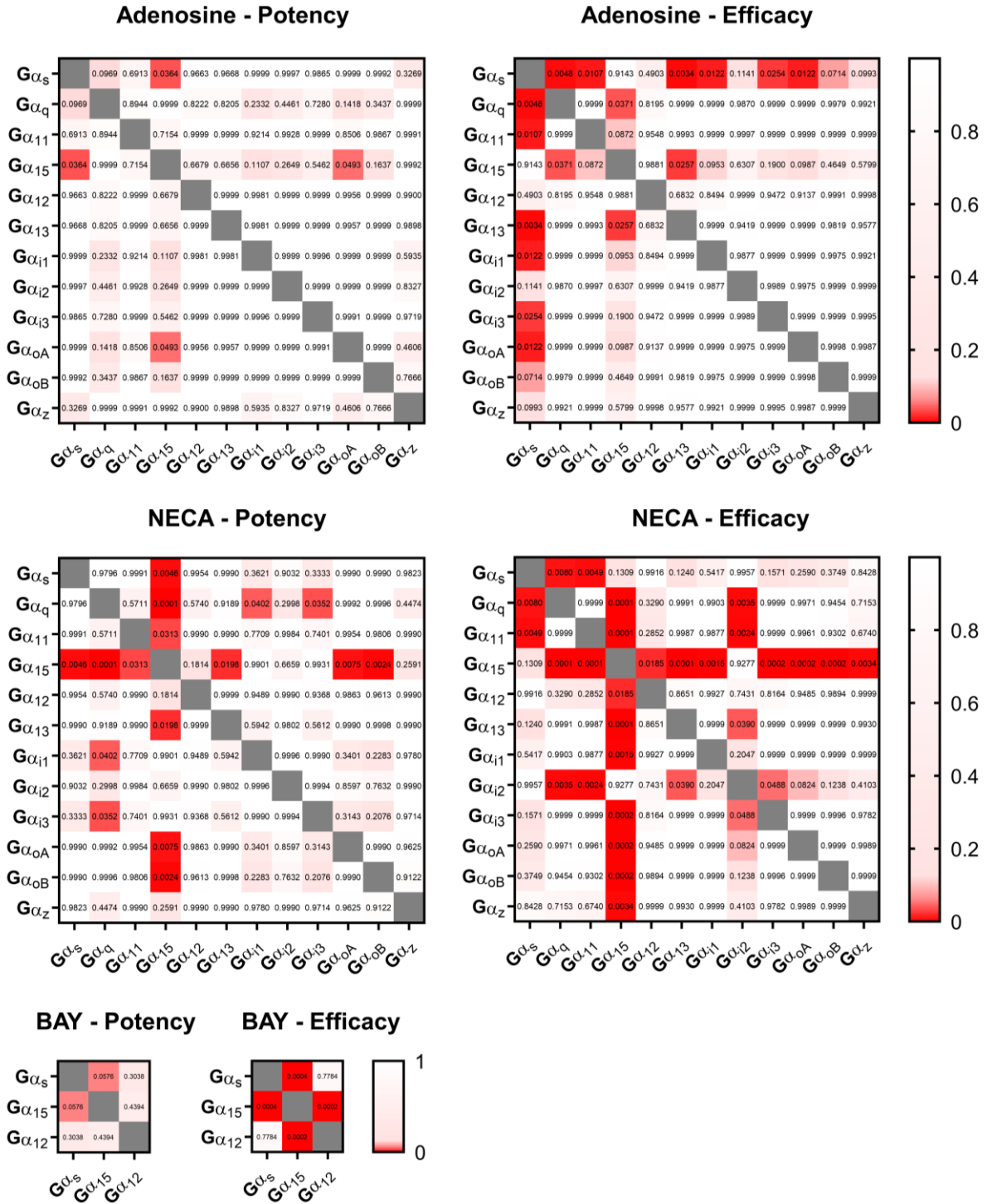


**Figure S1:** Structures of the investigated A<sub>2B</sub>AR agonists, and assay principles of the calcium mobilization assay and the TRUPATH BRET2 assay. **A.** Structural formulas of investigated A<sub>2B</sub>AR agonists. **B.** Calcium mobilization assay: Fluo-4-acetoxymethylester (Fluo-4-AM) penetrates the cell membrane. In the cytosol, the ester bonds are cleaved by esterases and the resulting anionic Fluo-4 is thereby trapped in the cytosol. Upon GPCR activation, G<sub>q/11</sub>-family proteins release GDP and bind GTP instead, leading to calcium release from the endoplasmic reticulum via the phospholipase C-β (PLC-β) – inositol trisphosphate (IP<sub>3</sub>) pathway. Free Ca<sup>2+</sup> binds to Fluo-4 forming a fluorescent complex, which allows the monitoring of changes in the intracellular calcium concentration by fluorescence measurement at an emission wavelength of 520 nm after excitation of the complex at 488 nm. **C.** TRUPATH BRET2 assay: in the inactive receptor state, the RLuc8-tagged Gα subunit is bound to GDP and associated to the Gβγ dimer. The RLuc8 domain oxidizes coelenterazine 400a, emitting a constant signal of blue light, which in turn excites the proximate Gγ-GFP2 fusion protein resulting in a high BRET2 ratio (luminescence / fluorescence). In the active receptor state, the active, GTP-bound Gα-RLuc8 fusion protein separates from the Gβγ-GFP2 dimer. Due to the increased distance between RLuc8 and GFP2, energy transfer is reduced and the BRET2 ratio decreases.



**Figure S2:** Establishment of TRUPATH assay: concentration-response curves of standard agonists at control receptors. **A.**  $G\alpha_q$  biosensor, **B.**  $G\alpha_{i1}$  biosensor, transiently expressed in HEK293 cells together with the investigated control GPCR.  $pEC_{50}$  values:  $G\alpha_q$  - U46619 - TP = 8.02;  $G\alpha_{i1}$  - NECA - A<sub>3</sub>AR = 7.72. Data points are means  $\pm$  SEM of three independent experiments performed in duplicates.





**Figure S3:** Statistical analysis of potency and efficacy values obtained from TRUPATH BRET2 assays. P-values were obtained from one-way analysis of variance (corrected for multiple comparisons using Turkey's test) comparing mean potencies and efficacies of adenosine (A, B), NECA (C, D), and BAY 60-6583 (E, F) at each investigated  $G\alpha$  subunit. High significance levels between means are indicated in red color.