

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- | | | |
|-------------------------------------|-------------------------------------|--|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The statistical test(s) used AND whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | A description of all covariates tested |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
<i>Give P values as exact values whenever suitable.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection	For x-ray diffraction data, data collection used software available through the X-ray generator (Bruker, SC-XRD) or at the beamline (Blu-Ice, remote access). Binding data did not use software for data collection. Data collection software including version numbers is listed in the supplemental tables describing data collection.
Data analysis	Prism 9.3.0 was used for statistical analysis. For diffraction data processing and analysis, we used XDS (version June 1, 2017), HKL2000 v 712, COOT v 0.9, and PHENIX 1.18.2. The Phaser subroutine is within phenix and does not have a separate version number. Program version numbers are now included in the materials and methods in the main text and/or in the supplementary information in legends to the data collection tables, as appropriate.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

Source data are provided with this paper, consisting of raw data for ELISAs and uncropped gels. Atomic coordinates and structure factors have been deposited into the RCSB Protein Data Bank at www.rcsb.org and may be accessed via accession codes or direct hyperlink to the DOI. The codes are 6EFA (doi.org/10.2210/

pdb6EFA/pdb), 6EFB (doi.org/10.2210/pdb6EFB/pdb), 6EFC (doi.org/10.2210/pdb6EFC/pdb), 6EFD (doi.org/10.2210/pdb6EFD/pdb), 6EFF (doi.org/10.2210/pdb6EFF/pdb), 6EFI (doi.org/10.2210/pdb6EFI/pdb), 6EF7 (doi.org/10.2210/pdb6EF7/pdb), 6EF9 (doi.org/10.2210/pdb6EF9/pdb), 6X3Q (doi.org/10.2210/pdb6X3Q/pdb), 6X3K (doi.org/10.2210/pdb6X3K/pdb), 7KMJ (doi.org/10.2210/pdb7KMJ/pdb). Raw X-ray diffraction data have been deposited into SGRID (data.sbgrid.org) with accession codes and direct hyperlinks of 328 (doi.org/10.15785/SBGRID/328), 329 (doi.org/10.15785/SBGRID/329), 507 (doi.org/10.15785/SBGRID/507), 508 (doi.org/10.15785/SBGRID/508), 509 (doi.org/10.15785/SBGRID/509), 510 (doi.org/10.15785/SBGRID/510), 601 (doi.org/10.15785/SBGRID/601), 604 (doi.org/10.15785/SBGRID/604), 787 (doi.org/10.15785/SBGRID/787), 788 (doi.org/10.15785/SBGRID/788), 812 (doi.org/10.15785/SBGRID/812), 813 (doi.org/10.15785/SBGRID/813). A summary of these accession codes is also listed in Tables S1, S2, and S3. Glycomic data were deposited in MassIVE (<https://massive.ucsd.edu/>) with the dataset identifier MSV000088327 and direct hyperlink doi.org/10.25345/C5G577.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	sample size of n=3 was used as the minimum number to allow for the calculation of standard deviation. It was not pre-determined by statistical methods.
Data exclusions	no data were excluded from the biophysical analyses.
Replication	all experiments were performed with a minimum of three independent measurements and all replicates were successful
Randomization	not applicable - biological samples were given numeric identifiers. ELISAs were measured in a logical concentration series.
Blinding	biological samples were given numeric identifiers.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Included in the study
<input type="checkbox"/>	<input checked="" type="checkbox"/> Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input checked="" type="checkbox"/>	<input type="checkbox"/> Animals and other organisms
<input type="checkbox"/>	<input checked="" type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Included in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Antibodies

Antibodies used	Anti-GST antibody was from Invitrogen (A5800) was used at a 1:500 dilution. Peroxidase-conjugated goat anti-rabbit IgG was from Sigma (A0545) and used at a 1:5,000 dilution
Validation	The anti-GST antibody recognizes the GST affinity tag and is broadly used. The ThermoFisher web site provides 67 references demonstrating its validation; three of these references are for ELISA, the technique used here. See https://www.thermofisher.com/antibody/product/GST-Tag-Antibody-Polyclonal/A-5800 . The goat anti-rabbit IgG recognizes rabbit IgG and is similarly widely-used. The Sigma web site points to 1,348 papers that use this antibody.

Human research participants

Policy information about [studies involving human research participants](#)

Population characteristics

Salivary samples were a from a banked collection that we accessed for free. We did not collect samples. This is an older collection and we do not have access to demographic information for the donors.

Recruitment

We did not recruit participants.

Ethics oversight

De-identified samples of banked SMSL saliva were provided by S. Fisher (UCSF). Because these specimens were de-identified prior to gifting, our use of this material was exempt from approval by the UCSF Institutional Review Board. This is now stated in the methods.

Note that full information on the approval of the study protocol must also be provided in the manuscript.