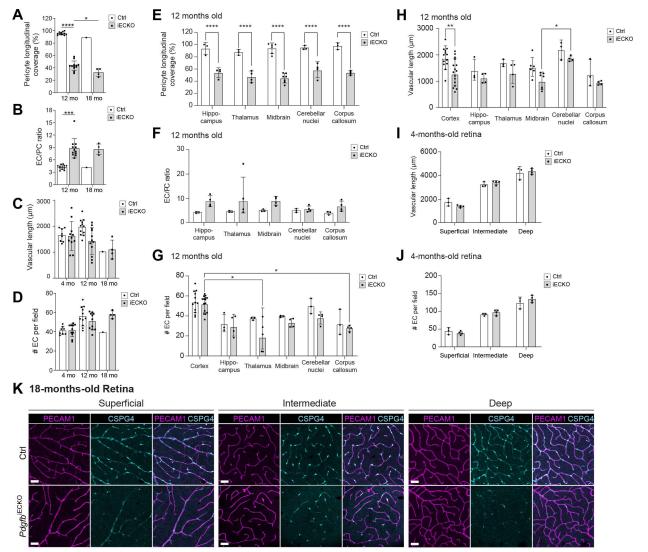
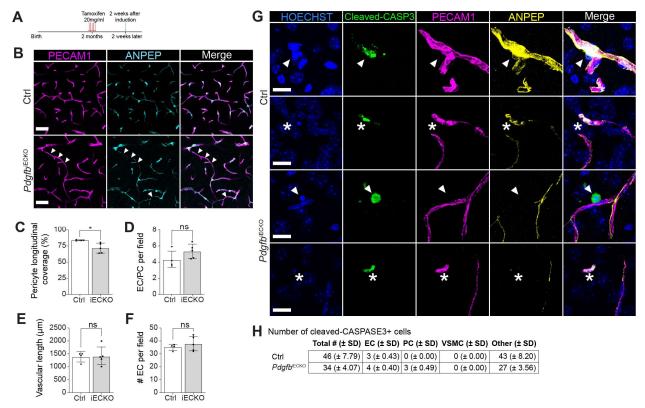


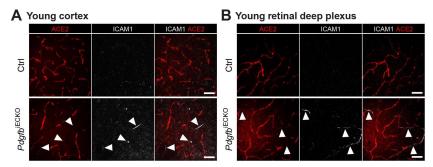
Supplementary figure 1. (A, B) RNA in situ hybridization on 3-months-old C57BL6 mouse cerebral cortex (n=3). *Pdgfb* (cyan) expression colocalizes with *Pecam1* (magenta) positive endothelium (A) and *Tubb3* (magenta) positive neurons (B). Cell nuclei are stained with DAPI (white). Scale bars 25 µm and 50 µm. (C) Representative overview images from the cortex of uninduced 2-months-old mice. Two litters were analyzed (n=6). Co-immunolabeling of PECAM1 (magenta) and ANPEP (cyan). Scale bars 50 µm. (D) Pericyte longitudinal coverage in uninduced *Pdgfb* cov and controls (two litters, n=6). (E) Quantification of the endothelial cell (ERG+) to pericyte (ANPEP+, DAPI+) ratio per field (two litters, n=6). (F) The skeletal length of PECAM1 positive capillaries was quantified and considered as the vascular length per field (two litters, n=6). (G) The total number of endothelial cells (ERG+) detected per field (two litters, n=6). (H) Representative images from the cortex of uninduced 2-months-old mice. One litter was analyzed (n=3). Co-immunolabeling of PECAM1 (magenta), ANPEP (cyan) and MCAM (green). The white arrowhead marks an MCAM-expressing aaVSMC and the yellow arrowhead marks a faint MCAM-expressing pericyte. Scale bar 50 µm. E, Normality tests revealed that the data was unevenly distributed so nonparametric Mann-Whitney U test was used to evaluate significance. D, F and G, The significance of evenly distributed data was evaluated using unpaired 2-tailed t test with Welch correction. ns= not significant. Data is presented as geometric mean with geometric SD.



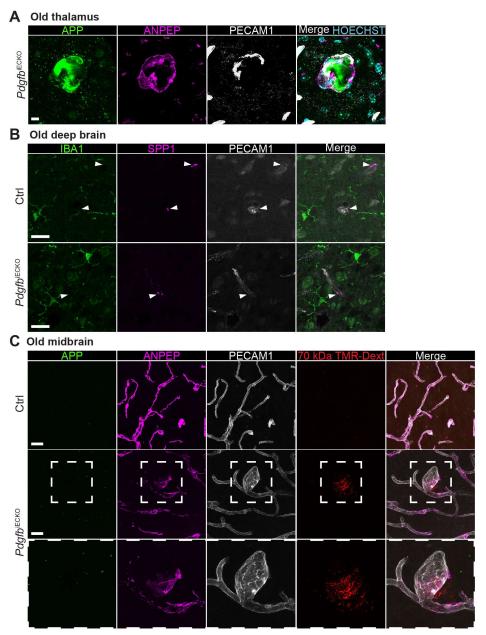
Supplementary figure 2. (A-D) Comparison between 12- and 18-months-old mice. 4 litters of 12-moths-old mice (Pdqfb|ECKO n=12, Ctrl n=12), 1 litter of 18-months-old mice (Pdgfb|ECKO n=4, Ctrl n=1). Not significant data is not labeled. (A) Pericyte longitudinal coverage of the vasculature per field in cerebral cortex. (B) Endothelial cell to pericyte ratio per field in cerebral cortex. (C) Vascular longitudinal length per field in cerebral cortex. (D) Endothelial cell numbers per field in cerebral cortex. (E) Pericyte longitudinal coverage per field in hippocampus, thalamus, midbrain, cerebellar nuclei and corpus callosum of 12-months-old mice. Two litters of 12-months-old mice (Pdgfb)ECKO n=4, Ctrl n=3). In midbrain, four litters of 12-months-old mice (Pdgfbjecko n=11, Ctrl n=7). (F) Endothelial cell to pericyte ratio per field in hippocampus, thalamus, midbrain, cerebellar nuclei and corpus callosum of 12-months-old mice. Two litters of 12-months-old mice (Pdgfb^{ECKO} n=4, Ctrl n=3). (G) Endothelial cell numbers per field in cerebral cortex, hippocampus, thalamus, midbrain, cerebellar nuclei and corpus callosum of 12-months-old mice. Two litters of 12-months-old mice (Pdgfbjecko n=4, Ctrl n=3). In cerebral cortex, five litters of 12-months-old mice (Pdgfbjecko n=16, Ctrl n=13). In midbrain, four litters of 2-months-old mice (Pdgfbiecko n=11, Ctrl n=7). (H) Vascular longitudinal length per field in cerebral cortex, hippocampus, thalamus, midbrain, cerebellar nuclei and corpus callosum of 12-months-old mice (two litters, Pdgfb^{ECKO} n=4, Ctrl n=3). In the cerebral cortex, five litters of 2-months-old mice (Pdgfb^{ECKO} n=16, Ctrl n=13). In midbrain, four litters of 12-months-old mice (Pdgfb^{ECKO} n=11, Ctrl n=7). (I) Vascular longitudinal length per field in Superficial, intermediate and deep retinal plexuses of 4-months-old mice (one litter, Pdgfb|ECKO n=4, Ctrl n=3). (J) Endothelial cell numbers per field in Superficial, intermediate and deep retinal plexuses of 4-months-old mice (one litter, Pdgfb)ECKO n=4, Ctrl n=3). (K) Representative images of 18-months-old retinal superficial, intermediate and deep plexuses. One litter was analyzed (Pdgfb^{ECKO} n=2, Ctrl n=1). Co-immunolabeling of PECAM1 (magenta) and CSPG4 (cyan). Scale bars 50 µm. A and B, Normality tests revealed that the data was unevenly distributed so nonparametric Kruskal-Wallis multiple comparison test was used to evaluate significance. C and D, The significance of evenly distributed data was evaluated using Tukey's multiple comparison test. G and I Intermediate, Normality tests revealed that the data was unevenly distributed so nonparametric Mann-Whitney U test was used to evaluate significance. E, F, H, I Superficial and Deep and J, The significance of evenly distributed data was evaluated using unpaired 2-tailed t test with Welch correction. Data is presented as geometric mean with geometric SD. *p<0.05, **p<0.01, ***p=0.001, ****p<0.0001, ns=not significant.



Supplementary figure 3. (A) Endothelial specific Pdgfb deletion was induced via 5 doses of Tamoxifen administration at 2 months of age. Two weeks after the mice were analyzed. (B) Representative overview images from the cortex of mice analyzed 2 weeks after induction. Co-immunolabeling of PECAM1 (magenta) and ANPEP (cyan). Arrowheads mark places where the expression of ANPEP is lacking. One litter was analyzed (Ctrl n=4, Pdgfb^{ECKO} n=5). Scale bars 50 µm. (C) The pericyte longitudinal coverage in Pdgfb^{iECKO} and controls (one litter, Ctrl n=4, Pdgfb^{iECKO} n=5). (D) Quantification of the endothelial cell (ERG+) to pericyte (ANPEP+, HOECHST+) ratio (one litter, Ctrl n=4, Pdgfb|ECKO n=5). (E) The skeletal length of PECAM1 positive capillaries was quantified and considered as the vascular length per field (one litter, Ctrl n=4, Pdgfb|ECKO n=5). (F) The total number of endothelial cells (ERG+) detected per field (one litter, Ctrl n=4, PdgfbiECKO n=5). (G) Representative images from the cortex of PdgfbiECKO and littermate control mice analyzed 2 weeks after induction. Hoechst (white), Cleaved-CASPASE3 (green), PECAM1 (magenta), ANPEP (yellow). Arrowheads show cells undergoing apoptosis and asterisks show potential endothelial cells undergoing apoptosis. One litter was analyzed (Ctrl n=4, PdgfbECKO n=5). Scale bars 25 µm. (H) Quantification of total number of cleaved-CASPASE3 positive cells per two 50 µm-thick sections. Breakdown of the total number of apoptotic cells into endothelial cells, pericytes, VSMCs and other cell types (one litter, Ctrl n=4, Pdgfb^{ECKO} n=5). Total numbers per group are shown with ± SD. E, Normality tests revealed that the data was unevenly distributed so nonparametric Mann-Whitney U test was used to evaluate significance. C, D and F, The significance of evenly distributed data was evaluated using unpaired 2-tailed t test with Welch correction. *p<0.05, ns= not significant. Data is presented as geometric mean with geometric SD.



Supplementary figure 4. (A) Representative images of the cortex of young *Pdgfb*|ECKO and littermate controls. Co-immunolabeling of ACE2 (red) and ICAM1 (white). White arrowheads mark capillaries with ICAM1 expression that lack pericyte coverage. Scale bar 50 μm. *Pdgfb*|ECKO n=11, Ctrl n=10. (B) Representative images of the retinal plexuses of young *Pdgfb*|ECKO and littermate controls. Co-immunolabeling of ACE2 (red) and ICAM1 (white). White arrowheads mark capillaries with ICAM1 expression that lack pericyte coverage. Scale bar 50 μm. *Pdgfb*|ECKO n=4, Ctrl n=3.



Supplementary figure 5. (A) Calcified nodule in the thalamus of an old *Pdgfb*^{ECKO} mouse. Co-immunolabeling of amyloid precursor protein, APP (green), ANPEP (magenta), PECAM1 (white) and labelling of nuclei with Hoechst (cyan). Scale bar 10 μm. Four independent experiments, *Pdgfb*^{ECKO} n=16, Ctrl n=12. **(B)** Representative images of SPP1 nodules (white arrowheads) on the endothelium of old *Pdgfb*^{ECKO} mice and littermate controls (two litters, *Pdgfb*^{ECKO} n=7, Ctrl n=6). Co-immunolabeling of IBA1 (green), SPP1 (magenta) and PECAM1 (white). Scale bars 25 μm. **(C)** Representative images of the midbrain of old mice. Co-immunolabeling of APP (green), ANPEP (magenta), PECAM1 (white) and 70 kDa TMR-dextran (red). Dashed inset shows a 70 kDa TMR-dextran hotspot on an enlarged capillary without any signs of APP+ calcification. One litter, *Pdgfb*^{ECKO} n=3, Ctrl n=3. Scale bars 25 μm.