

Fig. S1 Purification and cryo-EM data processing of SST14 bound SSTR2-Gi complex.

a Size-exclusion chromatography profile and SDS-PAGE analysis of SSTR2-Gi complex bound with SST14. **b** Representative micrograph after motion correction and dose weighting. **c** 2D class averages of SSTR2-Gi complex bound with SST14. **d** Flow chart of cryo-EM data processing using cryoSPARC. **e** Gold-standard FSC validation curves from cryoSPARC. **f** Density map of SSTR2-Gi/SST14 complex colored by local resolution estimation.







Fig. S3 Purification and cryo-EM data processing of lanreotide bound SSTR2-Gi complex. a Size-exclusion chromatography profile and SDS-PAGE analysis of SSTR2-Gi complex bound with lanreotide. **b** Representative micrograph after motion correction and dose weighting. **c** 2D class averages of SSTR2-Gi complex bound with lanreotide. **d** Flow chart of cryo-EM data processing using cryoSPARC. **e** Gold-standard FSC validation curves from cryoSPARC. **f** Density map of SSTR2-Gi/lanreotide complex colored by local resolution estimation.



Fig. S4 Cryo-EM densities of representative segments of SSTR2-Gi complex.

a-c Cryo-EM density maps and models of transmembrane domain of SSTR2-Gi/SST14 complex(**a**), SSTR2-Gi/octreotide(**b**) and SSTR2-Gi/lanreotide complex(**c**). The cryo-EM maps are shown in mesh and molecular models as stick representation.



Fig. S5 The ligand binding pockets of SSTR2 bound with SST14, Octreotide or Lanreotide. Side views of the orthosteric binding pockets in the SST14-bound SSTR2(**a**), ortreotide-bound SSTR2(**b**) and lanreotide-bound SSTR2(**c**) structures. Residues within 4 Å of both ligands are shown as sticks and the hydrogen bond interactions are represented by dotted lines. In both panels, red and blue sticks represent oxygen and nitrogen, respectively.





a-c Space-filling models and cross-section views of SST14-bound SSTR2(a), ortreotide-bound

SSTR2(b) and lanreotide-bound SSTR2(c).

d Comparision of ECL2 of lanreotide or ortreotide or SST14 bound SSTR2.



Fig. S7 Sequence alignment of SSTR2 and µ-opioid receptor.



Fig. S8 Conformational comparison of SSTR2 upon activation by different ligands.

Side (a) and top (b) views of alignment of active SST14-bound SSTR2 (blue), ortreotide-bound SSTR2(green) and lanreotide-bound SSTR2(purple).

Data Collection			
Protein	SSTR2/SST14	SSTR2/OCT	SSTR2/LAN
Microscope	FEI Titan Krios	FEI Titan Krios	FEI Titan Krios
Voltage (kV)	300	300	300
Detector	Gatan K3 Summit	Gatan K3 Summit	Gatan K3 Summit
Detector mode	Super	Super	Super
Pixel size (Å)	0.535	0.535	0.535
Defocus range (µm)	-1.2 ~ -2.2	-1.2 ~ -2.2	-1.2 ~ -2.2
Electron dose (e ⁻ /Å ²)	55	55	55
Frames per image	30	30	30
Exposure time (s)	3.5	3.5	3.5
Number of images	4,189	4,837	7,377
3D reconstruction			
Final Particle number	457,712	666,575	743,506
Symmetry	C1	C1	C1
Overall resolution (Å)	2.85	2.97	2.87
Model refinement			
Model composition			
Chains	6	6	6
Ligands	DTR:1	DTR:1	DTR:1
Non-hydrogen atoms	8,440	8,443	8,410
Protein residues	1,152	1,189	1,140
Bonds (RMSD)			
Length (Å)	0.014	0.011	0.009
Angles (°)	1.252	1.189	1.051

Table S1 | Statistics of cryo-EM data collection, 3D reconstruction and model refinement.

Ramachandran plot (%)

Outliers	0.09	0.00	0.00
Allowed	7.23	8.12	8.30
Favored	92.68	91.88	91.70
Rotamer outliers (%)	0.83	1.17	0.71
MolProbity score	1.81	1.94	1.93
Clash score	6.35	7.24	7.84

Table S2 | Summary of EC50 values, Emax for the wild-type SSTR2 and SST22 mutants activation by SST14, Octreotide and Lanreotide. pEC50 values represent the negative logarithm of agonist concentration that produces half maximal response. Emax values are maximal response as percentage of wild-type SSTR2 response. All values are expressed as mean \pm s.e.m. of three independent experiments conducted in triplicate. Data were performed using one-way analysis of variance followed by Dunnett's test using wild-type receptors as the control (*P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001). n.d., not determined.

	Ligand	Ca ²⁺	Ca ²⁺	Surface
		pEC50	Emax	expression
				(%WT)
	Octreotide	$7.7 \pm 0.04, (22)$	/	
SSTR2	SST14	$8.0 \pm 0.1, (20)$	/	100
WT	Lanreotide	$7.3 \pm 0.05, (10)$	/	
	Octreotide	$7.5 \pm 0.1, (3)$	$82.6 \pm 7.4^{**}$	
SSTR2(1-359)-	SST14	$7.9 \pm 0.1, (3)$	$77.8 \pm 6.1^{****}$	113.5 ± 7.1
Largebit	Lanreotide	$7.2 \pm 0.1, (3)$	101.8 ± 9.1	
	Octreotide	$5.9 \pm 0.2, (3)$ *****	103.8 ± 12.1	
Q102A	SST14	$7.2 \pm 0.2, (3)$ *	95.3 ± 6.0	47.1 ± 1.7
	Lanreotide	$6.4 \pm 0.1, (3)$ ****	$56.1 \pm 4.2^{****}$	
	Octreotide	n.d. (3)	$5.6 \pm 0.5^{****}$	
D122A	SST14	n.d. (3)	$6.2 \pm 0.3^{****}$	114.3 ± 12.2
	Lanreotide	n.d. (3)	$11.7 \pm 4.3^{****}$	
	Octreotide	$7.0 \pm 0.1, (3)$	$83.5 \pm 5.9^{*}$	
Q126A	SST14	$7.0 \pm 0.1, (3)$ **	92.9 ± 8.1	35.9 ± 1.3
	Lanreotide	n.d. (3)	$25.2 \pm 1.8^{****}$	
	Octreotide	n.d. (3)	n.d.	
F272A	SST14	n.d. (3)	n.d.	55.5 ± 6.5
	Lanreotide	n.d. (3)	$12.38 \pm 1.1^{****}$	
	Octreotide	$5.9 \pm 0.1, (3)$	98.7 ± 6.9	
F208A	SST14	$6.8 \pm 0.1, (3)$ ***	97.9 ± 3.6	68.6 ± 8.5
	Lanreotide	n.d. (3)	$27.9 \pm 4.0^{****}$	
	Octreotide	n.d. (3)	$22.3 \pm 5.4^{****}$	
N276A	SST14	6.4 ± 0.1 , (3) ****	$67.7 \pm 2.4^{****}$	52.7 ± 5.7
	Lanreotide	n.d. (3)	$13.8 \pm 1.6^{****}$	
	Octreotide	n.d. (3)	$13.7 \pm 3.2^{****}$	
Y302A	SST14	n.d. (3)	$28.3 \pm 4.4^{****}$	37.2 ± 3.5
	Lanreotide	n.d. (3)	$14.0 \pm 2.2^{****}$	
	Octreotide	5.9 ± 0.1, (3) ****	91.9 ± 6.5	
Q102S	Lanreotide	6.3 ± 0.1, (3) ****	$54.8 \pm 1.6^{****}$	80.1 ± 2.1
	Octreotide	n.d. (3)	n.d.	
F294S	Lanreotide	n.d. (3)	$11.45 \pm 0.8^{****}$	192.5 ± 5.3

	Octreotide	n.d. (3)	$17.9 \pm 2.9^{****}$	
N276Q	Lanreotide	n.d. (3)	$17.0 \pm 2.6^{****}$	4.0 ± 0.3