Opening opportunities for K_d determination and screening of MHC peptide complexes

Janine-Denise Kopicki^{1,2}, Ankur Saikia³, Stephan Niebling⁴, Christian Günther⁴, Raghavendra Anjanappa³, Maria Garcia-Alai⁴, Sebastian Springer^{3*}, Charlotte Uetrecht^{1,2,5*}

¹Centre for Structural Systems Biology (CSSB), Leibniz Institute of Virology (LIV), Notkestraße 85, 22607 Hamburg, Germany.

²University of Siegen, Am Eichenhang 50, 57076 Siegen, Germany.

³ Life Sciences and Chemistry, Jacobs University, Campus Ring 1, 28759 Bremen, Germany.

⁴Centre for Structural Systems Biology (CSSB), European Molecular Biology Laboratory – Hamburg Outstation, Notkestraße 85, Hamburg 22607, Germany.

⁵European XFEL GmbH, Holzkoppel 4, 22869 Schenefeld, Germany

* corresponding authors, email: charlotte.uetrecht@cssb-hamburg.de; s.springer@jacobs-university.de

MHC class I, native MS, peptide binding, peptide affinity, iDSF, antigen presentation, erucamide

THIS PDF FILE INCLUDES:

Figure S1. Small molecule tandem MS analysis verifies that erucamide accounts for a large proportion of the contaminants found in dsA2 samples.

Figure S2. Raw spectrum of dsA2/NV9.

Figure S3. Melting temperature (T_m) of dsA2 in presence of two corresponding truncated NV9 variants.

Figure S4. Overall area under the curve (AUC) for the detected dsA2 mass species at 25 V acceleration voltage.

Figure S5. Overall area under the curve (AUC) for the detected dsA2 mass species at 50 V acceleration voltage.

Table S1. Experimental masses and FWHM for dsA2 and different peptides obtained by native mass spectrometry.

Table S2. Overall area under the curve (AUC) for the detected dsA2 mass species at different acceleration voltages.

Table S3. Apparent dissociation constants (K_d) for dsA2 and different peptides obtained by native MS and iDSF.

Table S4. Melting temperatures (T_m) for dsA2 and different peptides obtained by nDSF.



Figure S1. Small molecule tandem MS analysis verifies that erucamide accounts for a large proportion of the contaminants found in dsA2 samples. Displayed is a spectrum resulting from the subtraction of the spectrum of plastic ware contaminants (empty vial) from the one of the dsA2 sample itself. Erucamide (337 Da) is hereby identified as the contaminant adducted to dsA2. Although oleamide is also found in the sample, there was no evidence of binding to the protein in the native MS analysis, unlike for erucamide.



Figure S2. Raw spectrum of dsA2/NV9. A native mass spectrum dsA2 in presence of NV9 (protein-peptide ratio 1:5) recorded at an acceleration voltage of 25 V is shown. The dsA2/NV9 is the predominant species (light blue). In addition, peptide-free dsA2 (yellow), dissociated β_2 m (olive) and heavy chain (dark green) as well as free NV9 can be seen.



Figure S3. Melting temperature (T_m) of dsA2 in presence of two corresponding truncated NV9 variants. Nanoscale differential scanning fluorimetry is employed to study thermal denaturation of dsA2 in presence of two peptides at once. 2 µM dsA2 are combined with either exclusively the *N*- (teal) or *C*-terminal peptide (blue) or both peptides together (purple) at different concentrations. Results for ΔT_m along with respective standard deviations are displayed.



Figure S4. Overall area under the curve (AUC) for the detected dsA2 mass species at 25 V acceleration voltage. The AUC is determined over the entire spectrum for the respective mass species at 25 V. The mean value of the AUC in absence or presence of the different peptides (protein-peptide ratio 1:5, 1:10:10 in dual peptide approach) from at least three independent measurements is depicted along with error bars that represent the corresponding standard deviation. "dsA2" (yellow bars) corresponds to the empty HLA-A*02:01(Y84C/A139C) disulfide mutant complex, "dsA2/pep" (light blue bars) to dsA2 bound to one peptide, "dsA2/pep" to dsA2 bound to two molecules of this certain peptide (dark blue bars), "dsA2/pep2" to dsA2 bound to two molecules of the second peptide (dark red bars), "dsA2/pep1/pep2" to dsA2 bound to two molecules of the second peptide (dark red bars), "dsA2/pep1/pep2" to dsA2 bound to two molecules (dark green bars) and "dsA2/erucamide" to dsA2 bound to the erucamide (coral bars) respectively.



Figure S5. Overall area under the curve (AUC) for the detected dsA2 mass species at 50 V acceleration voltage. The AUC is determined over the entire spectrum for the respective mass species 50 V. The mean value of the AUC in absence or presence of the different peptides (protein-peptide ratio 1:5, 1:10:10 in dual peptide approach) from at least three independent measurements is depicted along with error bars that represent the corresponding standard deviation. "dsA2" (yellow bars) corresponds to the empty HLA-A*02:01(Y84C/A139C) disulfide mutant complex, "dsA2/pep" (light blue bars) to dsA2 bound to one peptide, "dsA2/pepp" to dsA2 bound to two molecules of this certain peptide (dark blue bars), "dsA2/pep2" to dsA2 bound to two molecules of the second peptide (dark red bars), "dsA2/pep1/pep2" to dsA2 bound to two molecules of the second peptide (dark red bars), "dsA2/pep1/pep2" to dsA2 bound to two molecules (dark green bars) and "dsA2/erucamide" to dsA2 bound to the erucamide (coral bars) respectively.

Table S1. Experimental masses and FWHM for dsA2 and different peptides obtained by native mass spectrometry. Experimental masses (m_{exp}) of the different protein species of disulfide stabilized HLA-A*02:01(Y84C/A139C) disulfide mutant (dsA2) in absence or presence of the different peptides (protein-peptide ratio 1:5, 1:10:10 in dual peptide approach) are determined from at least three independent mass spectrometry measurements. They are listed together with the respective values for standard deviation *s* and average full width of the peak at half maximum (FWHM) along with the theoretically calculated molecular weight (*M*). FWHM values are given for the whole peak area where individual species are not fully resolved. ¹NV9 – high affinity control, ²YF9 – low affinity control, ³GV9 – minimal binding motif, ⁴Ac-NV9 – modified *N*-terminus, ⁵Ac-NV9 NH₂ – modified *N*- and *C*-terminus, ⁶NV9-NH₂ – modified *C*-terminus.

mass species	<i>M</i> /Da	m _{exp} /Da	s(m _{exp})/Da	FWHM/Da	s(m _{exp})/Da
dsA2 - M1	43,608	43,603	4	20	20
dsA2	43,739	43,733	4	3	2
heavy chain (hc)	31,877	31,873	2	2.1	0.3
β₂m chain - M1	11,731	11,729	2	1.1	0.4
β₂m chain	11,862	11,860	1	1.4	0.2
dsA2/NV9 ¹	44,682	44,678	1	3	2
dsA2/2×NV9	45,625	45,624	4	12	6
hc/NV9	32,820	32,816.9	0.8	2.2	0.3
dsA2/YF9 ²	44,856	44,849	1	7	3
dsA2/GV9 ³	44,369	44,363.8	0.7	2	1
dsA2/2×GV9	44,999	44,995	2	3	2
hc/GV9	32,507	32,503.0	0.5	2.08	0.09
dsA2/Ac-NV9 ⁴	44,724	44,718.3	0.5	1.6	0.3
dsA2/2×Ac-NV9	45,709	45,704	2	3	2
hc/Ac-NV9	32,862	32,857.3	0.5	1.9	0.1
dsA2/Ac-NV9-NH ₂ ⁵	44,723	44,717.4	1.0	1.4	0.1
dsA2/2×Ac-NV9-NH ₂	45,707	45,701.00	1.00	1.8	0.6
hc/Ac-NV9-NH ₂	32,861	32,857.0	0.9	1.7	0.2
dsA2/NV9-NH ₂ ⁶	44,681	44,676.4	0.5	1.38	0.06
dsA2/2×NV9-NH ₂	45,623	45,619.6	0.9	1.6	0.6
hc/NV9-NH ₂	32,819	32,815.1	0.9	1.72	0.10

mass species	<i>M</i> /Da	<i>m</i> _{exp} ∕Da	s(<i>m</i> _{exp})/Da	FWHM/Da	s(m _{exp})/Da
dsA2/NP4	44,181	44,176	3	10	10
dsA2/2×NP4	44,623	44,619	4	30	40
hc/NP4	32,319	32,317	2	20	40
dsA2/NM5	44,312	44,309	2	6	3
dsA2/2×NM5	44,885	44,883	2	60	50
hc/NM5	32,450	32,449	1	2.5	0.6
dsA2/NV6	44,411	44411	1	7	2
dsA2/2×NV6	45,083	45,082	3	145	6
hc/NV6	32,549	32,547	3	6	7
dsA2/NA7	44,481	44,476.4	0.5	5	3
dsA2/2×NA7	45,223	45,225	4	30	60
hc/NA7	32,619	32,615	2	2.4	0.5
dsA2/NT8	44,583	44,579.33	1.00	3	2
dsA2/2×NT8	45,427	45,425	2	7	5
hc/NT8	32,721	32,718.1	0.3	2.4	0.7
dsA2/LV8	44,568	44,565.8	0.7	2.6	0.7
dsA2/2×LV8	45,397	45,396.1	0.3	3.4	1.0
hc/LV8	32,706	32,704.8	0.4	2.18	0.04
dsA2/VV7	44,455	44,447.8	0.8	7	3
dsA2/2×VV7	45,171	45,166	3	3	1
hc/VV7	32,593	32,586	3	3	2
dsA2/PV6	44,356	44,350	3	5	3
dsA2/2×PV6	44,973	44969	4	100	50
hc/PV6	32,194	32,487	3	2.3	0.4
dsA2/MV5	44,259	44,254	3	8	12
dsA2/2×MV5	44,779	44,774	4	20	30

mass species	<i>M</i> /Da	<i>m</i> _{exp} /Da	s(m _{exp})/Da	FWHM/Da	s(m _{exp})/Da
hc/MV5	32,397	32,388.4	1.0	2.1	0.2
dsA2/VV4	44,127	44,125	5	10	10
dsA2/2×VV4	44,515	44,512	8	20	30
hc/VV4	32,265	32,255	1	10	3
dsA2/erucamide	44,077	44,071	5	10	20
dsA2/NP4/VV4	44,569	44,565	2	30	40
dsA2/NP4/MV5	44,701	44,697	2	4	5
dsA2/NP4/PV6	44,798	44,792	3	7	6
dsA2/NM5/VV4	44,700	44,699	2	30	40
dsA2/NM5/MV5	44,832	44,827	2	12	3
dsA2/NM5/PV6	44,929	44,927	2	100	40
dsA2/NV6/VV4	44,799	44,797	4	30	40
dsA2/NV6/MV5	45,451	44927	5	100	60

Table S2. Overall area under the curve (AUC) for the detected dsA2 mass species at different acceleration voltages. The AUC is determined over the entire spectrum for the respective mass species at 10 V, 25 V and 50 V. The mean value of the AUC in absence or presence of the different peptides (protein-peptide ratio 1:5, 1:10:10 in dual peptide approach) from at least three independent measurements is listed together with their corresponding standard deviation s. "dsA2" corresponds to the empty HLA-A*02:01(Y84C/A139C) disulfide mutant complex, "dsA2/pep" to dsA2 bound to one peptide, "dsA2/pep/pep" to dsA2 bound to two molecules of this certain peptide, "dsA2/pep2" to dsA2 bound to another peptide when two different peptides where present, "dsA2/pep2/pep2" to dsA2 bound to two molecules of the second peptide, "dsA2/pep1/pep2" to dsA2 bound to one molecule of each of both peptides and "dsA2/erucamide" to dsA2 bound to the erucamide respectively. ¹NV9 – high affinity control, ²YF9 – low affinity control, ³GV9 – minimal binding motif, ⁴Ac-NV9 – modified *N*-terminus, ⁵Ac-NV9 NH2 – modified *N*- and *C*-terminus, ⁶NV9-NH2 – modified *C*-terminus.

nontido	acc.	ds	42	dsA2	/pep	dsA2/p	ep/pep	dsA2	pep2	dsA2/pe	p2/pep2	dsA2/pe	p1/pep2	dsA2/eru	ucamide
peptide	volt.	AUC	S	AUC	S	AUC	S	AUC	s	AUC	S	AUC	S	AUC	S
	10 V	31%	2%	64%	3%	4%	4%							2%	2%
NV9 ¹	25 V	28%	4%	64%	5%	7%	6%							0.5%	0.5%
	50 V	59%	5%	40%	4%	1%	2%							0%	0%
	10 V	56%	3%	4%	2%	0%	0%							39%	2%
YF9 ²	25 V	51%	4%	5%	2%	0%	0%							44%	3%
	50 V	95%	5%	5%	2%	0%	0%							2%	2%
	10 V	52%	2%	43%	2%	1.5%	0.4%							4.3%	0.6%
GV9 ³	25 V	50%	1%	43%	3%	2.2%	0.9%							5%	2%
	50 V	62%	3%	35%	3%	1.1%	0.3%							1.7%	0.2%
	10 V	24%	2%	63%	3%	11%	2%							2.1%	0.5%
Ac-NV9 ⁴	25 V	23%	2%	64%	1%	9%	1%							3%	2%
	50 V	33%	2%	57%	2%	8%	1%							1.7%	0.9%
	10 V	55%	2%	27.9%	0.5%	2.4%	0.6%							15%	1%
Ac-NV9-NH2 ⁵	25 V	57%	2%	28%	1%	1%	1%							14%	1%
	50 V	72%	3%	25%	3%	1%	1%							2.2%	0.4%
	10 V	36%	2%	59%	2%	5%	1%							0.4%	0.2%
NV9-NH2 ⁶	25 V	36%	2%	58%	2%	5%	1%							0.3%	0.3%
	50 V	61%	1%	38%	1%	1.6%	0.3%							0.1%	0.2%

nontido	acc.	ds/	42	dsA2	/pep	dsA2/p	ep/pep	dsA2	/pep2	dsA2/pe	ep2/pep2	dsA2/pe	p1/pep2	dsA2/eru	ucamide
peptide	volt.	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S
	10 V	55%	2%	11%	1%	0.6%	0.2%							33.8%	0.6%
NP4	25 V	57.8%	0.9%	10.4%	0.5%	0.69%	0.06%							31.1%	0.5%
	50 V	92.4%	0.5%	2.6%	0.1%	0.8%	0.1%							4.3%	0.3%
	10 V	62%	2%	17.9%	0.6%	1.0%	0.2%							19%	1%
NM5	25 V	64%	3%	18.0%	0.7%	1.2%	0.2%							17%	2%
	50 V	90.3%	0.6%	8.1%	0.3%	0.3%	0.6%							1.3%	0.1%
	10 V	66%	1%	10%	2%	0.5%	0.1%							24%	1%
NV6	25 V	66%	1%	11%	2%	0.69%	0.08%							22%	2%
	50 V	91.2%	0,5%	4.1%	0.8%	1.9%	0.3%							2.9%	0.4%
	10 V	75%	3%	16%	2%	0.8%	0.3%							8%	1%
NA7	25 V	68%	3%	19%	2%	1.4%	0.7%							11%	1%
	50 V	86%	4%	12%	3%	0.5%	0.5%							1.8%	0.2%
	10 V	47.3%	0.9%	28%	1%	0.6%	0.5%							24%	1%
NT8	25 V	50%	3%	28.2%	0.9%	1%	1%							21%	1%
	50 V	75%	2%	23%	1%	0.2%	0.3%							1.6%	0.3%
	10 V	42%	6%	54%	5%	2.8%	0.3%							1.7%	0.9%
LV8	25 V	40%	3%	54%	3%	3.9%	0.8%							1.5%	0.7%
	50 V	53%	2%	45%	2%	1.6%	0.2%							0.4%	0.4%
	10 V	61.8%	0.5%	17%	2%	1.7%	0.3%							19%	3%
VV7	25 V	62%	3%	17%	3%	1.16%	0.06%							20.2%	0.7%
	50 V	96%	3%	3%	2%	0%	0%							1.3%	0.6%
	10 V	68%	6%	12%	2%	0.5%	0.6%							19%	3%
PV6	25 V	68%	6%	13%	3%	0.5%	0.2%							19%	3%
	50 V	90%	3%	5%	1%	0.1%	0.1%							5%	1%

nontido	acc.	ds	42	dsA2	/pep	dsA2/p	pep/pep	dsA2/	pep2	dsA2/pe	ep2/pep2	dsA2/pe	ep1/pep2	dsA2/eru	Icamide
peptide	volt.	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S
	10 V	64%	3%	23%	2%	3.2%	0.7%							10.4%	0.2%
MV5	25 V	63%	4%	23%	2%	3.7%	1.0%							10.4%	0.4%
	50 V	80%	3%	15%	2%	1.5%	0.6%							3.3%	0.3%
	10 V	68%	1%	14%	1%	0%	0%							18.093%	0.007%
VV4	25 V	68%	2%	14%	1%	0%	0%							17.6%	1.0%
	50 V	92%	1%	4.1%	1.0%	0%	0%							4.0%	0.4%
NP4	10 V	54%	2%	9%	3%	1.1%	0.5%	10%	1%	1.2%	1.0%	2.1%	0.5%	22%	4%
+	25 V	56%	2%	8%	2%	0.5%	0.5%	10%	1%	0.7%	0.7%	1.6%	0.5%	22%	4%
VV4	50 V	87%	2%	5%	1%	0%	0%	4.4%	0.6%	0%	0%	0%	0%	3.1%	0.4%
NP4	10 V	42.9%	0.9%	6%	2%	1,8%	0.3%	27%	1%	5.6%	0.3%	5.2%	0.8%	10%	1%
+	25 V	43%	2%	5%	1%	1%	2%	27%	2%	6.4%	0.6%	5.3%	0.9%	10.5%	0.8%
MV5	50 V	76%	5%	3%	1%	0.5%	0.9%	14%	2%	1.9%	0.8%	1%	1%	1.8%	0.3%
NP4	10 V	48%	3%	8%	1%	1.7%	0.3%	13%	2%	0.9%	0.8%	2.8%	0.2%	25%	2%
+	25 V	49%	1%	8.5%	0.6%	1.4%	0.2%	13.6%	0.5%	0.4%	0.7%	2.1%	0.1%	24.5%	0.9%
PV6	50 V	90.8%	0.3%	3.0%	0.3%	0%	0%	3.5%	0.1%	0%	0%	0%	0%	2.5%	0.1%
NM5	10 V	56%	3%	13%	2%	1%	1%	8.9%	0.8%	0.5%	0.8%	1.8%	1.6%	18%	4%
+	25 V	55.7%	0.7%	14.2%	0.9%	0%	0%	8.6%	0.4%	0%	0%	2.0%	1.9%	19%	2%
VV4	50 V	86%	4%	8%	1%	0%	0%	2.8%	1.0%	0%	0%	0%	0%	4%	2%
NM5	10 V	46%	7%	8%	3%	5%	4%	20%	1%	4.9%	0.8%	6%	1%	9%	1%
+	25 V	51%	9%	6%	3%	3%	3%	20%	3%	4.5%	0.5%	5%	3%	9%	2%
MV5	50 V	75%	7%	6%	5%	0%	0%	11.7%	0.4%	4%	2%	2%	2%	1%	1%
NM5	10 V	58%	2%	11.9%	0.6%	1%	2%	9%	2%	0%	0%	0.4%	0.7%	18%	1%
+	25 V	60.3%	0.5%	12.0%	0.8%	1%	1%	8.9%	0.4%	0%	0%	0.4%	0.7%	17.3%	0.4%
PV6	50 V	91.1%	0.7%	5.6%	0.2%	0%	0%	2.4%	0.2%	0%	0%	0%	0%	0.9%	0.8%

nontido	acc.	ds	42	dsA2	/pep	dsA2/p	ep/pep	dsA2/	pep2	dsA2/pe	p2/pep2	dsA2/pe	p1/pep2	dsA2/eru	ucamide
peptide	volt.	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S	AUC	S
NV6	10 V	47%	8%	6%	1%	1%	0%	17%	4%	7%	4%	3%	0%	18%	1%
+	25 V	48%	4%	6%	0%	1%	2%	16%	2%	5%	1%	2%	1%	21%	3%
VV4	50 V	81%	7%	3%	1%	0%	0%	7%	4%	2%	1%	0%	0%	7%	2%
NV6	10 V	47%	5%	8%	2%	4%	2%	21%	1%	6.5%	0.8%	3.0%	0.3%	10%	2%
+	25 V	42%	8%	9%	3%	4%	2%	20%	3%	8%	2%	5%	3%	10.2%	0.8%
MV5	50 V	71%	5%	6%	2%	1%	1%	12.5%	0.9%	2.1%	0.7%	1.4%	0.6%	6%	1%

Table S3. Apparent dissociation constants (*K***_d) for dsA2 and different peptides obtained by native MS and iDSF.** The *K*_d is calculated from the respective area under the curve values at 10 V acceleration voltage (protein-peptide ratio 1:5). *K*_{d,high} is an affinity determined on the basis of a real experiment at a cone voltage of 150 V, while a theoretical cone voltage of 36 V is assumed for *K*_{d,low} to correct for ion-source decay. *K*_{d,iDSF} is derived by two independent measurements. Protein concentration is 2.2 µM. For each ligand, a two-fold serial dilution series is prepared using 11 concentrations depending on their predicted or assumed *K*_d range. The listed standard deviation *s* for *K*_d is determined using common equations, which estimate the propagation of uncertainty. ¹NV9 – high affinity control, ²GV9 – minimal binding motif, ³Ac-NV9 – modified *N*-terminus, ⁴Ac-NV9 NH2 – modified *N*- and *C*-terminus, ⁵NV9-NH2 – modified *C*-terminus; *iDSF reaches its limits at affinities below 200 nm, hence the values of grayed-out peptides are not reliable.

nontido		K _{d,hi}	gh	K d,I	ow	K _{d,iDSF}		
peptide	Sequence	<i>K</i> _d /μM	s/μM	<i>K</i> _d /μM	s/μM	<i>K</i> _d /μM	s/μM	
NV9 ¹	NLVPMVATV	8	2	0.06	0.08	0.04*	0.01	
GV9 ²	GLGGGGGGV	35	2	0.5	0.2	0.36	0.06	
Ac-NV9 ³		4.5	1.0	0.11	0.05	0.61	0.08	
Ac-NV9-NH2 ⁴		80	7	7	3	4	1	
NV9-NH2⁵		10	1	0.004	0.003	0.001*	0.001	
NP4	NLVP	350	60	100	70	50	20	
NM5	NLVPM	180	30	15	6	11	3	
NV6	NLVPMV	380	70	30	10	9	2	
NA7	NLVPMVA	210	40	2.0	0.7	3.6	0.5	
NT8	NLVPMVAT	92	5	30	10	2.6	0.4	
LV8	LVPMVATV	17	3	0.07	0.06	0.008*	0.005	
VV7	VPMVATV	180	30	15	7	7.8	1.0	
PV6	PMVATV	300	200	15	7	15	2	
MV5	MVATV	110	10	3	1	1.6	0.2	
VV4	VATV	270	30	13	4	50	20	
NP4+VV4				90	30			
NP4+MV5				11	3			
NP4+PV6				130	50			
NM5+VV4				50	20			
NM5+MV5				9	3			

peptide	peptide sequence	K _{d,hi}	gh	K _{d,l}	ow	<i>K</i> d,iDSF		
peptide		<i>K</i> _d /μM	s/μM	<i>K</i> ₀/μM	s/µM	<i>K</i> ₀/μM	s/μM	
NM5+PV6				50	10			
NV6+VV4				50	10			
NV6+MV5				13	4			

Table S4. Melting temperatures (T_m **) for dsA2 and different peptides obtained by nDSF.** The T_m as well as the resulting s for dsA2 in absence or presence of the different peptides is defined by at least two independent measurements. Protein concentration is 2 μ M. ¹NV9 – high affinity control, ²YF9 – low affinity control, ³GV9 – minimal binding motif, ⁴Ac-NV9 – modified *N*-terminus, ⁵Ac-NV9 NH2 – modified *N*- and *C*-terminus, ⁶NV9-NH2 – modified *C*-terminus.

nontido	0.2	JM	2 µ	Μ	20 µ	M	1 m	Μ	2 m	M
peptide	T _m /°C	s/°C	T _m /°C	s/°C	<i>T</i> _m /°C	s/°C	<i>T</i> _m /°C	s/°C	<i>T</i> _m /°C	s/°C
empty	35.7	0.6								
NV9 ¹	35.8	0.8	58.96	0.07	59.1	0.1	60.698	0.007		
YF9 ²	36.1	0.8	36.1	0.8	36.2	0.8				
GV9 ³	36.4	0.6	38.8	0.4	42.1	0.2	47.6	0.1		
Ac-NV9 ⁴	36.6	0.7	39.6	0.6	43.2	0.6	48.420	0.001		
Ac-NV9-NH2 ⁵	36.0	0.8	36.6	0.4	38.4	0.3				
NV9-NH2 ⁶	36.0	0.9	46.9	0.2	50.3	0.3	55.53	0.04		
NP4	35.6	0.4	35.7	0.5	35.8	0.4	38.95	0.04	42.10	0.06
NM5	35.6	0.5	35.9	0.4	37.3	0.3	45.04	0.07	45.93	0.03
NV6	35.5	0.1	35.9	0.2	37.61	0.05	44.1	0.6	44.6	0.2
NA7	35.51	0.09	35.9	0.1	37.95	0.06	45.442	0.007		
NT8	35.5	0.1	36.02	0.09	38.78	0.02	46.89	0.02		
LV8	35.40	0.08	43.5	0.1	47.28	0.04	53.30	0.04		
VV7	35.5	0.1	35.7	0.2	37.0	0.1	42.29	0.01		
PV6	35.5	0.2	35.62	0.09	36.40	0.02	42.2	0.3	43.6	0.5
MV5	35.64	0.08	36.5	0.1	39.31	0.07	47.90	0.01	49.253	0.003
VV4	35.475	0.002	35.1	0.6	35.5	0.2	41.47	0.03	42.6	0.1
NP4+VV4							44.99	0.05	47.93	0.10
NP4+MV5							47.55	0.06	50.35	0.05
NP4+PV6							43.88	0.05	46.47	0.08
NM5+VV4							47.22	0.05	47.26	0.08
NM5+MV5							47.59	0.03	49.900	0.002
NM5+PV6							45.45	0.02	47.295	0.010

nontido	0.2	JM	2 µM		20 µ	IM	1 m	М	2 mM	
peptide	T _m /°C	s/°C	<i>T</i> _m /°C	s/°C	T _m /°C	s/°C	T _m /°C	s/°C	T _m /°C	s/°C
NV6+VV4							44.14	0.05	46.3	0.2
NV6+MV5							47.36	0.04	49.84	0.02
NV6+PV6							44.8	0.1	46.6	0.1