Fine-Mapping Studies Distinguish Genetic Risks for Childhood- and Adult-Onset Asthma in the HLA Region

Supplementary Methods, Figures, and Tables

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Table of Contents

(3) Supplementary Methods

- (4) Fine Mapping the HLA Region
- (4) HLA Fine-Mapping Simulations
- (6) Gene Expression and eQTL Studies

(8) Supplementary Figures

- (9) Fig. S1. Ancestry PCs for the Replication and Discovery Cohorts
- (10) Fig. S2. HLA Allele Associations
- (11) Fig. S3. Amino Acid Associations
- (13) Fig. S4. Fine-Mapping Simulations in the HLA Region
- (15) Fig. S5. Expression of HLA-DQB2 and HLA-DQA2
- (16) Fig. S6. ENCODE ChromHMM Results for SNPs in the Childhood-Onset and Adult-Onset Credible Sets
- (18) Fig. S7. Replication Results

(20) Supplementary Tables

- (21) Table S1. RNA-seq Sample Composition
- (22) Table S2. HLA Allele Associations*
- (23) Table S3. Allele Associations: Additive vs. Dominant Model*
- (24) Table S4. HLA Heterogeneity Test*
- (25) Table S5. HLA Amino Acid Polymorphism Associations*
- (26) Table S6. Putatively Causal Variants and Allergy
- (27) Table S7. Putatively Causal Variants and Sex
- (28) Table S8. SuSiE Credible Set Results*
- (29) Table S9. HLA Allele Frequencies by Study*
- (30) Table S10. List of SNPs in the Credible Sets Excluded from eQTL Analyses*
- (31) Table S11. eQTL Results for All Credible Set SNPs*
- (32) Table S12. HLA Region eQTLs
- (33) Table S13. eQTL Fine-Mapping Results*
- (34) Table S14. Amino Acid Associations with the Highest PIPs in Each Credible Set
- (35) Table S15. Amino Acids in the Credible Sets and Their Corresponding HLA Alleles*
- (36) Table S16. Average r² Between Childhood-Onset and Adult-Onset Asthma SNPs
- (37) Table S17. Marginal vs. Conditional Association Results
- (38) Table S18. Sample Composition of the Replication Cohort
- (39) Table S19. Self-Reported Ethnic Composition of the Replication Cohort
- (40) Table S20. Replication Meta-Analysis Results*

* Tables are available in Additional File 2

(41) Supplementary References

Supplementary Methods

Fine Mapping the HLA Region

We used Sum of Single Effects (SuSiE(1)) (susieR R package version 0.9.0) to fine map the HLA loci for childhood-onset asthma (COA) and adult-onset asthma (AOA). The susieR R package does not currently allow for the inclusion of covariates, so sex and the first 10 ancestral principal components (PCs) were regressed out of the genotype matrix and phenotype vector using linear regression; we used the residuals of the genotype matrix and phenotype vector as inputs to SuSIE. The SuSiE method is based on a linear regression, and so when applied to binary data, it will estimate and test for effects in terms of risk differences, rather than the more conventional odds ratio (OR). Applying linear methods to binary data is justified here because the estimated ORs were all small (<1.3), the allele frequencies were not too extreme, and the sample size (here, limited by the smaller number of cases) was large (2-4). See Pirinen et al. Section 3 for detailed discussion of applying linear methods to binary data and the relationship between estimated risk differences and ORs. We assumed at most L = 10 causal variants and set susieR to estimate the residual and prior variances. We retained only level-95% credible sets (coverage = 0.95). We took the additional step of discarding credible sets in which the "purity" (smallest absolute correlation among all pairs of variants within the credible set) was less than 0.50. We only considered credible sets that contained at least one variant reaching genome-wide significance to avoid any possible artifacts.

HLA Fine-Mapping Simulations

Because the HLA region is extraordinarily complex, we assessed the performance of SuSiE in this region by simulation. Existing genotype and covariate data were used to leverage the true correlation structure in the class I and class II regions to simulate both binary (e.g. case/control status) and quantitative (e.g. gene expression) outcomes.

To simulate binary (e.g. case/control status) outcomes, we used the genotype matrix X (HLA class I or class II loci defined by Pividori *et al.*(5)) and covariate matrix Z from the UK Biobank and set the individual-level log-odds of asthma to be

$$\ln \frac{p_i}{1-p_i} = \sum_j \beta_j X_{ij} + \sum_k \delta_k Z_{ik} + \alpha$$

for individual *i*, SNPs *j*, covariates *k*, fixed effect vectors β and δ , and a fixed intercept α . We used the true matrix of covariates and covariate effects δ estimated from a logistic regression, separately for COA and AOA simulations. β_j was set to 0 for all non-causal variants. For causal variants, β_j was set using effect sizes similar to what was found in the Pividori COA and AOA GWASs(5). We randomly selected 0-3 variants from a random uniform distribution to be causal (with non-zero effects) for both the class I and class II regions using both COA and AOA effect sizes. We simulated case/control status for each individual as $Y_i \sim Bernoulli(p_i)$ independently and regressed out the covariates in Z from X and Y.

For quantitative outcomes (e.g. gene expression), we used *X* and *Z* in the HLA class I and HLA class II regions from the nasal epithelial cell (NEC) dataset from URECA described below. We set the individual-level mean to be

$$\mu_i = \sum_j \beta_j X_{ij} + \sum_k \delta_k Z_{ik} + \alpha$$

and we used the true matrix of covariates and effects δ estimated from a linear regression. β_j was set to 0 for all non-causal variants, and causal β_j were set using effect sizes similar to what was found from the NEC eQTL studies (described below). We similarly randomly selected 0-3 causal signals in both the class I and class II regions and set the individual level response to be $Y_i \sim N(\mu_i, \sigma^2)$. We similarly regressed out the covariates in Z from X and Y. These simulations were used to test how well SuSiE recovers the causal effects over each simulation.

Gene Expression and eQTL Studies

Lymphoblastoid Cell Lines (LCLs)

We examined RNA-seq data previously collected from LCLs from 398 Hutterites(6). The Hutterites are a founder population of European descent with well characterized HLA types for the polymorphic *HLA-A*, *HLA-B*, *HLA-C*, *HLA-E*, *HLA-G*, *HLA-DPB1*, *HLA-DRB1*, *HLA-DQB1*, and *HLA-DQA1* genes(7). The sample was composed of 191 males and 207 females who were between the ages of 10 and 60 at the time of sample collection. Informed consent was obtained from all participants under University of Chicago IRB-approved protocols.

Standard RNA-seq pipelines that map reads to a reference genome can provide biased expression estimates at the highly polymorphic HLA loci due to the potentially large number of differences between the sequence of an individual's HLA type and the reference sequence used for mapping(8,9). Expression estimates can be improved by mapping RNA-seq reads to the sequences for each individual's known HLA type(8). For the polymorphic HLA genes, we aligned RNA-seq reads to reference sequences from the IMGT database(10) for each individual's known HLA type, removing duplicate reads with WASP(11). Sequencing reads were mapped and quantified using STAR/2.6.1(12) for other genes. Samples with >7M uniquely mapped reads underwent trimmed means of M-value (TMM) normalization and voom transformation(13). We corrected for extraction date and sequencing batch with limma(14).

To perform eQTL mapping, associations between SNPs and expression of genes in the HLA class I and class II regions were performed with Genome-wide Efficient Mixed Model Association (GEMMA)(15) using a kinship matrix to correct for relatedness between Hutterite individuals. We used a linear mixed model (LMM) including age and sex as covariates and considered all variants within 1 Mb of the transcription start site (TSS) of each expressed gene.

Peripheral Blood Mononuclear Cells (PBMCs)

We examined unstimulated PBMC RNA-seq data from 132 (78 males, 54 females) African-American children from the URban Environment and Childhood Asthma (URECA) birth cohort who were 2 years old at the time of sample collection(16,17). Whole genome sequencing (WGS) was performed using the Illumina NovaSEQ6000 with 150 bp paired-end reads. Reads were aligned to the GRCh38 human reference genome (including alternate loci and decoy contigs) using BWA-MEM(18) (Burrows-Wheeler Aligner; v0.7.17). Aligned reads underwent duplicate removal (Picard MarkDuplicates v2.8.1) and base quality score recalibration (GATK BaseRecalibrator; v3.8) against known sites (dbSNP138, known indels, and Mills and 1 KG gold standard indels) provided in the GATK resource bundle(19). Reads that mapped to the primary HLA region (chr6:28510120-33480577), reads that mapped to the GRCh38 HLA contigs, and unmapped reads were used for WGS HLA typing. We used HLA-LA(20) to infer HLA types from WGS for *HLA-A, HLA-B, HLA-C, HLA-E, HLA-F, HLA-DRB1, HLA-DQB1, HLA-DQA1, HLA-DPB1*, and *HLA-DPA1*. Reads were mapped and normalized as previously described. To perform eQTL mapping, we examined linear regressions with QTLtools(21), using a nominal pass and *cis*-window size of 1 Mb around the TSS. We included sex, collection site, the first three ancestral PCs, and 19 latent factors(23) to account for unwanted variation as covariates in the analysis.

Nasal Epithelial Cells (NECs)

We examined NEC RNA-seq data from 188 (92 females, 96 males) African-American children (age 11 at time of sample collection) from the URECA cohort(24). As described above for the PBMCs, we used HLA-LA to infer HLA types from whole-genome sequences, mapped RNA-seq reads as described above, and used QTLtools to perform eQTL mapping, using sex, the first three ancestral PCs, collection site, epithelial cell proportion, sequencing batch, and seven latent factors(23) as covariates in the analysis.

Supplementary Figures



Fig. S1. Ancestry PCs for the Replication and Discovery Cohorts.

Ancestry PC1, PC2, and PC3 are shown for the discovery cohort ("White British (discovery)")

and the replication cohort, with the colors corresponding to self-reported ancestry.



Fig. S2. HLA Allele Associations.

Odds ratios and 95% confidence intervals are shown for the HLA alleles that were significantly associated ($p<5.0x10^{-8}$) with either childhood-onset asthma (COA, blue) and/or adult-onset asthma (AOA, red). The results for all alleles for the six HLA loci are shown in Table S2.



Fig. S3. Amino Acid Associations.

Odds ratios and 95% confidence intervals are shown for HLA amino acid polymorphisms that were significantly associated ($p < 5x10^{-8}$) with either childhood-onset asthma (COA, blue) and/or adult-onset asthma (AOA, red).





Each panel is a simulation. The top four rows are the simulated binary ("GWAS") traits and the bottom two rows are the simulated quantitative ("eQTL") traits. Simulations were performed for both the class I and class II regions separately. The binary outcomes were also simulated using covariate effects estimated for either childhood-onset asthma (COA) or adult-onset asthma (AOA) (see Supplementary Methods for more details). C refers to the number of causal variants (0-3). The colors represent the credible sets detected by SuSiE, with the designated causal

effect variant(s) in red. SuSiE correctly identified the accurate number of causal signals and reported a true causal signal in each credible set in all the simulations.



Fig. S5. Expression of *HLA-DQB2* and *HLA-DQA2*.

Normalized expression of each gene by the number of asthma-risk alleles for rs9272346 (A) for lymphoblastoid cell lines (LCLs) and peripheral blood mononuclear cells (PBMCs) and rs9274660 (G) for nasal epithelial cells (NECs), which were representative class II AOA CS1 SNPs.



b Class I COA CS2



2 kb⊣

32,885,000 l

32,855,000 |

c Class II COA CS1



d Class II COA CS2



e Class II AOA CS1



f Class II AOA CS2



Fig. S6. ENCODE ChromHMM Results for SNPs in the Childhood-Onset Asthma and

Adult-Onset Asthma Credible Sets.

Vertical red line indicates the location of each SNP. Layered H3K4Me1, H3K4ME3, and H3K27Ac marks and ChromHMM states are shown for the GM12878 cells. Red: active promoter, light red: weak promoter, purple: inactive/poised promoter, orange: strong enhancer, yellow: weak/poised enhancer, blue: insulator, dark green: transcriptional transition/elongation, light green: weak transcribed, gray: polycomb-repressed, light gray: heterochromatin/low signal. Asterisk denotes rsid with the highest PIP. **a)** rs2428494 (shared class I CS) was predicted to reside in a weak promoter, **b)** rs28481932 (class I COA CS2) in a weakly transcribed region, and **c)** rs28407950 (class II COA CS1) in an strong enhancer. **d)** Class II COA CS2 SNPs were predicted to reside in polycomb-repressed, active promoter, polycomb-repressed, insulator, and weakly transcribed regions (from left to right). **e)** Class II AOA CS1 SNPs (see main manuscript for a discussion of these results). The red or orange mark next to the rsid indicates it is predicted to reside in an active promoter or strong enhancer, respectively. Magenta ^ indicates if it was an eQTL in our study. **f)** Class II AOA CS2 SNPs. Figures created from http://genome.uscs.edu(25).



Fig. S7. Replication Results.

Odds ratios and 95% confidence intervals are shown for the candidate variant allele or amino acid polymorphism in the discovery COA and AOA CSs for self-reported White (British, Irish, White, Any other White background), self-reported Black or Black British, self-reported Asian or

Asian British, entire replication cohort (consisting of White, Black or Black British, Asian or Asian British), and the White British discovery cohort.

Supplementary Tables

Table S1. RNA-seq Sample Composition.

Cells	Study	Ancestry	Sample Size	Age (yrs)	Sex
LCLs (EBV-transformed B cells)	Hutterites	European American	398	10-60	191 M, 207 F
Peripheral blood mononuclear cells (PBMCs)	URECA	African American	132	2	78 M, 54 F
Nasal epithelial cells (NECs)	URECA	African American	188	11	96 M, 92 F

Sample composition from RNA-seq data collected from LCLs (Hutterites(6)) and the

PBMCs(16,17) and NECs(24) from URECA (URban Environment and Childhood Asthma).

Table S2. HLA Allele Associations

Table S3. Allele Associations: Additive vs. Dominant Model

Table S4. HLA Heterogeneity Test

Table S5. HLA Amino Acid Polymorphism Associations

CS	Var	Original	Original	Allergy Cov	Allergy	No allergy	No allergy	Interaction
		p-value	OR	p-value	Cov	p-value	OR	p-value
					OR			
		1	Child	nood-Onset Astl	hma			1
Class I CS1	rs2428494	8.77x10 ⁻²³	1.157	4.97x10 ⁻²¹	1.151	6.11x10 ⁻¹⁶	1.160	0.494
Class I CS2	HLA-C p.11	3.12x10- ¹⁹	1.241	2.12x10 ⁻¹⁸	1.236	7.84x10 ⁻¹²	1.224	0.562
Class II CS1	rs28407950	1.37x10- ⁵⁹	1.355	1.31x10 ⁻⁵⁴	1.339	7.74x10 ⁻⁴¹	1.361	0.219
Class II CS2	rs35571244	1.07x10 ⁻¹⁷	1.253	3.21x10 ⁻¹⁶	1.242	2.78x10 ⁻¹¹	1.243	0.9995
			Adı	ult-Onset Asthm	a	1		
Class I CS1	rs2428494	4.52x10 ⁻²³	1.104	3.13x10 ⁻²¹	1.100	1.66x10 ⁻¹⁶	1.099	0.939
Class II CS1	rs9272346	1.98x10 ⁻⁴⁷	1.163	1.42x10 ⁻⁴⁷	1.164	1.82x10 ⁻³⁷	1.166	0.77
Class II CS2	DQA1*0301	6.91x10 ⁻⁴⁷	1.187	1.72x10 ⁻⁴⁸	1.202	1.62x10 ⁻⁴⁰	1.211	0.352

Table S6. Putatively Causal Variants and Allergy

Each row is a result for each variant reflecting each credible set (CS). The original p-value and odds ratio (OR) for the risk allele are shown, then the p-value and OR when allergy was included as a covariate in the regression ("Allergy Cov"), when all individuals with allergy were excluded ("No allergy"), and the interaction between the variant and allergy status are shown.

CS	Var	Original p-	Original	Females p-	Females	Males p-	Males	Interaction
		value	OR	value	OR	value	OR	p-value
		•	Childho	ood-Onset Asth	nma			·
Class I CS1	rs2428494	8.77x10 ⁻²³	1.157	3.46x10 ⁻⁰⁹	1.146	3.71x10 ⁻¹⁵	1.164	0.613
Class I CS2	HLA-C p.11	3.12x10 ⁻¹⁹	1.241	4.59x10 ⁻¹²	1.302	3.68x10 ⁻⁰⁹	1.200	0.098
Class II CS1	rs28407950	1.37x10 ⁻⁵⁹	1.355	2.87x10 ⁻³³	1.426	6.08x10 ⁻²⁹	1.308	0.024
Class II CS2	rs35571244	1.07x10- ¹⁷	1.253	3.85x10 ⁻⁰⁸	1.253	4.94x10 ⁻¹¹	1.253	0.996
		·	Adul	t-Onset Asthm	a			·
Class I CS1	rs2428494	4.52x10 ⁻²³	1.104	6.63x10 ⁻¹⁴	1.099	9.69x10 ⁻¹¹	1.112	0.575
Class II CS1	rs9272346	1.98x10 ⁻⁴⁷	1.163	3.02x10 ⁻³¹	1.165	7.18x10 ⁻¹⁸	1.159	0.812
Class II CS2	DQA1*0301	6.91x10 ⁻⁴⁷	1.187	1.45x10 ⁻³⁴	1.202	2.32x10 ⁻¹⁴	1.162	0.174

Table S7. Putatively Causal Variants and Sex

For each variant, the p-value and ORs for the risk allele are shown for the original analysis ("Original"), in just female participants, and in just males. The p-value for the interaction between sex and the variant is also shown.

Table S8. SuSiE Credible Set Results

Table S9. HLA Allele Frequencies by Study

Table S10. List of SNPs in the Credible Sets Excluded from eQTL Analyses

Table S11. eQTL Results for All Credible Set SNPs

Table S12. HLA Region eQTLs

CS	LCLs (n=398, European American)	PBMCs (n=133, African American)	NECs (n=189, African American)
Class I COA CS1/AOA CS1	HLA-B MIR6891	No eQTLs	HLA-B ¥
Class I COA CS2	CCHCR1 * AL662844.4 *	No eQTLs	No eQTLs
Class II COA CS1	HLA-DQA2 * HLA-DQB2 * HLA-DRB5 * HLA-DQB1-AS1 *	HLA-DQA2 HLA-DQB2	HLA-DQB2
Class II COA CS2	HLA-DQA2 HLA-DPB2 HLA-DRB9 PSMB9 TAP2	No eQTLs	No eQTLs
Class II AOA CS1	HLA-DQA2 HLA-DQB2 HLA-DRB6 HLA-DRB9 TAP2	HLA-DQA2 HLA-DQB2 HLA-DRB6	HLA-DQB1 HLA-DQA2 HLA-DQB2 HLA-DRB6
Class II AOA CS2	HLA-DQA2 HLA-DQB2 HLA-DRB5 HLA-DRB6 PPT2 TAP1	HLA-DQA2	HLA-DQA1 HLA-DQA2

For all SNPs in each credible set (CS) and each gene within 1 Mb of the transcription start site (TSS) examined across the three datasets (lymphoblastoid cell lines [LCLs], peripheral blood mononuclear cells [PBMCs], and nasal epithelial cells [NECs], the genes with eQTLs at FDR < 0.05 are shown. n.i., no information for the SNP (Table S6). These genes were then included in the eQTL fine-mapping studies. * analyses performed in a subset of individuals who had genotypes for all SNPs in the CS. * analyses performed in a subset of individuals with genotypes from MEGA array

Table S13. eQTL Fine-Mapping Results

CS	HLA Locus	Pos	Ref	Alt	Frequency	p-value	OR	95% CI	PIP	Secondary Structure
Class I COA CS2	HLA-C	11	Ala	Ser	0.13	3.12x10 ⁻¹⁹	0.806	0.768-0.844	0.573	Beta Strand
Class II AOA CS1	HLA-DQB1	55	Lys, Pro	Arg	0.41	4.50x10 ⁻⁴⁹	0.858	0.841-0.876	0.039	Alpha Helix
		26	Thr	Ser	0.20	3.63x10 ⁻⁴⁷	1.187	1.160-1.215	0.066	Beta Strand
Class II		47	Cys	Gln	0.20	3.63x10 ⁻⁴⁷	1.187	1.160-1.215	0.066	-
AOA CS2	HLA-DQA1	56	Gly,x	Arg	0.20	3.63x10 ⁻⁴⁷	1.187	1.160-1.215	0.066	-
		76	lle	Val	0.20	3.63x10 ⁻⁴⁷	1.187	1.160-1.215	0.066	Alpha Helix
		187	Ala	Thr	0.20	3.63x10 ⁻⁴⁷	1.187	1.160-1.215	0.066	-

Table S14. Amino Acid Associations with the Highest PIPs in Each Credible Set.

For each of the putatively causal amino acid polymorphisms in the credible sets (CSs), the position (Pos), reference (Ref) or alternative (Alt) amino acid, frequency, p-value, odds ratio (OR), 95% confidence interval (CI), posterior inclusion probability (PIP), and secondary structure are described. A dash ("-") indicates that the variant is unlikely to reside in a functional domain.

Table S15. Amino Acids in the Credible Sets and Their Corresponding HLA Alleles

						HLA-
	rs2428494	HLA-C p.11	rs28407950	rs35571244	rs9272346	DQA1*0301
	(Class I COA	(Class I COA	(Class II	(Class II	(Class II	(Class II
Variant	AOA CS1)	CS2)	COA CS1)	COA CS2)	AOA CS1)	AOA CS2)
rs2428494	1.00	0.0085	0.0154	0.0112	0.0739	0.0088
HLA-C p.11		1.00	0.0522	0.0017	0.0162	0.0055
rs28407950			1.00	0.0097	0.5013	0.0814
rs35571244				1.00	0.0240	0.1585
rs9272346					1.00	0.1622
HLA-						1 00
DQA1*0301						1.00

Variant	Marginal								
variant	warginai	Association	Conditiona	Association					
Childhood-Onset Asthma									
	p-value	OR [95% Cl]	p-value	OR 95% CI]	Conditioned on				
rs2428292 (class I CS1)	8.77x10 ⁻²³	1.16 [1.12-1.19]	5.05x10 ⁻¹³	1.11 [1.08-1.15]	rs28407950 + rs35571244				
HLA-C p.11 (class I CS2)	3.12x10 ⁻¹⁹	0.81 [0.77-0.84]	4.89x10 ⁻⁰⁸	0.87 [0.83-0.92]	rs28407950 + rs35571244				
rs28407950 (class II CS1)	1.37x10 ⁻⁵⁹	0.74 [0.71-0.77]	4.22x10 ⁻⁴⁴	0.77 [0.74-0.80]	rs2428494 + HLA-C p.11				
rs35571244 (class II CS2)	1.07x10 ⁻¹⁷	1.25 [1.19-1.32]	5.59x10 ⁻¹³	1.21 [1.15-1.28]	rs2428494 + HLA-C p.11				
		Adu	It-Onset Asth	nma					
rs2428292 (class I CS1)	4.52x10 ⁻²³	1.10 [1.08-1.13]	2.03x10 ⁻¹⁰	1.07 [1.05-1.09]	rs9272346 + HLA-DQA1*0301				
rs9272346 (class II CS1)	1.98x10 ⁻⁴⁷	1.16 [1.14-1.19]	1.09x10 ⁻³⁴	1.14 [1.12-1.17]	rs2428494				
HLA-DQA1*0301 (class II CS2)	6.91x10 ⁻⁴⁷	1.19 [1.16-1.21]	1.99x10 ⁻⁴¹	1.18 [1.15-1.20]	rs2428494				

P-value, odds ratio (OR), and 95% confidence interval (CI) shown for the original, marginal association and for the association when conditioning on either the class I or class II signals.

Table S18. Sample Composition of the Replication Cohort

	Childhood-onset asthma	Adult-onset asthma	Controls
Sample size	1,686	3666	56,063
Age of asthma onset in years	Range: 0-11 Mean (SD): 6 (3)	Range: 26-65 Mean (SD): 43 (10)	NA
Female Sex	46.1%	64.4%	54.6%
Allergic Disease (ever)	30.5%	26.4%	10.3%

Group	COA	AOA	Ctl
White	14	26	391
British	537	1063	16533
Irish	314	689	9651
Any other white background	395	830	13008
White and Black Caribbean	18	28	459
White and Black African	11	18	308
Black or Black British	0	1	22
Caribbean	114	266	3294
African	60	115	2826
Any other Black background	6	7	84
White and Asian	35	45	617
Asian or Asian British	0	5	30
Chinese	29	53	1311
Indian	91	284	4542
Pakistani	28	127	1352
Bangladeshi	3	10	180
Any other Asian background	31	99	1455

 Table S19. Self-Reported Ethnic Composition of the Replication Cohort.

Number of individuals in each self-reported ethnic group are shown for childhood-onset asthma

(COA), adult-onset asthma (AOA), and the non-asthmatic controls (Ctl).

Table S20. Replication Meta-Analysis Results

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