Supplementary materials

A Minimalist and Robust Chemo-Photothermal Nanoplatform Capable of Augmenting Autophagy-Modulated Immune Response Against Breast Cancer

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Figure S1. Preparation and stability of IC/IR820 NPs. (A) Solubility of itraconazole in water. (B) Stability of IC/IR820 NPs with different ratios. (C) Diameter of IC/IR820 nanoparticles at indicated time points. (D) Photographs of IC/IR820 NPs at indicated time points. (E) Zeta potential of IC/IR820 NPs. (F) Comparison of the Zeta potential of IC/IR820 NPs at 0 h and 24 h. (G) Photothermal activity of IR820 dispersed in water at various concentrations ($\lambda = 808$ nm, P = 1.0

 W/cm^2 ; the irradiation time = 5 min). (H) Temperature profile of IC/IR820 NPs in water after 300 s irradiation ($\lambda = 808$ nm, P=1.0 W/cm²) (I) Linear time data versus -ln(θ) obtained from the cooling period of IC/IR820 NPs. (J) Photothermal stability characterization using 808 nm laser irradiation (1 W/cm²) of the IC/IR820 NPs for four heating-cooling cycles.

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Calculation of the photothermal conversion efficiency

The photothermal conversion efficiency (η) of IC/IR820 nanoparticles was calculated according 10 the following method.

$$\eta(\%) = \frac{hS\Delta T_{max} - Q_{Dis}}{I(1-10^{-A_{\lambda}})} \times 100\%$$

In detail, 15

hS=
$$\frac{mC}{\tau_s}$$

$$\tau_{s} = -\frac{t}{\ln(\theta)}$$

 ΔT_{max}

$$\theta = \frac{\mathrm{T} - \mathrm{T}_{\mathrm{surr}}}{\Delta \mathrm{T}_{\mathrm{max}}}$$

QDis was measured to be mW using water without IC/IR820 nanoparticles. m is the mass of the solution (1.0 g), C is the heat capacity of the water solution (4.2 J/g). τ_s is the associated time constant, which was showed in Figure S1J-K.

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Figure S2. Intracellular uptake of IC/IR820 NPs in MCF-7 and 4T1 cell line. (A-D) Fluorescence microscopy images and flow cytometric results show the uptake of IC/IR820 nanoparticles on indicated time points in MCF-7 and 4T1 cells. Scale bar: 20 µm. (E-H) Fluorescence microscopy images and flow cytometric results show the uptake of free IR820 and IC/IR820 nanoparticles with different concentrations in MCF-7 and 4T1 cells. Scale bar: 20 µm.

 IC50 of IC/IR820 NPs + L (μM)

 Cell line
 Equivalent to IR820 (μM)
 Equivalent to IC (μM)

 MDA-MB-231
 11.60
 13.92

 MCF-7
 5.088
 6.105

 4T1
 5.021
 6.026

А



Figure S3. Cytotoxicity of IR820 and IC/IR820 NPs without laser treatment. (A) IC₅₀ value of IC/IR820 NPs in different breast cancer cell lines. (B-D) Viability of MDA-MB-231, MCF-

7,4T1 cells co-cultured with IR820, IC/IR820 nanoparticles without laser irradiation. (E-H) Representative images for colony formation of MDA-MB-231, MCF-7,4T1 cells co-cultured with IR820, IC/IR820 nanoparticles without laser irradiation. (I-L) Quantification of the positive ratio of MDA-MB-231, MCF-7, and 4T1 cells co-cultured with IR820, IC/IR820 nanoparticles without laser irradiation in EdU assay. Scale bar: 20 μm.



Figure S4. Cytotoxicity of IC/IR820 NPs is dependent on ROS. (A-C) Immunofluorescence images to show subcellular localization of IR820, IC/IR820 nanoparticles. Scale bar: 20 μ m. (D, F) Fluorescence microscopy images and (E, G) flow cytometry analysis for intracellular ROS generation of MCF-7 and 4T1 cells using DCFH-DA as a probe. Scale bar: 20 μ m. (λ = 808 nm, P = 1.0 W/cm²; the irradiation time = 2 min). (H-I) Immunoblot analysis of antioxidant proteins for MDA-MB-231 and MCF-7 cells treated IC, IR820, and IC/IR820 with or without NAC treatment. (λ = 808 nm, P = 1.0 W/cm²; the irradiation time = 2 min). (J-M) Flow cytometry analysis for intracellular ROS generation of MDA-MB-231 and MCF-7 cells treated IC/IR820 with or without NAC. (λ = 808 nm, P = 1.0 W/cm²; the irradiation time = 2 min).

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Figure S5. IC/IR820 NPs induce apoptosis in breast cancer cells. (A-D) Apoptosis of IR820, IC, and IC/IR820 NPs were evaluated by annexin V–FITC/PI staining in MDA-MB-231, MCF-7, and 4T1 cells, quantification of apoptotic cell ratio. ($\lambda = 808$ nm, P = 1.0 W/cm²; the irradiation time = 2 min).



Figure S6. IC/IR820 NPs promote autophagy in breast cancer cells. (A-C) Immunoblot analysis of autophagic markers for MDA-MB-231, 4T1, and MCF-7 cells treated IC, IR820, and IC/IR820. ($\lambda = 808$ nm, P = 1.0 W/cm²; the irradiation time = 2 min). (D-F, H-J) Immunofluorescence assays display subcellular localization of LC3B in MDA-MB-231, 4T1, and MCF-7 cells treated IC, IR820, and IC/IR820 NPs, quantification of LC3B puncta per cell. Scale bar: 10 µm. ($\lambda = 808$ nm, P = 1.0 W/cm²; the irradiation time = 30 s). (G, K) Immunofluorescence assays display subcellular localization of LC3B puncta in 4T1 cells treated IC/IR820 NPs with or without NAC treatment, quantification of LC3B puncta per cell. Scale bar: 10 µm. ($\lambda = 808$ nm, P = 1.0 W/cm²; the irradiation time = 30 s).



Figure S7. Evaluation of anti-breast cancer effect of IC/IR820 NPs *in vivo*. (A) Fluorescence images of tumors and main organs at 24 h after injection of IR820 and IC/IR820 nanoparticles. (B) Representative images showing tumor size in mice bearing 4T1 orthotopic breast tumor xenografts treated with vehicle, IR820 + Laser, IC, IC/IR820 NPs + Laser at the end of the treatment period. (C) Representative heart, liver, spleen, lung, and kidney HE staining of mice bearing 4T1 orthotopic breast tumor xenografts treated with vehicle, IR820 NPs + Laser, IC, IC/IR820 NPs + Laser at the end of the treatment period. Scar bar: 200 μm. (D-G) Blood biochemical markers: ALT (D); AST (E); BUN (F); CR (G); Each group was tested three times in parallel.



Figure S8. IC/IR820 NPs inhibit migration and invasion of MCF-7 and 4T1 cell line. (A-C) Representative images of migration, invasion, and scratch test of MCF-7 cells, quantification of migration/invasion cell number and migratory distance. Scale bar for migration and invasion assay: 50 μm. Scale bar for scratch test: 20 μm. (D-F) Representative images of migration, invasion, and scratch test of 4T1 cells, quantification of migration/invasion cell number and migratory distance. Scale bar for scratch test: 20 μm. (D-F) Representative images of migration, invasion, and scratch test of 4T1 cells, quantification of migration/invasion cell number and migratory distance. Scale bar for migration and invasion assay: 50 μm. Scale bar for scratch test: 20 μm.



Figure S9. IC/IR820 NPs activate the immune response and inhibit breast cancer recurrence. (A) The body weight of mice in each group was measured at the indicated time points. (B) Primary tumor volumes. (C-E) Representative results of CD4⁺ T cells and CD8⁺ T cells. Quantification of the percentage of CD4⁺ T cells and CD8⁺ T cells in distant tumor. (F) Immunoblot analysis of E-cadherin, caspase3, cleaved-caspase3 for primary tumor treated with NS, IC, IR820, and IC/IR820 NPs. (G-H) Representative images showing tumors and spleens in mice before and after surgery treated with NS or IC/IR820 NPs. (I) The body weight of mice in each group was measured at the indicated time points. (J-L) Representative results of CD4⁺ T cells and CD8⁺ T cells and CD8⁺ T cells. Quantification of the percentage of CD4⁺ T cells and CD8⁺ T cells in recurrent tumor. (M) Immunoblot analysis of E-cadherin, claudin 1, HSP70, ZEB1 for recurrent tumor treated with NS or IC/IR820 NPs.