

## **Supplementary Material**

### **Selenium nanoparticles alleviate ischemia reperfusion injury-induced acute kidney injury by modulating GPx-1/NLRP3/Caspase-1 pathway**

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#### **Table of contents:**

**Table S1.** Antibodies applied in the immunofluorescence staining

**Table S2.** Antibodies applied in the immunohistochemistry staining

**Table S3.** Sequences of primers used in q-PCR

**Table S4.** Antibodies applied in the Western blotting

**Figure S1.** SDS-PAGE determining the existence of BSA in the NPs

**Figure S2.** Zeta potential and particle size of different batches of Se@BSA NPs

**Figure S3.** Changes in body weight at day 7 post Se@BSA NP treatment

**Figure S4.** Changes in ALT and AST levels at day 7 post Se@BSA NP treatment

**Figure S5.** Changes in mouse hemogram at day 7 post Se@BSA NP treatment

**Figure S6.** ICP-MS detecting the Se contents in serum post Se@BSA NP treatment

**Figure S7.** ICP-MS detecting the Se contents in HK-2 cells post Se@BSA NP treatment

**Figure S8.** Tubular injury score of IRI-AKI mouse

**Figure S9.** NLRP3 mRNA expression in HK-2 cells

**Figure S10.** Induced expression of GPx-1 in HK-2 cells by Se@BSA NPs

**Table S1.** Antibodies applied in the immunofluorescence staining

<b>Proteins</b>	<b>Primary Antibodies</b>	<b>Secondary Antibodies</b>
Mouse AQP1	1:100, sc-25287 ( Santa Cruz, Shanghai, CHN)	Cy3, A0521 (Beyotime)
NLRP3	1:200, bs-23722R (Bioss, Beijing, CHN)	Alexa Fluor 488, A0423 (Beyotime)
GPx-1	1:100, AF7017 (Beyotime)	Cy3, A0516 (Beyotime)
Fibronectin	1:100, ab2413 (Abcam, Shanghai, CHN)	Cy3, A0516 (Beyotime)
$\alpha$ -Sma	1:200, 19245 (CST, Shanghai, CHN)	Cy3, A0516 (Beyotime)

**Table S2.** Antibodies applied in the immunohistochemistry staining

<b>Proteins</b>	<b>Primary Antibodies</b>	<b>Secondary Antibodies</b>
Kim-1	1:100, ab78494 (Abcam)	ZLI-9019 (ZSGB-BIO, Beijing, CHN)
IL-1 $\beta$	1:100, ab283818 (Abcam)	ZLI-9019 (ZSGB-BIO)
IL-18	1:100, ab191860 (Abcam)	ZLI-9019 (ZSGB-BIO)
Collagen I	1:1000, 81375 (CST)	ZLI-9019 (ZSGB-BIO)

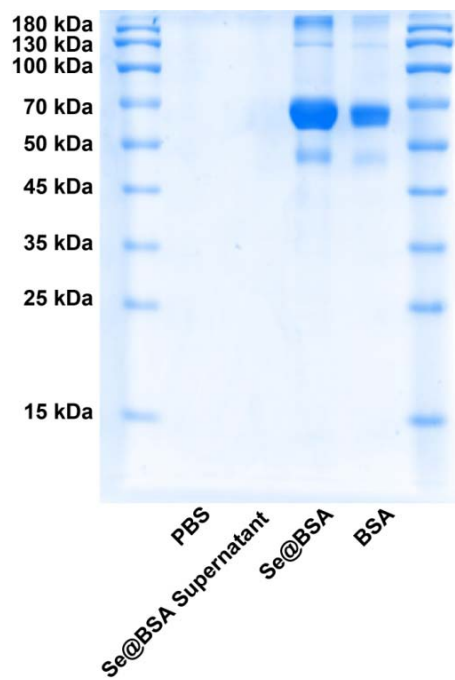
**Table S3.** Sequences of primers used in q-PCR

Species	Name	Sequence (5'-3')	Product length	
Mouse	Kim-1	Forward	CAGGGTCTCCTTCACAGCAG	172
		Reverse	CCACCACCCCCTTTACTTCC	
	GPx-1	Forward	AATGTCGCGTCTCTCTGAGG	117
		Reverse	TCCGAAGTATTGCACGGG	
	NLRP3	Forward	ATCAACAGGCGAGACCTCTG	96
		Reverse	GTCCTCCTGGCATAACCATAGA	
	IL-1 $\beta$	Forward	TTCAGGCAGGCAGTATCACTC	166
		Reverse	CCAGCAGGTTATCATCATCA	
	IL-18	Forward	CCAGACCAGACTGATAATATAC	110
		Reverse	TCTTGTTCTTACAGGAGAGG	
	$\beta$ -actin	Forward	CACTGTCGAGTCGCGTCC	89
		Reverse	TCATCCATGGCGAACTGGTG	
Human	Kim-1	Forward	CTTCTAACTAACCGGCTGATCC	244
		Reverse	TTCCGATAGGTGACGTGGGT	
	GPx-1	Forward	CAGTCGGTGTATGCCTTCTCG	105
		Reverse	GAGGGACGCCACATTCTCG	
	NLRP3	Forward	GATCTTCGCTGCGATCAACAG	81
		Reverse	CGTGCATTATCTGAACCCAC	
	IL-1 $\beta$	Forward	AGCCATGGCAGAAGTACCTG	116
		Reverse	CCTGGAAGGAGCACTTCATCT	
	IL-18	Forward	ATCGCTTCCTCTCGCAACAA	223
		Reverse	GAGGCCGATTTCTTGGTCA	
	$\beta$ -actin	Forward	GATTCCTATGTGGGCGACGA	229
		Reverse	AGGTCTCAAACATGATCTGGGT	

**Table S4.** Antibodies applied in the Western blotting

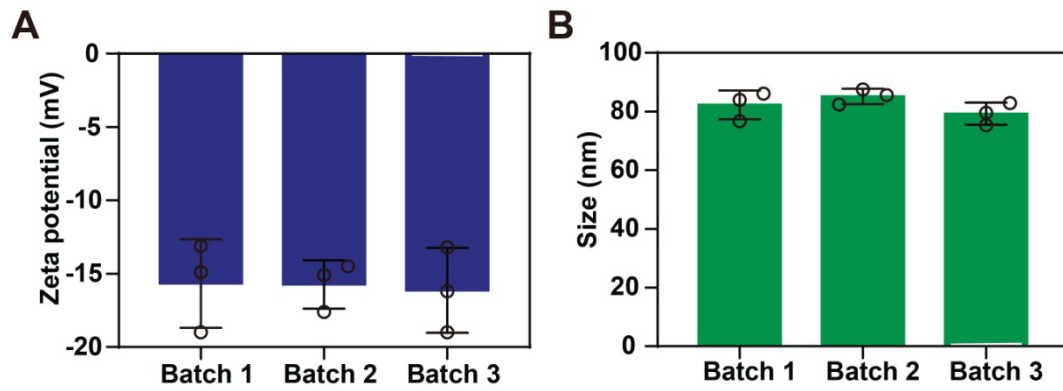
<b>Proteins</b>	<b>Primary Antibodies</b>	<b>Secondary Antibodies</b>
$\beta$ -actin	1:1000, AF50017 (Beyotime)	A0216 (Beyotime)
Kim-1	1:1000, ab78494 (Abcam)	A0208 (Beyotime)
NLRP3	1:1000, bs-23722R (Bioss)	A0208 (Beyotime)
IL-1 $\beta$	1:1000, ab283818 (Abcam)	A0208 (Beyotime)
Mouse IL-18	1:1000, ab191860 (Abcam)	A0208 (Beyotime)
Human IL-18	1:1000, ab243295 (Abcam)	A0208 (Beyotime)
Caspase-1	1:1000, ab138483 (Abcam)	A0208 (Beyotime)
GPx-1	1:1000, AF7017 (Beyotime)	A0208 (Beyotime)
Fibronectin	1:1000, ab2413 (Abcam)	A0208 (Beyotime)
$\alpha$ -Sma	1:1000, 19245 (CST)	A0208 (Beyotime)
Collagen I	1:1000, 81375 (CST)	A0208 (Beyotime)

**Figure S1.** SDS-PAGE determining the existence of BSA in the NPs. A total of 200  $\mu\text{g}$  of Se@BSA NPs was dissolved in 25  $\mu\text{L}$  PBS. After a high speed centrifugation at 12 000 g for 15 min, the supernatant was collected. The precipitate was further dissolved in 25  $\mu\text{L}$  PBS and heated for denaturation. Proteins were resolved by 10% SDS-PAGE and stained with Coomassie brilliant blue. BSA (5  $\mu\text{g}$ ) was employed as the positive control.

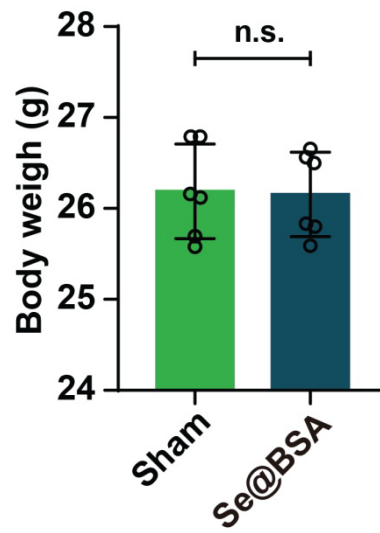


**Figure S2.** Zeta potential and particle size of different batches of Se@BSA NPs. (A) Zeta potential determination. (B) DLS detection of the hydrodynamic diameters of Se@BSA NPs.

The results are presented as the mean  $\pm$  SD.

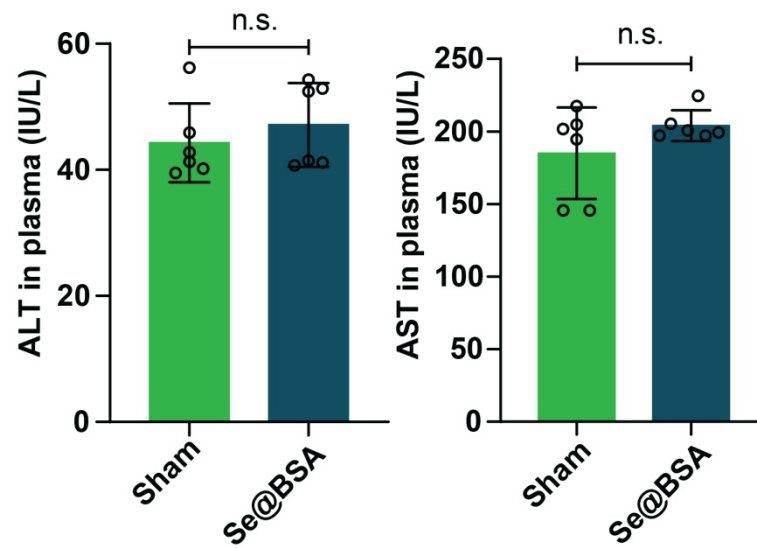


**Figure S3.** Changes in body weight at day 7 post Se@BSA NP treatment. Intravenous injection of 20 mg/kg Se@BSA NPs was innocuous to C57 mice. Mice in the sham group were treated with sterile PBS. n.s., not significantly.

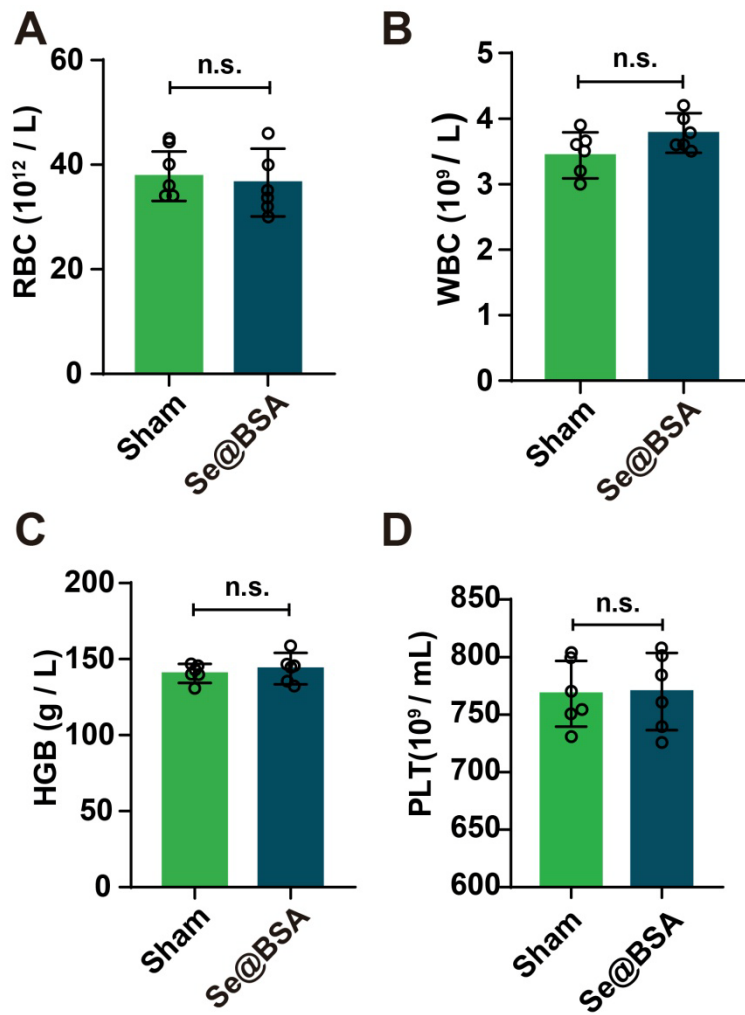




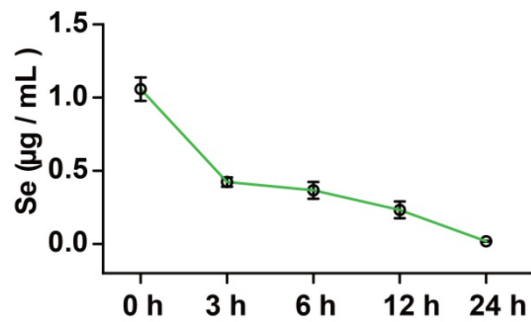
**Figure S4.** Changes in ALT and AST levels at day 7 post Se@BSA NP treatment. ALT and AST contents in mouse blood were determined by ELISA (SEA207Mu and SEB214Mu, CLOUD-CLONE CORP., Katy, TX, USA). n.s., not significantly.



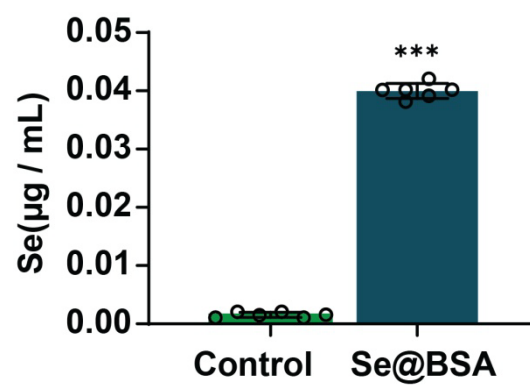
**Figure S5.** Changes in mouse hemogram at day 7 post Se@BSA NP treatment. Mouse blood was obtained at day 7 post Se@BSA NP (20 mg/kg, I.V.) treatment. Shown are the changes of (A) red blood cells, (B) white blood cells, (C) platelets, and (D) hemoglobin. n.s., not significantly.



**Figure S6.** ICP-MS detecting the Se contents in serum post Se@BSA NP treatment. Mouse blood (20  $\mu$ L) was obtained at 3, 6, 12, and 24 h post Se@BSA NP (1 mg/kg, I.V.) treatment. Samples were processed with 0.5 mL nitrohydrochloric acid for 24 h and adjusted to 1 mL, in which the Se contents were determined by ICP-MS. The results are presented as the mean  $\pm$  SD.

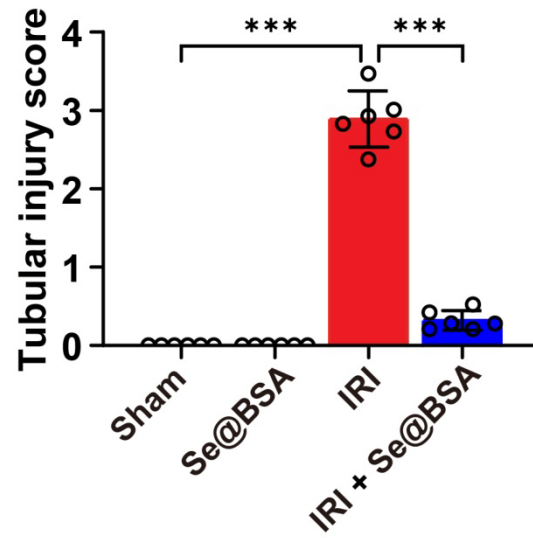


**Figure S7.** ICP-MS detecting the Se contents in HK-2 cells post Se@BSA NP treatment. Se@BSA NPs (50  $\mu\text{g}/\text{mL}$ ) were coincubated with  $2 \times 10^8$  CFU HK-2 cells at 37 °C for 6 h. The cells were collected and processed with 2 mL nitrohydrochloric acid for 24 h. After dilution to 5 mL with 2% salpeter solution, the Se contents in the samples were determined by ICP-MS. The results are presented as the mean  $\pm$  SD. **\*\*\***,  $P < 0.001$ .

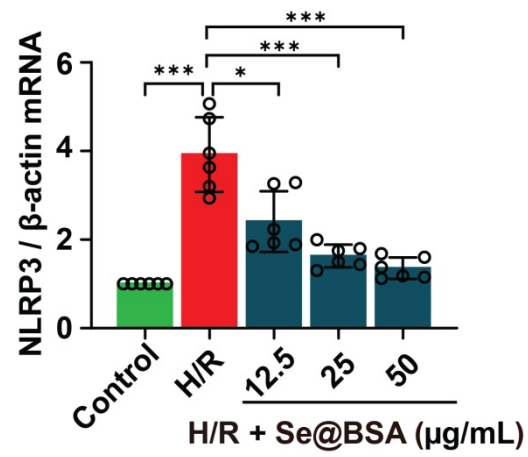


**Figure S8.** Tubular injury score of IRI-AKI mouse. The results are shown as the mean  $\pm$  SD.

\*\*\*,  $P < 0.001$ .



**Figure S9.** NLRP3 mRNA expression in HK-2 cells. H/R cells were exposed to different concentrations of NPs for 6 h.  $\beta$ -actin was used as the reference. The results are shown as the mean  $\pm$  SD. \*,  $P < 0.05$ ; \*\*\*,  $P < 0.001$ .



**Figure S10.** Induced expression of GPx-1 in HK-2 cells by Se@BSA NPs. (A) q-PCR detecting the mRNA expression of GPx-1. The results are shown as the mean  $\pm$  SD. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ . (B) Western blotting revealing the dose-dependent modulation of Se@BSA NPs on GPx-1 expression in HK-2 cells.  $\beta$ -actin was used as the reference.

