

SUPPLEMENTARY INFORMATION

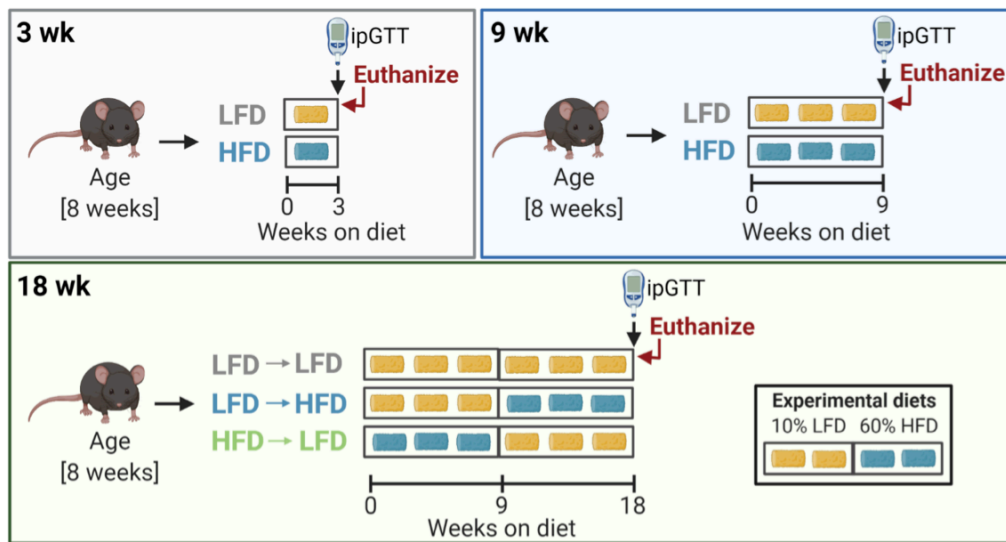
Multiomics reveals persistence of obesity-associated immune cell phenotypes in adipose tissue during weight loss and weight regain in mice

Cottam, M.A., Caslin, H.L., Winn, N.C., and Hasty A.H.

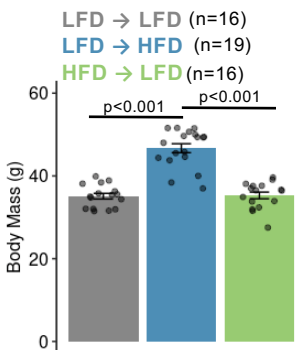
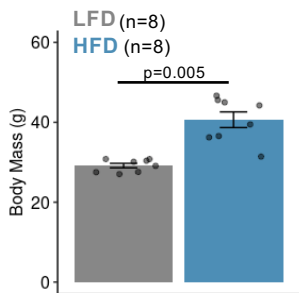
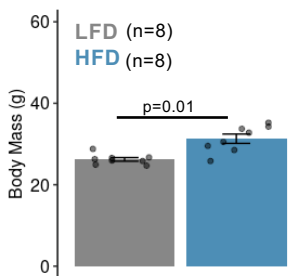
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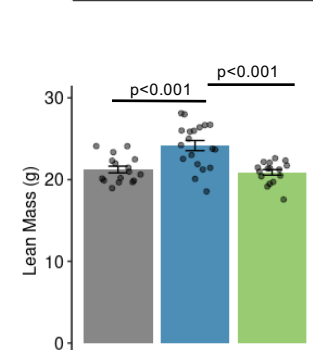
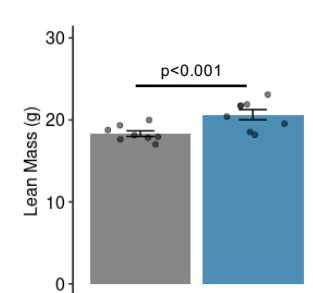
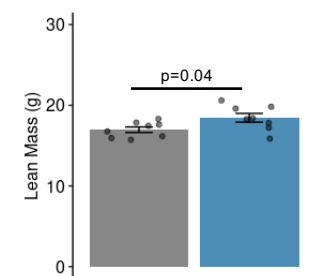
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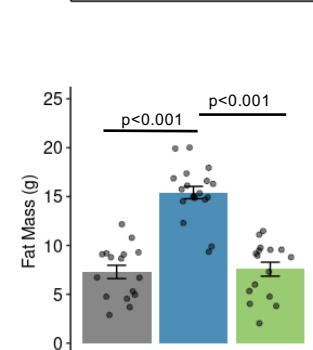
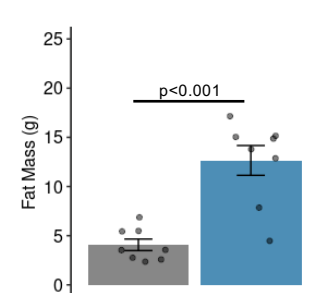
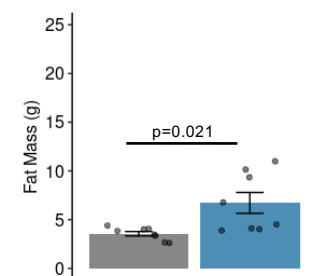
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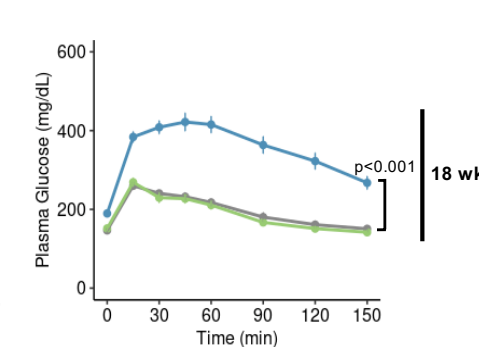
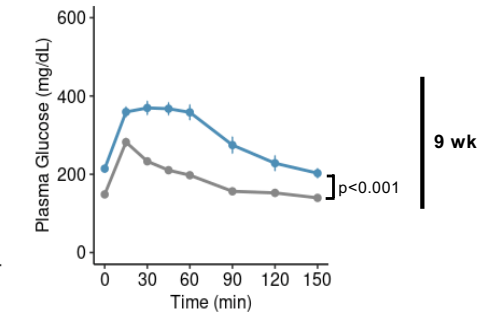
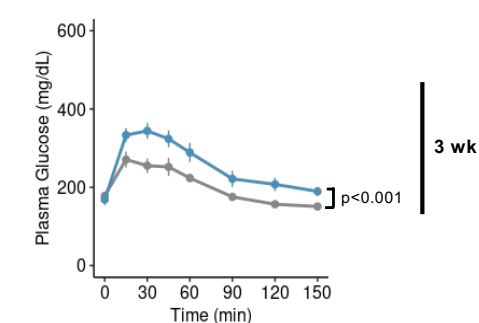
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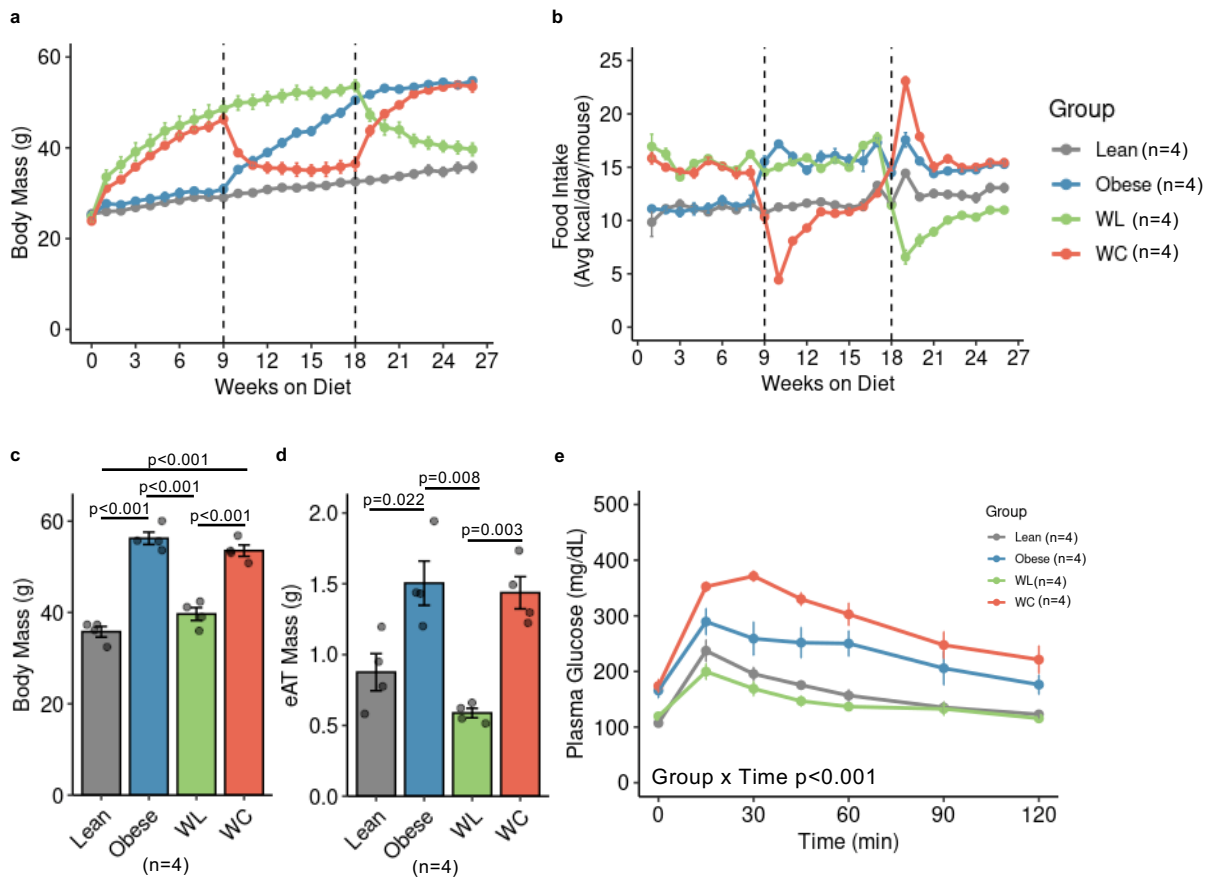
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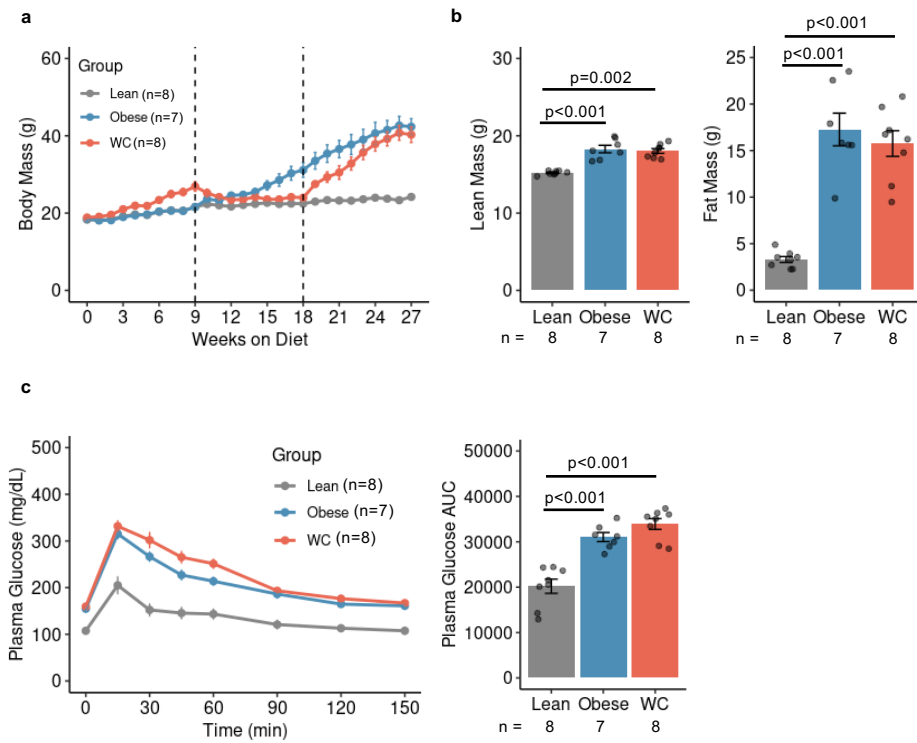


Supplementary Figure 1. Interim body mass and ipGTT for male mice. (a) Schematic of interim study timepoints. (b) Body mass, (c) lean mass, (d) fat mass, and (e) ipGTT at weeks 3, 9, and 18 for male mice (mean \pm SEM). Pairwise two-tailed T-tests (3 and 9-week timepoints) with Bonferroni correction for multiple comparisons (18-week timepoint only) were used to compare groups for body mass, body composition, and ipGTT area under the curve. Significant p values are shown. Data are not paired, mice at each timepoint represent independent cohorts that were sacrificed at the indicated time.

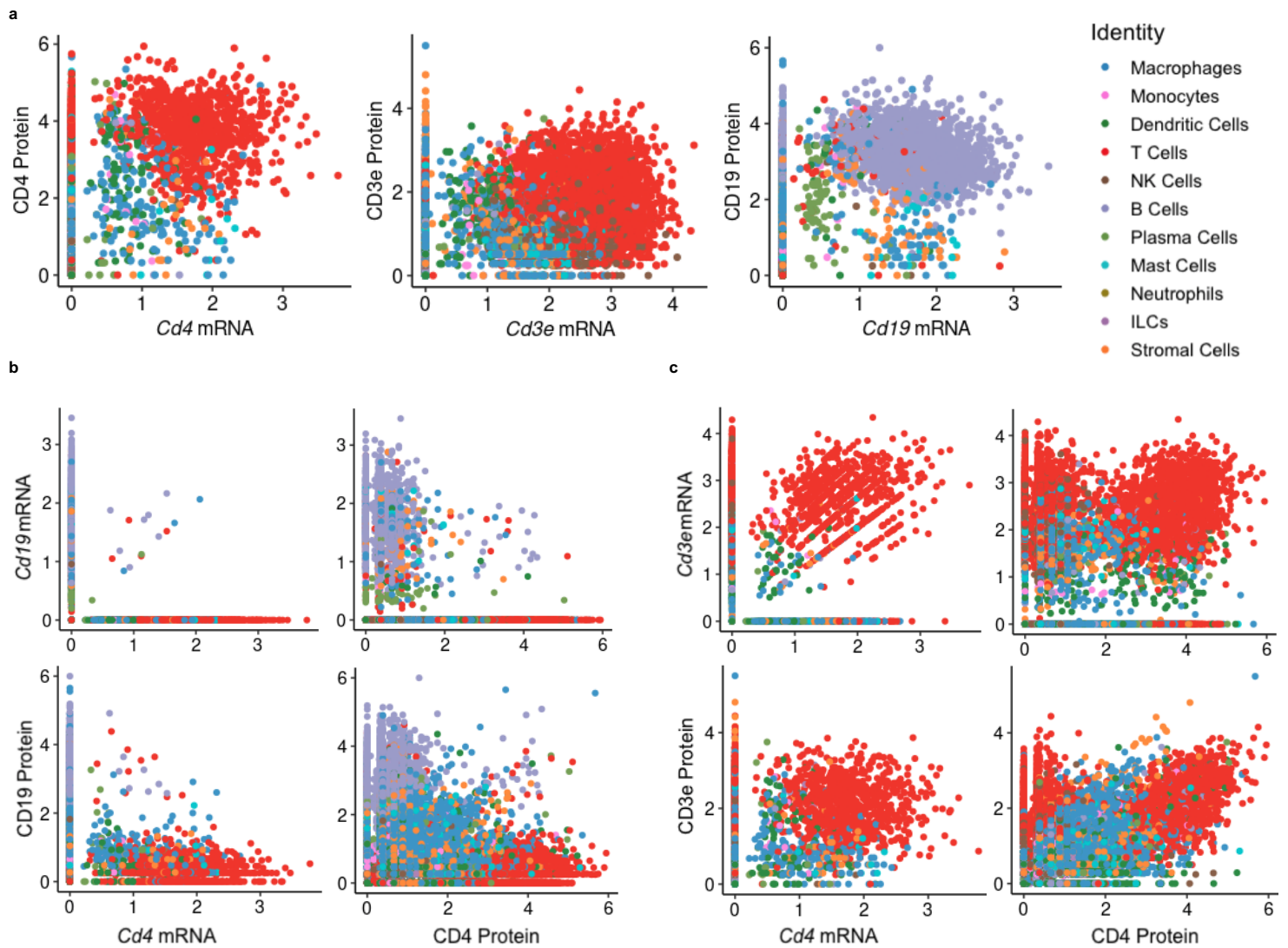


Supplementary Figure 2. Body mass and glucose tolerance for mice used for CITEseq analysis. (a) Body mass over time measured weekly. (b) Food intake over time measured weekly. (c) Body mass and (d) eAT mass measured at sacrifice. (e) Intraperitoneal glucose tolerance test dosed at 1.5 g dextrose/kg lean mass one week prior to sacrifice. Pairwise two-tailed T-tests with Bonferroni correction for multiple comparisons were used to compare groups for body mass and eAT mass and two-way ANOVA was used to compare groups for ipGTT; significant p values shown. Data are plotted as mean \pm SEM (n=4 mice).

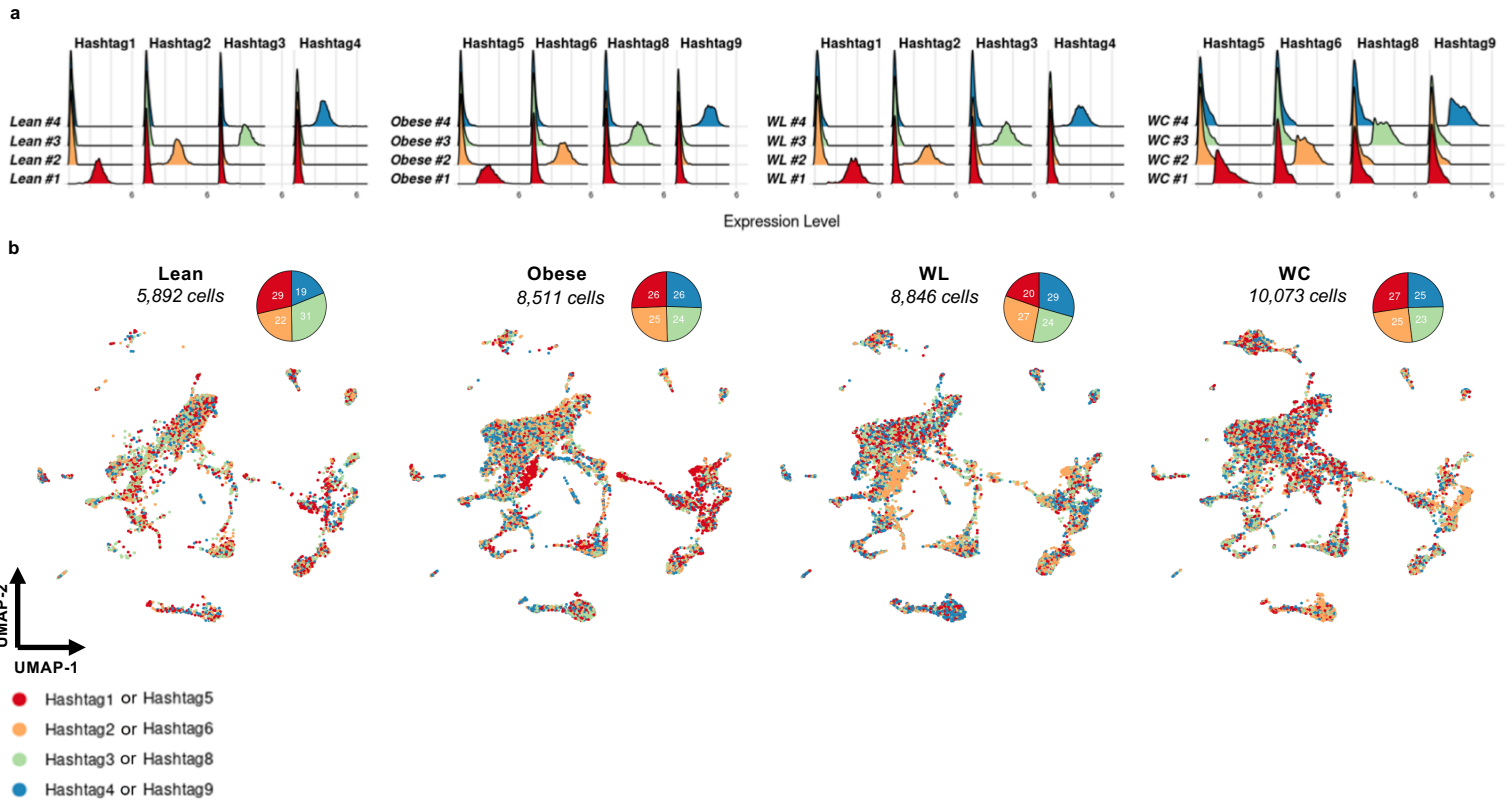
Female



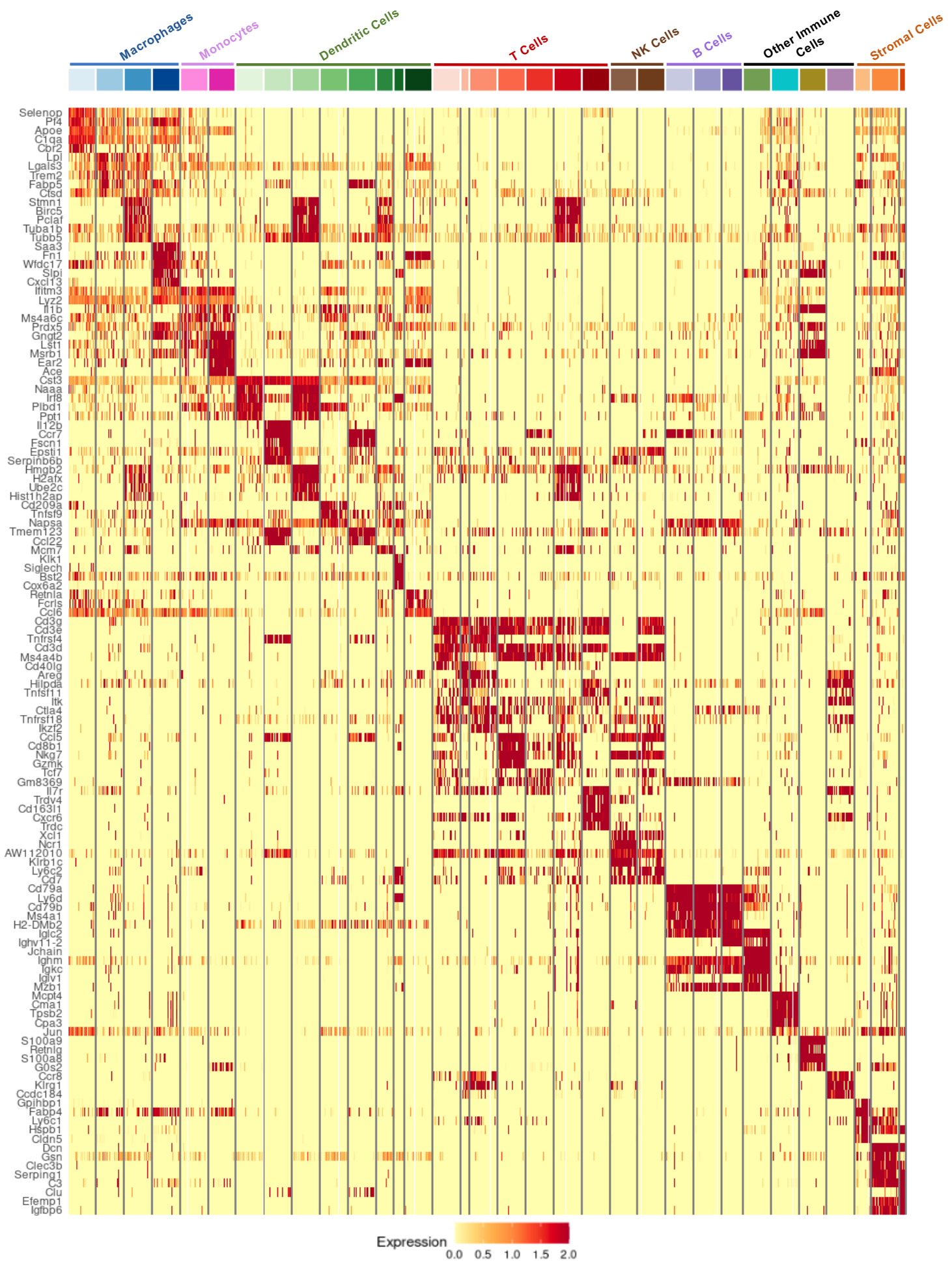
Supplementary Figure 3. Weight cycling in female C57BL/6J mice. (a) Body mass over time, (b) body composition after 27-weeks of diet, and (c) intraperitoneal glucose tolerance test and corresponding area under the curve (AUC) after 27-weeks of diet for female mice following our WC model (number of mice indicated for each panel; n). Pairwise two-tailed T-tests with Bonferroni correction for multiple comparisons were used to compare groups for body composition and ipGTT area under the curve; significant p values shown.



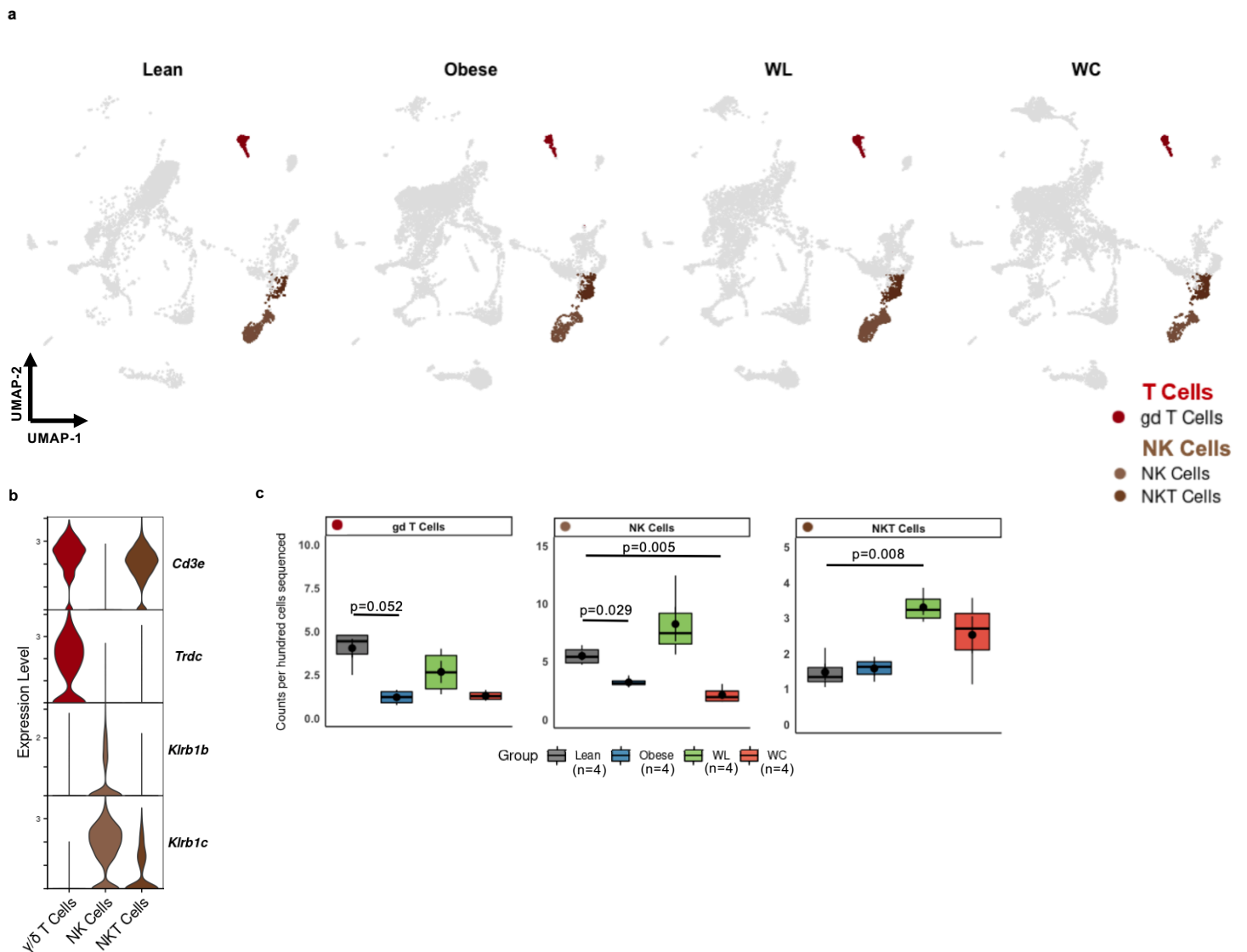
Supplementary Figure 4. Validation of CITE-seq antibodies. (a) Comparison of genes for *Cd3e*, and *Cd19* with their corresponding surface proteins measured using CITE-seq. Individual cells are colored by annotated cell type. (b) Comparison of cell type exclusive genes *Cd4* and *Cd19* with their corresponding surface proteins. (c) Comparison of co-expressed genes *Cd4* and *Cd3e* with their corresponding surface proteins.



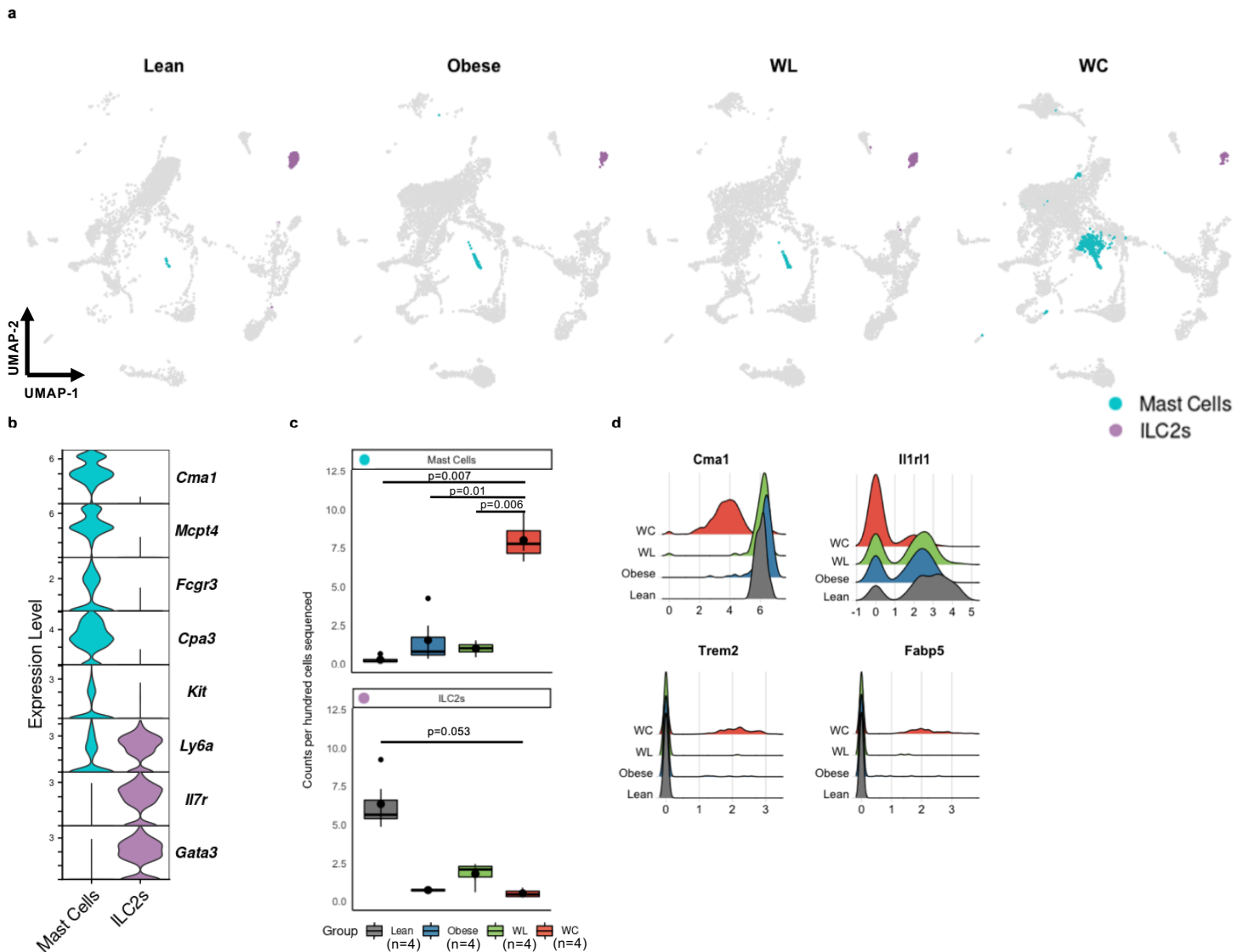
Supplementary Figure 5. Biological replicates identified by Hashtag demultiplexing are well represented within diet groups. (a) Hashtags demultiplexed for each individual mouse per diet group (n=4 per group; total of 16 mice sequenced). (b) Uniform manifold projection with total cell counts retained following QC for each diet group and distribution of hashtags measured within each diet group (inset).



Supplemental Figure 6. Heatmap of top five differentially expressed genes for each high-resolution cluster identity. Each row is a unique gene, colored red when expressed highly. Each thin vertical line indicates one subsampled cell (up to 100 cells per subcluster).



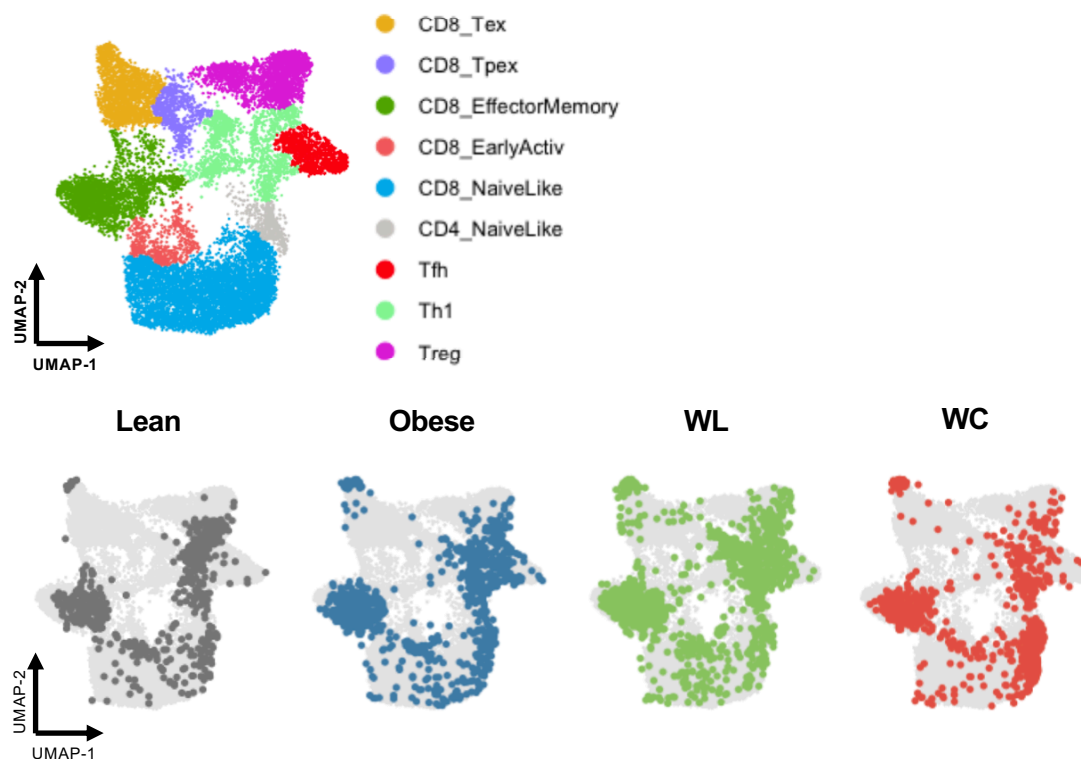
Supplementary Figure 7. Frequency and markers of adipose tissue γ/δ , NK, and NKT cells. (a) UMAP of γ/δ T cells and NK cell subclusters by diet group. (b) Expression of markers enriched in γ/δ T cells and NK cell subclusters. (c) Counts per hundred cells sequenced for γ/δ T cells and NK cell subclusters (mean \pm SEM; n=4 mice). Box indicates interquartile range (25th-75th percentile) with 50th percentile indicated by solid line and mean indicated by large circle. Range of whiskers indicates largest and smallest values within 1.5 times the interquartile range and values outside of the range are indicated by small circles. Pairwise two-tailed t-tests with Bonferroni correction for multiple comparisons were used to compare groups for cell counts; significant p values shown.



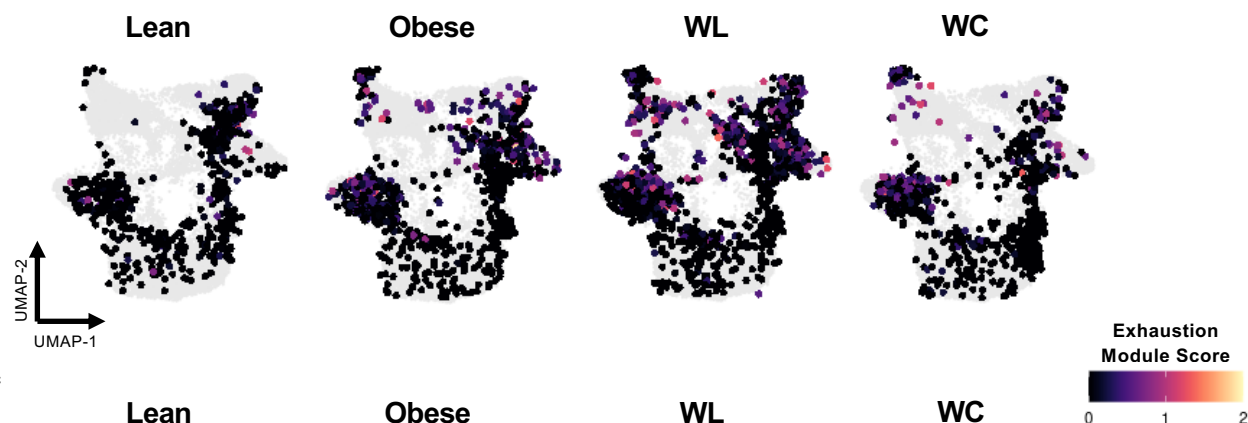
Supplementary Figure 8. Adipose tissue mast cells have reduced expression of *Il1r1* and ILC2s are reduced in frequency after obesity. (a) UMAP of mast cells and ILC2s by diet group. (b) Expression of markers enriched in mast cells and ILC2 subclusters. (c) Counts per hundred cells sequenced for mast cells and ILC2s (mean \pm sem; n=4 mice). Box indicates interquartile range (25th-75th percentile) with 50th percentile indicated by solid line and mean indicated by large circle. Range of whiskers indicates largest and smallest values within 1.5 times the interquartile range and values outside of the range are indicated by small circles. (d) Expression of *Cma1* coding for the secreted protease chymase, *Il1r1* coding for the IL33-R, and lipid-associated proteins *Trem2*, and *Fabp5* in mast cells by diet group. Pairwise two-tailed t-tests with Bonferroni correction for multiple comparisons were used to compare groups for cell counts; significant p values shown.

a

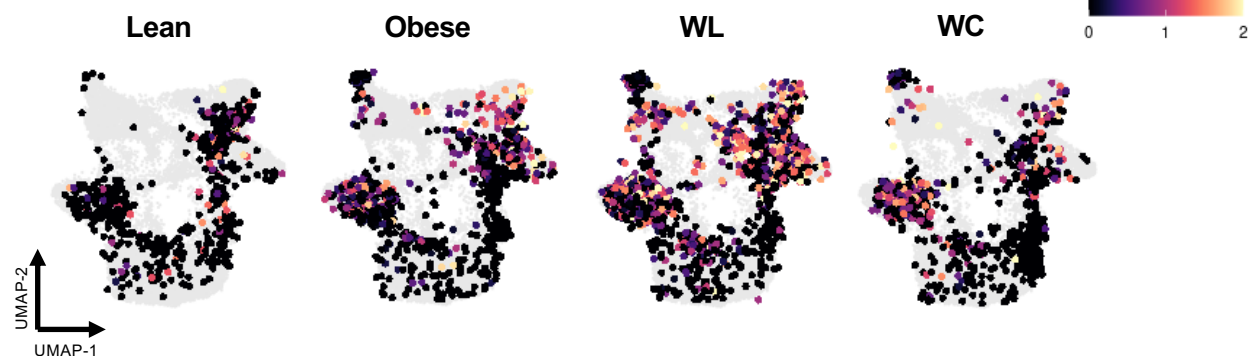
Tumor-infiltrating Lymphocyte Atlas



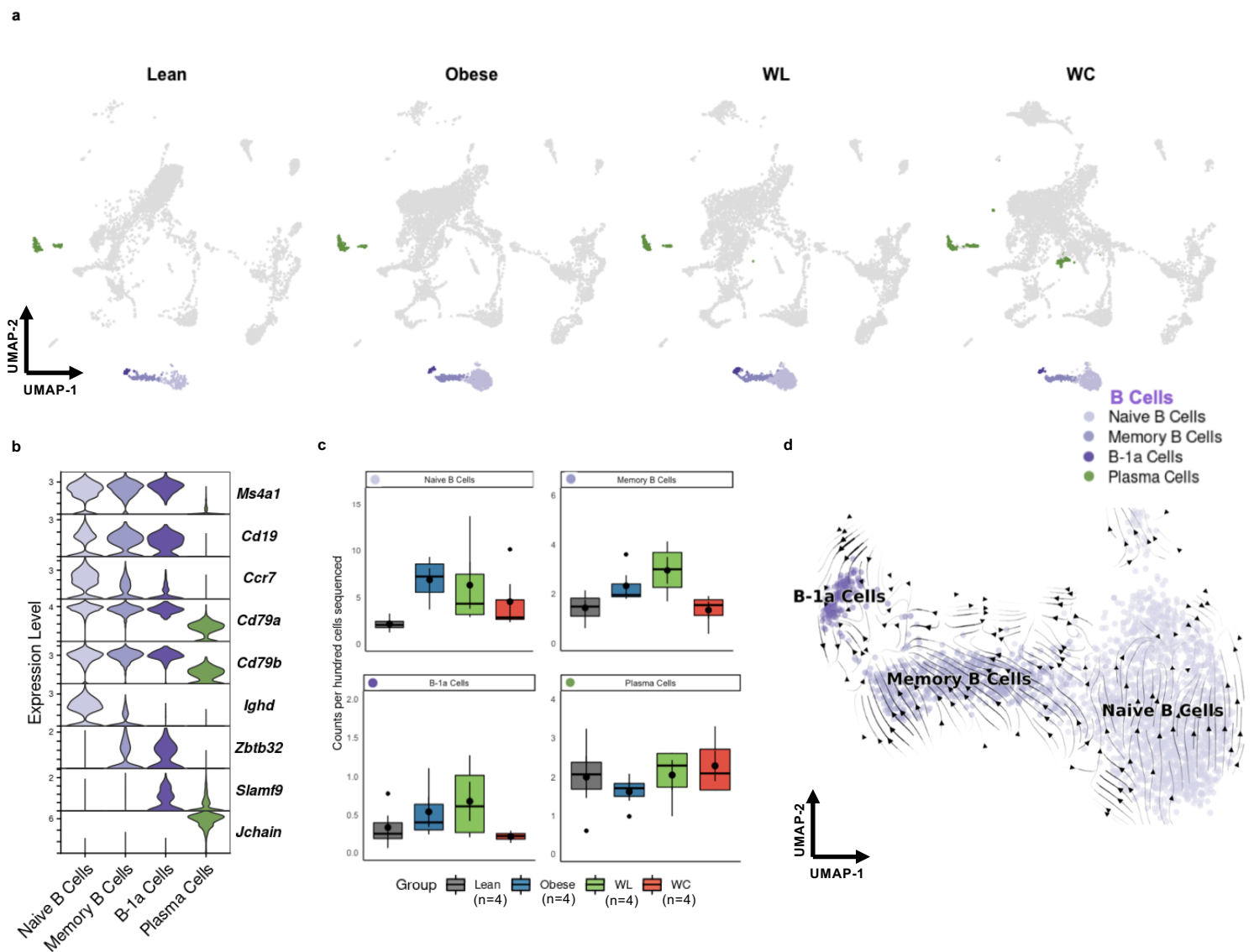
b



c



Supplementary Figure 9. Alignment of CD8+ T cells to a tumor-infiltrating lymphocyte reference atlas. (a) T cells separated by diet group plotted onto a tumor-infiltrating lymphocyte reference atlas using ProjectTILs. T cells, projected onto the TIL reference atlas, colored by an exhaustion module containing the (b) mRNA features *Pdcd1*, *Tox*, *Tigit*, *Lag3*, and *Entpd1* and (c) protein features PD-1 (CD279) and TIGIT. Note that although the reference atlas suggests the majority of cells are effector memory, they express high levels of genes within the T cell mRNA and protein exhaustion modules.



Supplementary Figure 10. Frequency and markers of adipose tissue B cells and plasma cells. (a) UMAP of B cell subclusters and plasma cells by diet group. (b) Expression of markers enriched in B cell subclusters and plasma cells. (c) Counts per hundred cells sequenced for B cell subclusters and plasma cells (mean \pm SEM; $n=4$ mice). Box indicates interquartile range (25th-75th percentile) with 50th percentile indicated by solid line and mean indicated by large circle. Range of whiskers indicates largest and smallest values within 1.5 times the interquartile range and values outside of the range are indicated by small circles. (d) Embedding of RNA velocity displayed on the UMAP for B cell subsets. Pairwise two-tailed t-tests with Bonferroni correction for multiple comparisons were used to compare groups for cell counts (no significant differences observed for $adj\ p < 0.05$).

Mouse Info	Hashing Antibody	Pooled Sample
Mouse 1, Lean	0301 anti-mouse Hashtag 1	1
Mouse 2, Lean	0302 anti-mouse Hashtag 2	1
Mouse 3, Lean	0303 anti-mouse Hashtag 3	1
Mouse 4, Lean	0304 anti-mouse Hashtag 4	1
Mouse 5, Obese	0305 anti-mouse Hashtag 5	2
Mouse 6, Obese	0306 anti-mouse Hashtag 6	2
Mouse 7, Obese	0308 anti-mouse Hashtag 8	2
Mouse 8, Obese	0309 anti-mouse Hashtag 9	2
Mouse 9, WL	0301 anti-mouse Hashtag 1	3
Mouse 10, WL	0302 anti-mouse Hashtag 2	3
Mouse 11, WL	0303 anti-mouse Hashtag 3	3
Mouse 12, WL	0304 anti-mouse Hashtag 4	3
Mouse 13, WC	0305 anti-mouse Hashtag 5	4
Mouse 14, WC	0306 anti-mouse Hashtag 6	4
Mouse 15, WC	0308 anti-mouse Hashtag 8	4
Mouse 16, WC	0309 anti-mouse Hashtag 9	4

Cell Sorting Reagents	Manufacturer	Catalog
Fc Block	BD Biosciences	553142
CD45 microbeads	Mitenyi	130-052-301
DAPI	Thermofischer	62247

Hashing Antibodies	Manufacturer	Catalog
0301 anti-mouse Hashtag 1	Biolegend	155861
0302 anti-mouse Hashtag 2	Biolegend	155863
0303 anti-mouse Hashtag 3	Biolegend	155865
0304 anti-mouse Hashtag 4	Biolegend	155867
0305 anti-mouse Hashtag 5	Biolegend	155869
0306 anti-mouse Hashtag 6	Biolegend	155871
0308 anti-mouse Hashtag 8	Biolegend	155875
0309 anti-mouse Hashtag 9	Biolegend	155877

Total Seq C Feature Antibodies	Manufacturer	Catalog
anti-mouse Mac2/gal3	Biolegend	125423
anti-mouse CD279.PD-1	Biolegend	109127
anti-mouse FCyR1	Biolegend	139327
anti-mouse CD4	Biolegend	100571
anti-mouse CCR7/CD197	Biolegend	120131
anti-mouse CD80	Biolegend	109127
anti-mouse CD11c	Biolegend	117361
anti-mouse CD44	Biolegend	103063
anti-mouse NK1.1	Biolegend	108765
anti-mouse TCRy/d	Biolegend	118141
anti-mouse CD39	Biolegend	143815
anti-mouse CD19	Biolegend	115571
anti-mouse CD11b	Biolegend	101275
anti-mouse CD3	Biolegend	100263
anti-mouse TIGIT/vstm3	Biolegend	142119
anti-mouse CD8a	Biolegend	100785

Supplementary Table 1: Ordering Information for antibodies and reagents